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NOTICE OF ENTITLEMENT

682508

We, THE REGENTS OF THE UNIVERSITY OF CALIFORNIA of 22nd Floor, 300 Lakeside Drive, Oakland, California, 94612-3550, UNITED STATES OF AMERICA state the following in connection with Australian Patent Application No. 65,875/94.

The person nominated for the grant of the patent (THE REGENTS OF THE UNIVERSITY OF CALIFORNIA) has entitlement from the actual inventors GEORGE J TURNER and MARY C BETLACH by Assignment.

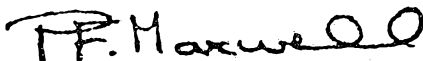
The person nominated for the grant of the patent (THE REGENTS OF THE UNIVERSITY OF CALIFORNIA) has entitlement from the applicants of the application listed in the declaration under Article 8 of the PCT on the basis of Assignment.

The basic application listed in the declaration under Article 8 of the PCT is the first application made in a Convention country in respect of the invention.

Dated this 17<sup>th</sup> day of April 1997

THE REGENTS OF THE UNIVERSITY OF CALIFORNIA

By their Patent Attorneys  
PETER MAXWELL & ASSOCIATES



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EXPRESSION OF HETEROLOGOUS POLYPEPTIDES IN HALOBACTERIA
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- (57) Claim

1. An expression vector useful for the production of heterologous polypeptide in a halobacterial host comprising:
- a) transcription and translation regulatory DNA;
  - b) DNA encoding a heterologous polypeptide; and
  - c) DNA encoding transcription and translation stop signals;
- wherein said DNA of a), b) and c) is operably linked, and wherein said heterologous polypeptide is other than a halobacteria polypeptide.



INTERN.

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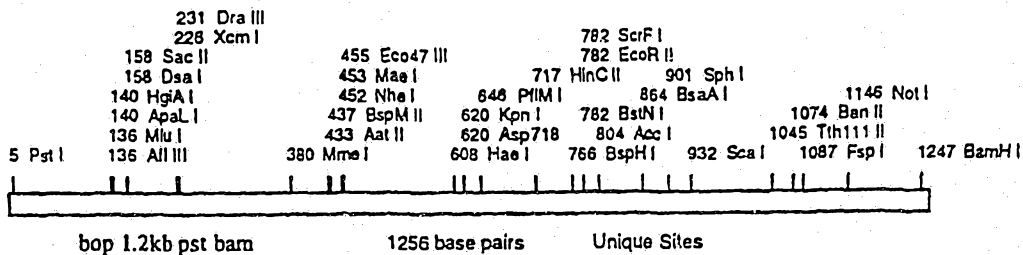
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(54) Title: EXPRESSION OF HETEROLOGOUS POLYPEPTIDES IN HALOBACTERIA



(57) Abstract

This invention relates to the preparation and use of expression systems capable of producing heterologous polypeptides in halobacterial hosts.

## EXPRESSION OF HETEROLOGOUS POLYPEPTIDES IN HALOBACTERIA

### Field of the Invention

The present invention is directed to the preparation and use of a halobacterial expression system that is capable of producing soluble and transmembrane heterologous polypeptides that are not endogenous to said halobacterium.

### Background of the Invention

Halobacteria are found in nature in evaporating salt water ponds under conditions of intense light and low oxygen saturation. They contain distinctive brightly colored pigments such as the orange-red pigment, bacterioruberin, or patches of "purple membrane". Halobacteria belong to a phylogenetically distinct group of prokaryotic organisms - the "archaebacteria" (Archaea) - that are as distantly related to the eubacteria as they are to the eukaryotes.

Archaebacteria possess some attributes in common with the eukaryotes and the eubacteria, as well as characteristics that are uniquely archaeal. For example, the archaebacteria possess a eukaryotic-like transcription apparatus with a 7-12 subunit RNA polymerase which is immunologically related to eukaryotic RNA polymerase (1) and promoter structures are similar to those of RNA Pol II (2). In contrast, the archaebacteria have prokaryotic cellular morphology and 23S, 16S and 5S rRNAs with the genes encoding the rRNAs arranged into eubacterial-like operons (3). Notably, the archaebacteria are unique in their membrane composition.

Bacteriorhodopsin (BR) is found as the sole protein in specialized crystalline patches of the "purple membrane" in halobacteria. Synthesis of BR is induced by high light intensity and low oxygen tension and the patches of purple membrane can constitute up to 50% of the archaebacterium *Halobacterium halobium* cell surface area.

BR consists of a complex of one protein (bacterio-opsin) along with the chromophore retinal in a 1:1 stoichiometric ratio (4). This complex is embedded in the lipid matrix as seven transmembrane hydrophobic  $\alpha$ -helices in a trimeric

configuration (5). Retinal is covalently attached at lysine at position 216 approximately one-third of the way across the transmembrane region of one of the  $\alpha$ -helices (6). The complex of bacterio-opsin with retinal was named bacteriorhodopsin (BR). The so-called *bop* gene encodes the light-driven protein pump bacteriorhodopsin (BR) in *H. halobium*.

There has been some reported research on expression of endogenous polypeptides in halobacteria (7, 8 and 9).

#### Summary of the Invention

The present invention is directed to the preparation and use of an expression system for heterologous polypeptide production in a halobacterial host.

In a first aspect, such systems in their broadest context would include transcription and translation regulatory DNA, DNA encoding a heterologous polypeptide that is not endogenous to the halobacterial host and DNA encoding transcription and translation stop signals.

Preferably such systems would include DNA encoding the pre-sequence of bacteriorhodopsin such that the polypeptide which is expressed is attached to the pre-sequence, thus allowing the heterologous polypeptide to be properly targeted to the membrane and either inserted into or secreted across the membrane.

Yet another preferred embodiment of the present invention uses the transcription and translation regulatory sequences and the translation and transcription stop sequences of the bacteriorhodopsin gene, either in the presence or absence of the bacteriorhodopsin pre-sequence. The use of the regulatory and stop sequences of the bacteriorhodopsin gene serves to allow high level expression of the heterologous polypeptide sequence.

In a second aspect, the present invention is also directed to utilizing the C-terminal domain of the bacteriorhodopsin polypeptide in order to enhance the separation of the mature heterologous polypeptide from the membrane of the halobacterial host following expression. In a preferred embodiment of this aspect, DNA encoding a unique protease site is introduced between said C-terminal sequence and the DNA encoding the heterologous polypeptide.

In a preferred embodiment of this aspect, high levels of expression of the heterologous polypeptide linked to the C-terminal region of bacteriorhodopsin are achieved by using DNA encoding the transcription and translation regulatory and stop sequences of the bacteriorhodopsin gene.

5 A further preferred embodiment of the invention is directed to the use of the bacteriorhodopsin pre-sequence to enhance expression of the heterologous polypeptide linked to the C-terminal region of bacteriorhodopsin.

The invention is directed to such systems in all their equivalent aspects, including expression vectors, halobacterial hosts transformed with such vectors  
10 and methods for producing, isolating and optionally further purifying heterologous polypeptides using such expression vectors.

#### Detailed Description

The present invention has been described herein by disclosing the preferred  
15 embodiments and best mode. It will be understood, however, that having detailed the method first used by the present inventors to produce the heterologous polypeptide expression system in halobacterium, it will be apparent to those skilled in the art that one could make modifications within the general skill of the art to produce expression systems that differ in one or more ways from that originally described.

#### 20 A) Brief Description of the Drawings

Figure 1 is a restriction map of the PstI/BamHI fragment containing the bacteriorhodopsin gene and about 400 bp of upstream sequences from *Halobacterium halobium* strain R1.

Figure 2 shows the nucleic acid sequence (SEQ ID NO:1) of the PstI/BamHI  
25 construct of Figure 1 containing the bacteriorhodopsin gene and about 400 bp of upstream sequences from *Halobacterium halobium* strain R1. Also shown is the amino acid sequence (SEQ ID NO:2) of the BR protein translation product.

Figure 3 shows the restriction map of pUBP2.

Figure 4 is a map of the secondary structure of the mature BR protein (SEQ ID NO:3).

5 Figure 5 is a restriction map of the *PstI/BamHI* fragment containing BR regulatory sequences and the gene for human muscarinic acetylcholine receptor (Type HM1) in pENDS-OM1.

Figure 6 shows the nucleic acid sequence (SEQ ID NO:6) of the *PstI/BamHI* fragment of Figure 5 containing the gene for human muscarinic acetylcholine receptor (Type HM1) of pENDS-OM1. Also shown is the amino acid sequence (SEQ ID NO:7) of HM1.

10 Figure 7 is a restriction map of the *PstI/BamHI* fragment containing the BR regulatory sequences and gene for human muscarinic acetylcholine receptor (Type HM1) in pENDS-OM2.

15 Figure 8 shows the nucleic acid sequence (SEQ ID NO:8) of the *PstI/BamHI* fragment of Figure 7 containing the gene for human muscarinic acetylcholine receptor (Type HM1) which lacks the I3 domain. The amino acid sequence (SEQ ID NO:9) of HM1 having a deleted I3 domain is shown.

Figure 9 is a restriction map of the *PstI/BamHI* fragment containing the BR regulatory sequences and the rat serotonin receptor (Type 1C) gene.

20 Figure 10 shows the nucleic acid sequence (SEQ ID NO:10) of the *PstI/BamHI* construct of Figure 9 containing the rat serotonin receptor gene and the amino acid sequence (SEQ ID NO:11) of the rat serotonin receptor.

25 Figure 11 is a Southern blot of DNA isolated from *H. halobium Bop* deficient strain L33 transformed with pUBP2 containing the rat serotonin receptor (Type 1C) gene. Lanes 1-10, 12-19, 21-24 and 27 contained DNA from strain L33 transformed with pUBP2 containing the *PstI/BamHI* fragment of Figs. 9 and 10

(SEQ ID NO:10). Lanes 11 and 25 are positive controls which contained purified plasmid DNA (i.e. pUBP2 containing serotonin receptor gene). Lane 29 contained DNA from strain L33. The arrow indicates the location of the *PstI/BamHI* fragment corresponding to serotonin DNA.

5 Figure 12 shows a Northern blot of total RNA isolated from *H. halobium Bop* deficient strain L33 transformed with pUBP2 containing the rat serotonin receptor gene. Lanes 2 and 5 contain RNA from wild type strain L33 transformed with the 1.2 kb *PstI/BamHI* fragment containing the *bop* gene in pUBP2 as a control. Lanes 1, 3 and 4 contain DNA from L33 transformed with  
10 the rat serotonin receptor gene. The 1.85 kb *PstI/BamHI* fragment of Figs. 9 and 10 was used as probe. The arrow shows the location of the rat serotonin receptor RNA.

Figure 13 is a restriction map of the *PstI/BamHI* fragment containing BR regulatory sequences and the human thrombin receptor gene.

15 Figure 14 shows the nucleic acid sequence (SEQ ID NO:12) of the *PstI/BamHI* fragment of Figure 13 containing the human thrombin receptor gene and the amino acid sequence (SEQ ID NO:13) of the human thrombin receptor.

Figure 15 shows the restriction maps of p $\beta$ gbop, pEK17, pBATC, p1.2KbBop and pBRAT.

20 Figure 16 shows a restriction map of the *PstI/BamHI* fragment containing BR regulatory sequences, the bacterio-opsin gene and the gene encoding the *Escherichia coli* catalytic subunit of aspartate transcarbamylase.

Figure 17 shows the nucleic acid sequence (SEQ ID NO:14) of the *PstI/BamHI* fragment of Figure 16 containing the bacterio-opsin and the *E. coli* aspartate  
25 transcarbamylase genes and the amino acid sequence (SEQ ID NO:15) of the BR/*E. coli* aspartate transcarbamylase fusion protein.



Figure 18 shows a Western blot of *H. halobium* transformed with pBRAT. Blots were probed with antibodies to the catalytic subunit of aspartate transcarbamylase. Lane 2 contains *E. coli* aspartate transcarbamylase. Lanes 6-9 and 11 contain protein from *H. halobium* transformed with pBRAT. The arrow in lane 8 indicates the position of the bacteriorhodopsin/aspartate transcarbamylase (BR/ATCase) fusion protein.

Figure 19 shows the localization of expression of the bacteriorhodopsin/aspartate transcarbamylase (BR/ATCase) fusion protein to the purple halobacterial cell membranes. Washed *H. halobium* whole cell membranes fractionated on sucrose density gradients (A) were electrophoresed on SDS-polyacrylamide gels and stained with Coomassie blue (B). Lanes in (B) contained the following protein samples: Molecular weight markers (lane 1); unfractionated total membranes from *H. halobium* strain L33 transformed with pBRAT (lane 2); purple membrane from *H. halobium* strain L33 transformed with a 1.2 Kb *PstI/BamHI* fragment containing the *bop* gene (lane 3) or with a 9 Kb genomic DNA fragment containing the *bop* gene (lane 4); total membranes from *H. halobium* strain L33 (lane 5); purple membrane from wild-type *H. halobium* strain R1 (lane 6); purple membrane of *H. halobium* strain L33 transformed with pBRAT (lanes 7-9).

