

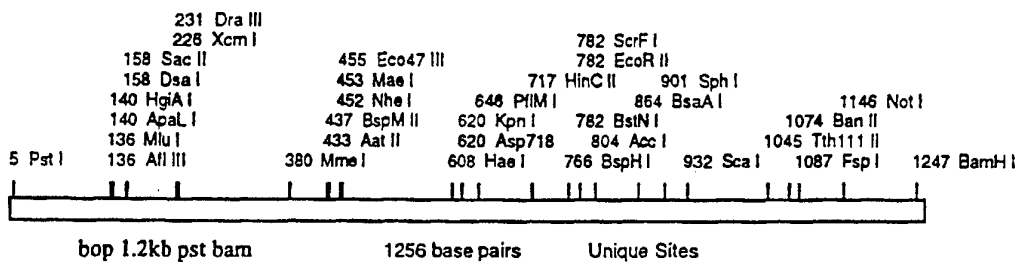


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(54) Title: EXPRESSION OF HETEROLOGOUS POLYPEPTIDES IN HALOBACTERIA



(57) Abstract

This invention relates to the preparation and use of expression systems capable of producing heterologous polypeptides in halobacterial hosts.

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EXPRESSION OF HETEROLOGOUS  
POLYPEPTIDES IN HALOBACTERIA

Field of the Invention

5 The present invention is directed to the preparation and use of a halobacterial expression system that is capable of producing soluble and transmembrane heterologous polypeptides that are not endogenous to said halobacterium.

Background of the Invention

10 Halobacteria are found in nature in evaporating salt water ponds under conditions of intense light and low oxygen saturation. They contain distinctive brightly colored pigments such as the orange-red pigment, bacterioruberin, or patches of "purple membrane". Halobacteria belong to a phylogenetically distinct group of prokaryotic organisms - the "archaebacteria" (Archaea) - that are as distantly related to the eubacteria as they are to the eukaryotes.

15 Archaeobacteria possess some attributes in common with the eukaryotes and the eubacteria, as well as characteristics that are uniquely archaeal. For example, the archaeobacteria possess a eukaryotic-like transcription apparatus with a 7-12 subunit RNA polymerase which is immunologically related to eukaryotic RNA polymerase (1) and promoter structures are similar to those of RNA Pol II (2).  
20 In contrast, the archaeobacteria have prokaryotic cellular morphology and 23S, 16S and 5S rRNAs with the genes encoding the rRNAs arranged into eubacterial-like operons (3). Notably, the archaeobacteria are unique in their membrane composition.

25 Bacteriorhodopsin (BR) is found as the sole protein in specialized crystalline patches of the "purple membrane" in halobacteria. Synthesis of BR is induced by high light intensity and low oxygen tension and the patches of purple membrane can constitute up to 50% of the archaeobacterium *Halobacterium halobium* cell surface area.

30 BR consists of a complex of one protein (bacterio-opsin) along with the chromophore retinal in a 1:1 stoichiometric ratio (4). This complex is embedded in the lipid matrix as seven transmembrane hydrophobic  $\alpha$ -helices in a trimeric

configuration (5). Retinal is covalently attached at lysine at position 216 approximately one-third of the way across the transmembraneous region of one of the  $\alpha$ -helices (6). The complex of bacterio-opsin with retinal was named bacteriorhodopsin (BR). The so-called *bop* gene encodes the light-driven protein pump bacteriorhodopsin (BR) in *H. halobium*.

There has been some reported research on expression of endogenous polypeptides in halobacteria (7, 8 and 9).

### Summary of the Invention

The present invention is directed to the preparation and use of an expression system for heterologous polypeptide production in a halobacterial host.

In a first aspect, such systems in their broadest context would include transcription and translation regulatory DNA, DNA encoding a heterologous polypeptide that is not endogenous to the halobacterial host and DNA encoding transcription and translation stop signals.

Preferably such systems would include DNA encoding the pre-sequence of bacteriorhodopsin such that the polypeptide which is expressed is attached to the pre-sequence, thus allowing the heterologous polypeptide to be properly targeted to the membrane and either inserted into or secreted across the membrane.

Yet another preferred embodiment of the present invention uses the transcription and translation regulatory sequences and the translation and transcription stop sequences of the bacteriorhodopsin gene, either in the presence or absence of the bacteriorhodopsin pre-sequence. The use of the regulatory and stop sequences of the bacteriorhodopsin gene serves to allow high level expression of the heterologous polypeptide sequence.

In a second aspect, the present invention is also directed to utilizing the C-terminal domain of the bacteriorhodopsin polypeptide in order to enhance the separation of the mature heterologous polypeptide from the membrane of the halobacterial host following expression. In a preferred embodiment of this aspect, DNA encoding a unique protease site is introduced between said C-terminal sequence and the DNA encoding the heterologous polypeptide.

In a preferred embodiment of this aspect, high levels of expression of the heterologous polypeptide linked to the C-terminal region of bacteriorhodopsin are achieved by using DNA encoding the transcription and translation regulatory and stop sequences of the bacteriorhodopsin gene.

5 A further preferred embodiment of the invention is directed to the use of the bacteriorhodopsin pre-sequence to enhance expression of the heterologous polypeptide linked to the C-terminal region of bacteriorhodopsin.

The invention is directed to such systems in all their equivalent aspects, including expression vectors, halobacterial hosts transformed with such vectors and methods for producing, isolating and optionally further purifying  
10 heterologous polypeptides using such expression vectors.

#### Detailed Description

The present invention has been described herein by disclosing the preferred embodiments and best mode. It will be understood, however, that having  
15 detailed the method first used by the present inventors to produce the heterologous polypeptide expression system in halobacterium, it will be apparent to those skilled in the art that one could make modifications within the general skill of the art to produce expression systems that differ in one or more ways from that originally described.

#### 20 A) Brief Description of the Drawings

Figure 1 is a restriction map of the PstI/BamHI fragment containing the bacteriorhodopsin gene and about 400 bp of upstream sequences from  
*Halobacterium halobium* strain R1.

Figure 2 shows the nucleic acid sequence (SEQ ID NO:1) of the PstI/BamHI  
25 construct of Figure 1 containing the bacteriorhodopsin gene and about 400 bp of upstream sequences from *Halobacterium halobium* strain R1. Also shown is the amino acid sequence (SEQ ID NO:2) of the BR protein translation product.

Figure 3 shows the restriction map of pUBP2.

Figure 4 is a map of the secondary structure of the mature BR protein (SEQ ID NO:3).

5 Figure 5 is a restriction map of the *PstI/BamHI* fragment containing BR regulatory sequences and the gene for human muscarinic acetylcholine receptor (Type HM1) in pENDS-OM1.

Figure 6 shows the nucleic acid sequence (SEQ ID NO:6) of the *PstI/BamHI* fragment of Figure 5 containing the gene for human muscarinic acetylcholine receptor (Type HM1) of pENDS-OM1. Also shown is the amino acid sequence (SEQ ID NO:7) of HM1.

10 Figure 7 is a restriction map of the *PstI/BamHI* fragment containing the BR regulatory sequences and gene for human muscarinic acetylcholine receptor (Type HM1) in pENDS-OM2.

15 Figure 8 shows the nucleic acid sequence (SEQ ID NO:8) of the *PstI/BamHI* fragment of Figure 7 containing the gene for human muscarinic acetylcholine receptor (Type HM1) which lacks the I3 domain. The amino acid sequence (SEQ ID NO:9) of HM1 having a deleted I3 domain is shown.

Figure 9 is a restriction map of the *PstI/BamHI* fragment containing the BR regulatory sequences and the rat serotonin receptor (Type 1C) gene.

20 Figure 10 shows the nucleic acid sequence (SEQ ID NO:10) of the *PstI/BamHI* construct of Figure 9 containing the rat serotonin receptor gene and the amino acid sequence (SEQ ID NO:11) of the rat serotonin receptor.

25 Figure 11 is a Southern blot of DNA isolated from *H. halobium Bop* deficient strain L33 transformed with pUBP2 containing the rat serotonin receptor (Type 1C) gene. Lanes 1-10, 12-19, 21-24 and 27 contained DNA from strain L33 transformed with pUBP2 containing the *PstI/BamHI* fragment of Figs. 9 and 10

(SEQ ID NO:10). Lanes 11 and 25 are positive controls which contained purified plasmid DNA (i.e. pUBP2 containing serotonin receptor gene). Lane 29 contained DNA from strain L33. The arrow indicates the location of the *PstI/BamHI* fragment corresponding to serotonin DNA.

5 Figure 12 shows a Northern blot of total RNA isolated from *H. halobium Bop* deficient strain L33 transformed with pUBP2 containing the rat serotonin receptor gene. Lanes 2 and 5 contain RNA from wild type strain L33 transformed with the 1.2 kb *PstI/BamHI* fragment containing the *bop* gene in pUBP2 as a control. Lanes 1, 3 and 4 contain DNA from L33 transformed with  
10 the rat serotonin receptor gene. The 1.85 kb *PstI/BamHI* fragment of Figs. 9 and 10 was used as probe. The arrow shows the location of the rat serotonin receptor RNA.

Figure 13 is a restriction map of the *PstI/BamHI* fragment containing BR regulatory sequences and the human thrombin receptor gene.

15 Figure 14 shows the nucleic acid sequence (SEQ ID NO:12) of the *PstI/BamHI* fragment of Figure 13 containing the human thrombin receptor gene and the amino acid sequence (SEQ ID NO:13) of the human thrombin receptor.

Figure 15 shows the restriction maps of p $\beta$ gbop, pEK17, pBATC, p1.2KbBop and pBRAT.

20 Figure 16 shows a restriction map of the *PstI/BamHI* fragment containing BR regulatory sequences, the bacterio-opsin gene and the gene encoding the *Escherichia coli* catalytic subunit of aspartate transcarbamylase.

Figure 17 shows the nucleic acid sequence (SEQ ID NO:14) of the *PstI/BamHI* fragment of Figure 16 containing the bacterio-opsin and the *E. coli* aspartate transcarbamylase genes and the amino acid sequence (SEQ ID NO:15) of the  
25 BR/*E. coli* aspartate transcarbamylase fusion protein.

Figure 18 shows a Western blot of *H. halobium* transformed with pBRAT. Blots were probed with antibodies to the catalytic subunit of aspartate transcarbamylase. Lane 2 contains *E. coli* aspartate transcarbamylase. Lanes 6-9 and 11 contain protein from *H. halobium* transformed with pBRAT. The arrow in lane 8 indicates the position of the bacteriorhodopsin/aspartate transcarbamylase (BR/ATCase) fusion protein.

Figure 19 shows the localization of expression of the bacteriorhodopsin/aspartate transcarbamylase (BR/ATCase) fusion protein to the purple halobacterial cell membranes. Washed *H. halobium* whole cell membranes fractionated on sucrose density gradients (A) were electrophoresed on SDS-polyacrylamide gels and stained with Coomassie blue (B). Lanes in (B) contained the following protein samples: Molecular weight markers (lane 1); unfractionated total membranes from *H. halobium* strain L33 transformed with pBRAT (lane 2); purple membrane from *H. halobium* strain L33 transformed with a 1.2 Kb *PstI/BamHI* fragment containing the *bop* gene (lane 3) or with a 9 Kb genomic DNA fragment containing the *bop* gene (lane 4); total membranes from *H. halobium* strain L33 (lane 5); purple membrane from wild-type *H. halobium* strain R1 (lane 6); purple membrane of *H. halobium* strain L33 transformed with pBRAT (lanes 7-9).

## 20 B) Definitions

The term "expression vector" herein has a functional definition and includes vectors capable of expressing DNA sequences contained therein, where such sequences are operably linked to other sequences capable of effecting their expression. In the present specification, "vector" and "plasmid" are used interchangeably as the plasmid is the most commonly used form of vector. However, the invention is intended to include such other forms of expression vectors capable of equivalent functions and which are or become known in the art.



By the term "operable" herein, and grammatical equivalents, is meant that the respective DNA sequences are operational and work for their intended purposes.

5 The term "heterologous polypeptide" herein refers to presently known or unknown polypeptides not endogenous to the host cell, or if endogenous to the host cell, are obtainable herein in amounts not achievable in native state. Included within the definition are the halobacterial non-retinal binding proteins. Examples of heterologous polypeptides include, but are not restricted to, polypeptides from eukaryotes, eubacteria, archaeobacteria, synthetic polypeptides and polypeptides containing bioequivalent amino acid analogs. Further included  
10 are other members of the 7-transmembrane crossing family such as muscarinic acetylcholine receptor, serotonin receptor, thrombin receptor,  $\beta$ -adrenergic receptor, and the like. Heterologous polypeptides also include membrane proteins, for example, cystic fibrosis transmembrane conductance regulator, and soluble proteins, such as various enzymes (e.g. proteases and aspartate  
15 transcarbamylase). Each is used in accord with their known or determined function biologically and is adapted for such in accord with procedures generally known in the art.

By the term "DNA encoding heterologous polypeptide" is meant a DNA  
20 sequence coding for a polypeptide that is not endogenous to the host wherein it is expressed. Because of the high GC content (i.e. about 58-68%) of the genome of halobacteria, it is preferred that the DNA sequence encoding the heterologous polypeptide be in this range, although sequences with higher and lower GC content than that usually found in halobacteria can be used. For example, we  
25 have been successful in expressing *Escherichia coli* aspartate transcarbamylase, having a GC content of about 50%, as a fusion protein to the C-terminus of BR.

The term "transcription and translation regulatory DNA" and equivalents, in its broadest sense refers to a DNA sequence responsible for the dual transcription and translation elements of expression. In a preferred embodiment  
30 the regulatory DNA is that of the bacteriorhodopsin gene (from -364 to +41 relative to the RNA start site, Fig.2 (SEQ ID NO:1)).

In an alternative embodiment, the regulatory DNA contains about 4000 bp of sequences (from about -4000 to +41) upstream of the *bop* gene and includes three other genes of the *bop* gene cluster, which include *brp* (13), *bat* (14) and *blp* (Gropp & Betlach, manuscript in preparation). Some or all of these genes may be regulatory genes.

By the term "transcription and translation stop signals" and equivalents, in its broadest scope is meant DNA which functions to terminate transcription and translation, respectively. It is preferred that the transcription and translation stop signals be those of the bacteriorhodopsin gene.

By the term "pre-sequence of bacteriorhodopsin gene" herein is meant a sequence of about 13 amino acids required to target bacteriorhodopsin to the membrane. The 13 amino acid pre-sequence is encoded by nucleotides +3 to +41 relative to the RNA start site depicted in Fig. 2 (SEQ ID NO:1).

By the term "halobacterium host" is meant strains belonging to Halobacterium, including species of extreme and moderate halophiles having a wild-type genotype. Examples of the extreme halophilic species having a wild type genotype include *Halobacterium saccharovorum* (ATCC 29252), *Halobacterium californica* (ATCC 38799), *Halobacterium halobium* (CCM 2090) and *Halobacterium valismortis* (ATCC 29-715). Wild type moderate halophiles are exemplified by the species *Halobacterium mediterranei* (ATCC 33500). It may be preferred that the halobacterial host species is bacteriorhodopsin deficient. Bacteriorhodopsin deficient species are either wild-type, such as *H. volcanii*, or mutants, such as L33 (15), S9F1x3 (16), IV-8 (17) and IV-14 (17). Bacteriorhodopsin deficient mutants derived from strains which express purple membrane constitutively, such as L33, or inducibly, are useful for different applications. Depending on the nature of the upstream regulatory regions in the expression vector construct, inducible strains permit regulated expression whereas constitutive strains do not.

The term "restriction site" herein refers to a DNA sequence recognizable by an endonuclease as a site of DNA cleavage.

By the term "C-terminal sequence", "C-terminal region" and equivalents, is meant the polypeptide sequence at the C-terminus of bacteriorhodopsin; See Fig. 4.

By the term "unique protease site" is meant an amino acid sequence recognizable by a protease as the site of cleavage of the polypeptide wherein it is disposed and which is absent from the heterologous polypeptide expression product. In a preferred embodiment, the protease site (Ile-Glu-Gly-Arg) (SEQ ID NO:4) of factor X<sub>a</sub> is used in view of the rarity of this sequence.

C) Examples

1. Cloning the DNA sequence encoding the heterologous polypeptide into a halobacterial expression vector

i. Constructs for expression of membrane proteins

All constructions are assembled using standard molecular techniques (12) including PCR. Expression vectors can be prepared in a variety of conventional ways. Although others may be used, a preferred halobacterial cloning vector to be adapted into an expression vector is plasmid pUBP2 (Fig.3) described by Blaseio et al. (7). The plasmid may be isolated using conventional techniques. For example, the plasmid may be purified using caesium chloride-ethidium bromide density gradients, electrophoresis from an agarose gel onto a dialysis membrane, use of commercially available chromatography columns for the separation of plasmids, such as magic minipreps DNA purification system (Promega Corp., Madison, WI), etc.

The expression vectors which will be employed will normally include a marker which allows for selection of cells into which the DNA has been integrated, as against cells which have not integrated the DNA construct. An example of commonly used selection markers is antibiotic resistance. Two markers are available for selection of halobacteria, including resistance to novobiocin (8) and mevinolin (7). It is preferred that the marker used be that for mevinolin resistance; mevinolin is a HMG CoA reductase inhibitor (7). This marker is present in the preferred cloning plasmid pUBP2 (Fig.3).

To convenience insertion of DNA sequences, plasmids will contain polylinker sequences containing various restriction sites. Several examples of polylinkers are known and available (12). A typical polylinker is polylinker 1 (12.3) (Fig.3.) which contains restriction sites for HindIII, SphI, MluI, XhoI, PstI, 5 Sall, XbaI, *Bam*HI, HindIII, XbaI and KpnI. Another typical polylinker is polylinker 2 (3.70) (Fig.3) with restriction sites for SphI, EcoR5, SstI, SmaI and EcoRI.

The DNA sequence encoding the heterologous polypeptide is inserted such that it is placed downstream from a transcription and translation regulatory 10 region containing a promoter and a ribosome binding site using standard techniques. It is preferred that the promoter used is inducible, allowing controlled expression of the heterologous polypeptide product. In a preferred embodiment of the invention, the transcription and translation regulatory sequences of the bacteriorhodopsin gene will be used. The bacteriorhodopsin 15 gene may be isolated from the genome of halobacteria using appropriate restriction enzymes. Transcription and translation regulatory sequences of the bacteriorhodopsin gene are located in the region of -365 to +41 relative to the RNA start site of the bacteriorhodopsin sequence depicted in Fig.2 (SEQ ID NO:1).

To effect appropriate termination of heterologous polypeptide synthesis, 20 DNA sequences encoding transcription and translation stop signals are placed downstream of the inserted DNA sequence encoding the heterologous polypeptide sequence using well known techniques (12). Preferably, the sequences downstream of the bacteriorhodopsin gene (Fig.2) (SEQ ID NO:1) 25 which includes the translational stop codon (TGA) followed by ~80 bp which include the transcriptional termination signal are employed as stop signals.

Where it is advantageous to produce a heterologous transmembrane polypeptide which is targeted to the halobacterial membrane, DNA encoding the heterologous polypeptide is ligated downstream of DNA encoding the pre- 30 sequence of BR.

The heterologous gene of interest may be cloned into the *E. coli* plasmid, pUC19 (20), along with BR regulatory sequences such that all cloned sequences will reside on a DNA fragment containing two unique restriction sites (choice of *Pst*I, *Bam*HI, *Sma*I). More specifically, the heterologous gene is ligated such that it is in frame with the BR pre-sequence, downstream of the bacteriorhodopsin regulatory sequences/promoter and upstream of the bacteriorhodopsin transcriptional and translational termination sequences. A specific unique protease site may be engineered into some constructions between the BR pre-sequence and the heterologous gene.

A 1.2 kbp fragment containing the *bop* gene and ~370 bp of upstream sequences was isolated from *H. halobium* strain R1 DNA using PCR and cloned into the *Pst*I/*Bam*HI sites of pUC19 (denoted p1.2K**bop**) (Fig.15B). Two endogenous *Alw*NI sites were removed from the cloned 1.2 kbp fragment: i) one site located 165 bp upstream of the *bop* gene start codon (SEQ ID NO:1) was removed by generating a G→T point mutation using the Kunkel method (29), and ii) the second *Alw*NI site located 7 bp upstream of the *bop* gene stop codon was removed using the Transformer Site-Directed Mutagenesis kit (Clontech Laboratories, Inc., Palo Alto, CA). Subsequently, a ~400 bp *Pst*I/*Alw*NI fragment (denoted "*bop* 5' fragment") containing the *bop* upstream sequences, DNA encoding the BR presequence and the first four (extrahelical) residues of BR was isolated by PCR from the mutated 1.2 kbp fragment. Concurrently, a ~100 bp *Not*I/*Bam*HI fragment (denoted "*bop* 3' fragment") containing DNA encoding six C-terminal residues of BR, the BR stop codon and the transcriptional termination sequences of BR (up to 44 bp downstream of the stop codon) was obtained from the 1.2 kbp *bop* gene fragment by preparative digestion and purification (Prep-A-Gene, BioRad, Richmond, CA). In addition, an endogenous *Alw*NI site located in pUC19 (position 1217) was removed using the Clontech Transformer kit and the mutated pUC19 was preparatively digested with *Pst*I/*Bam*HI and preparatively purified (denoted "vector fragment"). The three fragments (i.e., "*bop* 5' fragment", "*bop* 3' fragment" and "vector fragment") were ligated with DNA fragments containing various heterologous genes engineered to be in frame with the BR presequence and extrahelical

residues and to contain a single *AlwNI* site at the 5' terminus of the fragment and a single *NotI* site at the 3' terminus of the fragment as described below. In all of the heterologous genes, endogenous *AlwNI*, *NotI*, *BamHI* and *PstI* sites were first removed (if necessary) to facilitate the construction. Once the  
5 heterologous gene was cloned along with the BR 5' and 3' regulatory sequences into pUC19, this intermediate construct (denoted "pENDs") was preparatively digested with *PstI/BamHI*

Subsequently, the *PstI/BamHI* restriction fragment containing the heterologous gene with the regulatory sequences of BR was preparatively isolated  
10 away from pUC19 sequences by agarose gel electrophoresis, purified using Prep-A-Gene (Bio-Rad, Richmond, CA) and cloned into the *E. coli/H. halobium* shuttle vector, pUBP2 (7). pUBP2 carries the pBR322 replicon and ampicillin resistance marker, the halobacterial plasmid pHH1 origin of replication and a mevinolin resistance marker. Mevinolin resistance is encoded by an up-promoter  
15 mutation of the HMG-CoA reductase gene.

The construction was verified by restriction mapping and nucleotide sequencing across the junctions between 5' and 3' BR regulatory sequences and the heterologous gene.

a. Human muscarinic acetylcholine receptor (Type HM1)

20 Two different constructs were made with this gene. The first (denoted pENDs-OM1) contained the entire gene whereas the second (denoted pENDs-OM2) lacked the large internal cytoplasmic loop (i.e., I3 ) which is thought to be involved in signaling. Prior to the generation of the constructions described below, two endogenous *AlwNI* sites and one endogenous *PstI* site were removed  
25 from human muscarinic acetylcholine receptor (denoted HM1) cloned in pGEM3 (Promega Corp, Madison, WI) using either the Clontech Transformer kit or the Kunkel method (29). The positions of the removed sites are shown in Fig.6 (SEQ ID NO:6).

pENDs-OM1 was generated as follows. First, the HM1 gene was isolated  
30 by PCR from pGEM3/HM1 so as to contain an *AlwNI* site at the 5' terminus and a *NotI* site the 3' terminus of the PCR fragment. This PCR fragment was

ligated to the "bop 5' fragment", "bop 3' fragment" and "vector fragment" described above and transformed into *E. coli*. The resultant plasmid was named pENDs-OM1. pENDs-OM1 contains the methionine start codon of HM1 located 4 codons downstream from the BR 5' sequences. Nine extra base pairs generated by introduction of the *Alw*NI site encode 3 extra residues (i.e., gln, ala, leu) located in frame between the BR 5' sequences and the start codon of the HM1 gene. At the 3' terminus of the gene, the HM1 stop codon precedes the BR stop codon by 48 bp. From pENDs-OM1, the BR regulatory sequences with the HM1 gene were transferred to pUBP2 on a *Pst*I/*Bam*HI fragment (Fig.5 and Fig.6, SEQ ID NO:6) as described above.

pENDs-OM2 was generated in a similar manner as its sibling construct. First, however, deletions of the I3 domain were introduced after digestion of the HM1 gene at the unique *Stu*I restriction site (position 712 relative to the start codon of the HM1 gene, SEQ ID NO:6), followed by digestion with the exonuclease *Bal*-31 for varying times at 4°C. The blunt-ended product was self-ligated to yield mutants with deletions of varying size within the I3 domain. One of these was chosen for further study which lacked amino acid residues 231 through 357 of HM1 (SEQ ID NO:7). DNA from this mutant was used to generate a PCR fragment containing the HM1 gene (less I3 loop) with a 5' *Alw*NI site and a 3' *Not* I site. This PCR fragment was identical to the fragment described above except for the lack of the I3 loop and was used to generate pENDs-OM2 in a similar manner to the pENDs-OM1 construct. The sequence of the *Pst*I/*Bam*HI fragment containing the BR regulatory sequences and the HM1 gene (less I3 loop) is shown in Fig.8 (SEQ ID NO:8).

25                    b.     Rat serotonin receptor (Type 1 C)

The rat serotonin receptor gene (denoted "Ser") cloned as a 3 Kb *Eco*RI cDNA fragment on the plasmid pSR1c (27) was used as a basis for the following constructions. The Ser gene contains no endogenous *Alw*NI, *Not*I, *Bam*HI and *Pst*I sites and was adapted for expression in *H. halobium* as follows. *Alw*NI and *Not*I cloning sites were introduced within the 5' coding and 3' noncoding regions of the Ser gene, respectively. In addition, DNA encoding a poly-aspartic acid

peptide was placed in frame upstream of the Ser gene and downstream of the *AlwNI* site. Translation of this sequence generates a peptide epitope useful for subsequent detection of expressed protein (31). This fragment was isolated and ligated to the "bop 5' fragment", "bop 3' fragment", and "vector fragment" described above and transformed into *E. coli*. The resultant plasmid was named pENDs-Ser and contains the 36th codon of the rat serotonin receptor gene preceded by DNA encoding the peptide epitope and BR 5' sequences. Nine extra base pairs generated by the construction and encoding 3 extra residues (i.e., gln, ala, leu) are located in frame between the BR 5' sequences and the epitope sequences. At the 3' terminus of the gene, the Ser stop codon precedes the BR stop codon by 18 bp. Following the construction of pENDs-Ser, the BR regulatory sequences with the Ser gene were transferred to pUBP2 on a *PstI/BamHI* fragment (Fig.9 and Fig.10, SEQ ID NO:10).

c. Human thrombin receptor

A clone of the human thrombin receptor gene (denoted "Thromb") (33) was used as a basis for the following constructions. Four endogenous DNA restriction sites were removed from the gene using the Kunkel method (29). These included three *AlwNI* sites (291, 945, and 1038) and one *PstI* site (537). Positions are given relative to the first base of the start codon of the gene. "pENDs-Thromb" was generated as follows. An *AlwNI/NotI* fragment containing the gene was generated using oligonucleotide-directed-insertion-mutagenesis and PCR. Included on this fragment were additional nucleotide sequences encoding short peptides for use in the detection and purification of the expressed protein. The *AlwNI/NotI* fragment containing the gene along with epitope encoding sequences was ligated to the "bop 5' fragment", "bop 3' fragment" and "vector fragment" described above and transformed into *E. coli*. The resultant plasmid was named pENDs-Thromb. In pENDs-Thromb, thirty-three extra base pairs generated by the construction and encoding eleven extra amino acids are located in frame between the BR 5' sequences and the Thromb sequences. Twenty seven of the extra residues encode a poly-aspartic acid peptide sequence which when translated generates a peptide epitope useful for detection of expressed



protein (31). At the 3' terminus of the gene, six histidine codons have been inserted upstream of the Thromb stop codon. These histidine codons are intended to aid in the affinity purification of expressed protein (26). At the 3' terminus of the gene, the Thromb stop codon precedes the BR stop codon by 18  
5 bp.

