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## **Reporting Summary**

Nature Research wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Research policies, see Authors & Referees and the Editorial Policy Checklist.

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FOI	all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.
n/a	Confirmed
	$\square$ The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
	🔀 A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
	The statistical test(s) used AND whether they are one- or two-sided  Only common tests should be described solely by name; describe more complex techniques in the Methods section.
	A description of all covariates tested
$\boxtimes$	A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
	For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i> ) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted <i>Give P values as exact values whenever suitable.</i>
$\boxtimes$	For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
	For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
$\boxtimes$	$\square$ Estimates of effect sizes (e.g. Cohen's $d$ , Pearson's $r$ ), indicating how they were calculated
	Our web collection on <u>statistics for biologists</u> contains articles on many of the points above.
So	ftware and code

Policy information about availability of computer code

Data collection

The data was collected from public repositories (TCGA images) and our local biobank (DACHS images) and from a previous trial (KCCH) as described in the methods section. The data were preprocessed with the source codes at https://github.com/jnkather/MSIfromHE

Data analysis

All source codes are available at https://github.com/jnkather/MSIfromHE

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors/reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Research guidelines for submitting code & software for further information.

## Data

Policy information about availability of data

All manuscripts must include a <u>data availability statement</u>. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A list of figures that have associated raw data
- A description of any restrictions on data availability

All whole slide images for data sets are available at https://portal.gdc.cancer.gov/. Training images for tumor detection are available at http://dx.doi.org/10.5281/zenodo.2530789. Training images for MSI detection are available at http://dx.doi.org/10.5281/zenodo.2530835 and http://dx.doi.org/10.5281/zenodo.2530835.

Field-spe	ecific reporting			
Please select the o	ne below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.			
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For a reference copy of	the document with all sections, see <u>nature.com/documents/nr-reporting-summary-flat.pdf</u>			
Life scier	nces study design			
All studies must dis	sclose on these points even when the disclosure is negative.			
Sample size	We used histological images of cancer to predict molecular features. Prior to the study, the effect size was not known. Therefore, we aimed to include approximately as many patients as Coudray (doi: 10.1038/s41591-018-0177-5) and Kather (doi: 10.1371/journal.pmed.1002730) et al.			
Data exclusions	We downloaded all available histological images from TCGA but only included those patients with known MSI status as defined by Liu et al. (doi: 10.1016/j.ccell.2018.03.010) or hypermutation status as defined by Bailey et al. (doi: 10.1016/j.cell.2018.02.060). We excluded patients with missing MSI and hypermutation status and this criterion was pre-established			
Replication	We trained a deep neural network predictor on TCGA images and validated it in held-out patients and in external patient cohorts. Our experiments can be replicated by using the source codes we provided. We have used our source codes to repeat all experiments two times and all of these replication attempts were successful.			
Randomization	Participants were randomized to training and test sets by using MATLAB's random number generator with a constant seed.			
Blinding	All manual steps in our procedures were done in a blinded way (tissue handling, image handling).			

## Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Materials & experimental systems		Me	Methods	
n/a	Involved in the study	n/a	Involved in the study	
$\boxtimes$	Antibodies	$\boxtimes$	ChIP-seq	
$\boxtimes$	Eukaryotic cell lines	$\bowtie$	Flow cytometry	
$\boxtimes$	Palaeontology	$\bowtie$	MRI-based neuroimaging	
$\boxtimes$	Animals and other organisms			
$\boxtimes$	Human research participants			
$\boxtimes$	Clinical data			