## 20 B) Definitions

The term "expression vector" herein has a functional definition and includes vectors capable of expressing DNA sequences contained therein, where such sequences are operably linked to other sequences capable of effecting their expression. In the present specification, "vector" and "plasmid" are used interchangeably as the plasmid is the most commonly used form of vector. However, the invention is intended to include such other forms of expression vectors capable of equivalent functions and which are or become known in the art.

By the term "operable" herein, and grammatical equivalents, is meant that the respective DNA sequences are operational and work for their intended purposes.

5 The term "heterologous polypeptide" herein refers to presently known or unknown polypeptides not endogenous to the host cell, or if endogenous to the host cell, are obtainable herein in amounts not achievable in native state. Included within the definition are the halobacterial non-retinal binding proteins. Examples of heterologous polypeptides include, but are not restricted to, polypeptides from eukaryotes, eubacteria, archaeobacteria, synthetic polypeptides and polypeptides containing bioequivalent amino acid analogs. Further included  
10 are other members of the 7-transmembrane crossing family such as muscarinic acetylcholine receptor, serotonin receptor, thrombin receptor,  $\beta$ -adrenergic receptor, and the like. Heterologous polypeptides also include membrane proteins, for example, cystic fibrosis transmembrane conductance regulator, and soluble proteins, such as various enzymes (e.g. proteases and aspartate  
15 transcarbamylase). Each is used in accord with their known or determined function biologically and is adapted for such in accord with procedures generally known in the art.

By the term "DNA encoding heterologous polypeptide" is meant a DNA  
20 sequence coding for a polypeptide that is not endogenous to the host wherein it is expressed. Because of the high GC content (i.e. about 58-68%) of the genome of halobacteria, it is preferred that the DNA sequence encoding the heterologous polypeptide be in this range, although sequences with higher and lower GC content than that usually found in halobacteria can be used. For example, we  
25 have been successful in expressing *Escherichia coli* aspartate transcarbamylase, having a GC content of about 50%, as a fusion protein to the C-terminus of BR.

The term "transcription and translation regulatory DNA" and equivalents, in its broadest sense refers to a DNA sequence responsible for the dual transcription and translation elements of expression. In a preferred embodiment  
30 the regulatory DNA is that of the bacteriorhodopsin gene (from -364 to +41 relative to the RNA start site, Fig.2 (SEQ ID NO:1)).

In an alternative embodiment, the regulatory DNA contains about 4000 bp of sequences (from about -4000 to +41) upstream of the *bop* gene and includes three other genes of the *bop* gene cluster, which include *brp* (13), *bat* (14) and *blp* (Gropp & Betlach, manuscript in preparation). Some or all of these genes may be regulatory genes.

By the term "transcription and translation stop signals" and equivalents, in its broadest scope is meant DNA which functions to terminate transcription and translation, respectively. It is preferred that the transcription and translation stop signals be those of the bacteriorhodopsin gene.

By the term "pre-sequence of bacteriorhodopsin gene" herein is meant a sequence of about 13 amino acids required to target bacteriorhodopsin to the membrane. The 13 amino acid pre-sequence is encoded by nucleotides +3 to +41 relative to the RNA start site depicted in Fig. 2 (SEQ ID NO:1).

By the term "halobacterium host" is meant strains belonging to Halobacterium, including species of extreme and moderate halophiles having a wild-type genotype. Examples of the extreme halophilic species having a wild type genotype include *Halobacterium saccharovorium* (ATCC 29252), *Halobacterium californica* (ATCC 38799), *Halobacterium halobium* (CCM 2090) and *Halobacterium valismortis* (ATCC 29-715). Wild type moderate halophiles are exemplified by the species *Halobacterium mediterranei* (ATCC 33500). It may be preferred that the halobacterial host species is bacteriorhodopsin deficient. Bacteriorhodopsin deficient species are either wild-type, such as *H. volcanii*, or mutants, such as L33 (15), S9F1x3 (16), IV-8 (17) and IV-14 (17). Bacteriorhodopsin deficient mutants derived from strains which express purple membrane constitutively, such as L33, or inducibly, are useful for different applications. Depending on the nature of the upstream regulatory regions in the expression vector construct, inducible strains permit regulated expression whereas constitutive strains do not.

The term "restriction site" herein refers to a DNA sequence recognizable by an endonuclease as a site of DNA cleavage.

By the term "C-terminal sequence", "C-terminal region" and equivalents, is meant the polypeptide sequence at the C-terminus of bacteriorhodopsin; See Fig. 4.

5 By the term "unique protease site" is meant an amino acid sequence recognizable by a protease as the site of cleavage of the polypeptide wherein it is disposed and which is absent from the heterologous polypeptide expression product. In a preferred embodiment, the protease site (Ile-Glu-Gly-Arg) (SEQ ID NO:4) of factor X<sub>a</sub> is used in view of the rarity of this sequence.

C) Examples

- 10 1. Cloning the DNA sequence encoding the heterologous polypeptide into a halobacterial expression vector

i. Constructs for expression of membrane proteins

15 All constructions are assembled using standard molecular techniques (12) including PCR. Expression vectors can be prepared in a variety of conventional ways. Although others may be used, a preferred halobacterial cloning vector to be adapted into an expression vector is plasmid pUBP2 (Fig.3) described by Blaseio et al. (7). The plasmid may be isolated using conventional techniques. For example, the plasmid may be purified using caesium chloride-ethidium bromide density gradients, electrophoresis from an agarose gel onto a dialysis  
20 membrane, use of commercially available chromatography columns for the separation of plasmids, such as magic minipreps DNA purification system (Promega Corp., Madison, WI), etc.

25 The expression vectors which will be employed will normally include a marker which allows for selection of cells into which the DNA has been integrated, as against cells which have not integrated the DNA construct. An example of commonly used selection markers is antibiotic resistance. Two markers are available for selection of halobacteria, including resistance to novobiocin (8) and mevinolin (7). It is preferred that the marker used be that for mevinolin resistance; mevinolin is a HMG CoA reductase inhibitor (7). This  
30 marker is present in the preferred cloning plasmid pUBP2 (Fig.3).

To convenience insertion of DNA sequences, plasmids will contain polylinker sequences containing various restriction sites. Several examples of polylinkers are known and available (12). A typical polylinker is polylinker 1 (12.3) (Fig.3.) which contains restriction sites for HindIII, SphI, MluI, XhoI, PstI, 5 SalI, XbaI, *Bam*HI, HindIII, XbaI and KpnI. Another typical polylinker is polylinker 2 (3.70) (Fig.3) with restriction sites for SphI, EcoR5, SstI, SmaI and EcoRI.

The DNA sequence encoding the heterologous polypeptide is inserted such that it is placed downstream from a transcription and translation regulatory region containing a promoter and a ribosome binding site using standard 10 techniques. It is preferred that the promoter used is inducible, allowing controlled expression of the heterologous polypeptide product. In a preferred embodiment of the invention, the transcription and translation regulatory sequences of the bacteriorhodopsin gene will be used. The bacteriorhodopsin 15 gene may be isolated from the genome of halobacteria using appropriate restriction enzymes. Transcription and translation regulatory sequences of the bacteriorhodopsin gene are located in the region of -365 to +41 relative to the RNA start site of the bacteriorhodopsin sequence depicted in Fig.2 (SEQ ID NO:1).

To effect appropriate termination of heterologous polypeptide synthesis, 20 DNA sequences encoding transcription and translation stop signals are placed downstream of the inserted DNA sequence encoding the heterologous polypeptide sequence using well known techniques (12). Preferably, the sequences downstream of the bacteriorhodopsin gene (Fig.2) (SEQ ID NO:1) 25 which includes the translational stop codon (TGA) followed by ~80 bp which include the transcriptional termination signal are employed as stop signals.

Where it is advantageous to produce a heterologous transmembrane polypeptide which is targeted to the halobacterial membrane, DNA encoding the heterologous polypeptide is ligated downstream of DNA encoding the pre- 30 sequence of BR.

The heterologous gene of interest may be cloned into the *E. coli* plasmid, pUC19 (20), along with BR regulatory sequences such that all cloned sequences will reside on a DNA fragment containing two unique restriction sites (choice of *Pst*I, *Bam*HI, *Sma*I). More specifically, the heterologous gene is ligated such that  
5 it is in frame with the BR pre-sequence, downstream of the bacteriorhodopsin regulatory sequences/promoter and upstream of the bacteriorhodopsin transcriptional and translational termination sequences. A specific unique protease site may be engineered into some constructions between the BR pre-sequence and the heterologous gene.

10 A 1.2 kbp fragment containing the *bop* gene and ~370 bp of upstream sequences was isolated from *H. halobium* strain R1 DNA using PCR and cloned into the *Pst*I/*Bam*HI sites of pUC19 (denoted p1.2K**bop**) (Fig.15B). Two endogenous *Alw*NI sites were removed from the cloned 1.2 kbp fragment: i) one site located 165 bp upstream of the *bop* gene start codon (SEQ ID NO:1) was  
15 removed by generating a G→T point mutation using the Kunkel method (29), and ii) the second *Alw*NI site located 7 bp upstream of the *bop* gene stop codon was removed using the Transformer Site-Directed Mutagenesis kit (Clontech Laboratories, Inc., Palo Alto, CA). Subsequently, a ~400 bp *Pst*I/*Alw*NI fragment (denoted "*bop* 5' fragment") containing the *bop* upstream sequences,  
20 DNA encoding the BR presequence and the first four (extrahelical) residues of BR was isolated by PCR from the mutated 1.2 kbp fragment. Concurrently, a ~100 bp *Not*I/*Bam*HI fragment (denoted "*bop* 3' fragment") containing DNA encoding six C-terminal residues of BR, the BR stop codon and the transcriptional termination sequences of BR (up to 44 bp downstream of the stop  
25 codon) was obtained from the 1.2 kbp *bop* gene fragment by preparative digestion and purification (Prep-A-Gene, BioRad, Richmond, CA). In addition, an endogenous *Alw*NI site located in pUC19 (position 1217) was removed using the Clontech Transformer kit and the mutated pUC19 was preparatively digested with *Pst*I/*Bam*HI and preparatively purified (denoted "vector fragment"). The  
30 three fragments (i.e., "*bop* 5' fragment", "*bop* 3' fragment" and "vector fragment") were ligated with DNA fragments containing various heterologous genes engineered to be in frame with the BR presequence and extrahelical

residues and to contain a single *AlwNI* site at the 5' terminus of the fragment and a single *NotI* site at the 3' terminus of the fragment as described below. In all of the heterologous genes, endogenous *AlwNI*, *NotI*, *BamHI* and *PstI* sites were first removed (if necessary) to facilitate the construction. Once the  
5 heterologous gene was cloned along with the BR 5' and 3' regulatory sequences into pUC19, this intermediate construct (denoted "pENDs") was preparatively digested with *PstI/BamHI*

Subsequently, the *PstI/BamHI* restriction fragment containing the heterologous gene with the regulatory sequences of BR was preparatively isolated  
10 away from pUC19 sequences by agarose gel electrophoresis, purified using Prep-A-Gene (Bio-Rad, Richmond, CA) and cloned into the *E. coli/H. halobium* shuttle vector, pUBP2 (7). pUBP2 carries the pBR322 replicon and ampicillin resistance marker, the halobacterial plasmid pHH1 origin of replication and a mevinolin resistance marker. Mevinolin resistance is encoded by an up-promoter  
15 mutation of the HMG-CoA reductase gene.