The BR regulatory sequences with the human thrombin receptor gene may be transferred into pUBP2 on a *PstI/BamHI* fragment (Fig.13 and Fig.14, SEQ ID NO:12) as described above.

ii. Constructs for expression of soluble proteins

10 Where it is desired that heterologous soluble polypeptide be released extracellularly into the culture medium following expression, the DNA sequence encoding the heterologous polypeptide may be ligated to DNA encoding the pre-sequence of bacteriorhodopsin (Fig.2 (SEQ ID NO:1), from +3 to +41 relative to the RNA start site) using techniques well known to those skilled in the art  
15 (12).

Where it is advantageous to produce a heterologous soluble polypeptide that is targeted, following expression, to the halobacterial membrane, DNA encoding the heterologous polypeptide is ligated downstream of the DNA encoding the C-terminal region (Fig.2 and Fig.4 (SEQ ID NOs:1 and 3)) of bacteriorhodopsin or to fragments thereof.  
20

To facilitate subsequent purification of the heterologous polypeptide product, a DNA sequence encoding a unique protease site is engineered between DNA encoding the bacteriorhodopsin C-terminal region and DNA encoding the heterologous polypeptide. Sequences encoding unique protease cleavage sites are known and include, for example, subtilisin, thrombin, enterokinase, and factor  $X_a$ . In a preferred embodiment, a DNA sequence encoding the amino acid sequence Ile-Glu-Gly-Arg (SEQ ID NO:4) is used to encode a unique protease site which is recognized by Factor  $X_a$ .  
25

Design of the soluble protein expression vector and methods used are similar to that described above for membrane proteins. However, soluble proteins are expressed as in-frame fusions to the C-terminal region of BR. Thus,  
30

these fusion proteins will have membranous domain (i.e. BR or portions thereof) and a soluble domain (i.e. heterologous polypeptide). The heterologous gene is cloned at the C-terminus of BR, between the bacteriorhodopsin gene and the downstream transcriptional/translational termination sequences of BR. In addition, a unique protease site is engineered between BR and the heterologous gene to facilitate subsequent purification of the protein. The final construct is cloned into the *E. coli/H. halobium* shuttle vector, pUBP2 (7).

a. *E. coli* Aspartate Transcarbamylase (catalytic subunit)

The catalytic subunit of Aspartate Transcarbamylase, (denoted ATCase), a soluble protein, has been fused to the C-terminus of BR as follows. The *bop* gene containing plasmid, p $\beta$ gbop (32), was digested at the unique *NotI* site located near the 3' terminus of the *bop* gene (see Figure 15A). Subsequently, this *NotI* site was filled-in to create a blunt site (12). The resulting DNA was digested with *SphI* to generate two fragments, a large fragment (denoted fragment 1) containing the vector along with the N-terminus of the *bop* gene and a small fragment containing internal *bop* gene sequences. Fragment 1 was isolated and purified. A second aliquot of p $\beta$ gbop was digested with *SphI/HaeII* and a 217 bp fragment (denoted fragment 2) containing an internal portion of the *bop* gene was isolated and purified (Figure 15A).

The structural gene for the *E. coli* catalytic subunit of aspartate transcarbamylase was isolated from pEK17 (Fig.15A) (30). A 845 bp *MseI/NruI* fragment (denoted fragment 3) which contains all but the first 18 bp of the gene encoding ATCase was isolated and purified.

A synthetic fragment of DNA (denoted fragment 4) was constructed by annealing two complementary oligonucleotides and used to connect the *bop* and ATCase genes. The synthetic fragment was engineered to contain a *HaeII* site at the 5' terminus, a *MseI* site at the 3' terminus and an internal *NruI* site. Also included were nucleotides encoding: i) a unique protease site (i.e., blood clotting Factor X<sub>a</sub>) and ii) ATCase amino acids 6 and 7 (relative to ATCase start codon) Fig.17, SEQ ID NO:14.

All four DNA fragments were ligated together and used to transform *E. coli* strain D1210 (28) with selection for ampicillin resistance. Positive clones were identified by colony filter hybridization using P<sup>32</sup> radiolabeled random primed (25) ATCase *MseI/NruI* fragment as probe. Positive clones were verified by restriction mapping and nucleotide sequencing. One positive clone was chosen and denoted pBATC (Figure 15A).

Subsequently, the *bop*-ATCase fusion construct was adapted for *H. halobium* expression as follows. A fragment spanning the sequences in between and including the internal *SphI* site of the *bop* gene at the 5' terminus and the ATCase translational stop codon at the 3' terminus was isolated from pBATC by PCR (see Figure 15B). In addition, the oligonucleotide used to construct the 3' terminus of this PCR fragment was designed to be complementary to *bop* sequences downstream of the transcriptional termination sequences and to include a unique *BamHI* to facilitate subsequent cloning steps. The resultant PCR fragment was digested with *SphI/BamHI*, purified and used in the following construction.

The plasmid, p1.2Kbbop, containing the *bop* gene and upstream sequences cloned in pUC19 (described above) was digested with *SphI/BamHI* to yield two fragments, a large one containing the vector and the majority of the *bop* gene, and a 358 bp fragment containing the C-terminal half of the *bop* gene (Fig.15B). The larger of these two fragments was isolated, purified and ligated to the *SphI/BamHI* *bop*-ATCase PCR fragment. A positive clone was isolated and confirmed by restriction mapping and nucleotide sequencing. This clone was digested with *PstI/BamHI* and a fragment containing DNA encoding the BR/ATCase fusion along with *bop* upstream regulatory sequences (Fig.16) was cloned into the *E. coli/H. halobium* shuttle vector pUBP2. The resultant construct was named pBRAT (Fig.15B). The nucleotide sequence (SEQ ID NO:14) and the translated amino acid sequence (SEQ ID NO:15) of this *PstI/BamHI* fragment is shown in Fig.17.

## 2. Transformation of *Halobacterium halobium*

The *Pst*I/*Bam*HI fragments of the pENDs-Ser (Fig.9 and 10, SEQ ID NO:10) and pBRAT (Fig.15B, Fig.16 and Fig.17, SEQ ID NO:14) constructs containing the heterologous genes with the BR regulatory sequences were isolated and purified. Subsequently, these fragments were cloned into the  
5 *E. coli/H. halobium* shuttle vector pUBP2 (7) and transformed into *H. halobium* Bop deficient strain L33 as described (24).

Preferably, plasmids may be introduced into halobacteria using the polyethylene glycol (PEG) method (10, 11). Transformed halobacterial cells are then grown in culture in an appropriate nutrient medium sufficient to maintain  
10 the growth of halobacterial cells (7, 8).

*H. halobium* is prone to cell lysis during transformation procedures (7). Since surfactants are known to promote halobacterial lysis (21), all media and glassware used were soap-free. Transformation was performed according to  
15 Blaseio (7) and Cline (11) with modifications. Initially, cells were subcultured several times in soap-free complex (YET) medium. Subsequently, cells were subcultured to an OD<sub>660</sub> of about 0.01 and grown at 40°C until the early to mid-logarithmic stage of growth (OD<sub>660</sub> of 0.4 to 0.6). All succeeding manipulations were performed at room temperature. The culture was removed from the  
20 waterbath shaker and incubated without agitation for 4 h to overnight, followed by centrifugation of 2 ml of culture at 1000 x g for 15 min. The supernatant was carefully removed with a pipette and the interior of the centrifuge tube dried with absorbent tissue. The cell pellet was resuspended in 1/10 volume of spheroplasting solution (11), followed by addition of 1/100 volume of 0.5 M  
25 EDTA in Spheroplasting solution (11) and incubation for 2 min. One µg of DNA in 10 µl of spheroplasting solution was then added to the spheroplasted cells along with an equal volume of 60% PEG 600 (un-recrystallized) in spheroplasting solution. The combined solutions were gently but thoroughly mixed and then incubated for 20 min. Ten ml of 15% sucrose in complex (YET)  
30 medium was added followed by incubation overnight with no agitation at 42°C. The following day, cells were centrifuged at 3000 x g for 15 minutes and

resuspended in 300  $\mu$ l of 15% sucrose in complex (YET) medium. This solution was plated on solid complex (YET) selection medium.

3. Analysis of transformants, expression of the heterologous polypeptide and assays for expression

5 To establish that halobacterial cells have been successfully transformed, various techniques may be employed. Where the expression vector used to transform the halobacteria contains a dominant selectable marker, transformed cells can be selected by growing in the appropriate selection medium such that growth of halobacterial cells not harboring the recombinant plasmid is inhibited.  
10 For example, where a plasmid containing the mevinolin resistance marker is used, halobacterial cells which harbor this plasmid may be selected by growing on solid nutrient medium containing mevinolin at a concentration in the range of 5 to 25  $\mu$ M. Further, the plasmid may be isolated using standard techniques (12), restricted and used. The polymerase chain reaction, gel electrophoresis,  
15 restriction analysis, Southern, Northern, and Western blots may be employed, sequencing, or the like, may all be employed with advantage.

Depending upon the particular construct and the halobacterial background strain which have been employed for expression of the heterologous polypeptides, one may have constitutive or inducible expression of the heterologous  
20 polypeptide product. In the case of constitutive expression, the product will be continuously formed as the cells grow. By contrast, for inducible expression, one may provide for induction when the cells reach a predetermined cell density.

Where inducible promoters have been engineered into the expression vector containing the heterologous polypeptide DNA sequence, transcription may  
25 be induced using appropriate inducers under such conditions of concentration and duration as to effect induction of transcription. For example, if the regulatory sequences of the bacteriorhodopsin gene are used, transcription can be induced by low oxygen tension and high light intensity (18, 19) which are known to induce high level expression of BR. Low oxygen tensions are achieved  
30 in various ways such as by flushing culture flasks with oxygen-free nitrogen and sealing them, or by permitting cultures to reach the stationary phase of growth

in which oxygen limitation occurs naturally (18). High light intensity of greater than about 100 mW/cm<sup>2</sup> can be achieved using various light sources and apparatus as described (18, 19).

*H. halobium* transformed with pUBP2 containing the pBRAT *PstI/BamHI* fragment and with pUBP2 containing the pENDS-Ser *PstI/BamHI* fragment was  
5 plated on solid complex (YET) medium containing 25  $\mu$ M mevinolin. Plates were incubated for one to two weeks at 42°C to permit growth of transformants. Plasmid DNA was isolated from individual transformants using Magic Minipreps DNA Purification System (Promega Corp., Madison, WI). Southern analysis was  
10 used to verify the presence of the heterologous gene on pUBP2. Southern blot analysis using the *AlwNI/NotI* fragment containing the serotonin receptor gene as probe indicated the presence of serotonin receptor gene sequences in all assayed transformants (Figure 11). Total RNA was isolated from individual transformants using the RNazol procedure (Cinna Biotech) and subjected to  
15 Northern analysis (18). Northern blot analysis revealed that transcription of Ser gene sequences had occurred (Figure 12). Western analysis using both BR and ATCase antibodies demonstrated that the BR/ATCase fusion was expressed and localized to halobacterial membranes (Figure 18). Washed halobacterial whole cell membranes were fractionated on sucrose gradients (Fig.19A) and aliquots  
20 were subjected to SDS PAGE (Fig.19B). A band corresponding to the predicted molecular weight of the fusion protein (i.e., ~60 kDa, see Fig.19B) was observed which derived from a purple fraction. These data verify expression of the fusion and indicate that the BR portion of the fusion is folded correctly in the halobacterial membrane. The presence of the BR chromophore (extinction  
25 coefficient of 63,000; 31) affords an estimate of 5 mg/liter of fusion protein expression.

Transformants testing positive in Southern and Northern analyses are subjected to Western analysis if specific antibodies to the heterologous protein are available. If antibodies are not available, DNA encoding an epitope known  
30 to be antigenic may be engineered into the expression vector construction to aid in detection of expression. An example of such an epitope is the sequence encoding Glu-Glu-Glu-Glu-Tyr-Met-Pro-Met-Glu (SEQ ID NO:5) (22).

Alternatively, expression of the heterologous protein may be assayed functionally; for example, ligand binding assays for receptors, and assays for enzymic activity for soluble proteins using appropriate substrates.

#### 4. Purification of heterologous polypeptides

5 Production of the heterologous polypeptide may be stopped in a variety of ways. Where the heterologous polypeptide is released into the medium, it may be isolated in a soluble or insoluble form using physical e.g. mechanical or thermal, or chemical treatments. Treatments employed may include freezing ( $\leq 0^{\circ}$  C), heating, hydrodynamic shearing, drying, selective filtration or  
10 precipitation by addition of acid, base, salts or organic solvents.

Where the expressed heterologous polypeptide resides in the membrane or in the cytoplasm, cells are harvested to separate them from the culture medium. Various techniques may be used for harvesting, desirably using centrifugation. The supernatant may then be discarded and the cell pellet  
15 washed with an appropriate buffered aqueous medium to remove any residual culture medium components. Typically the buffered medium will be at a temperature in the range of about 1 to  $10^{\circ}$ C, more usually  $4^{\circ}$ C.

The cells may be lysed by any convenient means, such as freezing and mechanical, use of hypotonic solutions (23), and the like. The resulting  
20 dispersion of disrupted cells is then treated by such means as to substantially separate cell membranes from soluble proteins and other contaminants. Several techniques may be employed to advantage for isolating membranes including differential centrifugation, density gradient centrifugation, and the like. This membrane isolation separates the fusion protein from the bulk of the soluble  
25 proteins.

Heterologous polypeptides are purified according to procedures dependent on their individual properties and those of BR. Where the expressed soluble heterologous polypeptide is fused at the C-terminal region of BR, advantage may be taken of the likelihood that the BR domain will anchor the fusion protein in  
30 the membrane.

Where the heterologous polypeptide is expressed as a fusion polypeptide linked to the C-terminal region, or fragment thereof, of the bacteriorhodopsin gene with a unique protease site between said heterologous polypeptide and C-terminal region, the heterologous polypeptide may be isolated by incubating the halobacterial membranes with an appropriate unique protease to effect substantially complete cleavage at the protease cleavage site. For example, where the heterologous polypeptide is linked to the bacteriorhodopsin C-terminal region through the amino acid sequence Ile-Glu-Gly-Arg (SEQ ID N0:4), cell membranes are incubated with factor X<sub>a</sub> under conditions recommended by the manufacturers. Factor X<sub>a</sub> is dissolved in redistilled water to a final protein concentration of 1 mg/ml. The fusion protein to be cleaved is dissolved in 100 mM NaCl, 50 mM Tris-HCl, 1 mM CaCl<sub>2</sub>, pH 8.0. To increase the solubility of the substrate, urea or acetonitrile can be added up to a final concentration of 1 M and 10% (v/v), respectively without significant inhibition of the enzyme activity. The recommended amount of enzyme is 1/200 to 1/10 of the substrate by weight. Incubation should be carried out at 4°C to 25°C for 1-18 h. The optimum cleavage conditions have to be determined for each fusion protein. The release of the desired polypeptide from the fusion protein is influenced by the adjacent amino acid sequences at the cleavage site, the size of the two fused polypeptide components, and the accessibility of the cleavage site. Protease treatment is followed by standard purification protocols to remove the minor unique protease component.

If further purification of the heterologous polypeptide protein is desired, antibodies specific for the heterologous polypeptide, ligand affinity, electrophoresis, chromatography, zonal centrifugation, and the like, may be employed to advantage. The product may then be dried by any convenient means, such as freeze drying, spray drying, and the like, or alternatively suspended in an appropriate buffered aqueous solution. The heterologous polypeptide product is then ready for use.

30 5. Bioassays



The heterologous polypeptides may be assayed using protocols dependent on their individual properties. For example, receptors are assayed using ligand binding assays. Soluble proteins having enzyme activity are assayed using appropriate substrates.

## 5 Bibliography

For the sake of convenience, various documents referenced in the body of the present specification are grouped in the following bibliography by number that corresponds to the parenthetical number of that reference in the text. Each of these documents is hereby expressly incorporated by reference.

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15 Concluding Remarks

The foregoing description details specific methods that can be employed to practice the present invention. Having detailed specific methods initially used to construct and use vectors for the expression, isolation, detection and further purification of heterologous polypeptides in halobacteria, those skilled in the art will know how to devise alternative reliable methods for arriving at the same and equivalent systems described herein. The foregoing should not be construed as limiting the overall scope hereof; rather, the ambit of the present invention is to be governed only by the lawful interpretation of the appended claims.

20

The Halobacterium strains referred to above were deposited with the American Type Culture Collection, located at 12301 Parklawn Drive, Rockville, Maryland 20852-1776. The dates of the deposits were ATCC 29252 - February 3, 1976; ATCC 38799 - September 13, 1979; ATCC 29715 - September 19, 1977 and ATCC 33500 - February 23, 1981.

25

SEQUENCE LISTING

## (1) GENERAL INFORMATION:

- (i) APPLICANT: TURNER, George J.  
BETLACH, Mary C.
- (ii) TITLE OF INVENTION: EXPRESSION OF HETEROLOGOUS POLYPEPTIDES  
IN HALOBACTERIA
- (iii) NUMBER OF SEQUENCES: 15
- (iv) CORRESPONDENCE ADDRESS:
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  - (C) CITY: Los Angeles
  - (D) STATE: California
  - (E) COUNTRY: USA
  - (F) ZIP: 90012
- (v) COMPUTER READABLE FORM:
  - (A) MEDIUM TYPE: Floppy disk
  - (B) COMPUTER: IBM PC compatible
  - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
  - (D) SOFTWARE: PatentIn Release #1.0, Version #1.25
- (vi) CURRENT APPLICATION DATA:
  - (A) APPLICATION NUMBER: PCT
  - (B) FILING DATE:
  - (C) CLASSIFICATION:
- (viii) ATTORNEY/AGENT INFORMATION:
  - (A) NAME: Berliner, Robert
  - (B) REGISTRATION NUMBER: 20,121
  - (C) REFERENCE/DOCKET NUMBER: 5555-206-PCT
- (ix) TELECOMMUNICATION INFORMATION:
  - (A) TELEPHONE: (213) 977-1001
  - (B) TELEFAX: (213) 977-1003

## (2) INFORMATION FOR SEQ ID NO:1:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1254 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)

- (ix) FEATURE:
  - (A) NAME/KEY: misc\_feature
  - (B) LOCATION: 376..414
  - (D) OTHER INFORMATION: /note= "Bacteriorhodopsin pre-sequence."
  
- (ix) FEATURE:
  - (A) NAME/KEY: CDS
  - (B) LOCATION: 376..1161
  
- (ix) FEATURE:
  - (A) NAME/KEY: misc\_feature
  - (B) LOCATION: 3..8
  - (D) OTHER INFORMATION: /note= "PstI site."
  
- (ix) FEATURE:
  - (A) NAME/KEY: misc\_feature
  - (B) LOCATION: 1245..1250
  - (D) OTHER INFORMATION: /note= "BamHI site."
  
- (ix) FEATURE:
  - (A) NAME/KEY: misc\_signal
  - (B) LOCATION: 374
  - (D) OTHER INFORMATION: /note= "RNA start site."
  
- (ix) FEATURE:
  - (A) NAME/KEY: misc\_feature
  - (B) LOCATION: 9..414
  - (D) OTHER INFORMATION: /note= "Bacteriorhodopsin transcriptional and translational regulatory sequences are located in this region."

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

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TTCGAGTGGT AACACGCGTG CACGCATCGA CTTCACCGCG GGTGTTTCGA CGCCAGCCGG	180
CCGTTGAACC AGCAGGCAGC GGGCATTTC AAGCCGCTGT GGCCACACA CTCGGTGGGG	240
TGCGCTATTT TGGTATGGTT TGAATCCGC GTGTCGGCTC CGTGTCTGAC GGTTTCATCGG	300
TCTAAATTCC GTCACGAGCG TACCATACTG ATTGGGTCGT AGAGTTACAC ACATATCCTC	360
GTTAGGTACT GTTGC ATG TTG GAG TTA TTG CCA ACA GCA GTG GAG GGG GTA	411
Met Leu Glu Leu Leu Pro Thr Ala Val Glu Gly Val	
1 5 10	

TCG CAG GCC CAG ATC ACC GGA CGT CCG GAG TGG ATC TGG CTA GCG CTC Ser Gln Ala Gln Ile Thr Gly Arg Pro Glu Trp Ile Trp Leu Ala Leu 15 20 25	459
GGT ACG GCG CTA ATG GGA CTC GGG ACG CTC TAT TTC CTC GTG AAA GGG Gly Thr Ala Leu Met Gly Leu Gly Thr Leu Tyr Phe Leu Val Lys Gly 30 35 40	507
ATG GGC GTC TCG GAC CCA GAT GCA AAG AAA TTC TAC GCC ATC ACG ACG Met Gly Val Ser Asp Pro Asp Ala Lys Lys Phe Tyr Ala Ile Thr Thr 45 50 55 60	555
CTC GTC CCA GCC ATC GCG TTC ACG ATG TAC CTC TCG ATG CTG CTG GGG Leu Val Pro Ala Ile Ala Phe Thr Met Tyr Leu Ser Met Leu Leu Gly 65 70 75	603
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GCC GAA AGC ATG CGC CCC GAG GTC GCA TCC ACG TTC AAA GTA CTG CGT Ala Glu Ser Met Arg Pro Glu Val Ala Ser Thr Phe Lys Val Leu Arg 175 180 185	939
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225 230 235

CTG CGC AGT CGT GCG ATC TTC GGC GAA GCC GAA GCG CCG GAG CCG TCC 1131  
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240 245 250

GCC GGC GAC GGC GCG GCC GCG ACC AGC GAC TGATCGCACA CGCAGGACAG 1181  
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255 260

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CTCGGATCCT TTT 1254

(2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 262 amino acids
  - (B) TYPE: amino acid
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

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20 25 30

Met Gly Leu Gly Thr Leu Tyr Phe Leu Val Lys Gly Met Gly Val Ser  
35 40 45

Asp Pro Asp Ala Lys Lys Phe Tyr Ala Ile Thr Thr Leu Val Pro Ala  
50 55 60

Ile Ala Phe Thr Met Tyr Leu Ser Met Leu Leu Gly Tyr Gly Leu Thr  
65 70 75 80

Met Val Pro Phe Gly Gly Glu Gln Asn Pro Ile Tyr Trp Ala Arg Tyr  
85 90 95

Ala Asp Trp Leu Phe Thr Thr Pro Leu Leu Leu Leu Asp Leu Ala Leu  
100 105 110

Leu Val Asp Ala Asp Gln Gly Thr Ile Leu Ala Leu Val Gly Ala Asp  
115 120 125

Gly Ile Met Ile Gly Thr Gly Leu Val Gly Ala Leu Thr Lys Val Tyr  
 130 135 140  
 Ser Tyr Arg Phe Val Trp Trp Ala Ile Ser Thr Ala Ala Met Leu Tyr  
 145 150 155 160  
 Ile Leu Tyr Val Leu Phe Phe Gly Phe Thr Ser Lys Ala Glu Ser Met  
 165 170 175  
 Arg Pro Glu Val Ala Ser Thr Phe Lys Val Leu Arg Asn Val Thr Val  
 180 185 190  
 Val Leu Trp Ser Ala Tyr Pro Val Val Trp Leu Ile Gly Ser Glu Gly  
 195 200 205  
 Ala Gly Ile Val Pro Leu Asn Ile Glu Thr Leu Leu Phe Met Val Leu  
 210 215 220  
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## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 248 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

## (ix) FEATURE:

- (A) NAME/KEY: Region
- (B) LOCATION: 225..248
- (D) OTHER INFORMATION: /note= "Cytoplasmic C-terminal region of bacteriorhodopsin."

## ·(ix) FEATURE:

- (A) NAME/KEY: Region
- (B) LOCATION: 1
- (D) OTHER INFORMATION: /note= "Pyroglutamate."

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

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 50 55 60  
 Gly Leu Thr Met Val Pro Phe Gly Gly Glu Gln Asn Pro Ile Tyr Trp  
 65 70 75 80  
 Ala Arg Tyr Ala Asp Trp Leu Phe Thr Thr Pro Leu Leu Leu Leu Asp  
 85 90 95  
 Leu Ala Leu Leu Val Asp Ala Asp Gln Gly Thr Ile Leu Ala Leu Val  
 100 105 110  
 Gly Ala Asp Gly Ile Met Ile Gly Thr Gly Leu Val Gly Ala Leu Thr  
 115 120 125  
 Lys Val Tyr Ser Tyr Arg Phe Val Trp Trp Ala Ile Ser Thr Ala Ala  
 130 135 140  
 Met Leu Tyr Ile Leu Tyr Val Leu Phe Phe Gly Phe Thr Ser Lys Ala  
 145 150 155 160  
 Glu Ser Met Arg Pro Glu Val Ala Ser Thr Phe Lys Val Leu Arg Asn  
 165 170 175  
 Val Thr Val Val Leu Trp Ser Ala Tyr Pro Val Val Trp Leu Ile Gly  
 180 185 190  
 Ser Glu Gly Ala Gly Ile Val Pro Leu Asn Ile Glu Thr Leu Leu Phe  
 195 200 205  
 Met Val Leu Asp Val Ser Ala Lys Val Gly Phe Gly Leu Ile Leu Leu  
 210 215 220  
 Arg Ser Arg Ala Ile Phe Gly Glu Ala Glu Ala Pro Glu Pro Ser Ala  
 225 230 235 240  
 Gly Asp Gly Ala Ala Ala Thr Ser  
 245



## (2) INFORMATION FOR SEQ ID NO:4:

- (i) SEQUENCE CHARACTERISTICS:  
(A) LENGTH: 4 amino acids  
(B) TYPE: amino acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Ile Glu Gly Arg  
1

## (2) INFORMATION FOR SEQ ID NO:5:

- (i) SEQUENCE CHARACTERISTICS:  
(A) LENGTH: 9 amino acids  
(B) TYPE: amino acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

Glu Glu Glu Glu Tyr Met Pro Met Glu  
1 5

## (2) INFORMATION FOR SEQ ID NO:6:

- (i) SEQUENCE CHARACTERISTICS:  
(A) LENGTH: 1956 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

## (ix) FEATURE:

- (A) NAME/KEY: CDS  
(B) LOCATION: 376..1812

## (ix) FEATURE:

- (A) NAME/KEY: misc feature  
(B) LOCATION: 376..414  
(D) OTHER INFORMATION: /note= "Bacteriorhodopsin pre-sequence."

- (ix) FEATURE:  
(A) NAME/KEY: terminator  
(B) LOCATION: 1864..1866  
(D) OTHER INFORMATION: /note= "Bacteriorhodopsin stop codon."
- (ix) FEATURE:  
(A) NAME/KEY: mutation  
(B) LOCATION: replace(213, "")  
(D) OTHER INFORMATION: /note= "G to T mutation removes AlwNI restriction site."
- (ix) FEATURE:  
(A) NAME/KEY: misc feature  
(B) LOCATION: 427..435  
(D) OTHER INFORMATION: /note= "AlwNI cloning site."
- (ix) FEATURE:  
(A) NAME/KEY: mutation  
(B) LOCATION: replace(930, "")  
(D) OTHER INFORMATION: /note= "G to A mutation removes AlwNI restriction site."
- (ix) FEATURE:  
(A) NAME/KEY: mutation  
(B) LOCATION: replace(1179, "")  
(D) OTHER INFORMATION: /note= "T to A mutation removes AlwNI site."
- (ix) FEATURE:  
(A) NAME/KEY: mutation  
(B) LOCATION: replace(1245, "")  
(D) OTHER INFORMATION: /note= "G to A mutation removes PstI restriction site."
- (ix) FEATURE:  
(A) NAME/KEY: misc\_signal  
(B) LOCATION: 374  
(D) OTHER INFORMATION: /note= "RNA start site."
- (ix) FEATURE:  
(A) NAME/KEY: mutation  
(B) LOCATION: replace(1863, "")  
(D) OTHER INFORMATION: /note= "C to T mutation removes AlwNI restriction site."
- (ix) FEATURE:  
(A) NAME/KEY: terminator  
(B) LOCATION: 1813..1815  
(D) OTHER INFORMATION: /note= "Muscarinic "OM1" stop codon."

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

ATCTGCAGGA TGGGTGCAAC CGTGAAGTCC GTCACGGCTG CGTCACGACA GGAGCCGACC	60
AGCGACACCC AGAAGGTGCG AACGGTTGAG TGCCGCAACG ATCACGAGTT TTTCGTGCGC	120
TTTCAGTGGT AACACGCGTG CACGCATCGA CTTCACCGCG GGTGTTTCGA CGCCAGCCGG	180
CCGTTGAACC AGCAGGCAGC GGGCATTTC AATCCGCTGT GGCCACACA CTCGGTGGGG	240
TGCGCTATTT TGGTATGGTT TGAATCCGC GTGTCGGCTC CGTGTCTGAC GGTTTCATCGG	300
TCTAAATTCC GTCACGAGCG TACCATACTG ATTGGGTCGT AGAGTTACAC ACATATCCTC	360
GTTAGGTACT GTTGC ATG TTG GAG TTA TTG CCA ACA GCA GTG GAG GGG GTA	411
Met Leu Glu Leu Leu Pro Thr Ala Val Glu Gly Val	
1 5 10	
TCG CAG GCC CAG ATC CAG GCG CTG ATG AAC ACT TCA GCC CCA CCT GCT	459
Ser Gln Ala Gln Ile Gln Ala Leu Met Asn Thr Ser Ala Pro Pro Ala	
15 20 25	
GTC AGC CCC AAC ATC ACC GTC CTG GCA CCA GGA AAG GGT CCC TGG CAA	507
Val Ser Pro Asn Ile Thr Val Leu Ala Pro Gly Lys Gly Pro Trp Gln	
30 35 40	
GTG GCC TTC ATT GGG ATC ACC ACG GGC CTC CTG TCG CTA GCC ACA GTG	555
Val Ala Phe Ile Gly Ile Thr Thr Gly Leu Leu Ser Leu Ala Thr Val	
45 50 55 60	
ACA GGC AAC CTG CTG GTA CTC ATC TCT TTC AAG GTC AAC ACG GAG CTC	603
Thr Gly Asn Leu Leu Val Leu Ile Ser Phe Lys Val Asn Thr Glu Leu	
65 70 75	
AAG ACA GTC AAT AAC TAC TTC CTG CTG AGC CTG GCC TGT GCT GAC CTC	651
Lys Thr Val Asn Asn Tyr Phe Leu Leu Ser Leu Ala Cys Ala Asp Leu	
80 85 90	
ATC ATC GGT ACC TTC TCC ATG AAC CTC TAT ACC ACG TAC CTG CTC ATG	699
Ile Ile Gly Thr Phe Ser Met Asn Leu Tyr Thr Thr Tyr Leu Leu Met	
95 100 105	
GGC CAC TGG GCT CTG GGC ACG CTG GCT TGT GAC CTC TGG CTG GCC CTG	747
Gly His Trp Ala Leu Gly Thr Leu Ala Cys Asp Leu Trp Leu Ala Leu	
110 115 120	
GAC TAT GTG GCC AGC AAT GCC TCC GTC ATG AAT CTG CTG CTC ATC AGC	795
Asp Tyr Val Ala Ser Asn Ala Ser Val Met Asn Leu Leu Leu Ile Ser	
125 130 135 140	
TTT GAC CGC TAC TTC TCC GTG ACT CGG CCC CTG AGC TAC CGT GCC AAG	843
Phe Asp Arg Tyr Phe Ser Val Thr Arg Pro Leu Ser Tyr Arg Ala Lys	
145 150 155	

CGC Arg	ACA Thr	CCC Pro	CGC Arg 160	CGC Arg	GCA Ala	GCT Ala	CTG Leu	ATG Met 165	ATC Ile	GGC Gly	CTG Leu	GCC Ala 170	TGG Trp	CTG Leu	GTT Val	891
TCC Ser	TTT Phe	GTG Val 175	CTC Leu	TGG Trp	GCC Ala	CCA Pro	GCC Ala 180	ATC Ile	CTC Leu	TTC Phe	TGG Trp	CAA Gln 185	TAC Tyr	CTG Leu	GTA Val	939
GGG Gly 190	GAG Glu	CGG Arg	ACG Thr	ATG Met	CTA Leu	GCT Ala 195	GGG Gly	CAG Gln	TGC Cys	TAC Tyr	ATC Ile 200	CAG Gln	TTC Phe	CTC Leu	TCC Ser	987
CAG Gln 205	CCC Pro	ATC Ile	ATC Ile	ACC Thr	TTT Phe 210	GGC Gly	ACA Thr	GCC Ala	ATG Met	GCT Ala 215	GCC Ala	TTC Phe	TAC Tyr	CTC Leu	CCT Pro 220	1035
GTC Val	ACA Thr	GTC Val	ATG Met 225	TGC Cys	ACG Thr	CTC Leu	TAC Tyr	TGG Trp	CGC Arg 230	ATC Ile	TAC Tyr	CGG Arg	GAG Glu	ACA Thr 235	GAG Glu	1083
AAC Asn	CGA Arg	GCA Ala	CGG Arg 240	GAG Glu	CTG Leu	GCA Ala	GCC Ala	CTT Leu 245	CAG Gln	GGC Gly	TCC Ser	GAG Glu 250	ACG Thr	CCA Pro	GGC Gly	1131
AAA Lys	GGG Gly	GGT Gly 255	GGC Gly	AGC Ser	AGC Ser	AGC Ser	AGC Ser 260	TCA Ser	GAG Glu	AGG Arg	TCT Ser	CAG Gln 265	CCA Pro	GGG Gly	GCA Ala	1179
GAG Glu	GGC Gly 270	TCA Ser	CCA Pro	GAG Glu	ACT Thr	CCT Pro 275	CCA Pro	GGC Gly	CGC Arg	TGC Cys	TGT Cys 280	CGC Arg	TGC Cys	TGC Cys	CGG Arg	1227
GCC Ala 285	CCA Pro	AGG Arg	CTG Leu	CTG Leu	CAA Gln 290	GCC Ala	TAC Tyr	AGC Ser	TGG Trp	AAG Lys 295	GAA Glu	GAA Glu	GAG Glu	GAA Glu	GAG Glu 300	1275
GAC Asp	GAA Glu	GGC Gly	TCC Ser 305	ATG Met	GAG Glu	TCC Ser	CTC Leu	ACA Thr 310	TCC Ser	TCA Ser	GAG Glu	GGA Gly	GAG Glu	GAG Glu	CCT Pro 315	1323
GGC Gly	TCC Ser	GAA Glu	GTG Val 320	GTG Val	ATC Ile	AAG Lys	ATG Met	CCA Pro 325	ATG Met	GTG Val	GAC Asp	CCC Pro	GAG Glu 330	GCA Ala	CAG Gln	1371
GCC Ala	CCC Pro	ACC Thr 335	AAG Lys	CAG Gln	CCC Pro	CCA Pro	CGG Arg 340	AGC Ser	TCC Ser	CCA Pro	AAT Asn 345	ACA Thr	GTC Val	AAG Lys	AGG Arg	1419
CCG Pro	ACT Thr 350	AAG Lys	AAA Lys	GGG Gly	CGT Arg	GAT Asp 355	CGA Arg	GCT Ala	GGC Gly	AAG Lys	GGC Gly 360	CAG Gln	AAG Lys	CCC Pro	CGT Arg	1467

GGA AAG GAG CAG CTG GCC AAG CGG AAG ACC TTC TCG CTG GTC AAG GAG 1515  
 Gly Lys Glu Gln Leu Ala Lys Arg Lys Thr Phe Ser Leu Val Lys Glu 380  
 365 370 375

AAG AAG GCG GCT CGG ACC CTG AGT GCC ATC CTC CTG GCC TTC ATC CTC 1563  
 Lys Lys Ala Ala Arg Thr Leu Ser Ala Ile Leu Leu Ala Phe Ile Leu 395  
 385 390

ACC TGG ACA CCG TAC AAC ATC ATG GTG CTG GTG TCC ACC TTC TGC AAG 1611  
 Thr Trp Thr Pro Tyr Asn Ile Met Val Leu Val Ser Thr Phe Cys Lys 410  
 400 405

GAC TGT GTT CCC GAG ACC CTG TGG GAG CTG GGC TAC TGG CTG TGC TAC 1659  
 Asp Cys Val Pro Glu Thr Leu Trp Glu Leu Gly Tyr Trp Leu Cys Tyr 425  
 415 420

GTC AAC AGC ACC ATC AAC CCC ATG TGC TAC GCA CTC TGC AAC AAA GCC 1707  
 Val Asn Ser Thr Ile Asn Pro Met Cys Tyr Ala Leu Cys Asn Lys Ala 440  
 430 435

TTC CGG GAC ACC TTT CGC CTG CTG CTT TGC CGC TGG GAC AAG AGA CGC 1755  
 Phe Arg Asp Thr Phe Arg Leu Leu Leu Cys Arg Trp Asp Lys Arg Arg 460  
 445 450 455

TGG CGC AAG ATC CCC AAG CGC CCT GGC TCC GTG CAC CGC ACT CCC TCC 1803  
 Trp Arg Lys Ile Pro Lys Arg Pro Gly Ser Val His Arg Thr Pro Ser 475  
 465 470

CGC CAA TGC TGATAGTCCC CTCTCCTGCA TCCCTCCACC CCAGCGGCCG 1852  
 Arg Gln Cys

CGACCAGCGA TTGATCGCAC ACGCAGGACA GCCCCACAAC CGGCGCGGCT GTGTTCAACG 1912

ACACACGATG AGTCCCCAC TCGGTCTTGT ACTCGGATCC TTTT 1956

(2) INFORMATION FOR SEQ ID NO:7:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 479 amino acids
  - (B) TYPE: amino acid
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

Met Leu Glu Leu Leu Pro Thr Ala Val Glu Gly Val Ser Gln Ala Gln  
 1 5 10 15

Ile Gln Ala Leu Met Asn Thr Ser Ala Pro Pro Ala Val Ser Pro Asn  
 20 25 30

Ile Thr Val Leu Ala Pro Gly Lys Gly Pro Trp Gln Val Ala Phe Ile  
                   35                                  40                                  45  
 Gly Ile Thr Thr Gly Leu Leu Ser Leu Ala Thr Val Thr Gly Asn Leu  
           50                                  55                                  60  
 Leu Val Leu Ile Ser Phe Lys Val Asn Thr Glu Leu Lys Thr Val Asn  
   65                  70                                  75                                  80  
 Asn Tyr Phe Leu Leu Ser Leu Ala Cys Ala Asp Leu Ile Ile Gly Thr  
                   85                                  90                                  95  
 Phe Ser Met Asn Leu Tyr Thr Thr Tyr Leu Leu Met Gly His Trp Ala  
                  100                                 105                                 110  
 Leu Gly Thr Leu Ala Cys Asp Leu Trp Leu Ala Leu Asp Tyr Val Ala  
   115                 120                                 125  
 Ser Asn Ala Ser Val Met Asn Leu Leu Leu Ile Ser Phe Asp Arg Tyr  
   130                 135                                 140  
 Phe Ser Val Thr Arg Pro Leu Ser Tyr Arg Ala Lys Arg Thr Pro Arg  
   145                 150                                 155                                 160  
 Arg Ala Ala Leu Met Ile Gly Leu Ala Trp Leu Val Ser Phe Val Leu  
                  165                                 170                                 175  
 Trp Ala Pro Ala Ile Leu Phe Trp Gln Tyr Leu Val Gly Glu Arg Thr  
                  180                                 185                                 190  
 Met Leu Ala Gly Gln Cys Tyr Ile Gln Phe Leu Ser Gln Pro Ile Ile  
                  195                                 200                                 205  
 Thr Phe Gly Thr Ala Met Ala Ala Phe Tyr Leu Pro Val Thr Val Met  
   210                 215                                 220  
 Cys Thr Leu Tyr Trp Arg Ile Tyr Arg Glu Thr Glu Asn Arg Ala Arg  
   225                 230                                 235                                 240  
 Glu Leu Ala Ala Leu Gln Gly Ser Glu Thr Pro Gly Lys Gly Gly Gly  
                  245                                 250                                 255  
 Ser Ser Ser Ser Ser Glu Arg Ser Gln Pro Gly Ala Glu Gly Ser Pro  
                  260                                 265                                 270  
 Glu Thr Pro Pro Gly Arg Cys Cys Arg Cys Cys Arg Ala Pro Arg Leu  
                  275                                 280                                 285  
 Leu Gln Ala Tyr Ser Trp Lys Glu Glu Glu Glu Glu Asp Glu Gly Ser  
   290                 295                                 300  
 Met Glu Ser Leu Thr Ser Ser Glu Gly Glu Glu Pro Gly Ser Glu Val  
   305                 310                                 315                                 320



- (ix) FEATURE:  
 (A) NAME/KEY: terminator  
 (B) LOCATION: 1489..1491  
 (D) OTHER INFORMATION: /note= "Bacteriorhodopsin stop codon."
- (ix) FEATURE:  
 (A) NAME/KEY: mutation  
 (B) LOCATION: replace(213, "")  
 (D) OTHER INFORMATION: /note= "G to T mutation removes AlwNI restriction site."
- (ix) FEATURE:  
 (A) NAME/KEY: misc\_feature  
 (B) LOCATION: 427..435  
 (D) OTHER INFORMATION: /note= "AlwNI cloning site."
- (ix) FEATURE:  
 (A) NAME/KEY: mutation  
 (B) LOCATION: replace(930, "")  
 (D) OTHER INFORMATION: /note= "G to A mutation removes AlwNI site."
- (ix) FEATURE:  
 (A) NAME/KEY: misc\_signal  
 (B) LOCATION: 374  
 (D) OTHER INFORMATION: /note= "RNA start site."
- (ix) FEATURE:  
 (A) NAME/KEY: terminator  
 (B) LOCATION: 1438..1440  
 (D) OTHER INFORMATION: /note= "Muscarinic stop codon."
- (ix) FEATURE:  
 (A) NAME/KEY: mutation  
 (B) LOCATION: replace(1488, "")  
 (D) OTHER INFORMATION: /note= "C to T mutation removes AlwNI restriction site."

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

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ATCTGCAGGA TGGGTGCAAC CGTGAAGTCC GTCACGGCTG CGTCACGACA GGAGCCGACC      60
AGCGACACCC AGAAGGTGCG AACGGTTGAG TGCCGCAACG ATCACGAGTT TTTCGTGCGC      120
TTCGAGTGGT AACACGCGTG CACGCATCGA CTTACCCGCG GGTGTTTCGA CGCCAGCCGG      180
CCGTTGAACC AGCAGGCAGC GGGCATTTC AATCCGCTGT GGCCACACA CTCGGTGGGG      240
TGCGCTATTT TGGTATGGTT TGAATCCGC GTGTCGGCTC CGTGTCTGAC GGTTTCATCGG      300
TCTAAATTCC GTCACGAGCG TACCATACTG ATTGGGTCGT AGAGTTACAC ACATATCCTC      360

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GTTAGGTACT GTTGC ATG TTG GAG TTA TTG CCA ACA GCA GTG GAG GGG GTA	411
Met Leu Glu Leu Leu Pro Thr Ala Val Glu Gly Val	
1 5 10	
TCG CAG GCC CAG ATC CAG GCG CTG ATG AAC ACT TCA GCC CCA CCT GCT	459
Ser Gln Ala Gln Ile Gln Ala Leu Met Asn Thr Ser Ala Pro Pro Ala	
15 20 25	
GTC AGC CCC AAC ATC ACC GTC CTG GCA CCA GGA AAG GGT CCC TGG CAA	507
Val Ser Pro Asn Ile Thr Val Leu Ala Pro Gly Lys Gly Pro Trp Gln	
30 35 40	
GTG GCC TTC ATT GGG ATC ACC ACG GGC CTC CTG TCG CTA GCC ACA GTG	555
Val Ala Phe Ile Gly Ile Thr Thr Gly Leu Leu Ser Leu Ala Thr Val	
45 50 55 60	
ACA GGC AAC CTG CTG GTA CTC ATC TCT TTC AAG GTC AAC ACG GAG CTC	603
Thr Gly Asn Leu Leu Val Leu Ile Ser Phe Lys Val Asn Thr Glu Leu	
65 70 75	
AAG ACA GTC AAT AAC TAC TTC CTG CTG AGC CTG GCC TGT GCT GAC CTC	651
Lys Thr Val Asn Asn Tyr Phe Leu Leu Ser Leu Ala Cys Ala Asp Leu	
80 85 90	
ATC ATC GGT ACC TTC TCC ATG AAC CTC TAT ACC ACG TAC CTG CTC ATG	699
Ile Ile Gly Thr Phe Ser Met Asn Leu Tyr Thr Thr Tyr Leu Leu Met	
95 100 105	
GGC CAC TGG GCT CTG GGC ACG CTG GCT TGT GAC CTC TGG CTG GCC CTG	747
Gly His Trp Ala Leu Gly Thr Leu Ala Cys Asp Leu Trp Leu Ala Leu	
110 115 120	
GAC TAT GTG GCC AGC AAT GCC TCC GTC ATG AAT CTG CTG CTC ATC AGC	795
Asp Tyr Val Ala Ser Asn Ala Ser Val Met Asn Leu Leu Leu Ile Ser	
125 130 135 140	
TTT GAC CGC TAC TTC TCC GTG ACT CGG CCC CTG AGC TAC CGT GCC AAG	843
Phe Asp Arg Tyr Phe Ser Val Thr Arg Pro Leu Ser Tyr Arg Ala Lys	
145 150 155	
CGC ACA CCC CGC CGC GCA GCT CTG ATG ATC GGC CTG GCC TGG CTG GTT	891
Arg Thr Pro Arg Arg Ala Ala Leu Met Ile Gly Leu Ala Trp Leu Val	
160 165 170	
TCC TTT GTG CTC TGG GCC CCA GCC ATC CTC TTC TGG CAA TAC CTG GTA	939
Ser Phe Val Leu Trp Ala Pro Ala Ile Leu Phe Trp Gln Tyr Leu Val	
175 180 185	
GGG GAG CGG ACG ATG CTA GCT GGG CAG TGC TAC ATC CAG TTC CTC TCC	987
Gly Glu Arg Thr Met Leu Ala Gly Gln Cys Tyr Ile Gln Phe Leu Ser	
190 195 200	

CAG Gln 205	CCC Pro	ATC Ile	ATC Ile	ACC Thr	TTT Phe	GGC Gly	ACA Thr	GCC Ala	ATG Met	GCT Ala	GCC Ala	TTC Phe	TAC Tyr	CTC Leu	CCT Pro	1035
GTC Val	ACA Thr	GTC Val	ATG Met	TGC Cys	ACG Thr	CTC Leu	TAC Tyr	TGG Trp	CGC Arg	ATC Ile	TAC Tyr	CGG Arg	GAG Glu	ACA Thr	GAG Glu	1083
AAC Asn	CGA Arg	GCA Ala	CGG Arg	GAG Glu	CTG Leu	GCA Ala	GCC Ala	CTT Leu	CAG Gln	GGC Gly	TCC Ser	GAG Glu	ACG Thr	CCA Pro	GGC Gly	1131
AAA Lys	AAG Lys	GAG Glu	AAG Lys	AAG Lys	GCG Ala	GCT Ala	CGG Arg	ACC Thr	CTG Leu	AGT Ser	GCC Ala	ATC Ile	CTC Leu	CTG Leu	GCC Ala	1179
TTC Phe	ATC Ile	CTC Leu	ACC Thr	TGG Trp	ACA Thr	CCG Pro	TAC Tyr	AAC Asn	ATC Ile	ATG Met	GTG Val	CTG Leu	GTG Val	TCC Ser	ACC Thr	1227
TTC Phe	TGC Cys	AAG Lys	GAC Asp	TGT Cys	GTT Val	CCC Pro	GAG Glu	ACC Thr	CTG Leu	TGG Trp	GAG Glu	CTG Leu	GGC Gly	TAC Tyr	TGG Trp	1275
CTG Leu	TGC Cys	TAC Tyr	GTC Val	AAC Asn	AGC Ser	ACC Thr	ATC Ile	AAC Asn	CCC Pro	ATG Met	TGC Cys	TAC Tyr	GCA Ala	CTC Leu	TGC Cys	1323
AAC Asn	AAA Lys	GCC Ala	TTC Phe	CGG Arg	GAC Asp	ACC Thr	TTT Phe	CGC Arg	CTG Leu	CTG Leu	CTT Leu	TGC Cys	CGC Arg	TGG Trp	GAC Asp	1371
AAG Lys	AGA Arg	CGC Arg	TGG Trp	CGC Arg	AAG Lys	ATC Ile	CCC Pro	AAG Lys	CGC Arg	CCT Pro	GGC Gly	TCC Ser	GTG Val	CAC His	CGC Arg	1419
ACT Thr	CCC Pro	TCC Ser	CGC Arg	CAA Gln	TGC Cys	TGATAGTCCC CTCTCCTGCA TCCCTCCACC									1467	
CCAGCGGCCG CGACCAGCGA TTGATCGCAC ACGCAGGACA GCCCCACAAC CGGCGCGGCT																1527
GTGTTCAACG ACACACGATG AGTCCCCAC TCGGTCTTGT ACTCGGATCC TTTT																1581

## (2) INFORMATION FOR SEQ ID NO:9:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 354 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

Met Leu Glu Leu Leu Pro Thr Ala Val Glu Gly Val Ser Gln Ala Gln  
 1 5 10 15  
 Ile Gln Ala Leu Met Asn Thr Ser Ala Pro Pro Ala Val Ser Pro Asn  
 20 25 30  
 Ile Thr Val Leu Ala Pro Gly Lys Gly Pro Trp Gln Val Ala Phe Ile  
 35 40 45  
 Gly Ile Thr Thr Gly Leu Leu Ser Leu Ala Thr Val Thr Gly Asn Leu  
 50 55 60  
 Leu Val Leu Ile Ser Phe Lys Val Asn Thr Glu Leu Lys Thr Val Asn  
 65 70 75 80  
 Asn Tyr Phe Leu Leu Ser Leu Ala Cys Ala Asp Leu Ile Ile Gly Thr  
 85 90 95  
 Phe Ser Met Asn Leu Tyr Thr Thr Tyr Leu Leu Met Gly His Trp Ala  
 100 105 110  
 Leu Gly Thr Leu Ala Cys Asp Leu Trp Leu Ala Leu Asp Tyr Val Ala  
 115 120 125  
 Ser Asn Ala Ser Val Met Asn Leu Leu Leu Ile Ser Phe Asp Arg Tyr  
 130 135 140  
 Phe Ser Val Thr Arg Pro Leu Ser Tyr Arg Ala Lys Arg Thr Pro Arg  
 145 150 155 160  
 Arg Ala Ala Leu Met Ile Gly Leu Ala Trp Leu Val Ser Phe Val Leu  
 165 170 175  
 Trp Ala Pro Ala Ile Leu Phe Trp Gln Tyr Leu Val Gly Glu Arg Thr  
 180 185 190  
 Met Leu Ala Gly Gln Cys Tyr Ile Gln Phe Leu Ser Gln Pro Ile Ile  
 195 200 205  
 Thr Phe Gly Thr Ala Met Ala Ala Phe Tyr Leu Pro Val Thr Val Met  
 210 215 220  
 Cys Thr Leu Tyr Trp Arg Ile Tyr Arg Glu Thr Glu Asn Arg Ala Arg  
 225 230 235 240  
 Glu Leu Ala Ala Leu Gln Gly Ser Glu Thr Pro Gly Lys Lys Glu Lys  
 245 250 255  
 Lys Ala Ala Arg Thr Leu Ser Ala Ile Leu Leu Ala Phe Ile Leu Thr  
 260 265 270



- (ix) FEATURE:  
(A) NAME/KEY: misc\_feature  
(B) LOCATION: 736..807  
(D) OTHER INFORMATION: /note= "Helix III of rat serotonin receptor protein (Type 1C)."
- (ix) FEATURE:  
(A) NAME/KEY: misc\_feature  
(B) LOCATION: 868..939  
(D) OTHER INFORMATION: /note= "Helix IV of rat serotonin receptor protein (Type 1C)."
- (ix) FEATURE:  
(A) NAME/KEY: misc\_feature  
(B) LOCATION: 997..1059  
(D) OTHER INFORMATION: /note= "Helix V of rat serotonin receptor protein (Type 1C)."
- (ix) FEATURE:  
(A) NAME/KEY: misc\_feature  
(B) LOCATION: 1297..1362  
(D) OTHER INFORMATION: /note= "Helix VI of rat serotonin receptor protein (Type 1C)."
- (ix) FEATURE:  
(A) NAME/KEY: misc\_feature  
(B) LOCATION: 1411..1476  
(D) OTHER INFORMATION: /note= "Helix VII of rat serotonin receptor protein (Type 1C)."
- (ix) FEATURE:  
(A) NAME/KEY: mutation  
(B) LOCATION: replace(213, "")  
(D) OTHER INFORMATION: /note= "G to A mutation removes AlwNI restriction site."
- (ix) FEATURE:  
(A) NAME/KEY: misc\_feature  
(B) LOCATION: 1732..1734  
(D) OTHER INFORMATION: /note= "Codon encoding the C-terminal amino acid of the rat serotonin receptor protein (Type 1C)."
- (ix) FEATURE:  
(A) NAME/KEY: misc\_signal  
(B) LOCATION: 374  
(D) OTHER INFORMATION: /note= "RNA start site."
- (ix) FEATURE:  
(A) NAME/KEY: CDS  
(B) LOCATION: 376..1734

(ix) FEATURE:  
 (A) NAME/KEY: terminator  
 (B) LOCATION: 1735..1737  
 (D) OTHER INFORMATION: /note= "Serotonin stop codon."

(ix) FEATURE:  
 (A) NAME/KEY: repeat\_region  
 (B) LOCATION: 436..462  
 (D) OTHER INFORMATION: /note= "Sequence encoding polyaspartic acid."

(ix) FEATURE:  
 (A) NAME/KEY: mutation  
 (B) LOCATION: replace(1755, "")  
 (D) OTHER INFORMATION: /note= "C to T mutation removes AlwNI restriction site."