The construction was verified by restriction mapping and nucleotide sequencing across the junctions between 5' and 3' BR regulatory sequences and the heterologous gene.

a. Human muscarinic acetylcholine receptor (Type HM1)

20 Two different constructs were made with this gene. The first (denoted pENDs-OM1) contained the entire gene whereas the second (denoted pENDs-OM2) lacked the large internal cytoplasmic loop (i.e., I3) which is thought to be involved in signaling. Prior to the generation of the constructions described below, two endogenous *AlwNI* sites and one endogenous *PstI* site were removed  
25 from human muscarinic acetylcholine receptor (denoted HM1) cloned in pGEM3 (Promega Corp, Madison, WI) using either the Clontech Transformer kit or the Kunkel method (29). The positions of the removed sites are shown in Fig.6 (SEQ ID NO:6).

pENDs-OM1 was generated as follows. First, the HM1 gene was isolated  
30 by PCR from pGEM3/HM1 so as to contain an *AlwNI* site at the 5' terminus and a *NotI* site the 3' terminus of the PCR fragment. This PCR fragment was

ligated to the "bop 5' fragment", "bop 3' fragment" and "vector fragment" described above and transformed into *E. coli*. The resultant plasmid was named pENDs-OM1. pENDs-OM1 contains the methionine start codon of HM1 located 4 codons downstream from the BR 5' sequences. Nine extra base pairs generated by introduction of the *AlwNI* site encode 3 extra residues (i.e., gln, ala, leu) located in frame between the BR 5' sequences and the start codon of the HM1 gene. At the 3' terminus of the gene, the HM1 stop codon precedes the BR stop codon by 48 bp. From pENDs-OM1, the BR regulatory sequences with the HM1 gene were transferred to pUBP2 on a *PstI/BamHI* fragment (Fig.5 and Fig.6, SEQ ID NO:6) as described above.

pENDs-OM2 was generated in a similar manner as its sibling construct. First, however, deletions of the I3 domain were introduced after digestion of the HM1 gene at the unique *StuI* restriction site (position 712 relative to the start codon of the HM1 gene, SEQ ID NO:6), followed by digestion with the exonuclease *Bal-31* for varying times at 4°C. The blunt-ended product was self-ligated to yield mutants with deletions of varying size within the I3 domain. One of these was chosen for further study which lacked amino acid residues 231 through 357 of HM1 (SEQ ID NO:7). DNA from this mutant was used to generate a PCR fragment containing the HM1 gene (less I3 loop) with a 5' *AlwNI* site and a 3' *NotI* site. This PCR fragment was identical to the fragment described above except for the lack of the I3 loop and was used to generate pENDs-OM2 in a similar manner to the pENDs-OM1 construct. The sequence of the *PstI/BamHI* fragment containing the BR regulatory sequences and the HM1 gene (less I3 loop) is shown in Fig.8 (SEQ ID NO:8).

25                    b.     Rat serotonin receptor (Type 1 C)

The rat serotonin receptor gene (denoted "Ser") cloned as a 3 Kb *EcoRI* cDNA fragment on the plasmid pSR1c (27) was used as a basis for the following constructions. The Ser gene contains no endogenous *AlwNI*, *NotI*, *BamHI* and *PstI* sites and was adapted for expression in *H. halobium* as follows. *AlwNI* and *NotI* cloning sites were introduced within the 5' coding and 3' noncoding regions of the Ser gene, respectively. In addition, DNA encoding a poly-aspartic acid



peptide was placed in frame upstream of the Ser gene and downstream of the *AlwNI* site. Translation of this sequence generates a peptide epitope useful for subsequent detection of expressed protein (31). This fragment was isolated and ligated to the "bop 5' fragment", "bop 3' fragment", and "vector fragment" described above and transformed into *E. coli*. The resultant plasmid was named pENDs-Ser and contains the 36th codon of the rat serotonin receptor gene preceded by DNA encoding the peptide epitope and BR 5' sequences. Nine extra base pairs generated by the construction and encoding 3 extra residues (i.e., gln, ala, leu) are located in frame between the BR 5' sequences and the epitope sequences. At the 3' terminus of the gene, the Ser stop codon precedes the BR stop codon by 18 bp. Following the construction of pENDs-Ser, the BR regulatory sequences with the Ser gene were transferred to pUBP2 on a *PstI/BamHI* fragment (Fig.9 and Fig.10, SEQ ID NO:10).

c. Human thrombin receptor

A clone of the human thrombin receptor gene (denoted "Thromb") (33) was used as a basis for the following constructions. Four endogenous DNA restriction sites were removed from the gene using the Kunkel method (29). These included three *AlwNI* sites (291, 945, and 1038) and one *PstI* site (537). Positions are given relative to the first base of the start codon of the gene. "pENDs-Thromb" was generated as follows. An *AlwNI/NotI* fragment containing the gene was generated using oligonucleotide-directed-insertion-mutagenesis and PCR. Included on this fragment were additional nucleotide sequences encoding short peptides for use in the detection and purification of the expressed protein. The *AlwNI/NotI* fragment containing the gene along with epitope encoding sequences was ligated to the "bop 5' fragment", "bop 3' fragment" and "vector fragment" described above and transformed into *E. coli*. The resultant plasmid was named pENDs-Thromb. In pENDs-Thromb, thirty-three extra base pairs generated by the construction and encoding eleven extra amino acids are located in frame between the BR 5' sequences and the Thromb sequences. Twenty seven of the extra residues encode a poly-aspartic acid peptide sequence which when translated generates a peptide epitope useful for detection of expressed

protein (31). At the 3' terminus of the gene, six histidine codons have been inserted upstream of the Thromb stop codon. These histidine codons are intended to aid in the affinity purification of expressed protein (26). At the 3' terminus of the gene, the Thromb stop codon precedes the BR stop codon by 18 bp.

The BR regulatory sequences with the human thrombin receptor gene may be transferred into pUBP2 on a *PstI/BamHI* fragment (Fig.13 and Fig.14, SEQ ID NO:12) as described above.

ii. Constructs for expression of soluble proteins

Where it is desired that heterologous soluble polypeptide be released extracellularly into the culture medium following expression, the DNA sequence encoding the heterologous polypeptide may be ligated to DNA encoding the pre-sequence of bacteriorhodopsin (Fig.2 (SEQ ID NO:1), from +3 to +41 relative to the RNA start site) using techniques well known to those skilled in the art (12).

Where it is advantageous to produce a heterologous soluble polypeptide that is targeted, following expression, to the halobacterial membrane, DNA encoding the heterologous polypeptide is ligated downstream of the DNA encoding the C-terminal region (Fig.2 and Fig.4 (SEQ ID NOs:1 and 3)) of bacteriorhodopsin or to fragments thereof.

To facilitate subsequent purification of the heterologous polypeptide product, a DNA sequence encoding a unique protease site is engineered between DNA encoding the bacteriorhodopsin C-terminal region and DNA encoding the heterologous polypeptide. Sequences encoding unique protease cleavage sites are known and include, for example, subtilisin, thrombin, enterokinase, and factor  $X_a$ . In a preferred embodiment, a DNA sequence encoding the amino acid sequence Ile-Glu-Gly-Arg (SEQ ID NO:4) is used to encode a unique protease site which is recognized by Factor  $X_a$ .

Design of the soluble protein expression vector and methods used are similar to that described above for membrane proteins. However, soluble proteins are expressed as in-frame fusions to the C-terminal region of BR. Thus,

these fusion proteins will have membranous domain (i.e. BR or portions thereof) and a soluble domain (i.e. heterologous polypeptide). The heterologous gene is cloned at the C-terminus of BR, between the bacteriorhodopsin gene and the downstream transcriptional/translational termination sequences of BR. In addition, a unique protease site is engineered between BR and the heterologous gene to facilitate subsequent purification of the protein. The final construct is cloned into the *E. coli/H. halobium* shuttle vector, pUBP2 (7).

a. *E. coli* Aspartate Transcarbamylase (catalytic subunit)

The catalytic subunit of Aspartate Transcarbamylase, (denoted ATCase), a soluble protein, has been fused to the C-terminus of BR as follows. The *bop* gene containing plasmid, p $\beta$ gbop (32), was digested at the unique *NotI* site located near the 3' terminus of the *bop* gene (see Figure 15A). Subsequently, this *NotI* site was filled-in to create a blunt site (12). The resulting DNA was digested with *SphI* to generate two fragments, a large fragment (denoted fragment 1) containing the vector along with the N-terminus of the *bop* gene and a small fragment containing internal *bop* gene sequences. Fragment 1 was isolated and purified. A second aliquot of p $\beta$ gbop was digested with *SphI/HaeII* and a 217 bp fragment (denoted fragment 2) containing an internal portion of the *bop* gene was isolated and purified (Figure 15A).

The structural gene for the *E. coli* catalytic subunit of aspartate transcarbamylase was isolated from pEK17 (Fig.15A) (30). A 845 bp *MseI/NruI* fragment (denoted fragment 3) which contains all but the first 18 bp of the gene encoding ATCase was isolated and purified.

A synthetic fragment of DNA (denoted fragment 4) was constructed by annealing two complementary oligonucleotides and used to connect the *bop* and ATCase genes. The synthetic fragment was engineered to contain a *HaeII* site at the 5' terminus, a *MseI* site at the 3' terminus and an internal *NruI* site. Also included were nucleotides encoding: i) a unique protease site (i.e., blood clotting Factor X<sub>a</sub>) and ii) ATCase amino acids 6 and 7 (relative to ATCase start codon)

Fig.17, SEQ ID NO:14.

All four DNA fragments were ligated together and used to transform *E. coli* strain D1210 (28) with selection for ampicillin resistance. Positive clones were identified by colony filter hybridization using P<sup>32</sup> radiolabeled random primed (25) ATCase *MseI/NruI* fragment as probe. Positive clones were verified  
5 by restriction mapping and nucleotide sequencing. One positive clone was chosen and denoted pBATC (Figure 15A).

Subsequently, the *bop*-ATCase fusion construct was adapted for *H. halobium* expression as follows. A fragment spanning the sequences in between and including the internal *SphI* site of the *bop* gene at the 5' terminus and the  
10 ATCase translational stop codon at the 3' terminus was isolated from pBATC by PCR (see Figure 15B). In addition, the oligonucleotide used to construct the 3' terminus of this PCR fragment was designed to be complementary to *bop* sequences downstream of the transcriptional termination sequences and to include a unique *BamHI* to facilitate subsequent cloning steps. The resultant  
15 PCR fragment was digested with *SphI/BamHI*, purified and used in the following construction.

The plasmid, p1.2Kbbop, containing the *bop* gene and upstream sequences cloned in pUC19 (described above) was digested with *SphI/BamHI* to yield two fragments, a large one containing the vector and the majority of the *bop* gene,  
20 and a 358 bp fragment containing the C-terminal half of the *bop* gene (Fig.15B). The larger of these two fragments was isolated, purified and ligated to the *SphI/BamHI* *bop*-ATCase PCR fragment. A positive clone was isolated and confirmed by restriction mapping and nucleotide sequencing. This clone was digested with *PstI/BamHI* and a fragment containing DNA encoding the  
25 BR/ATCase fusion along with *bop* upstream regulatory sequences (Fig.16) was cloned into the *E. coli/H. halobium* shuttle vector pUBP2. The resultant construct was named pBRAT (Fig.15B). The nucleotide sequence (SEQ ID NO:14) and the translated amino acid sequence (SEQ ID NO:15) of this *PstI/BamHI* fragment is shown in Fig.17.

## 2. Transformation of *Halobacterium halobium*

The *Pst*I/*Bam*HI fragments of the pENDs-Ser (Fig.9 and 10, SEQ ID NO:10) and pBRAT (Fig.15B, Fig.16 and Fig.17, SEQ ID NO:14) constructs containing the heterologous genes with the BR regulatory sequences were isolated and purified. Subsequently, these fragments were cloned into the  
5 *E. coli*/*H. halobium* shuttle vector pUBP2 (7) and transformed into *H. halobium* Bop deficient strain L33 as described (24).

Preferably, plasmids may be introduced into halobacteria using the polyethylene glycol (PEG) method (10, 11). Transformed halobacterial cells are then grown in culture in an appropriate nutrient medium sufficient to maintain  
10 the growth of halobacterial cells (7, 8).

*H. halobium* is prone to cell lysis during transformation procedures (7). Since surfactants are known to promote halobacterial lysis (21), all media and glassware used were soap-free. Transformation was performed according to  
15 Blaseio (7) and Cline (11) with modifications. Initially, cells were subcultured several times in soap-free complex (YET) medium. Subsequently, cells were subcultured to an OD<sub>660</sub> of about 0.01 and grown at 40°C until the early to mid-logarithmic stage of growth (OD<sub>660</sub> of 0.4 to 0.6). All succeeding manipulations were performed at room temperature. The culture was removed from the  
20 waterbath shaker and incubated without agitation for 4 h to overnight, followed by centrifugation of 2 ml of culture at 1000 x g for 15 min. The supernatant was carefully removed with a pipette and the interior of the centrifuge tube dried with absorbent tissue. The cell pellet was resuspended in 1/10 volume of spheroplasting solution (11), followed by addition of 1/100 volume of 0.5 M  
25 EDTA in Spheroplasting solution (11) and incubation for 2 min. One µg of DNA in 10 µl of spheroplasting solution was then added to the spheroplasted cells along with an equal volume of 60% PEG 600 (un-recrystallized) in spheroplasting solution. The combined solutions were gently but thoroughly mixed and then incubated for 20 min. Ten ml of 15% sucrose in complex (YET)  
30 medium was added followed by incubation overnight with no agitation at 42°C. The following day, cells were centrifuged at 3000 x g for 15 minutes and

resuspended in 300  $\mu$ l of 15% sucrose in complex (YET) medium. This solution was plated on solid complex (YET) selection medium.