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

ATCTGCAGGA TGGGTGCAAC CGTGAAGTCC GTCACGGCTG CGTCACGACA GGAGCCGACC	60
AGCGACACCC AGAAGGTGCG AACGGTTGAG TGCCGCAACG ATCACGAGTT TTTCGTGCGC	120
TTCGAGTGGT AACACGCGTG CACGCATCGA CTTACCCGCG GGTGTTTCGA CGCCAGCCGG	180
CCGTTGAACC AGCAGGCAGC GGGCATTTC AATCCGCTGT GGCCACACA CTCGGTGGGG	240
TGCGCTATTT TGGTATGGTT TGGAAATCCGC GTGTCGGCTC CGTGTCTGAC GGTTTCATCGG	300
TCTAAATTCC GTCACGAGCG TACCATACTG ATTGGGTCGT AGAGTTACAC ACATATCCTC	360
GTTAGGTACT GTTGC ATG TTG GAG TTA TTG CCA ACA GCA GTG GAG GGG GTA	411
Met Leu Glu Leu Leu Pro Thr Ala Val Glu Gly Val	
1 5 10	
TCG CAG GCC CAG ATC CAG GCG CTG GAC TAC AAG GAC GAT GAT GAC GTC	459
Ser Gln Ala Gln Ile Gln Ala Leu Asp Tyr Lys Asp Asp Asp Val	
15 20 25	
GAC ACT TTT AAT TCC TCC GAT GGT GGA CGC TTG TTT CAA TTC CCG GAC	507
Asp Thr Phe Asn Ser Ser Asp Gly Gly Arg Leu Phe Gln Phe Pro Asp	
30 35 40	
GGG GTA CAA AAC TGG CCA GCA CTT TCA ATC GTC GTG ATT ATA ATC ATG	555
Gly Val Gln Asn Trp Pro Ala Leu Ser Ile Val Val Ile Ile Ile Met	
45 50 55 60	
ACA ATA GGG GGC AAC ATT CTT GTT ATC ATG GCA GTA AGC ATG GAG AAG	603
Thr Ile Gly Gly Asn Ile Leu Val Ile Met Ala Val Ser Met Glu Lys	
65 70 75	

AAA CTG CAC AAT GCA ACC AAT TAC TTC TTA ATG TCC CTA GCC ATT GCT Lys Leu His Asn Ala Thr Asn Tyr Phe Leu Met Ser Leu Ala Ile Ala 80 85 90	651
GAT ATG CTG GTG GGA CTA CTT GTC ATG CCC CTG TCC CTG CTT GCT ATT Asp Met Leu Val Gly Leu Leu Val Met Pro Leu Ser Leu Leu Ala Ile 95 100 105	699
CTT TAT GAT TAT GTC TGG CCT TTA CCT AGA TAT TTG TGC CCC GTC TGG Leu Tyr Asp Tyr Val Trp Pro Leu Pro Arg Tyr Leu Cys Pro Val Trp 110 115 120	747
ATT TCA CTA GAT GTG CTA TTT TCA ACT GCG TCC ATC ATG CAC CTC TGC Ile Ser Leu Asp Val Leu Phe Ser Thr Ala Ser Ile Met His Leu Cys 125 130 135 140	795
GCC ATA TCG CTG GAC CGG TAT GTA GCA ATA CGT AAT CCT ATT GAG CAT Ala Ile Ser Leu Asp Arg Tyr Val Ala Ile Arg Asn Pro Ile Glu His 145 150 155	843
AGC CGG TTC AAT TCG CGG ACT AAG GCC ATC ATG AAG ATT GCC ATC GTT Ser Arg Phe Asn Ser Arg Thr Lys Ala Ile Met Lys Ile Ala Ile Val 160 165 170	891
TGG GCA ATA TCA ATA GGA GTT TCA GTT CCT ATC CCT GTG ATT GGA CTG Trp Ala Ile Ser Ile Gly Val Ser Val Pro Ile Pro Val Ile Gly Leu 175 180 185	939
AGG GAC GAA AGC AAA GTG TTC GTG AAT AAC ACC ACG TGC GTG CTC AAT Arg Asp Glu Ser Lys Val Phe Val Asn Asn Thr Thr Cys Val Leu Asn 190 195 200	987
GAC CCC AAC TTC GTT CTC ATC GGG TCC TTC GTG GCA TTC TTC ATC CCG Asp Pro Asn Phe Val Leu Ile Gly Ser Phe Val Ala Phe Phe Ile Pro 205 210 215 220	1035
TTG ACG ATT ATG GTG ATC ACC TAC TTC TTA ACG ATC TAC GTC CTG CGC Leu Thr Ile Met Val Ile Thr Tyr Phe Leu Thr Ile Tyr Val Leu Arg 225 230 235	1083
CGT CAA ACT CTG ATG TTA CTT CGA GGT CAC ACC GAG GAG GAA CTG GCT Arg Gln Thr Leu Met Leu Leu Arg Gly His Thr Glu Glu Glu Leu Ala 240 245 250	1131
AAT ATG AGC CTG AAC TTT CTG AAC TGC TGC TGC AAG AAG AAT GGT GGT Asn Met Ser Leu Asn Phe Leu Asn Cys Cys Cys Lys Lys Asn Gly Gly 255 260 265	1179
GAG GAA GAG AAC GCT CCG AAC CCT AAT CCA GAT CAG AAA CCA CGT CGA Glu Glu Glu Asn Ala Pro Asn Pro Asn Pro Asp Gln Lys Pro Arg Arg 270 275 280	1227

AAG AAG AAA GAA AAG CGT CCC AGA GGC ACC ATG CAA GCT ATC AAC AAC Lys Lys Lys Glu Lys Arg Pro Arg Gly Thr Met Gln Ala Ile Asn Asn 285 290 295 300	1275
GAA AAG AAA GCT TCC AAA GTC CTT GGC ATT GTA TTC TTT GTG TTT CTG Glu Lys Lys Ala Ser Lys Val Leu Gly Ile Val Phe Phe Val Phe Leu 305 310 315	1323
ATC ATG TGG TGC CCG TTT TTC ATC ACC AAT ATC CTG TCG GTT CTT TGT Ile Met Trp Cys Pro Phe Phe Ile Thr Asn Ile Leu Ser Val Leu Cys 320 325 330	1371
GGG AAG GCC TGT AAC CAA AAG CTA ATG GAG AAG CTT CTC AAT GTG TTT Gly Lys Ala Cys Asn Gln Lys Leu Met Glu Lys Leu Leu Asn Val Phe 335 340 345	1419
GTG TGG ATT GGC TAT GTG TGT TCA GGC ATC AAT CCT CTG GTG TAC ACT Val Trp Ile Gly Tyr Val Cys Ser Gly Ile Asn Pro Leu Val Tyr Thr 350 355 360	1467
CTC TTT AAT AAA ATT TAC CGA AGG GCT TTC TCT AAA TAT TTG CGC TGC Leu Phe Asn Lys Ile Tyr Arg Arg Ala Phe Ser Lys Tyr Leu Arg Cys 365 370 375 380	1515
GAT TAT AAG CCA GAC AAA AAG CCT CCT GTT CGA CAG ATT CCT AGG GTT Asp Tyr Lys Pro Asp Lys Lys Pro Pro Val Arg Gln Ile Pro Arg Val 385 390 395	1563
GCT GCC ACT GCT TTG TCT GGG AGG GAG CTC AAT GTT AAC ATT TAT CGG Ala Ala Thr Ala Leu Ser Gly Arg Glu Leu Asn Val Asn Ile Tyr Arg 400 405 410	1611
CAT ACC AAT GAA CGT GTG GCT AGG AAA GCT AAT GAC CCT GAG CCT GGC His Thr Asn Glu Arg Val Ala Arg Lys Ala Asn Asp Pro Glu Pro Gly 415 420 425	1659
ATA GAG ATG CAG GTG GAG AAC TTA GAG CTG CCA GTC AAC CCC TCT AAT Ile Glu Met Gln Val Glu Asn Leu Glu Leu Pro Val Asn Pro Ser Asn 430 435 440	1707
GTG GTC AGC GAG AGG ATT AGT AGT GTG TGAGCGGCCG CGACCAGCGA Val Val Ser Glu Arg Ile Ser Ser Val 445 450	1754
TTGATCGCAC ACGCAGGACA GCCCCACAAC CGGCGCGGCT GTGTTCAACG ACACACGATG	1814
AGTCCCCCAC TCGGTCTTGT ACTCGGATCC TTTT	1848



## (2) INFORMATION FOR SEQ ID NO:11:

## (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 453 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

Met Leu Glu Leu Leu Pro Thr Ala Val Glu Gly Val Ser Gln Ala Gln  
 1 5 10 15  
 Ile Gln Ala Leu Asp Tyr Lys Asp Asp Asp Val Asp Thr Phe Asn  
 20 25 30  
 Ser Ser Asp Gly Gly Arg Leu Phe Gln Phe Pro Asp Gly Val Gln Asn  
 35 40 45  
 Trp Pro Ala Leu Ser Ile Val Val Ile Ile Ile Met Thr Ile Gly Gly  
 50 55 60  
 Asn Ile Leu Val Ile Met Ala Val Ser Met Glu Lys Lys Leu His Asn  
 65 70 75 80  
 Ala Thr Asn Tyr Phe Leu Met Ser Leu Ala Ile Ala Asp Met Leu Val  
 85 90 95  
 Gly Leu Leu Val Met Pro Leu Ser Leu Leu Ala Ile Leu Tyr Asp Tyr  
 100 105 110  
 Val Trp Pro Leu Pro Arg Tyr Leu Cys Pro Val Trp Ile Ser Leu Asp  
 115 120 125  
 Val Leu Phe Ser Thr Ala Ser Ile Met His Leu Cys Ala Ile Ser Leu  
 130 135 140  
 Asp Arg Tyr Val Ala Ile Arg Asn Pro Ile Glu His Ser Arg Phe Asn  
 145 150 155 160  
 Ser Arg Thr Lys Ala Ile Met Lys Ile Ala Ile Val Trp Ala Ile Ser  
 165 170 175  
 Ile Gly Val Ser Val Pro Ile Pro Val Ile Gly Leu Arg Asp Glu Ser  
 180 185 190  
 Lys Val Phe Val Asn Asn Thr Thr Cys Val Leu Asn Asp Pro Asn Phe  
 195 200 205  
 Val Leu Ile Gly Ser Phe Val Ala Phe Phe Ile Pro Leu Thr Ile Met  
 210 215 220

Val Ile Thr Tyr Phe Leu Thr Ile Tyr Val Leu Arg Arg Gln Thr Leu  
 225 230 235 240  
 Met Leu Leu Arg Gly His Thr Glu Glu Glu Leu Ala Asn Met Ser Leu  
 245 250 255  
 Asn Phe Leu Asn Cys Cys Cys Lys Lys Asn Gly Gly Glu Glu Glu Asn  
 260 265 270  
 Ala Pro Asn Pro Asn Pro Asp Gln Lys Pro Arg Arg Lys Lys Lys Glu  
 275 280 285  
 Lys Arg Pro Arg Gly Thr Met Gln Ala Ile Asn Asn Glu Lys Lys Ala  
 290 295 300  
 Ser Lys Val Leu Gly Ile Val Phe Phe Val Phe Leu Ile Met Trp Cys  
 305 310 315 320  
 Pro Phe Phe Ile Thr Asn Ile Leu Ser Val Leu Cys Gly Lys Ala Cys  
 325 330 335  
 Asn Gln Lys Leu Met Glu Lys Leu Leu Asn Val Phe Val Trp Ile Gly  
 340 345 350  
 Tyr Val Cys Ser Gly Ile Asn Pro Leu Val Tyr Thr Leu Phe Asn Lys  
 355 360 365  
 Ile Tyr Arg Arg Ala Phe Ser Lys Tyr Leu Arg Cys Asp Tyr Lys Pro  
 370 375 380  
 Asp Lys Lys Pro Pro Val Arg Gln Ile Pro Arg Val Ala Ala Thr Ala  
 385 390 395 400  
 Leu Ser Gly Arg Glu Leu Asn Val Asn Ile Tyr Arg His Thr Asn Glu  
 405 410 415  
 Arg Val Ala Arg Lys Ala Asn Asp Pro Glu Pro Gly Ile Glu Met Gln  
 420 425 430  
 Val Glu Asn Leu Glu Leu Pro Val Asn Pro Ser Asn Val Val Ser Glu  
 435 440 445  
 Arg Ile Ser Ser Val  
 450

## (2) INFORMATION FOR SEQ ID NO:12:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 1764 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

- (A) NAME/KEY: repeat region
- (B) LOCATION: 436..462
- (D) OTHER INFORMATION: /note= "Sequence encoding polyaspartic acid."

(ix) FEATURE:

- (A) NAME/KEY: misc feature
- (B) LOCATION: 463..465
- (D) OTHER INFORMATION: /note= "Codon encoding the N-terminal amino acid of the human thrombin receptor protein."

(ix) FEATURE:

- (A) NAME/KEY: misc feature
- (B) LOCATION: 1630..1632
- (D) OTHER INFORMATION: /note= "Codon encoding the C-terminal amino acid of the human thrombin receptor protein. "

(ix) FEATURE:

- (A) NAME/KEY: repeat region
- (B) LOCATION: 1633..1650
- (D) OTHER INFORMATION: /note= "Sequence encoding polyhistidine."

(ix) FEATURE:

- (A) NAME/KEY: misc feature
- (B) LOCATION: 648..656
- (D) OTHER INFORMATION: /note= "Deleted AlwNI restriction site."

(ix) FEATURE:

- (A) NAME/KEY: misc feature
- (B) LOCATION: 893..898
- (D) OTHER INFORMATION: /note= "Deleted PstI restriction site."

(ix) FEATURE:

- (A) NAME/KEY: misc feature
- (B) LOCATION: 1301..1309
- (D) OTHER INFORMATION: /note= "Deleted AlwNI restriction site."

(ix) FEATURE:

- (A) NAME/KEY: misc feature
- (B) LOCATION: 1394..1402
- (D) OTHER INFORMATION: /note= "Deleted AlwNI restriction site."

- (ix) FEATURE:
  - (A) NAME/KEY: misc\_signal
  - (B) LOCATION: 374
  - (D) OTHER INFORMATION: /note= "RNA start site."
- (ix) FEATURE:
  - (A) NAME/KEY: mutation
  - (B) LOCATION: replace(1671, "")
  - (D) OTHER INFORMATION: /note= "C to T mutation removes AlwNI site."
- (ix) FEATURE:
  - (A) NAME/KEY: CDS
  - (B) LOCATION: 376..1650
- (ix) FEATURE:
  - (A) NAME/KEY: misc\_feature
  - (B) LOCATION: 376..414
  - (D) OTHER INFORMATION: /note= "Bacteriorhodopsin pre-sequence."
- (ix) FEATURE:
  - (A) NAME/KEY: terminator
  - (B) LOCATION: 1672..1674
  - (D) OTHER INFORMATION: /note= "Bacteriorhodopsin stop codon."
- (ix) FEATURE:
  - (A) NAME/KEY: terminator
  - (B) LOCATION: 1651..1653
  - (D) OTHER INFORMATION: /note= "Thrombin stop codon."

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

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ATCTGCAGGA TGGGTGCAAC CGTGAAGTCC GTCACGGCTG CGTCACGACA GGAGCCGACC      60
AGCGACACCC AGAAGGTGCG AACGGTTGAG TGCCGCAACG ATCACGAGTT TTTCGTGCGC      120
TTCGAGTGGT AACACGCGTG CACGCATCGA CTTCACCGCG GGTGTTTCGA CGCCAGCCGG      180
CCGTTGAACC AGCAGGCAGC GGGCATTTC AATCCGCTGT GGCCACACA CTCGGTGGGG      240
TGCGCTATTT TGGTATGGTT TGAATCCGC GTGTCGGCTC CGTGTCTGAC GGTTTCATCGG      300
TCTAAATTCC GTCACGAGCG TACCATACTG ATTGGGTCGT AGAGTTACAC ACATATCCTC      360
GTTAGGTACT GTTGC ATG TTG GAG TTA TTG CCA ACA GCA GTG GAG GGG GTA      411
          Met Leu Glu Leu Leu Pro Thr Ala Val Glu Gly Val
           1           5           10
  
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TCG CAG GCC CAG ATC CAG GCG CTG GAC TAC AAG GAC GAT GAT GAC GTC Ser Gln Ala Gln Ile Gln Ala Leu Asp Tyr Lys Asp Asp Asp Asp Val 15 20 25	459
GAC GCC ACC TTA GAT CCC CGG TCA TTT CTT CTC AGG AAC CCC AAT GAT Asp Ala Thr Leu Asp Pro Arg Ser Phe Leu Leu Arg Asn Pro Asn Asp 30 35 40	507
AAA TAT GAA CCA TTT TGG GAG GAT GAG GAG AAA AAT GAA AGT GGG TTA Lys Tyr Glu Pro Phe Trp Glu Asp Glu Glu Lys Asn Glu Ser Gly Leu 45 50 55 60	555
ACT GAA TAC AGA TTA GTC TCC ATC AAT AAA AGC AGT CCT CTT CAA AAA Thr Glu Tyr Arg Leu Val Ser Ile Asn Lys Ser Ser Pro Leu Gln Lys 65 70 75	603
CAA CTT CCT GCA TTC ATC TCA GAA GAT GCC TCC GGA TAT TTG ACC AGC Gln Leu Pro Ala Phe Ile Ser Glu Asp Ala Ser Gly Tyr Leu Thr Ser 80 85 90	651
TCC TGG CTG ACA CTC TTT GTC CCA TCT GTG TAC ACC GGA GTG TTT GTA Ser Trp Leu Thr Leu Phe Val Pro Ser Val Tyr Thr Gly Val Phe Val 95 100 105	699
GTC AGC CTC CCA CTA AAC ATC ATG GCC ATC GTT GTG TTC ATC CTG AAA Val Ser Leu Pro Leu Asn Ile Met Ala Ile Val Val Phe Ile Leu Lys 110 115 120	747
ATG AAG GTC AAG AAG CCG GCG GTG GTG TAC ATG CTG CAC CTG GCC ACG Met Lys Val Lys Lys Pro Ala Val Val Tyr Met Leu His Leu Ala Thr 125 130 135 140	795
GCA GAT GTG CTG TTT GTG TCT GTG CTC CCC TTT AAG ATC AGC TAT TAC Ala Asp Val Leu Phe Val Ser Val Leu Pro Phe Lys Ile Ser Tyr Tyr 145 150 155	843
TTT TCC GGC AGT GAT TGG CAG TTT GGG TCT GAA TTG TGT CGC TTC GTC Phe Ser Gly Ser Asp Trp Gln Phe Gly Ser Glu Leu Cys Arg Phe Val 160 165 170	891
ACT GCA GCA TTT TAC TGT AAC ATG TAC GCC TCT ATC TTG CTC ATG ACA Thr Ala Ala Phe Tyr Cys Asn Met Tyr Ala Ser Ile Leu Leu Met Thr 175 180 185	939
GTC ATA AGC ATT GAC CGG TTT CTG GCT GTG GTG TAT CCC ATG CAG TCC Val Ile Ser Ile Asp Arg Phe Leu Ala Val Val Tyr Pro Met Gln Ser 190 195 200	987
CTC TCC TGG CGT ACT CTG GGA AGG GCT TCC TTC ACT TGT CTG GCC ATC Leu Ser Trp Arg Thr Leu Gly Arg Ala Ser Phe Thr Cys Leu Ala Ile 205 210 215 220	1035

TGG GCT TTG GCC ATC GCA GGG GTA GTG CCT CTC GTC CTC AAG GAG CAA	1083
Trp Ala Leu Ala Ile Ala Gly Val Val Pro Leu Val Leu Lys Glu Gln	
225 230 235	
ACC ATC CAG GTG CCC GGG CTC AAC ATC ACT ACC TGT CAT GAT GTG CTC	1131
Thr Ile Gln Val Pro Gly Leu Asn Ile Thr Thr Cys His Asp Val Leu	
240 245 250	
AAT GAA ACC CTG CTC GAA GGC TAC TAT GCC TAC TAC TTC TCA GCC TTC	1179
Asn Glu Thr Leu Leu Glu Gly Tyr Tyr Ala Tyr Tyr Phe Ser Ala Phe	
255 260 265	
TCT GCT GTC TTC TTT TTT GTG CCG CTG ATC ATT TCC ACG GTC TGT TAT	1227
Ser Ala Val Phe Phe Phe Val Pro Leu Ile Ile Ser Thr Val Cys Tyr	
270 275 280	
GTG TCT ATC ATT CGA TGT CTT AGC TCT TCC GCA GTT GCC AAC CGC AGC	1275
Val Ser Ile Ile Arg Cys Leu Ser Ser Ser Ala Val Ala Asn Arg Ser	
285 290 300	
AAG AAG TCC CGG GCT TTG TTC CTG TCA GCT GCT GTT TTC TGC ATC TTC	1323
Lys Lys Ser Arg Ala Leu Phe Leu Ser Ala Ala Val Phe Cys Ile Phe	
305 310 315	
ATC ATT TGC TTC GGA CCC ACA AAC GTC CTC CTG ATT GCG CAT TAC TCA	1371
Ile Ile Cys Phe Gly Pro Thr Asn Val Leu Leu Ile Ala His Tyr Ser	
320 325 330	
TTC CTT TCT CAC ACT TCC ACC ACA GAG GCT GCC TAC TTT GCC TAC CTC	1419
Phe Leu Ser His Thr Ser Thr Thr Glu Ala Ala Tyr Phe Ala Tyr Leu	
335 340 345	
CTC TGT GTC TGT GTC AGC AGC ATA AGC TCG TGC ATC GAC CCC CTA ATT	1467
Leu Cys Val Cys Val Ser Ser Ile Ser Ser Cys Ile Asp Pro Leu Ile	
350 355 360	
TAC TAT TAC GCT TCC TCT GAG TGC CAG AGG TAC GTC TAC AGT ATC TTA	1515
Tyr Tyr Tyr Ala Ser Ser Glu Cys Gln Arg Tyr Val Tyr Ser Ile Leu	
365 370 375 380	
TGC TGC AAA GAA AGT TCC GAT CCC AGC AGT TAT AAC AGC AGT GGG CAG	1563
Cys Cys Lys Glu Ser Ser Asp Pro Ser Ser Tyr Asn Ser Ser Gly Gln	
385 390 395	
TTG ATG GCA AGT AAA ATG GAT ACC TGC TCT AGT AAC CTG AAT AAC AGC	1611
Leu Met Ala Ser Lys Met Asp Thr Cys Ser Ser Asn Leu Asn Asn Ser	
400 405 410	
ATA TAC AAA AAG CTG TTA ACT CAC CAC CAC CAC CAC TGAGCGGCCG	1660
Ile Tyr Lys Lys Leu Leu Thr His His His His His His	
415 420 425	

CGACCAGCGA TTGATCGCAC ACGCAGGACA GCCCCACAAC CGGCGCGGCT GTGTTCAACG 1720  
 ACACACGATG AGTCCCCCAC TCGGTCTTGT ACTCGGATCC TTTT 1764

(2) INFORMATION FOR SEQ ID NO:13:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 425 amino acids
  - (B) TYPE: amino acid
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

Met Leu Glu Leu Leu Pro Thr Ala Val Glu Gly Val Ser Gln Ala Gln  
 1 5 10 15  
 Ile Gln Ala Leu Asp Tyr Lys Asp Asp Asp Val Asp Ala Thr Leu  
 20 25 30  
 Asp Pro Arg Ser Phe Leu Leu Arg Asn Pro Asn Asp Lys Tyr Glu Pro  
 35 40 45  
 Phe Trp Glu Asp Glu Glu Lys Asn Glu Ser Gly Leu Thr Glu Tyr Arg  
 50 55 60  
 Leu Val Ser Ile Asn Lys Ser Ser Pro Leu Gln Lys Gln Leu Pro Ala  
 65 70 75 80  
 Phe Ile Ser Glu Asp Ala Ser Gly Tyr Leu Thr Ser Ser Trp Leu Thr  
 85 90 95  
 Leu Phe Val Pro Ser Val Tyr Thr Gly Val Phe Val Val Ser Leu Pro  
 100 105 110  
 Leu Asn Ile Met Ala Ile Val Val Phe Ile Leu Lys Met Lys Val Lys  
 115 120 125  
 Lys Pro Ala Val Val Tyr Met Leu His Leu Ala Thr Ala Asp Val Leu  
 130 135 140  
 Phe Val Ser Val Leu Pro Phe Lys Ile Ser Tyr Tyr Phe Ser Gly Ser  
 145 150 155 160  
 Asp Trp Gln Phe Gly Ser Glu Leu Cys Arg Phe Val Thr Ala Ala Phe  
 165 170 175  
 Tyr Cys Asn Met Tyr Ala Ser Ile Leu Leu Met Thr Val Ile Ser Ile  
 180 185 190

Asp Arg Phe Leu Ala Val Val Tyr Pro Met Gln Ser Leu Ser Trp Arg  
 195 200 205  
 Thr Leu Gly Arg Ala Ser Phe Thr Cys Leu Ala Ile Trp Ala Leu Ala  
 210 215 220  
 Ile Ala Gly Val Val Pro Leu Val Leu Lys Glu Gln Thr Ile Gln Val  
 225 230 235 240  
 Pro Gly Leu Asn Ile Thr Thr Cys His Asp Val Leu Asn Glu Thr Leu  
 245 250 255  
 Leu Glu Gly Tyr Tyr Ala Tyr Tyr Phe Ser Ala Phe Ser Ala Val Phe  
 260 265 270  
 Phe Phe Val Pro Leu Ile Ile Ser Thr Val Cys Tyr Val Ser Ile Ile  
 275 280 285  
 Arg Cys Leu Ser Ser Ser Ala Val Ala Asn Arg Ser Lys Lys Ser Arg  
 290 295 300  
 Ala Leu Phe Leu Ser Ala Ala Val Phe Cys Ile Phe Ile Ile Cys Phe  
 305 310 315 320  
 Gly Pro Thr Asn Val Leu Leu Ile Ala His Tyr Ser Phe Leu Ser His  
 325 330 335  
 Thr Ser Thr Thr Glu Ala Ala Tyr Phe Ala Tyr Leu Leu Cys Val Cys  
 340 345 350  
 Val Ser Ser Ile Ser Ser Cys Ile Asp Pro Leu Ile Tyr Tyr Tyr Ala  
 355 360 365  
 Ser Ser Glu Cys Gln Arg Tyr Val Tyr Ser Ile Leu Cys Cys Lys Glu  
 370 375 380  
 Ser Ser Asp Pro Ser Ser Tyr Asn Ser Ser Gly Gln Leu Met Ala Ser  
 385 390 395 400  
 Lys Met Asp Thr Cys Ser Ser Asn Leu Asn Asn Ser Ile Tyr Lys Lys  
 405 410 415  
 Leu Leu Thr His His His His His His  
 420 425

## (2) INFORMATION FOR SEQ ID NO:14:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 2147 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear



- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
  - (A) NAME/KEY: misc\_signal
  - (B) LOCATION: 378..380
  - (D) OTHER INFORMATION: /note= "Bacteriorhodopsin start codon."
- (ix) FEATURE:
  - (A) NAME/KEY: misc\_feature
  - (B) LOCATION: 378..416
  - (D) OTHER INFORMATION: /note= "Bacteriorhodopsin pre-sequence."
- (ix) FEATURE:
  - (A) NAME/KEY: CDS
  - (B) LOCATION: 378..2054
- (ix) FEATURE:
  - (A) NAME/KEY: misc\_feature
  - (B) LOCATION: 417..419
  - (D) OTHER INFORMATION: /note= "Codon encoding N-terminal amino acid of mature bacteriorhodopsin."
- (ix) FEATURE:
  - (A) NAME/KEY: misc\_feature
  - (B) LOCATION: 1122..1124
  - (D) OTHER INFORMATION: /note= "Codon encoding amino acid number 236 of bacteriorhodopsin."
- (ix) FEATURE:
  - (A) NAME/KEY: misc\_feature
  - (B) LOCATION: 1137..1139
  - (D) OTHER INFORMATION: /note= "Codon encoding amino acid number 6 of the catalytic subunit of E. coli Aspartate Transcarbamylase."
- (ix) FEATURE:
  - (A) NAME/KEY: misc\_feature
  - (B) LOCATION: 1125..1178
  - (D) OTHER INFORMATION: /note= "Synthetic DNA fragment."
- (ix) FEATURE:
  - (A) NAME/KEY: misc\_feature
  - (B) LOCATION: 1125..1136
  - (D) OTHER INFORMATION: /note= "Sequence encoding Factor Xa proteolytic site."
- (ix) FEATURE:
  - (A) NAME/KEY: misc\_feature
  - (B) LOCATION: 2037..2039
  - (D) OTHER INFORMATION: /note= "Codon encoding amino acid number 306 of E. coli Aspartate Transcarbamylase."