3. Analysis of transformants, expression of the heterologous polypeptide and assays for expression

5 To establish that halobacterial cells have been successfully transformed, various techniques may be employed. Where the expression vector used to transform the halobacteria contains a dominant selectable marker, transformed cells can be selected by growing in the appropriate selection medium such that growth of halobacterial cells not harboring the recombinant plasmid is inhibited.  
10 For example, where a plasmid containing the mevinolin resistance marker is used, halobacterial cells which harbor this plasmid may be selected by growing on solid nutrient medium containing mevinolin at a concentration in the range of 5 to 25  $\mu$ M. Further, the plasmid may be isolated using standard techniques (12), restricted and used. The polymerase chain reaction, gel electrophoresis,  
15 restriction analysis, Southern, Northern, and Western blots may be employed, sequencing, or the like, may all be employed with advantage.

Depending upon the particular construct and the halobacterial background strain which have been employed for expression of the heterologous polypeptides, one may have constitutive or inducible expression of the heterologous  
20 polypeptide product. In the case of constitutive expression, the product will be continuously formed as the cells grow. By contrast, for inducible expression, one may provide for induction when the cells reach a predetermined cell density.

Where inducible promoters have been engineered into the expression vector containing the heterologous polypeptide DNA sequence, transcription may  
25 be induced using appropriate inducers under such conditions of concentration and duration as to effect induction of transcription. For example, if the regulatory sequences of the bacteriorhodopsin gene are used, transcription can be induced by low oxygen tension and high light intensity (18, 19) which are known to induce high level expression of BR. Low oxygen tensions are achieved  
30 in various ways such as by flushing culture flasks with oxygen-free nitrogen and sealing them, or by permitting cultures to reach the stationary phase of growth

in which oxygen limitation occurs naturally (18). High light intensity of greater than about 100 mW/cm<sup>2</sup> can be achieved using various light sources and apparatus as described (18, 19).

5 *H. halobium* transformed with pUBP2 containing the pBRAT *PstI/BamHI* fragment and with pUBP2 containing the pENDS-Ser *PstI/BamHI* fragment was plated on solid complex (YET) medium containing 25 μM mevinolin. Plates were incubated for one to two weeks at 42°C to permit growth of transformants. Plasmid DNA was isolated from individual transformants using Magic Minipreps DNA Purification System (Promega Corp., Madison, WI). Southern analysis was used to verify the presence of the heterologous gene on pUBP2. Southern blot analysis using the *AlwNI/NotI* fragment containing the serotonin receptor gene as probe indicated the presence of serotonin receptor gene sequences in all 10 assayed transformants (Figure 11). Total RNA was isolated from individual transformants using the RNazol procedure (Cinna Biotech) and subjected to Northern analysis (18). Northern blot analysis revealed that transcription of Ser 15 gene sequences had occurred (Figure 12). Western analysis using both BR and ATCase antibodies demonstrated that the BR/ATCase fusion was expressed and localized to halobacterial membranes (Figure 18). Washed halobacterial whole cell membranes were fractionated on sucrose gradients (Fig.19A) and aliquots 20 were subjected to SDS PAGE (Fig.19B). A band corresponding to the predicted molecular weight of the fusion protein (i.e., ~60 kDa, see Fig.19B) was observed which derived from a purple fraction. These data verify expression of the fusion and indicate that the BR portion of the fusion is folded correctly in the halobacterial membrane. The presence of the BR chromophore (extinction 25 coefficient of 63,000; 31) affords an estimate of 5 mg/liter of fusion protein expression.

Transformants testing positive in Southern and Northern analyses are subjected to Western analysis if specific antibodies to the heterologous protein are available. If antibodies are not available, DNA encoding an epitope known 30 to be antigenic may be engineered into the expression vector construction to aid in detection of expression. An example of such an epitope is the sequence encoding Glu-Glu-Glu-Glu-Tyr-Met-Pro-Met-Glu (SEQ ID NO:5) (22).

Alternatively, expression of the heterologous protein may be assayed functionally; for example, ligand binding assays for receptors, and assays for enzymic activity for soluble proteins using appropriate substrates.

#### 4. Purification of heterologous polypeptides

5 Production of the heterologous polypeptide may be stopped in a variety of ways. Where the heterologous polypeptide is released into the medium, it may be isolated in a soluble or insoluble form using physical e.g. mechanical or thermal, or chemical treatments. Treatments employed may include freezing ( $\leq 0^\circ \text{C}$ ), heating, hydrodynamic shearing, drying, selective filtration or  
10 precipitation by addition of acid, base, salts or organic solvents.

Where the expressed heterologous polypeptide resides in the membrane or in the cytoplasm, cells are harvested to separate them from the culture medium. Various techniques may be used for harvesting, desirably using centrifugation. The supernatant may then be discarded and the cell pellet  
15 washed with an appropriate buffered aqueous medium to remove any residual culture medium components. Typically the buffered medium will be at a temperature in the range of about 1 to  $10^\circ \text{C}$ , more usually  $4^\circ \text{C}$ .

The cells may be lysed by any convenient means, such as freezing and mechanical, use of hypotonic solutions (23), and the like. The resulting  
20 dispersion of disrupted cells is then treated by such means as to substantially separate cell membranes from soluble proteins and other contaminants. Several techniques may be employed to advantage for isolating membranes including differential centrifugation, density gradient centrifugation, and the like. This membrane isolation separates the fusion protein from the bulk of the soluble  
25 proteins.

Heterologous polypeptides are purified according to procedures dependent on their individual properties and those of BR. Where the expressed soluble heterologous polypeptide is fused at the C-terminal region of BR, advantage may be taken of the likelihood that the BR domain will anchor the fusion protein in  
30 the membrane.



Where the heterologous polypeptide is expressed as a fusion polypeptide linked to the C-terminal region, or fragment thereof, of the bacteriorhodopsin gene with a unique protease site between said heterologous polypeptide and C-terminal region, the heterologous polypeptide may be isolated by incubating the halobacterial membranes with an appropriate unique protease to effect substantially complete cleavage at the protease cleavage site. For example, where the heterologous polypeptide is linked to the bacteriorhodopsin C-terminal region through the amino acid sequence Ile-Glu-Gly-Arg (SEQ ID N0:4), cell membranes are incubated with factor  $X_a$  under conditions recommended by the manufacturers. Factor  $X_a$  is dissolved in redistilled water to a final protein concentration of 1 mg/ml. The fusion protein to be cleaved is dissolved in 100 mM NaCl, 50 mM Tris-HCl, 1 mM  $CaCl_2$ , pH 8.0. To increase the solubility of the substrate, urea or acetonitrile can be added up to a final concentration of 1 M and 10% (v/v), respectively without significant inhibition of the enzyme activity. The recommended amount of enzyme is 1/200 to 1/10 of the substrate by weight. Incubation should be carried out at 4°C to 25°C for 1-18 h. The optimum cleavage conditions have to be determined for each fusion protein. The release of the desired polypeptide from the fusion protein is influenced by the adjacent amino acid sequences at the cleavage site, the size of the two fused polypeptide components, and the accessibility of the cleavage site. Protease treatment is followed by standard purification protocols to remove the minor unique protease component.

If further purification of the heterologous polypeptide protein is desired, antibodies specific for the heterologous polypeptide, ligand affinity, electrophoresis, chromatography, zonal centrifugation, and the like, may be employed to advantage. The product may then be dried by any convenient means, such as freeze drying, spray drying, and the like, or alternatively suspended in an appropriate buffered aqueous solution. The heterologous polypeptide product is then ready for use.

## 30 5. Bioassays

The heterologous polypeptides may be assayed using protocols dependent on their individual properties. For example, receptors are assayed using ligand binding assays. Soluble proteins having enzyme activity are assayed using appropriate substrates.

## 5 Bibliography

For the sake of convenience, various documents referenced in the body of the present specification are grouped in the following bibliography by number that corresponds to the parenthetical number of that reference in the text. Each of these documents is hereby expressly incorporated by reference.

- 10 1. Gropp Syst. Appl. Microbiol. 7, 95 (1986).
2. Zillig Eur. J. Biochem. 173, 473 (1988).
3. Dennis J. Bacteriol. 168, 471 (1986).
4. Oesterhelt Proc. Nat. Acad. Sci. USA 70, 2853 (1971).
5. Henderson Annu. Rev. Biophys. Bioenerg. 6, 87 (1977).
- 15 6. Katre Proc. Natl. Acad. Sci. USA 78, 4068 (1981)
7. Blasieo Proc. natl. Acad. Sci. USA 87, 6772 (1990)
8. Holmes J. Bacteriol. 172, 756 (1990)
9. Ni Gene 90, 169 (1990).
10. Charlebois Proc. Natl. Acad. Sci. USA 84, 8530 (1987)
- 20 11. Cline J. bacteriol. 169, 1341 (1987)
12. Maniatis "Molecular Cloning: A Laboratory Manual". Cold Spring Harbor Laboratory, CSH, N.Y. (1989)
13. Betlach Nucl. Acids Res. 12, 7949 (1984)
14. Leong J. bacteriol. 170, 4903 (1988)
- 25 15. Wagner FEBS Letters 131, 341 (1983)
16. Spudich Proc. Natl. Acad. Sci. USA, 79 4398 (1982)
17. Pfeifer J. Bacteriol. 145, 375 (1981)
18. Shand J. Bacteriol. 173, 4692 (1991)
19. Betlach, In: "Protocols for Archael Research". Robb & Das Sarma (Eds.),  
30 Cold Spring Harbor Laboratory. Cold Spring Harbor, N.Y., in press (1993)

20. Yanisch-Perron Gene 33, 103 (1985)
21. Kamekura Appl. Environmental Microbiol. 54, 990 (1988)
22. Grussenmeyer Proc. Natl. Acad.Sci. USA 82 7952 (1985)
23. Turner Biochemistry 32 1332 (1993)
- 5 24. Cline Can. J. Microbiol., 35 148 (1989)
25. Feinberg Analytical Biochemistry 132 6 (1983)
26. Hoffmann Nucleic Acids Research 19 6337 (1991)
27. Julius Science 241 558 (1988)
28. Kuhn Gene 44 253 (1986)
- 10 29. Kunkel Methods Enzymol. 154 367 (1987)
30. Nowlan J. Biol. Chem. 260 14712 (1985)
31. Power Gene 113 95 (1992)
32. Shand Biochemistry 30 3082 (1991)
33. Vu Cell 64 1057 (1991)

#### 15 Concluding Remarks

The foregoing description details specific methods that can be employed to practice the present invention. Having detailed specific methods initially used to construct and use vectors for the expression, isolation, detection and further purification of heterologous polypeptides in halobacteria, those skilled in the art will know how to devise alternative reliable methods for arriving at the same and equivalent systems described herein. The foregoing should not be construed as limiting the overall scope hereof; rather, the ambit of the present invention is to be governed only by the lawful interpretation of the appended claims.

25 The Halobacterium strains referred to above were deposited with the American Type Culture Collection, located at 12301 Parklawn Drive, Rockville, Maryland 20852-1776. The dates of the deposits were ATCC 29252 - February 3, 1976; ATCC 38799 - September 13, 1979; ATCC 29715 - September 19, 1977 and ATCC 33500 - February 23, 1981.

SEQUENCE LISTING

## (1) GENERAL INFORMATION:

- (i) APPLICANT: TURNER, George J.  
BETLACH, Mary C.
- (ii) TITLE OF INVENTION: EXPRESSION OF HETEROLOGOUS POLYPEPTIDES  
IN HALOBACTERIA
- (iii) NUMBER OF SEQUENCES: 15
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  - (D) STATE: California
  - (E) COUNTRY: USA
  - (F) ZIP: 90012
- (v) COMPUTER READABLE FORM:
  - (A) MEDIUM TYPE: Floppy disk
  - (B) COMPUTER: IBM PC compatible
  - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
  - (D) SOFTWARE: PatentIn Release #1.0, Version #1.25
- (vi) CURRENT APPLICATION DATA:
  - (A) APPLICATION NUMBER: PCT
  - (B) FILING DATE:
  - (C) CLASSIFICATION:
- (viii) ATTORNEY/AGENT INFORMATION:
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  - (B) REGISTRATION NUMBER: 20,121
  - (C) REFERENCE/DOCKET NUMBER: 5555-206-PCT
- (ix) TELECOMMUNICATION INFORMATION:
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  - (B) TELEFAX: (213) 977-1003

## (2) INFORMATION FOR SEQ ID NO:1:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1254 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)

- (ix) FEATURE:
  - (A) NAME/KEY: misc feature
  - (B) LOCATION: 376..414
  - (D) OTHER INFORMATION: /note= "Bacteriorhodopsin pre-sequence."
  