(ix) FEATURE:

- (A) NAME/KEY: misc feature
- (B) LOCATION: 2040..2054
- (D) OTHER INFORMATION: /note= "Sequence encoding bacteriorhodopsin C-terminal amino acid numbers 245 through 249. "

(ix) FEATURE:

- (A) NAME/KEY: terminator
- (B) LOCATION: 2055..2057
- (D) OTHER INFORMATION: /note= "Bacteriorhodopsin stop codon."

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

TAATCTGCAG GATGGGTGCA ACCGTGAAGT CCGTCACGGC TCGTCACGA CAGGAGCCGA	60
CCAGCGACAC CCAGAAGGTG CGAACGGTTG AGTGCCGCAA CGATCACGAG TTTTTCGTGC	120
GCTTCGAGTG GTAACACGCG TGCACGCATC GACTTCACCG CGGGTGTTC GACGCCAGCC	180
GGCCGTTGAA CCAGCAGGCA GCGGGCATT CACAGCCGCT GTGGCCACA CACTCGGTGG	240
GGTGCCTAT TTTGGTATGG TTTGGAATCC GCGTGTGGC TCCGTGTCTG ACGTTCATC	300
GGTCTAAATT CCGTCACGAG CGTACCATAC TGATTGGGTC GTAGAGTTAC ACACATATCC	360
TCGTTAGGTA CTGTTGC ATG TTG GAG TTA TTG CCA ACA GCA GTG GAG GGG	410
Met Leu Glu Leu Leu Pro Thr Ala Val Glu Gly	
1 5 10	
GTA TCG CAG GCC CAG ATC ACC GGA CGT CCG GAG TGG ATC TGG CTA GCG	458
Val Ser Gln Ala Gln Ile Thr Gly Arg Pro Glu Trp Ile Trp Leu Ala	
15 20 25	
CTC GGT ACG GCG CTA ATG GGA CTC GGG ACG CTC TAT TTC CTC GTG AAA	506
Leu Gly Thr Ala Leu Met Gly Leu Gly Thr Leu Tyr Phe Leu Val Lys	
30 35 40	
GGG ATG GGC GTC TCG GAC CCA GAT GCA AAG AAA TTC TAC GCC ATC ACG	554
Gly Met Gly Val Ser Asp Pro Asp Ala Lys Lys Phe Tyr Ala Ile Thr	
45 50 55	
ACG CTC GTC CCA GCC ATC GCG TTC ACG ATG TAC CTC TCG ATG CTG CTG	602
Thr Leu Val Pro Ala Ile Ala Phe Thr Met Tyr Leu Ser Met Leu Leu	
60 65 70 75	
GGG TAT GGC CTC ACA ATG GTA CCG TTC GGT GGG GAG CAG AAC CCC ATC	650
Gly Tyr Gly Leu Thr Met Val Pro Phe Gly Gly Glu Gln Asn Pro Ile	
80 85 90	

TAC Tyr	TGG Trp	GCG Ala	CGG Arg 95	TAC Tyr	GCT Ala	GAC Asp	TGG Trp	CTG Leu	TTC Phe	ACC Thr	ACG Thr	CCG Pro	CTG Leu	TTG Leu	TTG Leu	698
TTA Leu	GAC Asp	CTC Leu	GCG Ala 110	TTG Leu	CTC Leu	GTT Val	GAC Asp 115	GCG Ala	GAT Asp	CAG Gln	GGA Gly	ACG Thr	ATC Ile	CTT Leu	GCG Ala	746
CTC Leu	GTC Val	GGT Gly	GCC Ala	GAC Asp	GGC Gly	ATC Ile	ATG Met	ATC Ile	GGG Gly	ACC Thr	GGC Gly	CTG Leu	GTC Val	GGC Gly	GCA Ala	794
CTG Leu	ACG Thr	AAG Lys	GTC Val	TAC Tyr	TCG Ser	TAC Tyr	CGC Arg	TTC Phe	GTG Val	TGG Trp	TGG Trp	GCG Ala	ATC Ile	AGC Ser	ACC Thr	842
GCA Ala	GCG Ala	ATG Met	CTG Leu	TAC Tyr	ATC Ile	CTG Leu	TAC Tyr	GTG Val	CTG Leu	TTC Phe	TTC Phe	GGG Gly	TTC Phe	ACC Thr	TCG Ser	890
AAG Lys	GCC Ala	GAA Glu	AGC Ser	ATG Met	CGC Arg	CCC Pro	GAG Glu	GTC Val	GCA Ala	TCC Ser	ACG Thr	TTC Phe	AAA Lys	GTA Val	CTG Leu	938
CGT Arg	AAC Asn	GTT Val	ACC Thr	GTT Val	GTG Val	TTG Leu	TGG Trp	TCC Ser	GCG Ala	TAT Tyr	CCC Pro	GTC Val	GTG Val	TGG Trp	CTG Leu	986
ATC Ile	GGC Gly	AGC Ser	GAA Glu	GGT Gly	GCG Ala	GGA Gly	ATC Ile	GTG Val	CCG Pro	CTG Leu	AAC Asn	ATC Ile	GAG Glu	ACG Thr	CTG Leu	1034
CTG Leu	TTC Phe	ATG Met	GTG Val	CTT Leu	GAC Asp	GTG Val	AGC Ser	GCG Ala	AAG Lys	GTC Val	GGC Gly	TTC Phe	GGG Gly	CTC Leu	ATC Ile	1082
CTC Leu	CTG Leu	CGC Arg	AGT Ser	CGT Arg	GCG Ala	ATC Ile	TTC Phe	GGC Gly	GAA Glu	GCC Ala	GAA Glu	GCG Ala	CCG Pro	ATC Ile	GAA Glu	1130
GGT Gly	CGT Arg	CAG Gln	AAA Lys	CAT His	ATC Ile	ATT Ile	TCC Ser	ATA Ile	AAC Asn	GAC Asp	CTT Leu	AGT Ser	CGC Arg	GAT Asp	GAC Asp	1178
CTT Leu	AAT Asn	CTG Leu	GTG Val	CTG Leu	GCG Ala	ACA Thr	GCG Ala	GCG Ala	AAA Lys	CTG Leu	AAA Lys	GCA Ala	AAC Asn	CCG Pro	CAA Gln	1226
CCA Pro	GAG Glu	CTG Leu	TTG Leu	AAG Lys	CAC His	AAA Lys	GTC Val	ATT Ile	GCC Ala	AGC Ser	TGT Cys	TTC Phe	TTC Phe	GAA Glu	GCC Ala	1274

TCT ACC CGT ACC CGC CTC TCT TTT CAA ACA TCT ATG CAC CGC CTG GGG Ser Thr Arg Thr Arg Leu Ser Phe Gln Thr Ser Met His Arg Leu Gly 300 305 310 315	1322
GCC AGC GTG GTG GGC TTC TCC GAC AGC GCC AAT ACA TCA CTG GGT AAA Ala Ser Val Val Gly Phe Ser Asp Ser Ala Asn Thr Ser Leu Gly Lys 320 325 330	1370
AAA GGC GAA ACG CTT GCC GAT ACC ATT TCA GTT ATC AGC ACT TAC GTC Lys Gly Glu Thr Leu Ala Asp Thr Ile Ser Val Ile Ser Thr Tyr Val 335 340 345	1418
GAT GCG ATA GTG ATG CGT CAT CCG CAG GAA GGT GCG GCG CGC CTG GCC Asp Ala Ile Val Met Arg His Pro Gln Glu Gly Ala Ala Arg Leu Ala 350 355 360	1466
ACC GAG TTT TCC GGC AAT GTA CCG GTA CTG AAT GCC GGT GAT GGC TCC Thr Glu Phe Ser Gly Asn Val Pro Val Leu Asn Ala Gly Asp Gly Ser 365 370 375	1514
AAC CAA CAT CCG ACG CAA ACC TTG CTG GAC TTA TTC ACT ATT CAG GAA Asn Gln His Pro Thr Gln Thr Leu Leu Asp Leu Phe Thr Ile Gln Glu 380 385 390 395	1562
ACC CAG GGG CGT CTG GAC AAT CTC CAC GTC GCA ATG GTT GGT GAC CTG Thr Gln Gly Arg Leu Asp Asn Leu His Val Ala Met Val Gly Asp Leu 400 405 410	1610
AAA TAT GGT CGC ACC GTT CAC TCC CTG ACT CAG GCG TTA GCT AAG TTC Lys Tyr Gly Arg Thr Val His Ser Leu Thr Gln Ala Leu Ala Lys Phe 415 420 425	1658
GAC GGC AAC CGT TTT TAC TTC ATC GCG CCG GAC GCG CTG GCA ATG CCG Asp Gly Asn Arg Phe Tyr Phe Ile Ala Pro Asp Ala Leu Ala Met Pro 430 435 440	1706
CAA TAC ATT CTG GAT ATG CTC GAT GAA AAA GGG ATC GCA TGG AGT CTG Gln Tyr Ile Leu Asp Met Leu Asp Glu Lys Gly Ile Ala Trp Ser Leu 445 450 455	1754
CAC AGC TCT ATT GAA GAA GTG ATG GTG GAA GTA GAC ATC CTG TAC ATG His Ser Ser Ile Glu Glu Val Met Val Glu Val Asp Ile Leu Tyr Met 460 465 470 475	1802
ACC CGC GTG CAA AAA GAG CGT CTG GAC CCG TCC GAG TAC GCC AAC GTG Thr Arg Val Gln Lys Glu Arg Leu Asp Pro Ser Glu Tyr Ala Asn Val 480 485 490	1850
AAA GCG CAG TTT GTT CTT CGC GCC AGT GAT CTC CAC AAC GCC AAA GCC Lys Ala Gln Phe Val Leu Arg Ala Ser Asp Leu His Asn Ala Lys Ala 495 500 505	1898

AAT ATG AAA GTG CTG CAT CCG TTG CCG CGT GTT GAT GAG ATT GCG ACG 1946  
 Asn Met Lys Val Leu His Pro Leu Pro Arg Val Asp Glu Ile Ala Thr  
 510 515 520

GAT GTT GAT AAA ACG CCA CAC GCC TGG TAC TTC CAG CAG GCA GGC AAC 1994  
 Asp Val Asp Lys Thr Pro His Ala Trp Tyr Phe Gln Gln Ala Gly Asn  
 525 530 535

GGG ATT TTC GCT CTG CAA GCG TTA CTG GCA CTG GTT CTG AAT CGG GCC 2042  
 Gly Ile Phe Ala Leu Gln Ala Leu Leu Ala Leu Val Leu Asn Arg Ala  
 540 545 550 555

GCG ACC AGC GAC TGATCGCACA CGCAGGACAG CCCACAACC GCGCGGCTG 2094  
 Ala Thr Ser Asp

TGTTCAACGA CACACGATGA GTCCCCACT CGGTCTTGTA CTCGGATCCT TTT 2147

(2) INFORMATION FOR SEQ ID NO:15:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 559 amino acids
  - (B) TYPE: amino acid
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

Met Leu Glu Leu Leu Pro Thr Ala Val Glu Gly Val Ser Gln Ala Gln  
 1 5 10 15

Ile Thr Gly Arg Pro Glu Trp Ile Trp Leu Ala Leu Gly Thr Ala Leu  
 20 25 30

Met Gly Leu Gly Thr Leu Tyr Phe Leu Val Lys Gly Met Gly Val Ser  
 35 40 45

Asp Pro Asp Ala Lys Lys Phe Tyr Ala Ile Thr Thr Leu Val Pro Ala  
 50 55 60

Ile Ala Phe Thr Met Tyr Leu Ser Met Leu Leu Gly Tyr Gly Leu Thr  
 65 70 75 80

Met Val Pro Phe Gly Gly Glu Gln Asn Pro Ile Tyr Trp Ala Arg Tyr  
 85 90 95

Ala Asp Trp Leu Phe Thr Thr Pro Leu Leu Leu Asp Leu Ala Leu  
 100 105 110

Leu Val Asp Ala Asp Gln Gly Thr Ile Leu Ala Leu Val Gly Ala Asp  
 115 120 125

Gly Ile Met Ile Gly Thr Gly Leu Val Gly Ala Leu Thr Lys Val Tyr  
 130 135 140  
 Ser Tyr Arg Phe Val Trp Trp Ala Ile Ser Thr Ala Ala Met Leu Tyr  
 145 150 155 160  
 Ile Leu Tyr Val Leu Phe Phe Gly Phe Thr Ser Lys Ala Glu Ser Met  
 165 170 175  
 Arg Pro Glu Val Ala Ser Thr Phe Lys Val Leu Arg Asn Val Thr Val  
 180 185 190  
 Val Leu Trp Ser Ala Tyr Pro Val Val Trp Leu Ile Gly Ser Glu Gly  
 195 200 205  
 Ala Gly Ile Val Pro Leu Asn Ile Glu Thr Leu Leu Phe Met Val Leu  
 210 215 220  
 Asp Val Ser Ala Lys Val Gly Phe Gly Leu Ile Leu Leu Arg Ser Arg  
 225 230 235 240  
 Ala Ile Phe Gly Glu Ala Glu Ala Pro Ile Glu Gly Arg Gln Lys His  
 245 250 255  
 Ile Ile Ser Ile Asn Asp Leu Ser Arg Asp Asp Leu Asn Leu Val Leu  
 260 265 270  
 Ala Thr Ala Ala Lys Leu Lys Ala Asn Pro Gln Pro Glu Leu Leu Lys  
 275 280 285  
 His Lys Val Ile Ala Ser Cys Phe Phe Glu Ala Ser Thr Arg Thr Arg  
 290 295 300  
 Leu Ser Phe Gln Thr Ser Met His Arg Leu Gly Ala Ser Val Val Gly  
 305 310 315 320  
 Phe Ser Asp Ser Ala Asn Thr Ser Leu Gly Lys Lys Gly Glu Thr Leu  
 325 330 335  
 Ala Asp Thr Ile Ser Val Ile Ser Thr Tyr Val Asp Ala Ile Val Met  
 340 345 350  
 Arg His Pro Gln Glu Gly Ala Ala Arg Leu Ala Thr Glu Phe Ser Gly  
 355 360 365  
 Asn Val Pro Val Leu Asn Ala Gly Asp Gly Ser Asn Gln His Pro Thr  
 370 375 380  
 Gln Thr Leu Leu Asp Leu Phe Thr Ile Gln Glu Thr Gln Gly Arg Leu  
 385 390 395 400  
 Asp Asn Leu His Val Ala Met Val Gly Asp Leu Lys Tyr Gly Arg Thr  
 405 410 415



CLAIMS

1. An expression vector useful for the production of heterologous polypeptide in a halobacterial host comprising:
  - a) transcription and translation regulatory DNA for operable expression of DNA in the 3'-position of said regulatory DNA;
  - b) DNA encoding a heterologous polypeptide 3' of said regulatory DNA; and
  - c) DNA encoding transcription and translation stop signals 3' of said heterologous DNA.
2. The vector according to Claim 1 having DNA encoding replication and selection capability for said halobacterial host.
3. The vector according to Claim 1 containing additional DNA encoding the pre-sequence of bacteriorhodopsin gene between said regulatory DNA and said DNA encoding said heterologous polypeptide for expression of a fusion polypeptide of said pre-sequence with said heterologous polypeptide.
4. The vector according to Claim 1 or 3 containing additional DNA encoding a C-terminal sequence of the bacteriorhodopsin gene and DNA operably encoding a unique protease cleavage site and a restriction site, in optional order, between said C-terminal sequence and said DNA encoding said heterologous polypeptide, said additional DNA being 3' of said DNA encoding said heterologous polypeptide..
5. The vector according to Claim 1 wherein said transcription and translation regulatory sequences and said transcription and translation stop signals are those of the bacteriorhodopsin gene.
6. The vector according to Claim 4 wherein said transcription and translation regulatory sequences and said transcription and translation stop signals are those of the bacteriorhodopsin gene.

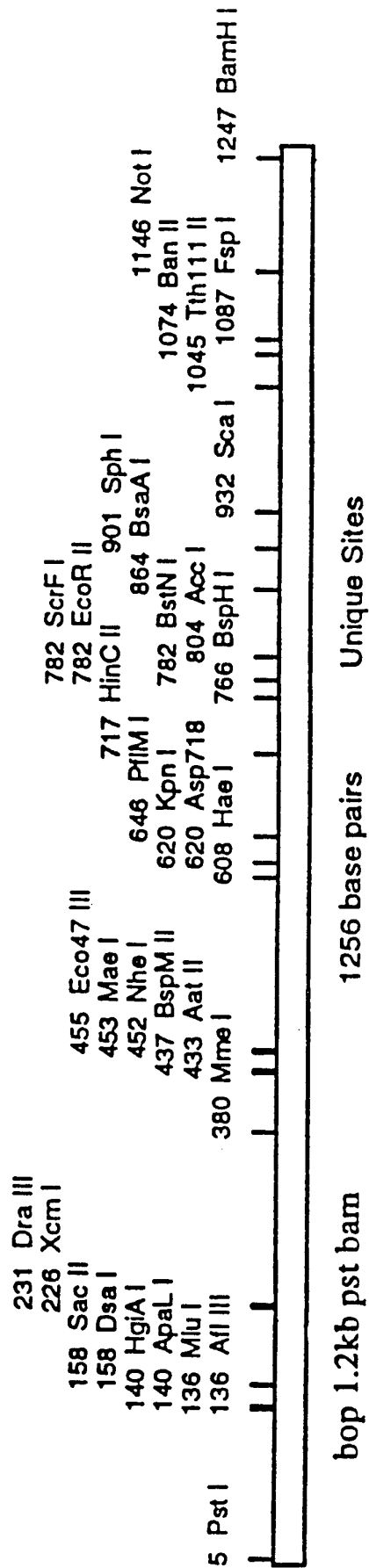


7. The vector according to Claim 1 or 2 wherein said heterologous polypeptide is Type HM1 human muscarinic acetylcholine receptor.
8. The vector according to Claim 4 wherein said heterologous polypeptide is the catalytic subunit of aspartate transcarbamylase from *Escherichia coli*.
- 5 9. The vector according to Claim 5 wherein said heterologous polypeptide is the catalytic subunit of aspartate transcarbamylase from *Escherichia coli*.
10. A halobacterial host transformed with a vector according to Claim 1.
11. A halobacterial host transformed with a vector according to Claim 4.
12. A halobacterial host transformed with a vector according to Claim 5.
- 10 13. A method for producing a heterologous polypeptide in a halobacterial host comprising:
  - a) obtaining the elements necessary for operable expression comprising:
    - 15 i) transcription and translation regulatory DNA for operable expression of DNA in the 3'-position of said regulatory DNA;
    - ii) DNA encoding a heterologous polypeptide 3' of said regulatory DNA; and
    - 20 iii) DNA encoding transcription and translation stop signals 3' of said heterologous DNA;
  - b) operably assembling the elements of a);
  - c) transforming a halobacterial host with said assembled elements;
  - d) causing expression of said DNA encoding the heterologous polypeptide;
  - 25 e) isolating said heterologous polypeptide; and
  - f) optionally further purifying said heterologous polypeptide.

14. The method according to Claim 13 wherein said elements include additional DNA encoding a C-terminal sequence of the bacteriorhodopsin gene and DNA operably encoding a unique protease cleavage site and a restriction site, in optional order, between said C-terminal sequence and said DNA encoding  
5 said heterologous polypeptide, said additional DNA being 3' of said DNA encoding said heterologous polypeptide.
15. The method according to Claim 13 wherein said transcription and translation regulatory sequences and said transcription and translation stop signals are those of the bacteriorhodopsin gene.
- 10 16. A method according to Claim 13 wherein said heterologous polypeptide is the catalytic subunit of aspartate transcarbamylase from *Escherichia coli*.
17. A method for producing a heterologous polypeptide in a halobacterial host comprising:
- 15 a) causing expression of DNA encoding said heterologous polypeptide within an operable expression vector transformed into said halobacterium;
- b) isolating said heterologous polypeptide; and
- c) optionally further purifying said heterologous polypeptide.
18. A method for producing a heterologous polypeptide in a halobacterial host  
20 comprising:
- a) causing expression of DNA encoding said heterologous polypeptide and a C-terminal sequence of the bacteriorhodopsin gene within an operable expression vector transformed into a halobacterium, said C-terminal sequence being 3' of said DNA encoding the  
25 heterologous polypeptide;
- b) separating the membrane of said halobacterium after expression of said DNA encoding a heterologous polypeptide and bacteriorhodopsin C-terminal region;

- c) isolating said heterologous polypeptide; and
- d) optionally further purifying said heterologous polypeptide.

19. The method according to Claim 15 wherein said DNA further comprises additional DNA operably encoding a unique protease cleavage site between said  
5 DNA encoding said heterologous polypeptide and said DNA encoding said bacteriorhodopsin C-terminal sequence, said additional DNA being 3' of said DNA encoding said heterologous polypeptide.



bop 1.2kb pst bam 1256 base pairs Unique Sites

FIG. 1

ATCTGCAGGA TGGGTGCAAC CGTGAAGTCC GTCACGGCTG CGTCACGACA GGAGCCGACC 60  
 AGCGACACCC AGAAGGTGCC AACGGTTGAG TGCCGCAACG ATCACGAGTT TTTCGTGCCG 120  
 TTCGAGTGGT AACACGGGTG CACGCATCGA CTTCACCGCG GGTGTTTCGA CGCCAGCCCGG 180  
 CCGTTGAACC AGCAGGCAGC GGGCATTICA CAGCCGCTGT GGCCACACACA CTCGGTGGGG 240  
 TGGGCTATTT TGGTATGGTT TGGAAATCCG GTGTCCGGCTC CGTGTCTGAC GGTTCATCGG 300  
 TCTAAATICC GTCACGAGCG TACCATACTG ATTGGGTCGT AGAGTTACAC ACATATCCTC 360  
 GTTAGGTA CTGTC ATG TTG GAG TTA TTG CCA ACA GCA GTG GAG GGG GTA 411  
 Met Leu Glu Leu Leu Pro Thr Ala Val Glu Gly Val  
 1 5 10  
 TCG CAG GCC CAG ATC ACC GGA CGT CCG GAG TGG ATC TGG CTA GCG CTC 459  
 Ser Gln Ala Gln Ile Thr Gly Arg Pro Glu Trp Ile Trp Leu Ala Leu  
 15 20 25  
 GGT ACG GCG CTA ATG GGA CTC GGG ACG CTC TAT TTC CTC GTG AAA GGG 507  
 Gly Thr Ala Leu Met Gly Leu Gly Thr Leu Tyr Phe Leu Val Lys Gly  
 30 35 40

FIG. 2(A)

ATG GGC GTC TCG GAC CCA GAT GCA AAG AAA TTC TAC GCC ATC ACG ACG	555
Met Gly Val Ser Asp Pro Asp Ala Lys Lys Phe Tyr Ala Ile Thr Thr	60
45	50
55	60
CTC GTC CCA GCC ATC GCG TTC ACG ATG TAC CTC TCG ATG CTG CTG GGG	603
Leu Val Pro Ala Ile Ala Phe Thr Met Tyr Leu Ser Met Leu Leu Gly	75
65	70
TAT GGC CTC ACA ATG GTA CCG TTC GGT GGG GAG CAG AAC CCC ATC TAC	651
Tyr Gly Leu Thr Met Val Pro Phe Gly Gly Glu Gln Asn Pro Ile Tyr	90
80	85
IGG GCG CCG TAC GCT GAC TGG CTG TTC ACC ACG CCG CTG TTG TTA	699
Trp Ala Arg Tyr Ala Asp Trp Leu Phe Thr Thr Pro Leu Leu Leu	105
95	100
GAC CTC GCG TTG CTC GTT GAC CCG GAT CAG GGA ACG ATC CTT GCG CTC	747
Asp Leu Ala Leu Leu Val Asp Ala Asp Gln Gly Thr Ile Leu Ala Leu	120
110	115
GTC GGT GCC GAC GGC ATC ATG GCG ACC GGC CTG GTC GGC GCA CTG	795
Val Gly Ala Asp Gly Ile Met Ile Gly Thr Gly Leu Val Gly Ala Leu	140
125	130
	135

FIG. 2(B)

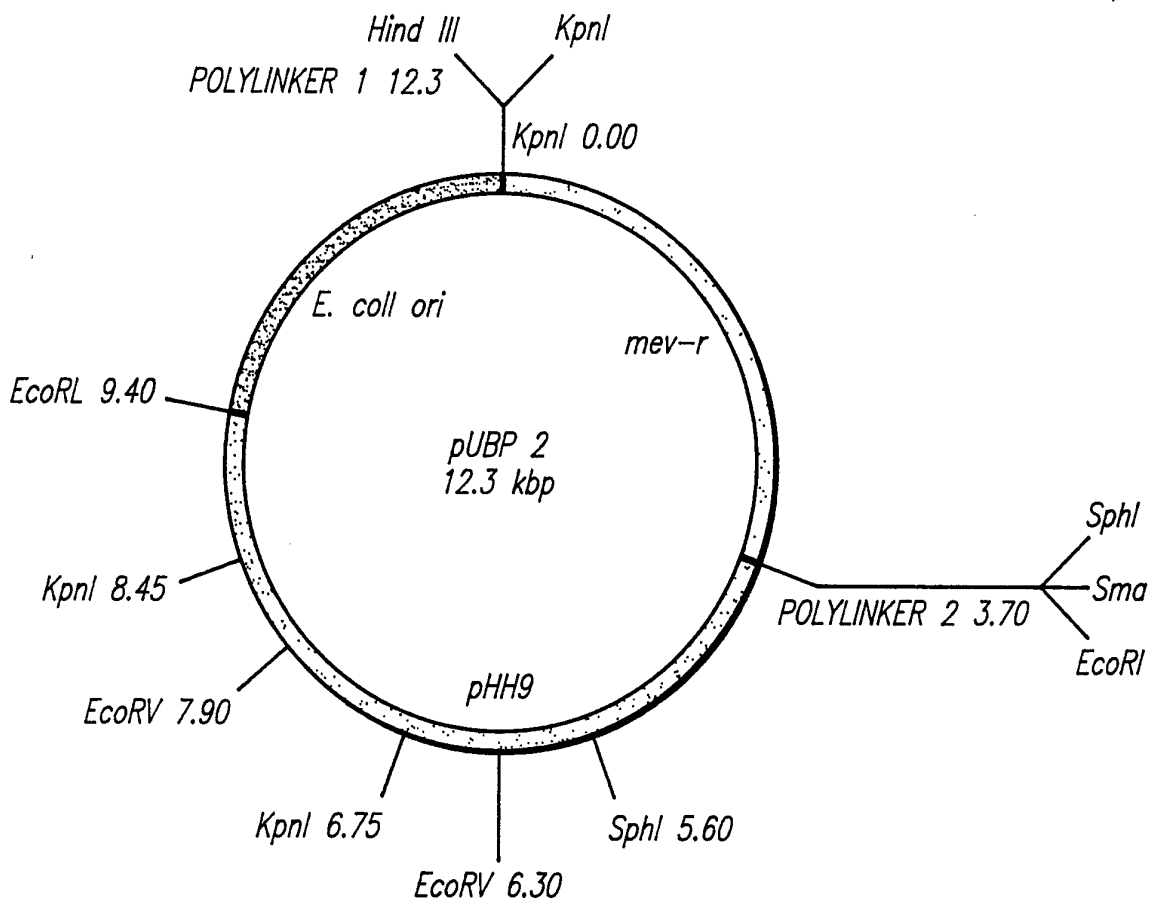
ACG AAG GTC TAC TCG TAC CGC TTC GTG TGG GCG ATC AGC ACC GCA	843
Thr Lys Val Tyr Ser Tyr Arg Phe Val Trp Trp Ala Ile Ser Thr Ala	155
	145
GCG ATG CTG TAC ATC CTG TAC GTG TTC TTC GGG TTC ACC TCG AAG	891
Ala Met Leu Tyr Ile Leu Tyr Val Leu Phe Phe Gly Phe Thr Ser Lys	170
	160
	165
GCC GAA AGC ATG CGC CCC GAG GTC GCA TCC ACG TTC AAA GTA CTG CGT	939
Ala Glu Ser Met Arg Pro Glu Val Ala Ser Thr Phe Lys Val Leu Arg	185
	175
	180
AAC GTT ACC GTG TTG TGG TCC GCG TAT CCC GTC GTG TGG CTG ATC	987
Asn Val Thr Val Val Leu Trp Ser Ala Tyr Pro Val Val Trp Leu Ile	200
	190
	195
GCC AGC GAA GGT GCG GGA ATC GTG CCG CTG AAC ATC GAG ACG CTG CTG	1035
Gly Ser Glu Gly Ala Gly Ile Val Pro Leu Asn Ile Glu Thr Leu Leu	220
	205
	210
	215
TTC ATG GTG CTT GAC GTG AGC GCG AAG GTC GGC TTC GGG CTC ATC CTC	1083
Phe Met Val Leu Asp Val Ser Ala Lys Val Gly Phe Gly Leu Ile Leu	235
	225
	230

FIG. 2(C)

CTG CGC AGT CGT GCG ATC TTC GGC GAA GCC GAA GCG CCG GAG CCG TCC 1131  
 Leu Arg Ser Arg Ala Ile Phe Gly Ala Glu Ala Pro Glu Pro Ser  
 240 245 250  
 GCC GGC GAC GCG GCG ACC AGC GAC TGATCGCACA CGCAGGACAG 1181  
 Ala Gly Asp Gly Ala Ala Thr Ser Asp  
 255 260  
 CCCCACAACC GCGCGGCTG TGTTCAACGA CACACGATGA GTCCCCCACT CCGTCTTGTA 1241  
 CTCGGATCCT TTT 1254

FIG. 2(D)





POLYLINKER 1 : 12.3/HindIII.SphI.MluI.XhoI.PstI.Sall.XbaI.BamHI.HindIII.XbaI.KpnI.  
 POLYLINKER 2 : 3.7/SphI.EcoR5.SstI.SmaI.EcoRI.