- (ix) FEATURE:
  - (A) NAME/KEY: CDS
  - (B) LOCATION: 376..1161
  
- (ix) FEATURE:
  - (A) NAME/KEY: misc feature
  - (B) LOCATION: 3..8
  - (D) OTHER INFORMATION: /note= "PstI site."
  
- (ix) FEATURE:
  - (A) NAME/KEY: misc feature
  - (B) LOCATION: 1245..1250
  - (D) OTHER INFORMATION: /note= "BamHI site."
  
- (ix) FEATURE:
  - (A) NAME/KEY: misc\_signal
  - (B) LOCATION: 374
  - (D) OTHER INFORMATION: /note= "RNA start site."
  
- (ix) FEATURE:
  - (A) NAME/KEY: misc feature
  - (B) LOCATION: 9..414
  - (D) OTHER INFORMATION: /note= "Bacteriorhodopsin transcriptional and translational regulatory sequences are located in this region."

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

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TTCGAGTGGT AACACGCGTG CACGCATCGA CTTCACCGCG GGTGTTTCGA CGCCAGCCGG	180
CCGTTGAACC AGCAGGCAGC GGGCATTTC AAGCCGCTGT GGCCACACA CTCGGTGGGG	240
TGCGCTATTT TGGTATGGTT TGGAAATCCGC GTGTCGGCTC CGTGTCTGAC GGTTTCATCGG	300
TCTAAATTCC GTCACGAGCG TACCATACTG ATTGGGTCGT AGAGTTACAC ACATATCCTC	360
GTTAGGTACT GTTGC ATG TTG GAG TTA TTG CCA ACA GCA GTG GAG GGG GTA	411
Met Leu Glu Leu Leu Pro Thr Ala Val Glu Gly Val	
1 5 10	

TCG CAG GCC CAG ATC ACC GGA CGT CCG GAG TGG ATC TGG CTA GCG CTC Ser Gln Ala Gln Ile Thr Gly Arg Pro Glu Trp Ile Trp Leu Ala Leu 15 20 25	459
GGT ACG GCG CTA ATG GGA CTC GGG ACG CTC TAT TTC CTC GTG AAA GGG Gly Thr Ala Leu Met Gly Leu Gly Thr Leu Tyr Phe Leu Val Lys Gly 30 35 40	507
ATG GGC GTC TCG GAC CCA GAT GCA AAG AAA TTC TAC GCC ATC ACG ACG Met Gly Val Ser Asp Pro Asp Ala Lys Lys Phe Tyr Ala Ile Thr Thr 45 50 55 60	555
CTC GTC CCA GCC ATC GCG TTC ACG ATG TAC CTC TCG ATG CTG CTG GGG Leu Val Pro Ala Ile Ala Phe Thr Met Tyr Leu Ser Met Leu Leu Gly 65 70 75	603
TAT GGC CTC ACA ATG GTA CCG TTC GGT GGG GAG CAG AAC CCC ATC TAC Tyr Gly Leu Thr Met Val Pro Phe Gly Gly Glu Gln Asn Pro Ile Tyr 80 85 90	651
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GCG ATG CTG TAC ATC CTG TAC GTG CTG TTC TTC GGG TTC ACC TCG AAG Ala Met Leu Tyr Ile Leu Tyr Val Leu Phe Phe Gly Phe Thr Ser Lys 160 165 170	891
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 225 230 235

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 240 245 250

GCC GGC GAC GGC GCG GCC GCG ACC AGC GAC TGATCGCACA CGCAGGACAG 1181  
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 255 260

CCCCACAACC GCGCGGGCTG TGTTCAACGA CACACGATGA GTCCCCCACT CGGTCTTGTA 1241

CTCGGATCCT TTT 1254

(2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 262 amino acids
  - (B) TYPE: amino acid
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Met Leu Glu Leu Leu Pro Thr Ala Val Glu Gly Val Ser Gln Ala Gln  
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Ile Thr Gly Arg Pro Glu Trp Ile Trp Leu Ala Leu Gly Thr Ala Leu  
 20 25 30

Met Gly Leu Gly Thr Leu Tyr Phe Leu Val Lys Gly Met Gly Val Ser  
 35 40 45

Asp Pro Asp Ala Lys Lys Phe Tyr Ala Ile Thr Thr Leu Val Pro Ala  
 50 55 60

Ile Ala Phe Thr Met Tyr Leu Ser Met Leu Leu Gly Tyr Gly Leu Thr  
 65 70 75 80

Met Val Pro Phe Gly Gly Glu Gln Asn Pro Ile Tyr Trp Ala Arg Tyr  
 85 90 95

Ala Asp Trp Leu Phe Thr Thr Pro Leu Leu Leu Asp Leu Ala Leu  
 100 105 110

Leu Val Asp Ala Asp Gln Gly Thr Ile Leu Ala Leu Val Gly Ala Asp  
 115 120 125

Gly Ile Met Ile Gly Thr Gly Leu Val Gly Ala Leu Thr Lys Val Tyr  
 130 135 140  
 Ser Tyr Arg Phe Val Trp Trp Ala Ile Ser Thr Ala Ala Met Leu Tyr  
 145 150 155 160  
 Ile Leu Tyr Val Leu Phe Phe Gly Phe Thr Ser Lys Ala Glu Ser Met  
 165 170 175  
 Arg Pro Glu Val Ala Ser Thr Phe Lys Val Leu Arg Asn Val Thr Val  
 180 185 190  
 Val Leu Trp Ser Ala Tyr Pro Val Val Trp Leu Ile Gly Ser Glu Gly  
 195 200 205  
 Ala Gly Ile Val Pro Leu Asn Ile Glu Thr Leu Leu Phe Met Val Leu  
 210 215 220  
 Asp Val Ser Ala Lys Val Gly Phe Gly Leu Ile Leu Leu Arg Ser Arg  
 225 230 235 240  
 Ala Ile Phe Gly Glu Ala Glu Ala Pro Glu Pro Ser Ala Gly Asp Gly  
 245 250 255  
 Ala Ala Ala Thr Ser Asp  
 260

## (2) INFORMATION FOR SEQ ID NO:3:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 248 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

## (ix) FEATURE:

- (A) NAME/KEY: Region
- (B) LOCATION: 225..248
- (D) OTHER INFORMATION: /note= "Cytoplasmic C-terminal region of bacteriorhodopsin."

## (ix) FEATURE:

- (A) NAME/KEY: Region
- (B) LOCATION: 1
- (D) OTHER INFORMATION: /note= "Pyroglutamate."



## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

Xaa Ala Gln Ile Thr Gly Arg Pro Glu Trp Ile Trp Leu Ala Leu Gly  
 1                               5                               10                               15  
 Thr Ala Leu Met Gly Leu Gly Thr Leu Tyr Phe Leu Val Lys Gly Met  
                               20                               25                               30  
 Gly Val Ser Asp Pro Asp Ala Lys Lys Phe Tyr Ala Ile Thr Thr Leu  
                               35                               40                               45  
 Val Pro Ala Ile Ala Phe Thr Met Tyr Leu Ser Met Leu Leu Gly Tyr  
                               50                               55                               60  
 Gly Leu Thr Met Val Pro Phe Gly Gly Glu Gln Asn Pro Ile Tyr Trp  
 65                               70                               75                               80  
 Ala Arg Tyr Ala Asp Trp Leu Phe Thr Thr Pro Leu Leu Leu Leu Asp  
                               85                               90                               95  
 Leu Ala Leu Leu Val Asp Ala Asp Gln Gly Thr Ile Leu Ala Leu Val  
                               100                               105                               110  
 Gly Ala Asp Gly Ile Met Ile Gly Thr Gly Leu Val Gly Ala Leu Thr  
                               115                               120                               125  
 Lys Val Tyr Ser Tyr Arg Phe Val Trp Trp Ala Ile Ser Thr Ala Ala  
                               130                               135                               140  
 Met Leu Tyr Ile Leu Tyr Val Leu Phe Phe Gly Phe Thr Ser Lys Ala  
 145                               150                               155                               160  
 Glu Ser Met Arg Pro Glu Val Ala Ser Thr Phe Lys Val Leu Arg Asn  
                               165                               170                               175  
 Val Thr Val Val Leu Trp Ser Ala Tyr Pro Val Val Trp Leu Ile Gly  
                               180                               185                               190  
 Ser Glu Gly Ala Gly Ile Val Pro Leu Asn Ile Glu Thr Leu Leu Phe  
                               195                               200                               205  
 Met Val Leu Asp Val Ser Ala Lys Val Gly Phe Gly Leu Ile Leu Leu  
                               210                               215                               220  
 Arg Ser Arg Ala Ile Phe Gly Glu Ala Glu Ala Pro Glu Pro Ser Ala  
 225                               230                               235                               240  
 Gly Asp Gly Ala Ala Ala Thr Ser  
                               245

## (2) INFORMATION FOR SEQ ID NO:4:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 4 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Ile Glu Gly Arg  
1

## (2) INFORMATION FOR SEQ ID NO:5:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 9 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

Glu Glu Glu Glu Tyr Met Pro Met Glu  
1 5

## (2) INFORMATION FOR SEQ ID NO:6:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1956 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

## (ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 376..1812

## (ix) FEATURE:

- (A) NAME/KEY: misc feature
- (B) LOCATION: 376..414
- (D) OTHER INFORMATION: /note= "Bacteriorhodopsin pre-sequence."

- (ix) FEATURE:  
(A) NAME/KEY: terminator  
(B) LOCATION: 1864..1866  
(D) OTHER INFORMATION: /note= "Bacteriorhodopsin stop codon."
- (ix) FEATURE:  
(A) NAME/KEY: mutation  
(B) LOCATION: replace(213, "")  
(D) OTHER INFORMATION: /note= "G to T mutation removes AlwNI restriction site."
- (ix) FEATURE:  
(A) NAME/KEY: misc\_feature  
(B) LOCATION: 427..435  
(D) OTHER INFORMATION: /note= "AlwNI cloning site."
- (ix) FEATURE:  
(A) NAME/KEY: mutation  
(B) LOCATION: replace(930, "")  
(D) OTHER INFORMATION: /note= "G to A mutation removes AlwNI restriction site."
- (ix) FEATURE:  
(A) NAME/KEY: mutation  
(B) LOCATION: replace(1179, "")  
(D) OTHER INFORMATION: /note= "T to A mutation removes AlwNI site."
- (ix) FEATURE:  
(A) NAME/KEY: mutation  
(B) LOCATION: replace(1245, "")  
(D) OTHER INFORMATION: /note= "G to A mutation removes PstI restriction site."
- (ix) FEATURE:  
(A) NAME/KEY: misc\_signal  
(B) LOCATION: 374  
(D) OTHER INFORMATION: /note= "RNA start site."
- (ix) FEATURE:  
(A) NAME/KEY: mutation  
(B) LOCATION: replace(1863, "")  
(D) OTHER INFORMATION: /note= "C to T mutation removes AlwNI restriction site."
- (ix) FEATURE:  
(A) NAME/KEY: terminator  
(B) LOCATION: 1813..1815  
(D) OTHER INFORMATION: /note= "Muscarinic "OM1" stop codon."