FIG. 3



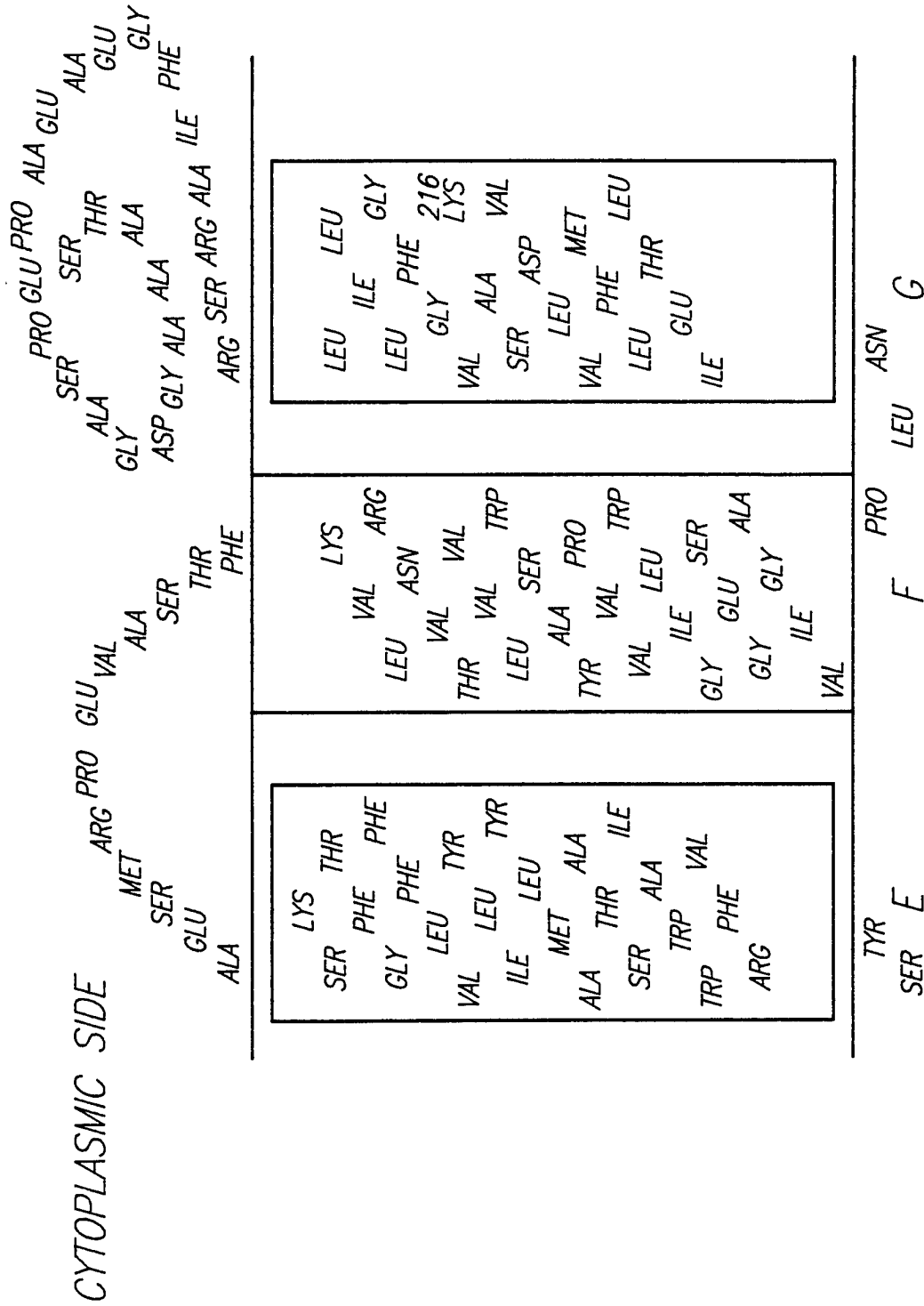


FIG. 4(B)

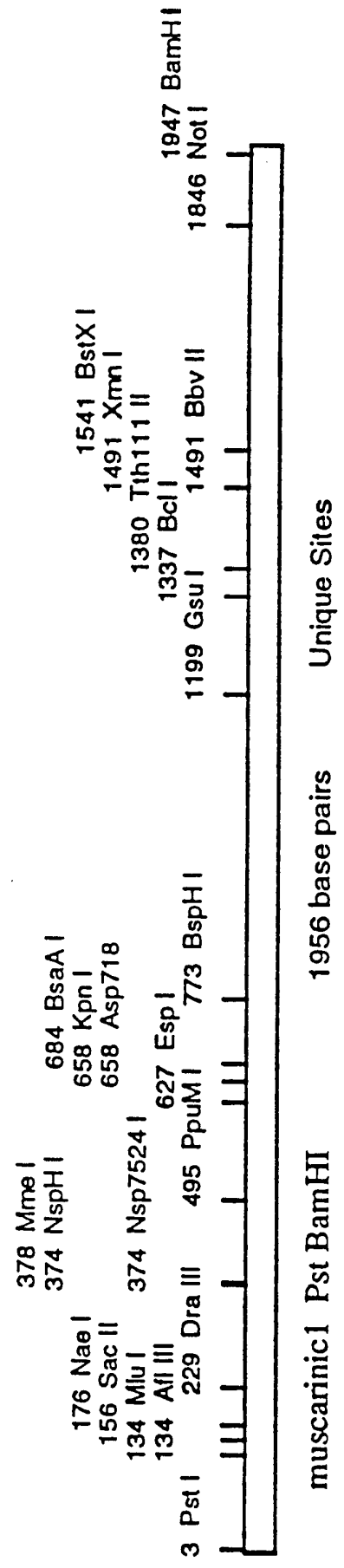


FIG. 5

ATCTGCAGGA TGGGTGCAAC CGTGAAGTCC GTCACGGGCTG CGTCACGACA GGAGCCGACC 60  
 AGCGACACCC AGAAGGTGCG AACGGTTGAG TGCCGCAACG ATCACGAGTT TTTCGGTGCGC 120  
 TTCGAGTGGT AACACGGCGTG CACGCATCGA CTTACCCGCG GGIGTTTCGA CGCCAGCCCGG 180  
 CCGTTGAACC AGCAGGCAGC GGGCATTCA CATCCGCTGT GGGCCACACA CTCGGTGCGG 240  
 TCGGCTATTT TGGTATGGTT TGGAATCCGC GTGTCGGCTC CGTGCTGAC GGTTCATCGG 300  
 TCCTAAATTC GTCACGAGCG TACCATACTG ATTGGTCTGT AGAGTTACAC ACATATCCTC 360  
 GTTAGGTAAT GTTGC ATG TTG GAG TTA TTG CCA ACA GCA GTG GAG GGG GTA 411  
 Met Leu Glu Leu Leu Pro Thr Ala Val Glu Gly Val  
 1 5 10  
 TCG CAG GCC CAG ATC CAG GCG CTG ATG AAC ACT TCA GCC CCA CCT GCT 459  
 Ser Gln Ala Gln Ile Gln Ala Leu Met Asn Thr Ser Ala Pro Pro Ala  
 15 20 25  
 GTC AGC CCC AAC ATC ACC GTC CTG GCA CCA GGA AAG GGT CCC TGG CAA 507  
 Val Ser Pro Asn Ile Thr Val Leu Ala Pro Gly Lys Gly Pro Trp Gln  
 30 35 40

FIG. 6(A)

GTC GCC TTC ATT GGG ATC ACC ACG GGC CTC TCG CTA GCC ACA GTG	555
Val Ala Phe Ile Gly Ile Thr Thr Gly Leu Ser Leu Ala Thr Val	60
45	55
ACA GGC AAC CTG CTG GTA CTC ATC TCT TTC AAG GTC AAC ACG GAG CTC	603
Thr Gly Asn Leu Leu Val Leu Ile Ser Phe Lys Val Asn Thr Glu Leu	75
65	70
AAG ACA GTC AAT AAC TAC TTC CTG CTG AGC CTG GCC TGT GCT GAC CTC	651
Lys Thr Val Asn Asn Tyr Phe Leu Leu Ser Leu Ala Cys Ala Asp Leu	90
80	85
ATC ATC GGT ACC TTC TCC ATG AAC CTC TAT ACC ACG TAC CTG CTC ATG	699
Ile Ile Gly Thr Phe Ser Met Asn Leu Tyr Thr Tyr Leu Leu Met	105
95	100
GGC CAC TGG GCT CTG GGC ACG CTG GCT TGT GAC CTC TGG CTG GCC CTG	747
Gly His Trp Ala Leu Gly Thr Leu Ala Cys Asp Leu Trp Leu Ala Leu	120
110	115
GAC TAT GTG GCC AGC AAT GCC TCC GTC ATG AAT CTG CTC ATC AGC	795
Asp Tyr Val Ala Ser Asn Ala Ser Val Met Asn Leu Leu Ile Ser	140
125	130
	135

FIG. 6(B)

TTT GAC CGC TAC TTC TCC GTG ACT CGG CCC CTG AGC TAC CGT GCC AAG 843  
 Phe Asp Arg Tyr Phe Ser Val Thr Arg Pro Leu Ser Tyr Arg Ala Lys  
 145 150 155  
 CGC ACA CCC CGC GCA GCT CTG ATG ATC GCC CTG GCC TGG CTG GTT 891  
 Arg Thr Pro Arg Arg Ala Ala Leu Met Ile Gly Leu Ala Trp Leu Val  
 160 165 170  
 TCC TTT GTG CTC TGG GCC CCA GCC ATC CTC TTC TGG CAA TAC CTG GTA 939  
 Ser Phe Val Leu Trp Ala Pro Ala Ile Leu Phe Trp Gln Tyr Leu Val  
 175 180 185  
 GGG GAG CGG ACG ATG CTA GCT GGG CAG TGC TAC ATC CAG TTC CTC TCC 987  
 Gly Glu Arg Thr Met Leu Ala Gly Gln Cys Tyr Ile Gln Phe Leu Ser  
 190 195 200  
 CAG CCC ATC ATC ACC TTT GGC ACA GCC ATG GCT GCC TTC TAC CTC CCT 1035  
 Gln Pro Ile Ile Thr Phe Gly Thr Ala Met Ala Ala Phe Tyr Leu Pro  
 205 210 215 220  
 GTC ACA GTC ATG TGC ACG CTC TAC TGG CGC ATC TAC CGG GAG ACA GAG 1083  
 Val Thr Val Met Cys Thr Leu Tyr Trp Arg Ile Tyr Arg Glu Thr Glu  
 225 230 235

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FIG. 6(C)

AAC CGA GCA CGG GAG CTG GCA GCC CTT CAG GGC TCC GAG ACC CCA GGC	1131
Asn Arg Ala Arg Glu Ala Ala Leu Gln Gly Ser Glu Thr Pro Gly	240 245 250
AAA GGG GGT GGC AGC AGC AGC TCA GAG AGG TCT CAG CCA GGG GCA	1179
Lys Gly Gly Ser Ser Ser Ser Glu Arg Ser Gln Pro Gly Ala	255 260 265
GAG GGC TCA CCA GAG ACT CCT CCA GGC CGC TGC TGT CGC TGC TGC CGG	1227
Glu Gly Ser Pro Glu Thr Pro Pro Gly Arg Cys Cys Arg Cys Arg	270 275 280
GCC CCA AGG CTG CTG CAA GCC TAC AGC TGG AAG GAA GAG GAA GAG	1275
Ala Pro Arg Leu Leu Gln Ala Tyr Ser Trp Lys Glu Glu Glu Glu	285 290 295 300
GAC GAA GGC TCC ATG GAG TCC CTC ACA TCC TCA GAG GGA GAG GAG CCT	1323
Asp Glu Gly Ser Met Glu Ser Leu Thr Ser Ser Ser Glu Gly Glu Pro	305 310 315
GGC TCC GAA GTG GTG ATC AAG ATG CCA ATG GTG GAC CCC GAG GCA CAG	1371
Gly Ser Glu Val Val Ile Lys Met Pro Met Val Asp Pro Glu Ala Gln	320 325 330

FIG. 6(D)



GCC CCC ACC AAG CAG CAG CCC CCA CGG AGC TCC CCA AAT ACA GTC AAG AGG 1419  
 Ala Pro Thr Lys Gln Pro Pro Arg Ser Ser Thr Val Lys Arg  
 335 340 345  
  
 CCG ACT AAG AAA GGG CGT GAT CGA GCT GGC AAG GGC CAG AAG CCC CGT 1467  
 Pro Thr Lys Lys Gly Arg Asp Arg Ala Gly Lys Gly Gln Lys Pro Arg  
 350 355 360  
  
 GGA AAG GAG CAG CTG GCC AAG CCG AAG ACC TTC TCG CTG GTC AAG GAG 1515  
 Gly Lys Glu Gln Leu Ala Lys Arg Lys Thr Phe Ser Leu Val Lys Glu  
 365 370 375 380  
  
 AAG AAG GCG GCT CCG ACC CTG AGT GCC ATC CTC CTG GCC TTC ATC CTC 1563  
 Lys Lys Ala Ala Arg Thr Leu Ser Ala Ile Leu Leu Ala Phe Ile Leu  
 385 390 395  
  
 ACC TGG ACA CCG TAC AAC ATC ATG GTG CTG GTG TCC ACC TTC TGC AAG 1611  
 Thr Trp Thr Pro Tyr Asn Ile Met Val Leu Val Ser Thr Phe Cys Lys  
 400 405 410  
  
 GAC TGT GTT CCC GAG ACC CTG TGG GAG CTG GGC TAC TGG CTG TGC TAC 1659  
 Asp Cys Val Pro Glu Thr Leu Trp Glu Leu Gly Tyr Trp Leu Cys Tyr  
 415 420 425

FIG. 6(E)

GTC AAC AGC ACC ATC AAC CCC ATG TGC TAC GCA CTC TGC AAC AAA GCC 1707  
 Val Asn Ser Thr Ile Asn Pro Met Cys Tyr Ala Leu Cys Asn Lys Ala  
 430 440  
 TTC CGG GAC ACC TTT CGC CTG CTG CTT TGC CGC TGG GAC AAG AGA CGC 1755  
 Phe Arg Asp Thr Phe Arg Leu Leu Cys Arg Trp Asp Lys Arg Arg  
 445 450 460  
 TGG CGC AAG ATC CCC AAG CGC CCT GGC TCC GTG CAC CGC ACT CCC TCC 1803  
 Trp Arg Lys Ile Pro Lys Arg Pro Gly Ser Val His Arg Thr Pro Ser  
 465 470 475  
 CGC CAA TGC TGATAGTCCC CTCCTCTGCA TCCCTCCACC CCAGCGGCCG 1852  
 Arg Gln Cys  
 CGACCAGCGA TTGATCGCAC ACGCAGGACA GCCCCACAAC CGGCGCGGCT GTGTTCAACG 1912  
 ACACACGATG AGTCCCCCCAC TCGGTCTTGT ACTCGGATCC TTTT 1956

15 / 50

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FIG. 6(F)

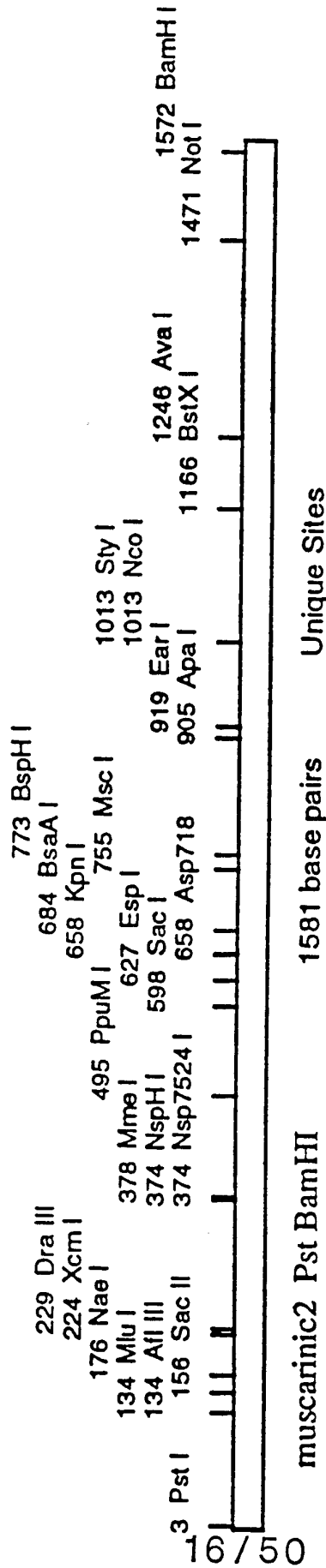


FIG. 7

ATCTGCAGGA TGGGTGCAAC CGTGAAGTCC GTCACGGCTG CGTCACGACA GGAGCCGACC 60  
 AGCGACACCC AGAAGGTGCG AACGGTTGAG TGCCGCAACG ATCACGAGTT TTTCGTGCGC 120  
 TTCGAGTGGT AACACGCGTG CACGCATCGA CTTACCCGG GGTGTTTCGA CGCCAGCCGG 180  
 CCGTTGAACC AGCAGGCAGC GGGCATTCA CATCCGCTGT GCCCCACACA CTCGGTGGGG 240  
 TCGGCTATTT TGGTATGGTT TGGAATCCGC GTGTCGGCTC CGTGTCTGAC GGTTCATCGG 300  
 TCTAAATTCC GTCACGAGCG TACCATACTG ATTGGGTCGT AGAGTTACAC ACATATCCTC 360  
 GTTAGGTACT GTTGC ATG TTG GAG TTA TTG CCA ACA GCA GTG GAG GGG GTA 411  
                   Met Leu Glu Leu Leu Pro Thr Ala Val Glu Gly Val  
                   1                  5                  10  
 TCG CAG GCC CAG ATC CAG GCG CTG ATG AAC ACT TCA GCC CCA CCT GCT 459  
 Ser Gln Ala Gln Ile Gln Ala Leu Met Asn Thr Ser Ala Pro Pro Ala  
                   15                  20                  25  
 GTC AGC CCC AAC ATC ACC GTC CTG GCA CCA GGA AAG GGT CCC TGG CAA 507  
 Val Ser Pro Asn Ile Thr Val Leu Ala Pro Gly Lys Gly Pro Trp Gln  
                   30                  35                  40

FIG. 8(A)

GTG GCC TTC ATT GGG ATC ACC ACG GGC CTC TCG CTA GCC ACA GTG 555  
 Val Ala Phe Ile Gly Ile Thr Thr Gly Leu Ser Leu Ala Thr Val  
 45 50 55 60  
 ACA GGC AAC CTG CTG GTA CTC ATC TCT TTC AAG GTC AAC ACG GAG CTC 603  
 Thr Gly Asn Leu Leu Val Leu Ile Ser Phe Lys Val Asn Thr Glu Leu  
 65 70 75  
 AAG ACA GTC AAT AAC TAC TTC CTG CTG AGC CTG GCC TGT GCT GAC CTC 651  
 Lys Thr Val Asn Asn Tyr Phe Leu Leu Ser Leu Ala Cys Ala Asp Leu  
 80 85 90  
 ATC ATC GGT ACC TTC TCC ATG AAC CTC TAT ACC ACG TAC CTG CTC ATG 699  
 Ile Ile Gly Thr Phe Ser Met Asn Leu Tyr Thr Tyr Leu Leu Met  
 95 100 105  
 GGC CAC TGG GCT CTG GGC ACG CTG GCT TGT GAC CTC TGG CTG GCC CTG 747  
 Gly His Trp Ala Leu Gly Thr Leu Ala Cys Asp Leu Trp Leu Ala Leu  
 110 115 120  
 GAC TAT GTG GCC AGC AAT GCC TCC GTC ATG AAT CTG CTG CTC ATC AGC 795  
 Asp Tyr Val Ala Ser Asn Ala Ser Val Met Asn Leu Leu Ile Ser  
 125 130 135 140

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FIG. 8(B)

TTT GAC CGC TAC TTC TCC GTG ACT CGG CCC CTG AGC TAC CGT GCC AAG 843  
 Phe Asp Arg Tyr Phe Ser Val Thr Arg Pro Leu Ser Tyr Arg Ala Lys  
 145 155  
 CGC ACA CCC CGC CGC GCA GCT CTG ATG ATC GGC CTG GCC TGG CTG GTT 891  
 Arg Thr Pro Arg Arg Ala Ala Leu Met Ile Gly Leu Ala Trp Leu Val  
 160 170  
 TCC TTT GTG CTC TGG GCC CCA GCC ATC CTC TTC TGG CAA TAC CTG GTA 939  
 Ser Phe Val Leu Trp Ala Pro Ala Ile Leu Phe Trp Gln Tyr Leu Val  
 175 180 185  
 GGG GAG CGG ACG ATG CTA GCT GGG CAG TGC TAC ATC CAG TTC CTC TCC 987  
 Gly Glu Arg Thr Met Leu Ala Gly Gln Cys Tyr Ile Gln Phe Leu Ser  
 190 195 200  
 CAG CCC ATC ATC ACC TTT GGC ACA GCC ATG GCT GCC TTC TAC CTC CCT 1035  
 Gln Pro Ile Ile Thr Phe Gly Thr Ala Met Ala Ala Phe Tyr Leu Pro  
 205 210 215 220  
 GTC ACA GTC ATG TGC ACG CTC TAC TGG CGC ATC TAC CGG GAG ACA GAG 1083  
 Val Thr Val Met Cys Thr Leu Tyr Trp Arg Ile Tyr Arg Glu Thr Glu  
 225 230 235

FIG. 8(C)

AAC CGA GCA CGG GAG CTG GCA GCC CTT CAG GGC TCC GAG ACG CCA GGC	1131
Asn Arg Ala Arg Glu Leu Ala Ala Leu Gln Gly Ser Glu Thr Pro Gly	240 245 250
AAA AAG GAG AAG AAG GCG GCT CGG ACC CTG AGT GCC ATC CTC CTG GCC	1179
Lys Lys Glu Lys Lys Ala Ala Arg Thr Leu Ser Ala Ile Leu Leu Ala	255 260 265
TTC ATC CTC ACC TGG ACA CCG TAC AAC ATC ATG GTG CTG GTG TCC ACC	1227
Phe Ile Leu Thr Trp Thr Pro Tyr Asn Ile Met Val Leu Val Ser Thr	270 275 280
TTC TGC AAG GAC TGT GTT CCC GAG ACC CTG TGG GAG CTG GGC TAC TGG	1275
Phe Cys Lys Asp Cys Val Pro Glu Thr Leu Trp Glu Leu Gly Tyr Trp	285 290 295 300
CTG TGC TAC GTC AAC AGC ACC ATC AAC CCC ATG TGC TAC GCA CTC TGC	1323
Leu Cys Tyr Val Asn Ser Thr Ile Asn Pro Met Cys Tyr Ala Leu Cys	305 310 315
AAC AAA GCC TTC CGG GAC ACC TTT CGC CTG CTG CTT TGC CGC TGG GAC	1371
Asn Lys Ala Phe Arg Asp Thr Phe Arg Leu Leu Cys Arg Trp Asp	320 325 330

FIG. 8(D)

AAG AGA CGC TGG CGC AAG ATC CCC AAG CGC CCT GGC TCC GTG CAC CGC	1419
Lys Arg Arg Trp Arg Lys Ile Pro Lys Arg Pro Gly Ser Val His Arg	
335	340
345	
ACT CCC TCC CGC CAA TGC TGATAGTCCC CTCCTCTGCA TCCCTCCACC	1467
Thr Pro Ser Arg Gln Cys	
350	
CCAGCGGCCG CGACCAGCGA TTGATCGCAC ACCCAGGACA GCCCACAAC CGGCGGGCT	1527
GTGTTCAACG ACACACGATG AGTCCCCCAC TCGGTCTTGT ACTCGGATCC TTTT	1581

FIG. 8(E)