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

ATCTGCAGGA TGGGTGCAAC CGTGAAGTCC GTCACGGCTG CGTCACGACA GGAGCCGACC	60
AGCGACACCC AGAAGGTGCG AACGGTTGAG TGCCGCAACG ATCACGAGTT TTTCGTGCGC	120
TTCGAGTGGT AACACGCGTG CACGCATCGA CTTACCGCG GGTGTTTCGA CGCCAGCCGG	180
CCGTTGAACC AGCAGGCAGC GGGCA <del>T</del> TCA CATCCGCTGT GGCCACACA CTCGGTGGGG	240
TGCGCTATTT TGGTATGGTT TGG <del>A</del> TCCGC GTGTCGGCTC CGTGTCTGAC GGTTTCATCGG	300
TCTAAATTCC GTCACGAGCG TACCATACTG ATTGGGTCTG AGAGTTACAC ACATATCCTC	360
GTTAGGTA <del>CT</del> GTTGC ATG TTG GAG TTA TTG CCA ACA GCA GTG GAG GGG GTA	411
Met Leu Glu Leu Leu Pro Thr Ala Val Glu Gly Val	
1 5 10	
TCG CAG GCC CAG ATC CAG GCG CTG ATG AAC ACT TCA GCC CCA CCT GCT	459
Ser Glr Ala Gln Ile Gln Ala Leu Met Asn Thr Ser Ala Pro Pro Ala	
15 20 25	
GTC AGC CCC AAC ATC ACC GTC CTG GCA CCA GGA AAG GGT CCC TGG CAA	507
Val Ser Pro Asn Ile Thr Val Leu Ala Pro Gly Lys Gly Pro Trp Gln	
30 35 40	
GTG GCC TTC ATT GGG ATC ACC ACG GGC CTC CTG TCG CTA GCC ACA GTG	555
Val Ala Phe Ile Gly Ile Thr Thr Gly Leu Leu Ser Leu Ala Thr Val	
45 50 55 60	
ACA GGC AAC CTG CTG GTA CTC ATC TCT TTC AAG GTC AAC ACG GAG CTC	603
Thr Gly Asn Leu Leu Val Leu Ile Ser Phe Lys Val Asn Thr Glu Leu	
65 70 75	
AAG ACA GTC AAT AAC TAC TTC CTG CTG AGC CTG GCC TGT GCT GAC CTC	651
Lys Thr Val Asn Asn Tyr Phe Leu Leu Ser Leu Ala Cys Ala Asp Leu	
80 85 90	
ATC ATC GGT ACC TTC TCC ATG AAC CTC TAT ACC ACG TAC CTG CTC ATG	699
Ile Ile Gly Thr Phe Ser Met Asn Leu Tyr Thr Thr Tyr Leu Leu Met	
95 100 105	
GGC CAC TGG GCT CTG GGC ACG CTG GCT TGT GAC CTC TGG CTG GCC CTG	747
Gly His Trp Ala Leu Gly Thr Leu Ala Cys Asp Leu Trp Leu Ala Leu	
110 115 120	
GAC TAT GTG GCC AGC AAT GCC TCC GTC ATG AAT CTG CTG CTC ATC AGC	795
Asp Tyr Val Ala Ser Asn Ala Ser Val Met Asn Leu Leu Leu Ile Ser	
125 130 135 140	
TTT GAC CGC TAC TTC TCC GTG ACT CGG CCC CTG AGC TAC CGT GCC AAG	843
Phe Asp Arg Tyr Phe Ser Val Thr Arg Pro Leu Ser Tyr Arg Ala Lys	
145 150 155	

CGC	ACA	CCC	CGC	CGC	GCA	GCT	CTG	ATG	ATC	GGC	CTG	GCC	TGG	CTG	GTT	891
Arg	Thr	Pro	Arg	Arg	Ala	Ala	Leu	Met	Ile	Gly	Leu	Ala	Trp	Leu	Val	
			160					165					170			
TCC	TTT	GTG	CTC	TGG	GCC	CCA	GCC	ATC	CTC	TTC	TGG	CAA	TAC	CTG	GTA	939
Ser	Phe	Val	Leu	Trp	Ala	Pro	Ala	Ile	Leu	Phe	Trp	Gln	Tyr	Leu	Val	
		175					180					185				
GGG	GAG	CGG	ACG	ATG	CTA	GCT	GGG	CAG	TGC	TAC	ATC	CAG	TTC	CTC	TCC	987
Gly	Glu	Arg	Thr	Met	Leu	Ala	Gly	Gln	Cys	Tyr	Ile	Gln	Phe	Leu	Ser	
	190					195					200					
CAG	CCC	ATC	ATC	ACC	TTT	GGC	ACA	GCC	ATG	GCT	GCC	TTC	TAC	CTC	CCT	1035
Gln	Pro	Ile	Ile	Thr	Phe	Gly	Thr	Ala	Met	Ala	Ala	Phe	Tyr	Leu	Pro	
205					210					215					220	
GTC	ACA	GTC	ATG	TGC	ACG	CTC	TAC	TGG	CGC	ATC	TAC	CGG	GAG	ACA	GAG	1083
Val	Thr	Val	Met	Cys	Thr	Leu	Tyr	Trp	Arg	Ile	Tyr	Arg	Glu	Thr	Glu	
				225					230					235		
AAC	CGA	GCA	CGG	GAG	CTG	GCA	GCC	CTT	CAG	GGC	TCC	GAG	ACG	CCA	GGC	1131
Asn	Arg	Ala	Arg	Glu	Leu	Ala	Ala	Leu	Gln	Gly	Ser	Glu	Thr	Pro	Gly	
			240					245						250		
AAA	GGG	GGT	GGC	AGC	AGC	AGC	AGC	TCA	GAG	AGG	TCT	CAG	CCA	GGG	GCA	1179
Lys	Gly	Gly	Gly	Ser	Ser	Ser	Ser	Ser	Glu	Arg	Ser	Gln	Pro	Gly	Ala	
		255					260					265				
GAG	GGC	TCA	CCA	GAG	ACT	CCT	CCA	GGC	CGC	TGC	TGT	CGC	TGC	TGC	CGG	1227
Glu	Gly	Ser	Pro	Glu	Thr	Pro	Pro	Gly	Arg	Cys	Cys	Arg	Cys	Cys	Arg	
	270					275					280					
GCC	CCA	AGG	CTG	CTG	CAA	GCC	TAC	AGC	TGG	AAG	GAA	GAA	GAG	GAA	GAG	1275
Ala	Pro	Arg	Leu	Leu	Gln	Ala	Tyr	Ser	Trp	Lys	Glu	Glu	Glu	Glu	Glu	
285					290					295					300	
GAC	GAA	GGC	TCC	ATG	GAG	TCC	CTC	ACA	TCC	TCA	GAG	GGA	GAG	GAG	CCT	1323
Asp	Glu	Gly	Ser	Met	Glu	Ser	Leu	Thr	Ser	Ser	Glu	Gly	Glu	Glu	Pro	
				305					310					315		
GGC	TCC	GAA	GTG	GTG	ATC	AAG	ATG	CCA	ATG	GTG	GAC	CCC	GAG	GCA	CAG	1371
Gly	Ser	Glu	Val	Val	Ile	Lys	Met	Pro	Met	Val	Asp	Pro	Glu	Ala	Gln	
			320					325					330			
GCC	CCC	ACC	AAG	CAG	CCC	CCA	CGG	AGC	TCC	CCA	AAT	ACA	GTC	AAG	AGG	1419
Ala	Pro	Thr	Lys	Gln	Pro	Pro	Arg	Ser	Ser	Pro	Asn	Thr	Val	Lys	Arg	
		335					340					345				
CCG	ACT	AAG	AAA	GGG	CGT	GAT	CGA	GCT	GGC	AAG	GGC	CAG	AAG	CCC	CGT	1467
Pro	Thr	Lys	Lys	Gly	Arg	Asp	Arg	Ala	Gly	Lys	Gly	Gln	Lys	Pro	Arg	
	350					355					360					

GGA AAG GAG CAG CTG GCC AAG CGG AAG ACC TTC TCG CTG GTC AAC GAG Gly Lys Glu Gln Leu Ala Lys Arg Lys Thr Phe Ser Leu Val Lys Glu 365 370 375 380	1515
AAG AAG GCG GCT CGG ACC CTG AGT GCC ATC CTC CTG GCC TTC ATC CTC Lys Lys Ala Ala Arg Thr Leu Ser Ala Ile Leu Leu Ala Phe Ile Leu 385 390 395	1563
ACC TGG ACA CCG TAC AAC ATC ATG GTG CTG GTG TCC ACC TTC TGC AAG Thr Trp Thr Pro Tyr Asn Ile Met Val Leu Val Ser Thr Phe Cys Lys 400 405 410	1611
GAC TGT GTT CCC GAG ACC CTG TGG GAG CTG GGC TAC TGG CTG TGC TAC Asp Cys Val Pro Glu Thr Leu Trp Glu Leu Gly Tyr Trp Leu Cys Tyr 415 420 425	1659
GTC AAC AGC ACC ATC AAC CCC ATG TGC TAC GCA CTC TGC AAC AAA GCC Val Asn Ser Thr Ile Asn Pro Met Cys Tyr Ala Leu Cys Asn Lys Ala 430 435 440	1707
TTC CGG GAC ACC TTT CGC CTG CTG CTT TGC CGC TGG GAC AAG AGA CGC Phe Arg Asp Thr Phe Arg Leu Leu Leu Cys Arg Trp Asp Lys Arg Arg 445 450 455 460	1755
TGG CGC AAG ATC CCC AAG CGC CCT GGC TCC CAC CGC ACT CCC TCC Trp Arg Lys Ile Pro Lys Arg Pro Gly Ser Val His Arg Thr Pro Ser 465 470 475	1803
CGC CAA TGC TGATAGTCCC CTCTCCTGCA TCCCTCCACC CCAGCGGCCG Arg Gln Cys	1852
CGACCAGCGA TTGATCGCAC ACGCAGGACA GCCCACAAC CGGCGCGGCT GTGTTCAACG	1912
ACACACGATG AGTCCCCAC TCGGTCTTGT ACTCGGATCC TTTT	1956

(2) INFORMATION FOR SEQ ID NO:7:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 479 amino acids
  - (B) TYPE: amino acid
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

Met Leu Glu Leu Leu Pro Thr Ala Val Glu Gly Val Ser Gln Ala Gln  
1 5 10 15  
Ile Gln Ala Leu Met Asn Thr Ser Ala Pro Pro Ala Val Ser Pro Asn  
20 25 30

Ile Thr Val Leu Ala Pro Gly Lys Gly Pro Trp Gln Val Ala Phe Ile  
 35 40 45

Gly Ile Thr Thr Gly Leu Leu Ser Leu Ala Thr Val Thr Gly Asn Leu  
 50 55 60

Leu Val Leu Ile Ser Phe Lys Val Asn Thr Glu Leu Lys Thr Val Asn  
 65 70 75 80

Asn Tyr Phe Leu Leu Ser Leu Ala Cys Ala Asp Leu Ile Ile Gly Thr  
 85 90 95

Phe Ser Met Asn Leu Tyr Thr Thr Tyr Leu Leu Met Gly His Trp Ala  
 100 105 110

Leu Gly Thr Leu Ala Cys Asp Leu Trp Leu Ala Leu Asp Tyr Val Ala  
 115 120 125

Ser Asn Ala Ser Val Met Asn Leu Leu Leu Ile Ser Phe Asp Arg Tyr  
 130 135 140

Phe Ser Val Thr Arg Pro Leu Ser Tyr Arg Ala Lys Arg Thr Pro Arg  
 145 150 155 160

Arg Ala Ala Leu Met Ile Gly Leu Ala Trp Leu Val Ser Phe Val Leu  
 165 170 175

Trp Ala Pro Ala Ile Leu Phe Trp Gln Tyr Leu Val Gly Glu Arg Thr  
 180 185 190

Met Leu Ala Gly Gln Cys Tyr Ile Gln Phe Leu Ser Gln Pro Ile Ile  
 195 200 205

Thr Phe Gly Thr Ala Met Ala Ala Phe Tyr Leu Pro Val Thr Val Met  
 210 215 220

Cys Thr Leu Tyr Trp Arg Ile Tyr Arg Glu Thr Glu Asn Arg Ala Arg  
 225 230 235 240

Glu Leu Ala Ala Leu Gln Gly Ser Glu Thr Pro Gly Lys Gly Gly Gly  
 245 250 255

Ser Ser Ser Ser Ser Glu Arg Ser Gln Pro Gly Ala Glu Gly Ser Pro  
 260 265 270

Glu Thr Pro Pro Gly Arg Cys Cys Arg Cys Cys Arg Ala Pro Arg Leu  
 275 280 285

Leu Gln Ala Tyr Ser Trp Lys Glu Glu Glu Glu Glu Asp Glu Gly Ser  
 290 295 300

Met Glu Ser Leu Thr Ser Ser Glu Gly Glu Glu Pro Gly Ser Glu Val  
 305 310 315 320

Val Ile Lys Met Pro Met Val Asp Pro Glu Ala Gln Ala Pro Thr Lys  
 325 330 335  
 Gln Pro Pro Arg Ser Ser Pro Asn Thr Val Lys Arg Pro Thr Lys Lys  
 340 345 350  
 Gly Arg Asp Arg Ala Gly Lys Gly Gln Lys Pro Arg Gly Lys Glu Gln  
 355 360 365  
 Leu Ala Lys Arg Lys Thr Phe Ser Leu Val Lys Glu Lys Lys Ala Ala  
 370 375 380  
 Arg Thr Leu Ser Ala Ile Leu Leu Ala Phe Ile Leu Thr Trp Thr Pro  
 385 390 395 400  
 Tyr Asn Ile Met Val Leu Val Ser Thr Phe Cys Lys Asp Cys Val Pro  
 405 410 415  
 Glu Thr Leu Trp Glu Leu Gly Tyr Trp Leu Cys Tyr Val Asn Ser Thr  
 420 425 430  
 Ile Asn Pro Met Cys Tyr Ala Leu Cys Asn Lys Ala Phe Arg Asp Thr  
 435 440 445  
 Phe Arg Leu Leu Leu Cys Arg Trp Asp Lys Arg Arg Trp Arg Lys Ile  
 450 455 460  
 Pro Lys Arg Pro Gly Ser Val His Arg Thr Pro Ser Arg Gln Cys  
 465 470 475

(2) INFORMATION FOR SEQ ID NO:8:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1581 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

- (ix) FEATURE:
  - (A) NAME/KEY: CDS
  - (B) LOCATION: 376..1437

- (ix) FEATURE:
  - (A) NAME/KEY: misc\_feature
  - (B) LOCATION: 376..414
  - (D) OTHER INFORMATION: /note= "Bacteriorhodopsin pre-sequence."



- (ix) FEATURE:  
 (A) NAME/KEY: terminator  
 (B) LOCATION: 1489..1491  
 (D) OTHER INFORMATION: /note= "Bacteriorhodopsin stop codon."
- (ix) FEATURE:  
 (A) NAME/KEY: mutation  
 (B) LOCATION: replace(213, "")  
 (D) OTHER INFORMATION: /note= "G to T mutation removes AlwNI restriction site."
- (ix) FEATURE:  
 (A) NAME/KEY: misc feature  
 (B) LOCATION: 427..435  
 (D) OTHER INFORMATION: /note= "AlwNI cloning site."
- (ix) FEATURE:  
 (A) NAME/KEY: mutation  
 (B) LOCATION: replace(930, "")  
 (D) OTHER INFORMATION: /note= "G to A mutation removes AlwNI site."
- (ix) FEATURE:  
 (A) NAME/KEY: misc\_signal  
 (B) LOCATION: 374  
 (D) OTHER INFORMATION: /note= "RNA start site."
- (ix) FEATURE:  
 (A) NAME/KEY: terminator  
 (B) LOCATION: 1438..1440  
 (D) OTHER INFORMATION: /note= "Muscarinic stop codon."
- (ix) FEATURE:  
 (A) NAME/KEY: mutation  
 (B) LOCATION: replace(1488, "")  
 (D) OTHER INFORMATION: /note= "C to T mutation removes AlwNI restriction site."

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

ATCTGCAGGA TGGGTGCAAC CGTGAAGTCC GTCACGGCTG CGTCACGACA GGAGCCGACC	60
AGCGACACCC AGAAGGTGCG AACGGTTGAG TGCCGCAACG ATCACGAGTT TTTCGTGCGC	120
TTCGAGTGGT AACACGCGTG CACGCATCGA CTTACCGCG GGTGTTTCGA CGCCAGCCGG	180
CCGTTGAACC AGCAGGCAGC GGGCATTTC AATCCGCTGT GGCCACACA CTCGGTGGGG	240
TGCGCTATTT TGGTATGGTT TGAATCCGC GTGTCGGCTC CGTGTCTGAC GGTTTCATCGG	300
TCTAAATTCC GTCACGAGCG TACCATACTG ATTGGGTCGT AGAGTTACAC ACATATCCTC	360

GTTAGGTACT GTTGC ATG TTG GAG TTA TTG CCA ACA GCA GTG GAG GGG GTA Met Leu Glu Leu Leu Pro Thr Ala Val Glu Gly Val 1 5 10	411
TCG CAG GCC CAG ATC CAG GCG CTG ATG AAC ACT TCA GCC CCA CCT GCT Ser Gln Ala Gln Ile Gln Ala Leu Met Asn Thr Ser Ala Pro Pro Ala 15 20 25	459
GTC AGC CCC AAC ATC ACC GTC CTG GCA CCA GGA AAG GGT CCC TGG CAA Val Ser Pro Asn Ile Thr Val Leu Ala Pro Gly Lys Gly Pro Trp Gln 30 35 40	507
GTG GCC TTC ATT GGG ATC ACC ACG GGC CTC CTG TCG CTA GCC ACA GTG Val Ala Phe Ile Gly Ile Thr Thr Gly Leu Leu Ser Leu Ala Thr Val 45 50 55 60	555
ACA GGC AAC CTG CTG GTA CTC ATC TCT TTC AAG GTC AAC ACG GAG CTC Thr Gly Asn Leu Leu Val Leu Ile Ser Phe Lys Val Asn Thr Glu Leu 65 70 75	603
AAG ACA GTC AAT AAC TAC TTC CTG CTG AGC CTG GCC TGT GCT GAC CTC Lys Thr Val Asn Asn Tyr Phe Leu Leu Ser Leu Ala Cys Ala Asp Leu 80 85 90	651
ATC ATC GGT ACC TTC TCC ATG AAC CTC TAT ACC ACG TAC CTG CTC ATG Ile Ile Gly Thr Phe Ser Met Asn Leu Tyr Thr Thr Tyr Leu Leu Met 95 100 105	699
GGC CAC TGG GCT CTG GGC ACG CTG GCT TGT GAC CTC TGG CTG GCC CTG Gly His Trp Ala Leu Gly Thr Leu Ala Cys Asp Leu Trp Leu Ala Leu 110 115 120	747
GAC TAT GTG GCC AGC AAT GCC TCC GTC ATG AAT CTG CTG CTC ATC AGC Asp Tyr Val Ala Ser Asn Ala Ser Val Met Asn Leu Leu Leu Ile Ser 125 130 135 140	795
TTT GAC CGC TAC TTC TCC GTG ACT CGG CCC CTG AGC TAC CGT GCC AAG Phe Asp Arg Tyr Phe Ser Val Thr Arg Pro Leu Ser Tyr Arg Ala Lys 145 150 155	843
CGC ACA CCC CGC CGC GCA GCT CTG ATG ATC GGC CTG GCC TGG CTG GTT Arg Thr Pro Arg Arg Ala Ala Leu Met Ile Gly Leu Ala Trp Leu Val 160 165 170	891
TCC TTT GTG CTC TGG GCC CCA GCC ATC CTC TTC TGG CAA TAC CTG GTA Ser Phe Val Leu Trp Ala Pro Ala Ile Leu Phe Trp Gln Tyr Leu Val 175 180 185	939
GGG GAG CGG ACG ATG CTA GCT GGG CAG TGC TAC ATC CAG TTC CTC TCC Gly Glu Arg Thr Met Leu Ala Gly Gln Cys Tyr Ile Gln Phe Leu Ser 190 195 200	987

CAG CCC ATC ATC ACC TTT GGC ACA GCC ATG GCT GCC TTC TAC CTC CCT Gln Pro Ile Ile Thr Phe Gly Thr Ala Met Ala Ala Phe Tyr Leu Pro 205 210 215 220	1035
GTC ACA GTC ATG TGC ACG CTC TAC TGG CGC ATC TAC CGG GAG ACA GAG Val Thr Val Met Cys Thr Leu Tyr Trp Arg Ile Tyr Arg Glu Thr Glu 225 230 235	1083
AAC CGA GCA CGG GAG CTG GCA GCC CTT CAG GGC TCC GAG ACG CCA GGC Asn Arg Ala Arg Glu Leu Ala Ala Leu Gln Gly Ser Glu Thr Pro Gly 240 245 250	1131
AAA AAG GAG AAG AAG GCG GCT CGG ACC CTG AGT GCC ATC CTC CTG GCC Lys Lys Glu Lys Lys Ala Ala Arg Thr Leu Ser Ala Ile Leu Leu Ala 255 260 265	1179
TTC ATC CTC ACC TGG ACA CCG TAC AAC ATC ATG GTG CTG GTG TCC ACC Phe Ile Leu Thr Trp Thr Pro Tyr Asn Ile Met Val Leu Val Ser Thr 270 275 280	1227
TTC TGC AAG GAC TGT GTT CCC GAG ACC CTG TGG GAG CTG GGC TAC TGG Phe Cys Lys Asp Cys Val Pro Glu Thr Leu Trp Glu Leu Gly Tyr Trp 285 290 295 300	1275
CTG TGC TAC GTC AAC AGC ACC ATC AAC CCC ATG TGC TAC GCA CTC TGC Leu Cys Tyr Val Asn Ser Thr Ile Asn Pro Met Cys Tyr Ala Leu Cys 305 310 315	1323
AAC AAA GCC TTC CGG GAC ACC TTT CGC CTG CTG CTT TGC CGC TGG GAC Asn Lys Ala Phe Arg Asp Thr Phe Arg Leu Leu Leu Cys Arg Trp Asp 320 325 330	1371
AAG AGA CGC TGG CGC AAG ATC CCC AAG CGC CCT GGC TCC GTG CAC CGC Lys Arg Arg Trp Arg Lys Ile Pro Lys Arg Pro Gly Ser Val His Arg 335 340 345	1419
ACT CCC TCC CGC CAA TGC TGATAGTCCC CTCTCCTGCA TCCCTCCACC Thr Pro Ser Arg Gln Cys 350	1467
CCAGCGGCCG CGACCAGCGA TTGATCGCAC ACGCAGGACA GCCCCACAAC CGGCGCGGCT	1527
GTGTTCAACG ACACACGATG AGTCCCCAC TCGGTCTTGT ACTCGGATCC TTTT	1581

## (2) INFORMATION FOR SEQ ID NO:9:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 354 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

Met Leu Glu Leu Leu Pro Thr Ala Val Glu Gly Val Ser Gln Ala Gln  
 1 5 10 15  
 Ile Gln Ala Leu Met Asn Thr Ser Ala Pro Pro Ala Val Ser Pro Asn  
 20 25 30  
 Ile Thr Val Leu Ala Pro Gly Lys Gly Pro Trp Gln Val Ala Phe Ile  
 35 40 45  
 Gly Ile Thr Thr Gly Leu Leu Ser Leu Ala Thr Val Thr Gly Asn Leu  
 50 55 60  
 Leu Val Leu Ile Ser Phe Lys Val Asn Thr Glu Leu Lys Thr Val Asn  
 65 70 75 80  
 Asn Tyr Phe Leu Leu Ser Leu Ala Cys Ala Asp Leu Ile Ile Gly Thr  
 85 90 95  
 Phe Ser Met Asn Leu Tyr Thr Thr Tyr Leu Leu Met Gly His Trp Ala  
 100 105 110  
 Leu Gly Thr Leu Ala Cys Asp Leu Trp Leu Ala Leu Asp Tyr Val Ala  
 115 120 125  
 Ser Asn Ala Ser Val Met Asn Leu Leu Leu Ile Ser Phe Asp Arg Tyr  
 130 135 140  
 Phe Ser Val Thr Arg Pro Leu Ser Tyr Arg Ala Lys Arg Thr Pro Arg  
 145 150 155 160  
 Arg Ala Ala Leu Met Ile Gly Leu Ala Trp Leu Val Ser Phe Val Leu  
 165 170 175  
 Trp Ala Pro Ala Ile Leu Phe Trp Gln Tyr Leu Val Gly Glu Arg Thr  
 180 185 190  
 Met Leu Ala Gly Gln Cys Tyr Ile Gln Phe Leu Ser Gln Pro Ile Ile  
 195 200 205  
 Thr Phe Gly Thr Ala Met Ala Ala Phe Tyr Leu Pro Val Thr Val Met  
 210 215 220  
 Cys Thr Leu Tyr Trp Arg Ile Tyr Arg Glu Thr Glu Asn Arg Ala Arg  
 225 230 235 240  
 Glu Leu Ala Ala Leu Gln Gly Ser Glu Thr Pro Gly Lys Lys Glu Lys  
 245 250 255  
 Lys Ala Ala Arg Thr Leu Ser Ala Ile Leu Leu Ala Phe Ile Leu Thr  
 260 265 270

Trp Thr Pro Tyr Asn Ile Met Val Leu Val Ser Thr Phe Cys Lys Asp  
 275 280 285

Cys Val Pro Glu Thr Leu Trp Glu Leu Gly Tyr Trp Leu Cys Tyr Val  
 290 300

Asn Ser Thr Ile Asn Pro Met Cys Tyr Ala Leu Cys Asn Lys Ala Phe  
 305 310 315 320

Arg Asp Thr Phe Arg Leu Leu Leu Cys Arg Trp Asp Lys Arg Arg Trp  
 325 330 335

Arg Lys Ile Pro Lys Arg Pro Gly Ser Val His Arg Thr Pro Ser Arg  
 340 345 350

Gln Cys

## (2) INFORMATION FOR SEQ ID NO:10:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1848 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: cDNA

## (ix) FEATURE:

- (A) NAME/KEY: misc feature
- (B) LOCATION: 376..414
- (D) OTHER INFORMATION: /note= "Bacteriorhodopsin pre-sequence."