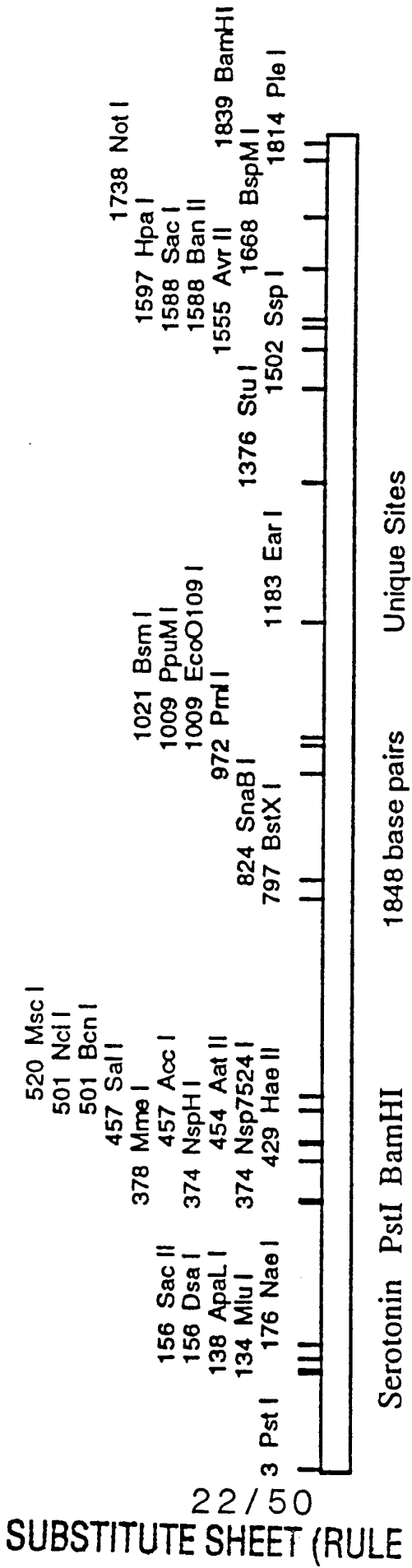


FIG. 9

ATCTGCAGGA TGGGTGCAAC CGTGAAGTCC GTCACGGCTG CGTCACGACA GGAGCCGACC 60  
 AGCGACACCC AGAAGGTGCG AACGGTTGAG TGCCGCAACG ATCACGAGTT TTTCGGTGCGC 120  
 TTCGAGTGGT AACACCGGTG CACGCATCGA CTTACCCGG GGTGTTTCGA CGCCAGCCCGG 180  
 CCGTTGAACC AGCAGGCAGC GGGCATTCA CATCCGCTGT GCCCCACACA CTCGGTGGGG 240  
 TCGGCTATTT TGGTATGGTT TGGAAATCCG GGTGCGGCTC CGTGCTGAC GGTTCATCGG 300  
 TCTAAATTC GTCACGAGCG TACCATACTG ATTGGGTCGT AGAGTTACAC ACATAATCCTC 360  
 GTTAGGTA CTGTG ATG TTG GAG TTA TTG CCA ACA GCA GTG GAG GGG GTA 411  
 Met Leu Glu Leu Leu Pro Thr Ala Val Glu Gly Val  
 1 5 10  
 TCG CAG GCC CAG ATC CAG GCG CTG GAC TAC AAG GAC GAT GAT GAC GTC 459  
 Ser Gln Ala Gln Ile Gln Ala Leu Asp Tyr Lys Asp Asp Asp Val  
 15 20 25  
 GAC ACT TTT AAT TCC TCC GAT GGT GGA CGC TTG TTT CAA TTC CCG GAC 507  
 Asp Thr Phe Asn Ser Ser Asp Gly Gly Arg Leu Phe Gln Phe Pro Asp  
 30 35 40

FIG. 10(A)

555 GGG GTA CAA AAC TGG CCA GCA CTT TCA ATC GTC GTG ATT ATA ATC ATG  
 Gly Val Gln Asn Trp Pro Ala Leu Ser Ile Val Val Ile Ile Ile Met  
 45 50 55 60  
 603 ACA ATA GGG GGC AAC ATT CTT GTT ATC ATG GCA GTA AGC ATG GAG AAG  
 Thr Ile Gly Gly Asn Ile Leu Val Ile Met Ala Val Ser Met Glu Lys  
 65 70 75  
 651 AAA CTG CAC AAT GCA ACC AAT TAC TTC TTA ATG TCC CTA GCC ATT GCT  
 Lys Leu His Asn Ala Thr Asn Tyr Phe Leu Met Ser Leu Ala Ile Ala  
 80 85 90  
 699 GAT ATG CTG GTG GGA CTA CTT GTC ATG CCC CTG TCC CTG CTT GCT ATT  
 Asp Met Leu Val Gly Leu Leu Val Met Pro Leu Ser Leu Ala Ile  
 95 100 105  
 747 CTT TAT GAT TAT GTC TGG CCT TTA CCT AGA TAT TTG TGC CCC GTC TGG  
 Leu Tyr Asp Tyr Val Trp Pro Leu Pro Arg Tyr Leu Cys Pro Val Trp  
 110 115 120  
 795 ATT TCA CTA GAT GTG CTA TTT TCA ACT GCG TCC ATC ATG CAC CTC TGC  
 Ile Ser Leu Asp Val Leu Phe Ser Thr Ala Ser Ile Met His Leu Cys  
 125 130 135 140

FIG. 10(B)

GCC ATA TCG CTG GAC CGG TAT GTA GCA ATA CGT AAT CCT ATT GAG CAT	843
Ala Ile Ser Leu Asp Arg Tyr Val Ala Ile Arg Asn Pro Ile Glu His	155
145	150
AGC CGG TTC AAT TCG CGG ACT AAG GCC ATC ATG AAG ATT GCC ATC GTT	891
Ser Arg Phe Asn Ser Arg Thr Lys Ala Ile Met Lys Ile Ala Ile Val	170
160	165
TGG GCA ATA TCA ATA GGA GTT TCA GTT CCT ATC CCT GTG ATT GGA CTG	939
Trp Ala Ile Ser Ile Gly Val Ser Val Pro Ile Pro Val Ile Gly Leu	185
175	180
AGG GAC GAA AGC AAA GTG TTC GTG AAT AAC ACC ACG TGC GTG CTC AAT	987
Arg Asp Glu Ser Lys Val Phe Val Asn Asn Thr Thr Cys Val Leu Asn	200
190	195
GAC CCC AAC TTC GTT CTC ATC GGG TCC TTC GTG GCA TTC TTC ATC CCG	1035
Asp Pro Asn Phe Val Leu Ile Gly Ser Phe Val Ala Phe Ile Pro	220
205	210
TTG ACG ATT ATG GTG ATC ACC TAC TTC TTA ACG ATC TAC GTC CTG CGC	1083
Leu Thr Ile Met Val Ile Thr Tyr Phe Leu Thr Ile Tyr Val Leu Arg	235
225	230

FIG. 10(C)

CGT CAA ACT CTG ATG TTA CTT CGA GGT CAC ACC GAG GAG GAA CTG GCT	1131
Arg Gln Thr Leu Met Leu Arg Gly His Thr Glu Glu Leu Ala	
240	245
AAT ATG AGC CTG AAC TTT CTG AAC TGC TGC AAG AAG AAT GGT GGT	1179
Asn Met Ser Leu Asn Phe Leu Asn Cys Cys Lys Lys Asn Gly Gly	
255	260
GAG GAA GAG AAC GCT CCG AAC CCT AAT CCA GAT CAG AAA CCA CGT CGA	1227
Glu Glu Glu Asn Ala Pro Asn Pro Asp Gln Lys Pro Arg Arg	
270	275
AAG AAG AAA GAA AAG CGT CCC AGA GGC ACC ATG CAA GCT ATC AAC AAC	1275
Lys Lys Lys Glu Lys Arg Pro Arg Gly Thr Met Gln Ala Ile Asn Asn	
285	290
GAA AAG AAA GCT TCC AAA GTC CTT GGC ATT GTA TTC TTT GTG TTT CTG	1323
Glu Lys Lys Ala Ser Lys Val Leu Gly Ile Val Phe Phe Val Phe Leu	
305	310
ATC ATG TGG TGC CCG TTT TTC ATC ACC AAT ATC CTG TCG GTT CTT TGT	1371
Ile Met Trp Cys Pro Phe Phe Ile Thr Asn Ile Leu Ser Val Leu Cys	
320	325
	330

FIG. 10(D)

GGG AAG GCC TGT AAC CAA AAG CTA ATG GAG AAG CTT CTC AAT GTG TTT 1419  
 Gly Ala Cys Asn Gln Lys Leu Met Glu Lys Leu Leu Asn Val Phe  
 335 340 345  
 GTG TGG ATT GGC TAT GTG TGT TCA GGC ATC AAT CCT CTG GTG TAC ACT 1467  
 Val Trp Ile Gly Tyr Val Cys Ser Gly Ile Asn Pro Leu Val Tyr Thr  
 350 355 360  
 CTC TTT AAT AAA ATT TAC CGA AGG GCT TTC TCT AAA TAT TTG CGC TGC 1515  
 Leu Phe Asn Lys Ile Tyr Arg Arg Ala Phe Ser Lys Tyr Leu Arg Cys  
 365 370 375 380  
 GAT TAT AAG CCA GAC AAA AAG CCT CCT GTT CGA CAG ATT CCT AGG GTT 1563  
 Asp Tyr Lys Pro Asp Lys Lys Lys Pro Pro Val Arg Gln Ile Pro Arg Val  
 385 390 395  
 GCT GCC ACT GCT TTG TCT GGG AGG GAG CTC AAT GTT AAC ATT TAT CGG 1611  
 Ala Ala Thr Ala Leu Ser Gly Arg Glu Leu Asn Val Asn Ile Tyr Arg  
 400 405 410  
 CAT ACC AAT GAA CGT GTG GCT AGG AAA GCT AAT GAC CCT GAG CCT GGC 1659  
 His Thr Asn Glu Arg Val Ala Arg Lys Ala Asn Asp Pro Glu Pro Gly  
 415 420 425

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FIG. 10(E)

ATA GAG ATG CAG GTG GAG AAC TTA GAG CTG CCA GTC AAC CCC TCT AAT 1707  
 Ile Glu Met Gln Val Glu Asn Leu Glu Leu Pro Val Asn Pro Ser Asn  
 430 435 440  
 GTG GTC AGC GAG AGG ATT AGT AGT GTG TGAGCGGCGG CGACCAGCGA 1754  
 Val Val Ser Glu Arg Ile Ser Ser Val  
 445 450  
 TTGATCGGCAC ACGCAGGACA GCCCCACAAC CGGCGCGGCT GTGTTCAACG ACACACGATG 1814  
 AGTCCCCCCAC TCGGTCCTTGT ACTCGGATCC TTTT 1848

FIG. 10(F)

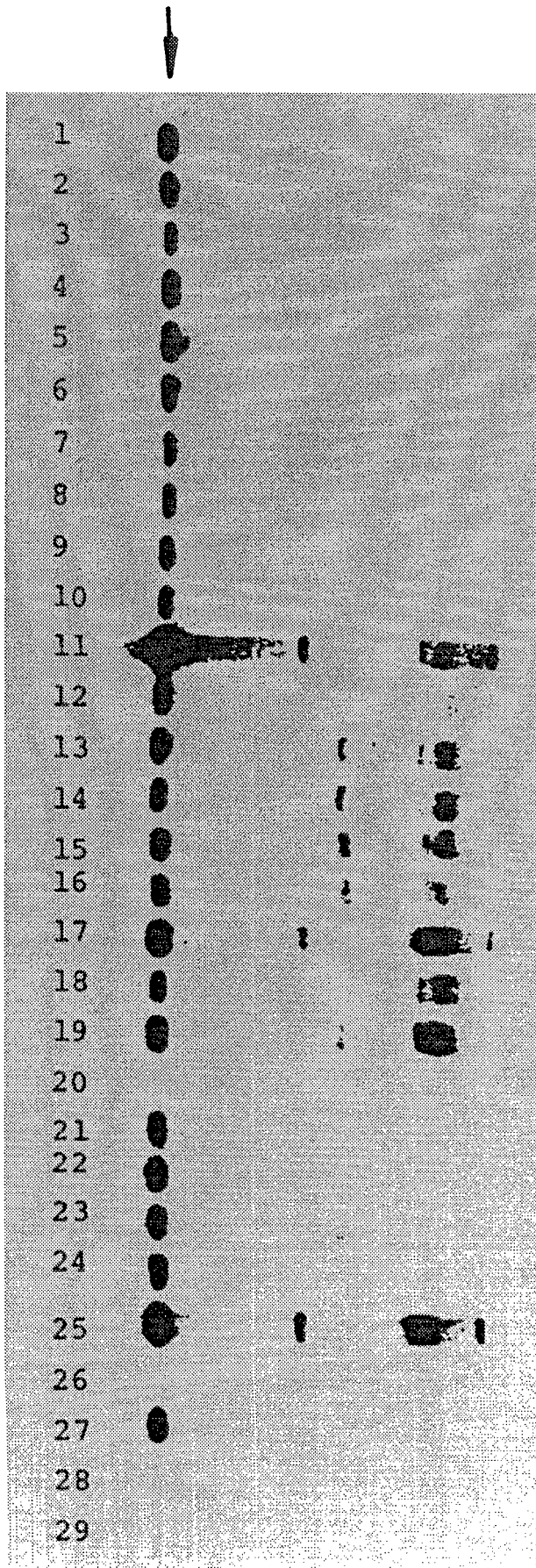


FIG. 11



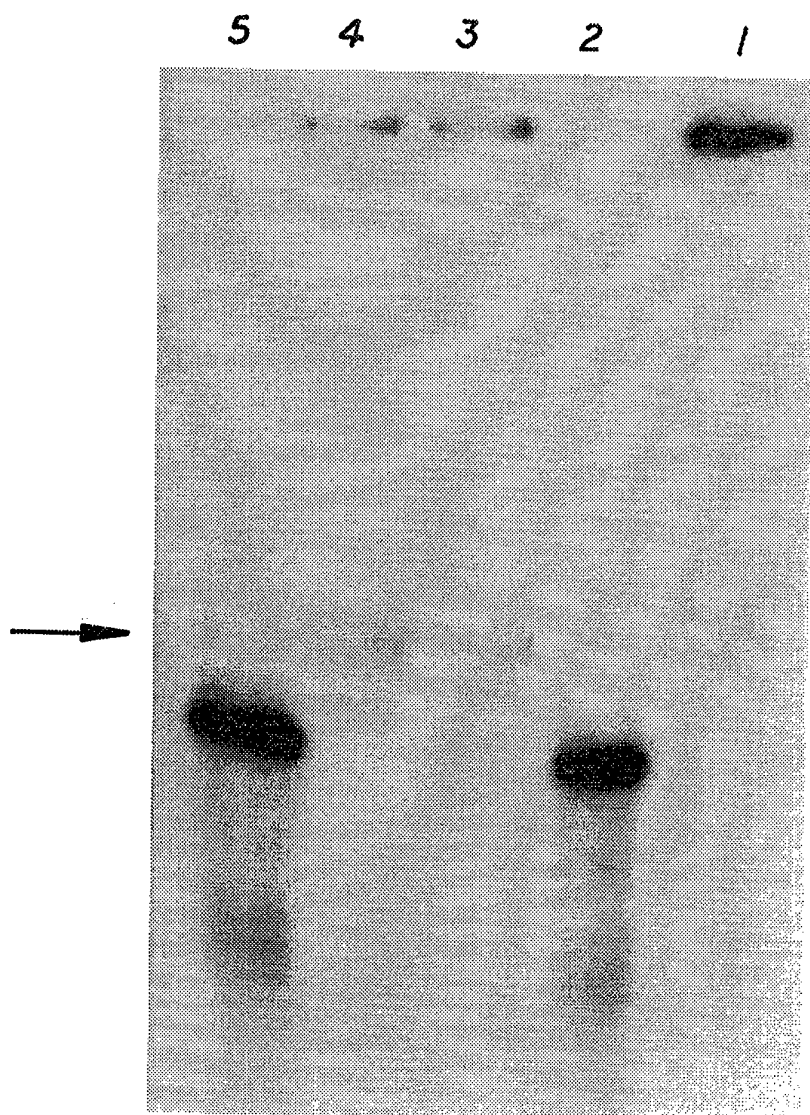


FIG. 12

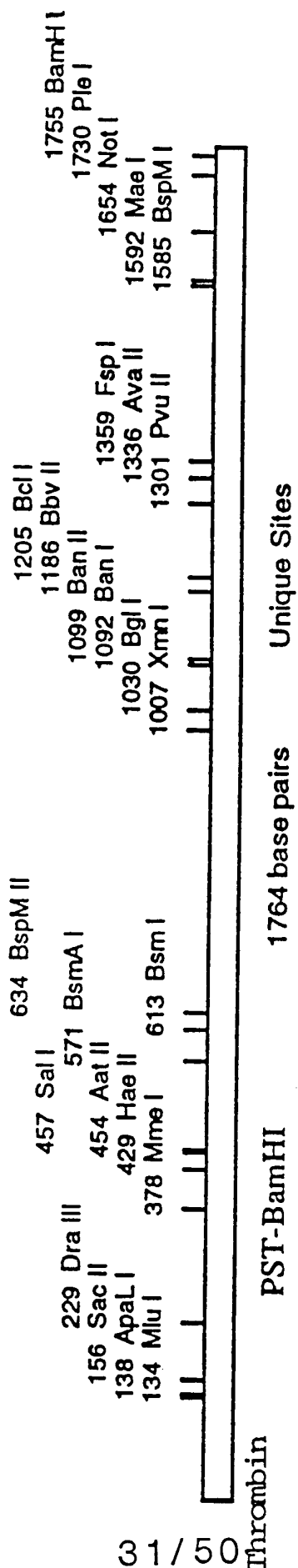


FIG. 13

ATCTGCAGGA TGGGTGCAAC CGTGAAGTCC GTCACGGCTG CGTCACGACA GGAGCCGACC 60  
 AGGACACCC AGAAGGTGG AACGGTTGAG TGCCGCAACG ATCACGAGTT TTTCGTGCGC 120  
 TTCGAGTGGT AACACGGGTG CACGCATCGA CTTACCCGG GGTGTTTCCA CGCCAGCCCG 180  
 CCGTTGAACC AGCAGGCAGC GGGCATTCA CATCCGCTGT GGCCACACA CTCGGTGGGG 240  
 TCGGCTATTT TGGTATGGTT TGGAATCCGC GTGTCGGCTC CGTGTCTGAC GGTTCATCCG 300  
 TCTAAATCC GTCACGAGCG TACCATACTG ATTGGTCTGT AGAGTIACAC ACATATCCTC 360  
 GTTAGTACT GTTGC ATG TTG GAG TTA TTG CCA ACA GCA GTG GAG GGG GTA 411  
 Met Leu Glu Leu Leu Pro Thr Ala Val Glu Gly Val  
 1 5 10  
 TCG CAG GCC CAG ATC CAG GCG CTG GAC TAC AAG GAC GAT GAT GAC GTC 459  
 Ser Gln Ala Gln Ile Gln Ala Leu Asp Tyr Lys Asp Asp Asp Val  
 15 20 25  
 GAC GCC ACC TTA GAT CCC CGG TCA TTT CTT CTC AGG AAC CCC AAT GAT 507  
 Asp Ala Thr Leu Asp Pro Arg Ser Phe Leu Leu Arg Asn Pro Asn Asp  
 30 35 40

FIG. 14(A)

AAA TAT GAA CCA TTT TGG GAG GAT GAG AAA AAT GAA AGT GGG TTA	555
Lys Tyr Glu Pro Phe Trp Glu Asp Glu Glu Lys Asn Glu Ser Gly Leu	60
45	
50	
55	
60	
ACT GAA TAC AGA TTA GTC TCC ATC AAT AAA AGC AGT CCT CTT CAA AAA	603
Thr Glu Tyr Arg Leu Val Ser Ile Asn Lys Ser Ser Pro Leu Gln Lys	75
65	
70	
75	
CAA CTT CCT GCA TTC ATC TCA GAA GAT GCC TCC GGA TAT TTG ACC AGC	651
Gln Leu Pro Ala Phe Ile Ser Glu Asp Ala Ser Gly Tyr Leu Thr Ser	90
80	
85	
TCC TGG CTG ACA CTC TTT GTC CCA TCT GTG TAC ACC GGA GTG TTT GTA	699
Ser Trp Leu Thr Leu Phe Val Pro Ser Val Tyr Thr Gly Val Phe Val	105
95	
100	
105	
GTC AGC CTC CCA CTA AAC ATC ATG GCC ATC GTT GTG TTC ATC CTG AAA	747
Val Ser Leu Pro Leu Asn Ile Met Ala Ile Val Val Phe Ile Leu Lys	120
110	
115	
120	
ATG AAG GTC AAG AAG CCG GCG GTG TAC ATG CTG CAC CTG GCC ACG	795
Met Lys Val Lys Lys Pro Ala Val Tyr Met Leu His Leu Ala Thr	140
125	
130	
135	
140	

FIG. 14(B)

GCA GAT GTG CTG TTT GTG TCT GTG CTC CCC TTT AAG ATC AGC TAT TAC 843  
 Ala Asp Val Leu Phe Val Ser Val Leu Pro Phe Lys Ile Ser Tyr Tyr  
 145 150 155  
 TTT TCC GGC AGT GAT TGG CAG TTT GGG TCT GAA TTG TGT CGC TTC GTC 891  
 Phe Ser Gly Ser Asp Trp Gln Phe Gly Ser Glu Leu Cys Arg Phe Val  
 160 165 170  
 ACT GCA GCA TTT TAC TGT AAC ATG TAC GCC TCT ATC TTG CTC ATG ACA 939  
 Thr Ala Ala Phe Tyr Cys Asn Met Tyr Ala Ser Ile Leu Leu Met Thr  
 175 180 185  
 GTC ATA AGC ATT GAC CCG TTT CTG GCT GTG TAT CCC ATG CAG TCC 987  
 Val Ile Ser Ile Asp Arg Phe Leu Ala Val Val Tyr Pro Met Gln Ser  
 190 195 200  
 CTC TCC TGG CGT ACT CTG GGA AGG GCT TCC TTC ACT TGT CTG GCC ATC 1035  
 Leu Ser Trp Arg Thr Leu Gly Arg Ala Ser Phe Thr Cys Leu Ala Ile  
 205 210 215 220  
 TGG GCT TTG GCC ATC GCA GGG GTA GTG CCT CTC GTC CTC AAG GAG CAA 1083  
 Trp Ala Leu Ala Ile Ala Gly Val Val Pro Leu Val Leu Lys Glu Gln  
 225 230 235

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FIG. 14(C)

ACC ATC CAG GTG CCC GGG CTC AAC ATC ACT ACC TGT CAT GAT GTG CTC 1131  
 Thr Ile Gln Val Pro Gly Leu Asn Ile Thr Thr Cys His Asp Val Leu  
 240 245 250

AAT GAA ACC CTG CTC GAA GGC TAC TAT GCC TAC TAC TTC TCA GCC TTC 1179  
 Asn Glu Thr Leu Leu Glu Gly Tyr Tyr Ala Tyr Tyr Phe Ser Ala Phe  
 255 260 265

TCT GCT GTC TTC TTT GTG CCG CTG ATC ATT TCC ACG GTC TGT TAT 1227  
 Ser Ala Val Phe Phe Val Pro Leu Ile Ile Ser Thr Val Cys Tyr  
 270 275 280

GTG TCT ATC ATT CGA TGT CTT AGC TCT TCC GCA GTT GCC AAC CGC AGC 1275  
 Val Ser Ile Ile Arg Cys Leu Ser Ser Ala Val Ala Asn Arg Ser  
 285 290 295 300

AAG AAG TCC CGG GCT TTG TTC CTG TCA GCT GCT GTT TTC TGC ATC TTC 1323  
 Lys Lys Ser Arg Ala Leu Phe Leu Ser Ala Ala Val Phe Cys Ile Phe  
 305 310 315

ATC ATT TGC TTC GGA CCC ACA AAC GTC CTC CTG ATT GCG CAT TAC TCA 1371  
 Ile Ile Cys Phe Gly Pro Thr Asn Val Leu Leu Ile Ala His Tyr Ser  
 320 325 330

35 / 50  
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FIG. 14(D)

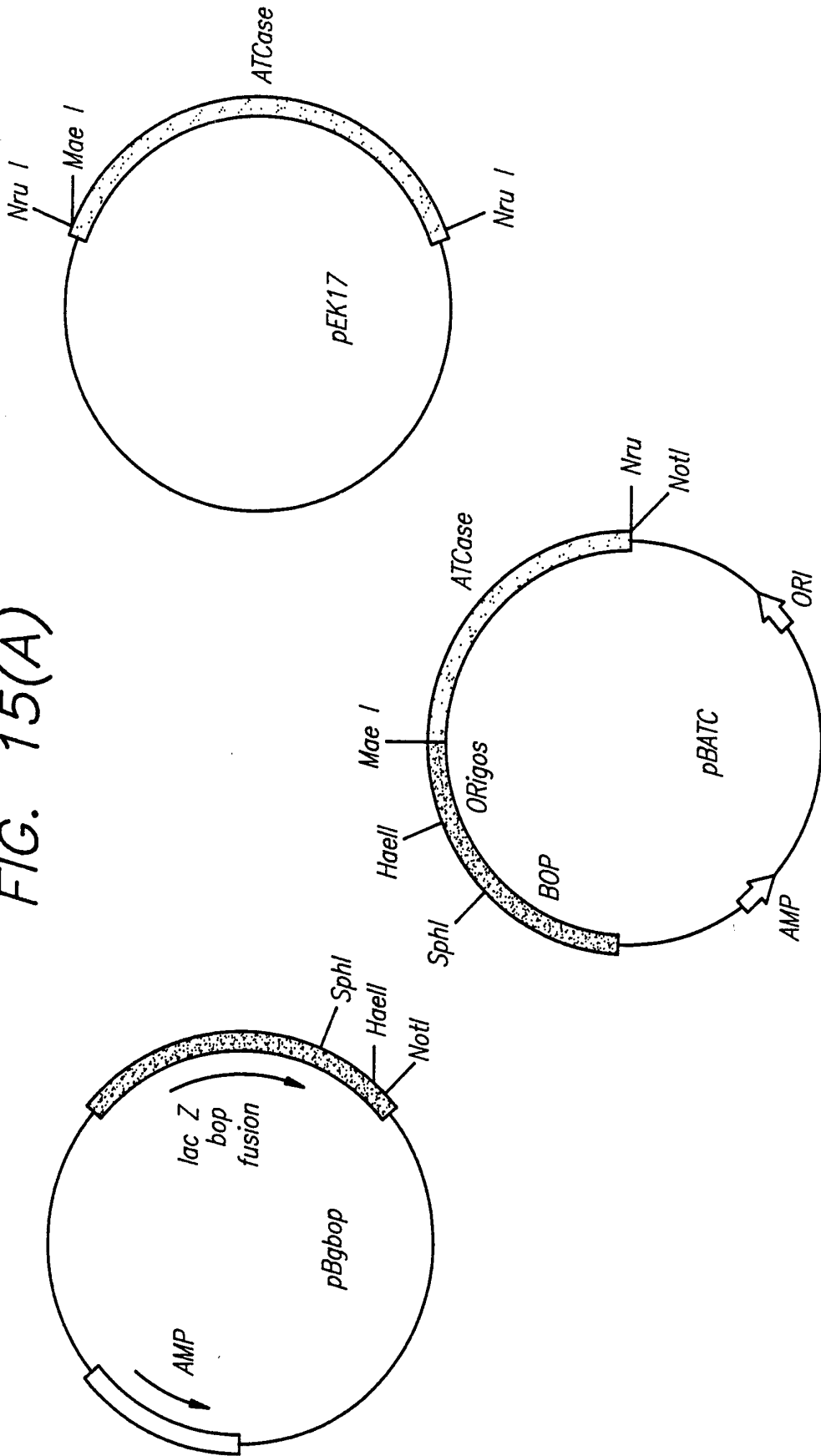
TTC CTT TCT CAC ACT TCC ACC ACA GAG GCT GCC TAC TTT GCC TAC CTC	1419
Phe Leu Ser His Thr Ser Thr Ser Thr Glu Ala Tyr Phe Ala Tyr Leu	
335 340 345	
CTC TGT GTC TGT GTC AGC AGC ATA AGC TCG TGC ATC GAC CCC CTA ATT	1467
Leu Cys Val Cys Val Ser Ser Ile Ser Ser Cys Ile Asp Pro Leu Ile	
350 355 360	
TAC TAT TAC GCT TCC TCT GAG TGC CAG AGG TAC GTC TAC AGT ATC TTA	1515
Tyr Tyr Tyr Ala Ser Ser Glu Cys Gln Arg Tyr Val Tyr Ser Ile Leu	
365 370 375 380	
TGC TGC AAA GAA AGT TCC GAT CCC AGC AGT TAT AAC AGC AGT GGG CAG	1563
Cys Cys Lys Glu Ser Ser Asp Pro Ser Ser Tyr Asn Ser Ser Gly Gln	
385 390 395	
TTG ATG GCA AGT AAA ATG GAT ACC TGC TCT AGT AAC CTG AAT AAC AGC	1611
Leu Met Ala Ser Lys Met Asp Thr Cys Ser Ser Asn Leu Asn Asn Ser	
400 405 410	

FIG. 14(E)





FIG. 15(A)



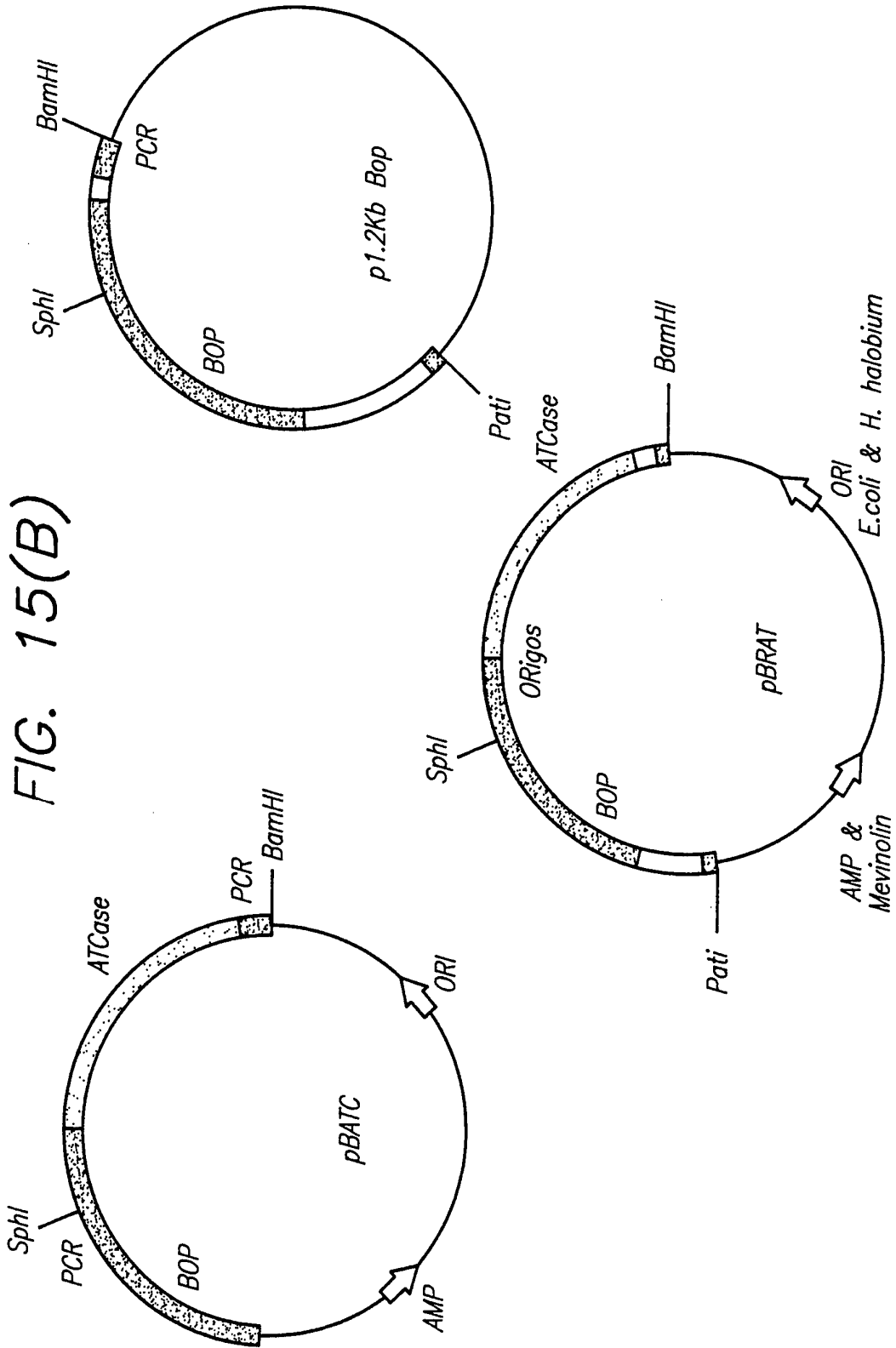


FIG. 15(B)

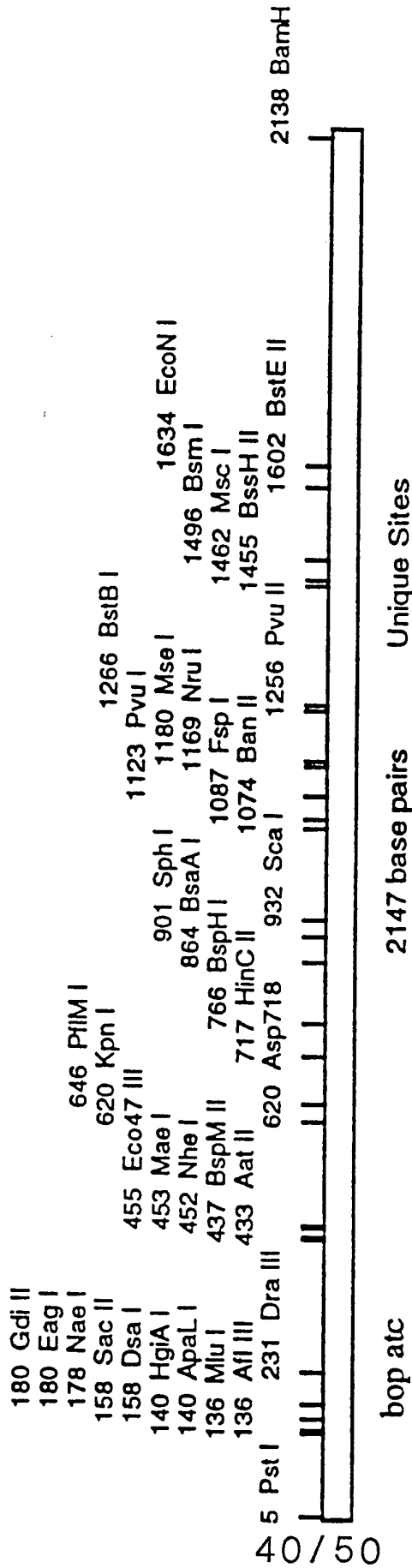


FIG. 16

TAATCTGCAG GATGGGTGCA ACCGTGAAGT CCGTCACGGC TGGTACCGA CAGGAGCCGA 60  
 CCAGCGACAC CCAGAAGGTG CGAACGGTTG AGTGCCGCAA CGATCACGAG TTTTTCGTGC 120  
 GCTTCGAGTG GTAACACGGG TGCACGCATC GACTTCACCG CGGGTGTTC GACGCCAGCC 180  
 GGCCGTTGAA CCAGCAGGCA GCGGGCATT CACAGCCGCT GTGGCCACACA CACTCGGTGG 240  
 GGTGCGCTAT TTTGGTATGG TTGGAAATC GCGTGTCCGC TCCGTGTCTG ACGTTCATC 300  
 GGTCTAAATT CCGTCACGAG CGTACCATAC TGATIGGGTC GTAGAGTTAC ACACATAATC 360  
 TCGTTAGGTA CTGTTGC ATG TTG GAG TTA TTG CCA ACA GCA GTG GAG GGG 410  
 Met Leu Glu Leu Leu Pro Thr Ala Val Glu Gly  
 1 5 10  
 GTA TCG CAG GCC CAG ATC ACC GGA CGT CCG GAG TGG ATC TGG CTA GCG 458  
 Val Ser Gln Ala Gln Ile Thr Gly Arg Pro Glu Trp Ile Trp Leu Ala  
 15 20 25  
 CTC GGT ACG GCG CTA ATG GGA CTC GGG ACG CTC TAT TTC CTC GTG AAA 506  
 Leu Gly Thr Ala Leu Met Gly Leu Gly Thr Leu Tyr Phe Leu Val Lys  
 30 35 40

FIG. 17(A)

GGG ATG GGC GTC TCG GAC CCA GAT GCA AAG AAA TTC TAC GCC ATC ACG 554  
 Gly Met Gly Val Ser Asp Pro Asp Ala Lys Lys Phe Tyr Ala Ile Thr  
 45 55  
 ACC CTC GTC CCA GCC ATC GCG TTC ACG ATG TAC CTC TCG ATG CTG CTG 602  
 Thr Leu Val Pro Ala Ile Ala Phe Thr Met Tyr Leu Ser Met Leu Leu  
 60 65 70 75  
 GGG TAT GGC CTC ACA ATG GTA CCG TTC GGT GGG GAG CAG AAC CCC ATC 650  
 Gly Tyr Gly Leu Thr Met Val Pro Phe Gly Gly Glu Gln Asn Pro Ile  
 80 85 90  
 TAC TGG GCG CCG TAC GCT GAC TGG CTG TTC ACC ACG CCG CTG TTG TTG 698  
 Tyr Trp Ala Arg Tyr Ala Asp Trp Leu Phe Thr Thr Pro Leu Leu Leu  
 95 100 105  
 TTA GAC CTC GCG TTG CTC GTT GAC GCG GAT CAG GGA ACG ATC CTT GCG 746  
 Leu Asp Leu Ala Leu Leu Val Asp Ala Asp Gln Gly Thr Ile Leu Ala  
 110 115 120  
 CTC GTC GGT GCC GAC GGC ATC ATG GGC ACC GGC CTG GTC GGC GCA 794  
 Leu Val Gly Ala Asp Gly Ile Met Ile Gly Thr Gly Leu Val Gly Ala  
 125 130 135

FIG. 17(B)

CTG	ACG	AAG	GTC	TAC	TCG	TAC	CGC	TTC	GTG	TGG	TGG	GCG	ATC	AGC	ACC	842
Leu	Thr	Lys	Val	Tyr	Ser	Tyr	Arg	Phe	Val	Trp	Trp	Ala	Ile	Ser	Thr	155
140					145			150								
GCA	GCG	ATG	CTG	TAC	ATC	CTG	TAC	GTG	CTG	TTC	TTC	GCG	TTC	ACC	TCG	890
Ala	Ala	Met	Leu	Tyr	Ile	Leu	Tyr	Val	Leu	Phe	Phe	Gly	Phe	Thr	Ser	170
			160					165								
AAG	GCC	GAA	AGC	ATG	CGC	CCC	GAG	GTC	GCA	TCC	ACG	TTC	AAA	GTA	CTG	938
Lys	Ala	Glu	Ser	Met	Arg	Pro	Glu	Val	Ala	Ser	Thr	Phe	Lys	Val	Leu	
			175					180								185
CGT	AAC	GTT	ACC	GTT	GTG	TTG	TGG	TCC	GCG	TAT	CCC	GTC	GTG	TGG	CTG	986
Arg	Asn	Val	Thr	Val	Val	Leu	Trp	Ser	Ala	Tyr	Pro	Val	Val	Trp	Leu	
			190					195								200
ATC	GCC	AGC	GAA	GGT	GCG	GGA	ATC	GTG	CCG	CTG	AAC	ATC	GAG	ACG	CTG	1034
Ile	Gly	Ser	Glu	Gly	Ala	Gly	Ile	Val	Pro	Leu	Asn	Ile	Glu	Thr	Leu	
			205					210								215
CTG	TTC	ATG	GTG	CTT	GAC	GTG	AGC	GCG	AAG	GTC	GCC	TTC	GGG	CTC	ATC	1082
Leu	Phe	Met	Val	Leu	Asp	Val	Ser	Ala	Lys	Val	Gly	Phe	Gly	Leu	Ile	235
220								225								230

FIG. 17(C)

CTC CTG CGC AGT CGT GCG ATC TTC GGC GAA GCC GAA GCG CCG ATC GAA 1130  
 Leu Leu Arg Ser Arg Ala Ile Phe Gly Glu Ala Glu Ala Pro Ile Glu 250  
 240  
 GGT CGT CAG AAA CAT ATC ATT TCC ATA AAC GAC CTT AGT CGC GAT GAC 1178  
 Gly Arg Gln Lys His Ile Ile Ser Ile Asn Asp Leu Ser Arg Asp Asp 265  
 255 260  
 CTT AAT CTG GTG CTG GCG ACA GCG GCG AAA CTG AAA GCA AAC CCG CAA 1226  
 Leu Asn Leu Val Leu Ala Thr Ala Ala Lys Leu Lys Ala Asn Pro Gln 280  
 270  
 CCA GAG CTG TTG AAG CAC AAA GTC ATT GCC AGC TGT TTC TTC GAA GCC 1274  
 Pro Glu Leu Leu Lys His Lys Val Ile Ala Ser Cys Phe Phe Glu Ala 295  
 285 290  
 TCT ACC CGT ACC CGC CTC TCT TTT CAA ACA TCT ATG CAC CGC CTG GCG 1322  
 Ser Thr Arg Thr Arg Leu Ser Phe Gln Thr Ser Met His Arg Leu Gly 315  
 300 305 310  
 GCC AGC GTG GTG GGC TTC TCC GAC AGC GCC AAT ACA TCA CTG GGT AAA 1370  
 Ala Ser Val Val Gly Phe Ser Asp Ser Ala Asn Thr Ser Leu Gly Lys 330  
 320 325 330

FIG. 17(D)

AAA GGC GAA ACG CTT GCC GAT ACC ATT TCA GTT ATC AGC ACT TAC GTC	1418
Lys Gly Glu Thr Leu Ala Asp Thr Ile Ser Val Ile Ser Thr Tyr Val	340 345
GAT GCG ATA GTG ATG CGT CAT CCG CAG GAA GGT GCG GCG CTG GCC	1466
Asp Ala Ile Val Met Arg His Pro Gln Glu Gly Ala Ala Arg Leu Ala	350 355 360
ACC GAG TTT TCC GGC AAT GTA CCG GTA CTG AAT GCC GGT GAT GGC TCC	1514
Thr Glu Phe Ser Gly Asn Val Pro Val Leu Asn Ala Gly Asp Gly Ser	365 370 375
AAC CAA CAT CCG ACG CAA ACC TTG CTG GAC TTA TTC ACT ATT CAG GAA	1562
Asn Gln His Pro Thr Gln Thr Leu Leu Asp Leu Phe Thr Ile Gln Glu	380 385 390 395
ACC CAG GGG CGT CTG GAC AAT CTC CAC GTC GCA ATG GTT GGT GAC CTG	1610
Thr Gln Gly Arg Leu Asp Asn Leu His Val Ala Met Val Gly Asp Leu	400 405 410
AAA TAT GGT CCG ACC GTT CAC TCC CTG ACT CAG GCG TTA GCT AAG TTC	1658
Lys Tyr Gly Arg Thr Val His Ser Leu Thr Gln Ala Leu Ala Lys Phe	415 420 425

FIG. 17(E)



GAC GGC AAC CGT TTT TAC TTC ATC GCG CCG GAC GCG CTG GCA ATG CCG 1706  
 Asp Gly Asn Arg Phe Tyr Phe Ile Ala Pro Asp Ala Leu Ala Met Pro  
 430 435 440

CAA TAC ATT CTG GAT ATG CTC GAT GAA AAA GGG ATC GCA TGG AGT CTG 1754  
 Gln Tyr Ile Leu Asp Met Leu Asp Glu Lys Gly Ile Ala Trp Ser Leu  
 445 450 455

CAC AGC TCT ATT GAA GAA GTG ATG GTG GAA GTA GAC ATC CTG TAC ATG 1802  
 His Ser Ser Ile Glu Glu Val Met Val Glu Val Asp Ile Leu Tyr Met  
 460 465 470 475

ACC CGC GTG CAA AAA GAG CGT CTG GAC CCG TCC GAG TAC GCC AAC GTG 1850  
 Thr Arg Val Gln Lys Glu Arg Leu Asp Pro Ser Glu Tyr Ala Asn Val  
 480 485 490

AAA GCG CAG TTT GTT CTT CGC GCC AGT GAT CTC CAC AAC GCC AAA GCC 1898  
 Lys Ala Gln Phe Val Leu Arg Ala Ser Asp Leu His Asn Ala Lys Ala  
 495 500 505

AAT ATG AAA GTG CTG CAT CCG TTG CCG CGT GTT GAT GAG ATT GCG ACG 1946  
 Asn Met Lys Val Leu His Pro Leu Pro Arg Val Asp Glu Ile Ala Thr  
 510 515 520

FIG. 17(F)

GAT GTT GAT AAA ACG CCA CAC GCC TGG TAC TTC CAG CAG GCA GGC AAC	1994
Asp Val Asp Lys Thr Pro His Ala Trp Tyr Phe Gln Gln Ala Gly Asn	
525	530
GGG ATT TTC GCT CTG CAA GCG TTA CTG GCA CTG GTT CTG AAT CCG GCC	2042
Gly Ile Phe Ala Leu Gln Ala Leu Ala Leu Val Leu Asn Arg Ala	
540	545
GGG ACC AGC GAC TGATCGCACA CGCAGGACAG CCCCACAACC GGCGGGCTG	2094
Ala Thr Ser Asp	
TGTTCAACGA CACACGATGA GTCCCCCACT CCGTCTTGTA CTCGGATCCT TTT	2147

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FIG. 17(G)

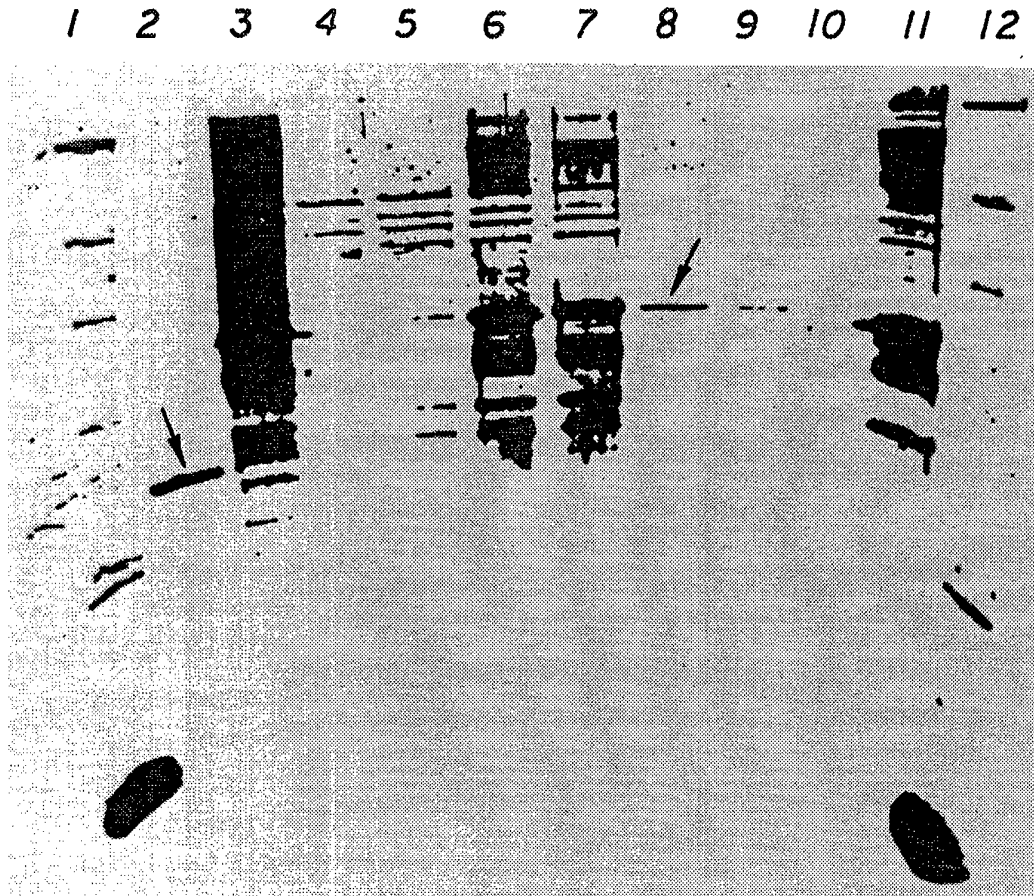


FIG. 18

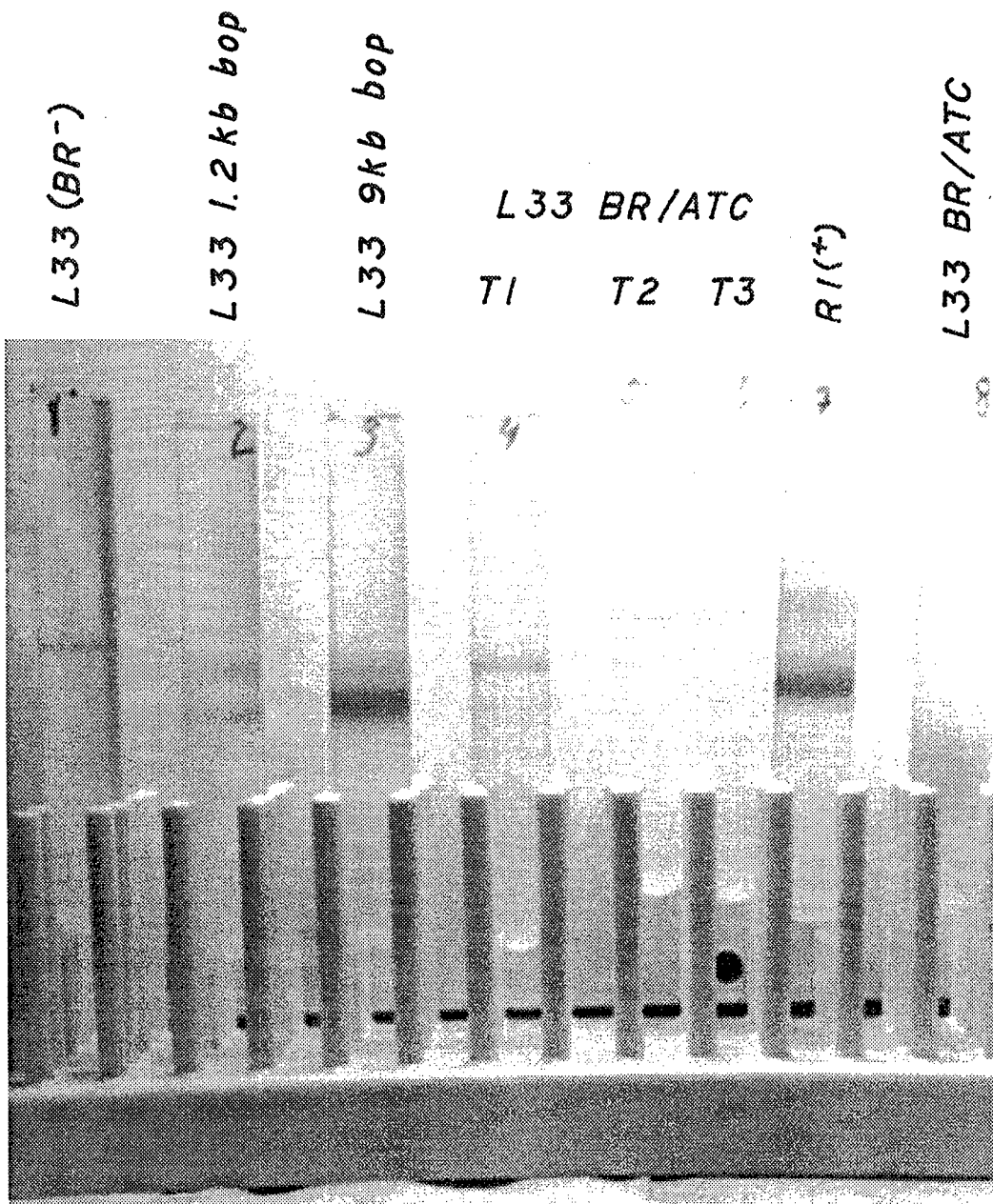
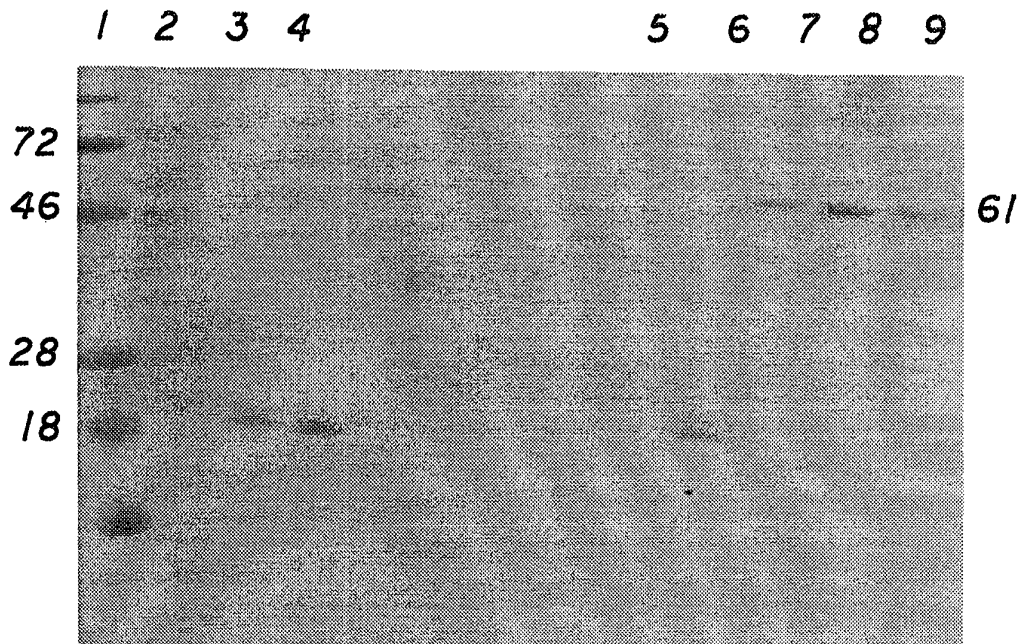


FIG. 19(A)



*FIG. 19(B)*

## INTERNATIONAL SEARCH REPORT

International Application No. PCT/US 94/02388

## A. CLASSIFICATION OF SUBJECT MATTER

C 12 N 15/11, C 12 N 1/21, C 12 N 15/00, C 12 P 21/00

According to International Patent Classification (IPC) or to both national classification and IPC<sup>5</sup>

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

C 12 N, C 12 P

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF THE UNITED STATES OF AMERICA, vol. 87, issued 1990, September U. BLASEIO et al. "Transformation of Halobacterium halobium: Development of vectors and investigation of gas vesicle synthesis", pages 6772-6776, the whole article.	1, 2, 10
A	PATENT ABSTRACTS OF JAPAN, unexamined applications, c field, vol. 8, no. 126, issued 1994, June 13 THE PATENT OFFICE JAPANESE GOVERNMENT,	1, 2, 10

 Further documents are listed in the continuation of box C. Patent family members are listed in annex.

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"&amp;" document member of the same patent family

Date of the actual completion of the international search

09 August 1994

Date of mailing of the international search report

26 -08- 1994

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## INTERNATIONAL SEARCH REPORT

-2-

International Application No  
PCT/US 94/02388

## C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
	page 30 C 228; & JP,A,59-36 700 (MITSUBISHI KASEI KOGYO). -----	