## (ix) FEATURE:

- (A) NAME/KEY: terminator
- (B) LOCATION: 1756..1758
- (D) OTHER INFORMATION: /note= "Bacteriorhodopsin stop codon."

## (ix) FEATURE:

- (A) NAME/KEY: misc feature
- (B) LOCATION: 517..591
- (D) OTHER INFORMATION: /note= "Helix I of rat serotonin receptor protein (Type 1C)."

## (ix) FEATURE:

- (A) NAME/KEY: misc feature
- (B) LOCATION: 625..690
- (D) OTHER INFORMATION: /note= "Helix II of rat serotonin receptor protein (Type 1C)."

- (ix) FEATURE:  
(A) NAME/KEY: misc feature  
(B) LOCATION: 736..807  
(D) OTHER INFORMATION: /note= "Helix III of rat serotonin receptor protein (Type 1C)."
- (ix) FEATURE:  
(A) NAME/KEY: misc feature  
(B) LOCATION: 868..939  
(D) OTHER INFORMATION: /note= "Helix IV of rat serotonin receptor protein (Type 1C)."
- (ix) FEATURE:  
(A) NAME/KEY: misc feature  
(B) LOCATION: 997..1059  
(D) OTHER INFORMATION: /note= "Helix V of rat serotonin receptor protein (Type 1C)."
- (ix) FEATURE:  
(A) NAME/KEY: misc feature  
(B) LOCATION: 1297..1362  
(D) OTHER INFORMATION: /note= "Helix VI of rat serotonin receptor protein (Type 1C)."
- (ix) FEATURE:  
(A) NAME/KEY: misc feature  
(B) LOCATION: 1411..1476  
(D) OTHER INFORMATION: /note= "Helix VII of rat serotonin receptor protein (Type 1C)."
- (ix) FEATURE:  
(A) NAME/KEY: mutation  
(B) LOCATION: replace(213, "")  
(D) OTHER INFORMATION: /note= "G to A mutation removes AlwNI restriction site."
- (ix) FEATURE:  
(A) NAME/KEY: misc feature  
(B) LOCATION: 1732..1734  
(D) OTHER INFORMATION: /note= "Codon encoding the C-terminal amino acid of the rat serotonin receptor protein (Type 1C)."
- (ix) FEATURE:  
(A) NAME/KEY: misc\_signal  
(B) LOCATION: 374  
(D) OTHER INFORMATION: /note= "RNA start site."
- (ix) FEATURE:  
(A) NAME/KEY: CDS  
(B) LOCATION: 376..1734

(ix) FEATURE:  
 (A) NAME/KEY: terminator  
 (B) LOCATION: 1735..1737  
 (D) OTHER INFORMATION: /note= "Serotonin stop codon."

(ix) FEATURE:  
 (A) NAME/KEY: repeat region  
 (B) LOCATION: 436..462  
 (D) OTHER INFORMATION: /note= "Sequence encoding polyaspartic acid."

(ix) FEATURE:  
 (A) NAME/KEY: mutation  
 (B) LOCATION: replace(1755, "")  
 (D) OTHER INFORMATION: /note= "C to T mutation removes AlwNI restriction site."

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

ATCTGCAGGA TGGGTGCAAC CGTGAAGTCC GTCACGGCTG CGTCACGACA GGAGCCGACC	60
AGCGACACCC AGAAGGTGCG AACGGTTTAG TGCCGCAACG ATCAGGAGTT TTTCGTGCGC	120
TTCGAGTGGT AACACGCGTG CACGCATCGA CTTACCAGCG GGTGTTTCGA CGCCAGCCGG	180
CCGTTGAACC AGCAGGCAGC GGGCATTTCAT CATCCGCTGT GGCCACACA CTCGGTGGGG	240
TGCGCTATTT TGGTATGGTT TGGAATCCGC GTGTCGGCTC CGTGTCTGAC GGTTTCATCGG	300
TCTAAATTCC GTCACGAGCG TACCATACTG ATTGGGTCGT AGAGTTACAC ACATATCCTC	360
GTTAGGTACT GTTGC ATG TTG GAG TTA TTG CCA ACA GCA GTG GAG GGG GTA	411
Met Leu Glu Leu Leu Pro Thr Ala Val Glu Gly Val	
1 5 10	
TCG CAG GCC CAG ATC CAG GCG CTG GAC TAC AAG GAC GAT GAT GAC GTC	459
Ser Gln Ala Gln Ile Gln Ala Leu Asp Tyr Lys Asp Asp Asp Asp Val	
15 20 25	
GAC ACT TTT AAT TCC TCC GAT GGT GGA CGC TTG TTT CAA TTC CCG GAC	507
Asp Thr Phe Asn Ser Ser Asp Gly Gly Arg Leu Phe Gln Phe Pro Asp	
30 35 40	
GGG GTA CAA AAC TGG CCA GCA CTT TCA ATC GTC GTG ATT ATA ATC ATG	555
Gly Val Gln Asn Trp Pro Ala Leu Ser Ile Val Val Ile Ile Ile Met	
45 50 55 60	
ACA ATA GGG GGC AAC ATT CTT GTT ATC ATG GCA GTA AGC ATG GAG AAG	603
Thr Ile Gly Gly Asn Ile Leu Val Ile Met Ala Val Ser Met Glu Lys	
65 70 75	

AAA CTG CAC AAT GCA ACC AAT TAC TTC TTA ATG TCC CTA GCC ATT GCT Lys Leu His Asn Ala Thr Asn Tyr Phe Leu Met Ser Leu Ala Ile Ala 80 85 90	651
GAT ATG CTG GTG GGA CTA CTT GTC ATG CCC CTG TCC CTG CTT GCT ATT Asp Met Leu Val Gly Leu Leu Val Met Pro Leu Ser Leu Leu Ala Ile 95 100 105	699
CTT TAT GAT TAT GTC TGG CCT TTA CCT AGA TAT TTG TGC CCC GTC TGG Leu Tyr Asp Tyr Val Trp Pro Leu Pro Arg Tyr Leu Cys Pro Val Trp 110 115 120	747
ATT TCA CTA GAT GTG CTA TTT TCA ACT GCG TCC ATC ATG CAC CTC TGC Ile Ser Leu Asp Val Leu Phe Ser Thr Ala Ser Ile Met His Leu Cys 125 130 135 140	795
GCC ATA TCG CTG GAC CGG TAT GTA GCA ATA CGT AAT CCT ATT GAG CAT Ala Ile Ser Leu Asp Arg Tyr Val Ala Ile Arg Asn Pro Ile Glu His 145 150 155	843
AGC CGG TTC AAT TCG CGG ACT AAG GCC ATC ATG AAG ATT GCC ATC GTT Ser Arg Phe Asn Ser Arg Thr Lys Ala Ile Met Lys Ile Ala Ile Val 160 165 170	891
TGG GCA ATA TCA ATA GGA GTT TCA GTT CCT ATC CCT GTG ATT GGA CTG Trp Ala Ile Ser Ile Gly Val Ser Val Pro Ile Pro Val Ile Gly Leu 175 180 185	939
AGG GAC GAA AGC AAA GTG TTC GTG AAT AAC ACC ACG TGC GTG CTC AAT Arg Asp Glu Ser Lys Val Phe Val Asn Asn Thr Thr Cys Val Leu Asn 190 195 200	987
GAC CCC AAC TTC GTT CTC ATC GGG TCC TTC GTG GCA TTC TTC ATC CCG Asp Pro Asn Phe Val Leu Ile Gly Ser Phe Val Ala Phe Phe Ile Pro 205 210 215 220	1035
TTG ACG ATT ATG GTG ATC ACC TAC TTC TTA ACG ATC TAC GTC CTG CGC Leu Thr Ile Met Val Ile Thr Tyr Phe Leu Thr Ile Tyr Val Leu Arg 225 230 235	1083
CGT CAA ACT CTG ATG TTA CTT CGA GGT CAC ACC GAG GAG GAA CTG GCT Arg Gln Thr Leu Met Leu Leu Arg Gly His Thr Glu Glu Glu Leu Ala 240 245 250	1131
AAT ATG AGC CTG AAC TTT CTG AAC TGC TGC TGC AAG AAG AAT GGT GGT Asn Met Ser Leu Asn Phe Leu Asn Cys Cys Cys Lys Lys Asn Gly Gly 255 260 265	1179
GAG GAA GAG AAC GCT CCG AAC CCT AAT CCA GAT CAG AAA CCA CGT CGA Glu Glu Glu Asn Ala Pro Asn Pro Asn Pro Asp Gln Lys Pro Arg Arg 270 275 280	1227



AAG AAG AAA GAA AAG CGT CCC AGA GGC ACC ATG CAA GCT ATC AAC AAC Lys Lys Lys Glu Lys Arg Pro Arg Gly Thr Met Gln Ala Ile Asn Asn 285 290 295 300	1275
GAA AAG AAA GCT TCC AAA GTC CTT GGC ATT GTA TTC TTT GTG TTT CTG Glu Lys Lys Ala Ser Lys Val Leu Gly Ile Val Phe Phe Val Phe Leu 305 310 315	1323
ATC ATG TGG TGC CCG TTT TTC ATC ACC AAT ATC CTG TCG GTT CTT TGT Ile Met Trp Cys Pro Phe Phe Ile Thr Asn Ile Leu Ser Val Leu Cys 320 325 330	1371
GGG AAG GCC TGT AAC CAA AAG CTA ATG GAG AAG CTT CTC AAT GTG TTT Gly Lys Ala Cys Asn Gln Lys Leu Met Glu Lys Leu Leu Asn Val Phe 335 340 345	1419
GTG TGG ATT GGC TAT GTG TGT TCA GGC ATC AAT CCT CTG GTG TAC ACT Val Trp Ile Gly Tyr Val Cys Ser Gly Ile Asn Pro Leu Val Tyr Thr 350 355 360	1467
CTC TTT AAT AAA ATT TAC CGA AGG GCT TTC TCT AAA TAT TTG CGC TGC Leu Phe Asn Lys Ile Tyr Arg Arg Ala Phe Ser Lys Tyr Leu Arg Cys 365 370 375 380	1515
GAT TAT AAG CCA GAC AAA AAG CCT CCT GTT CGA CAG ATT CCT AGG GTT Asp Tyr Lys Pro Asp Lys Lys Pro Pro Val Arg Gln Ile Pro Arg Val 385 390 395	1563
GCT GCC ACT GCT TTG TCT GGG AGG GAG CTC AAT GTT AAC ATT TAT CGG Ala Ala Thr Ala Leu Ser Gly Arg Glu Leu Asn Val Asn Ile Tyr Arg 400 405 410	1611
CAT ACC AAT GAA CGT GTG GCT AGG AAA GCT AAT GAC CCT GAG CCT GGC His Thr Asn Glu Arg Val Ala Arg Lys Ala Asn Asp Pro Glu Pro Gly 415 420 425	1659
ATA GAG ATG CAG GTG GAG AAC TTA GAG CTG CCA GTC AAC CCC TCT AAT Ile Glu Met Gln Val Glu Asn Leu Glu Leu Pro Val Asn Pro Ser Asn 430 435 440	1707
GTG GTC AGC GAG AGG ATT AGT AGT GTG TGAGCGGCCG CGACCAGCGA Val Val Ser Glu Arg Ile Ser Ser Val 445 450	1754
TTGATCGCAC ACGCAGGACA GCCCCACAAC CGGCGCGGCT GTGTTCAACG ACACACGATG	1814
AGTCCCCCAC TCGGTCTTGT ACTCGGATCC TTTT	1848

## (2) INFORMATION FOR SEQ ID NO:11:

- (i) SEQUENCE CHARACTERISTICS:  
 (A) LENGTH: 453 amino acids  
 (B) TYPE: amino acid  
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

Met Leu Glu Leu Leu Pro Thr Ala Val Glu Gly Val Ser Gln Ala Gln  
 1 5 10 15  
 Ile Gln Ala Leu Asp Tyr Lys Asp Asp Asp Val Asp Thr Phe Asn  
 20 25 30  
 Ser Ser Asp Gly Gly Arg Leu Phe Gln Phe Pro Asp Gly Val Gln Asn  
 35 40 45  
 Trp Pro Ala Leu Ser Ile Val Val Ile Ile Ile Met Thr Ile Gly Gly  
 50 55 60  
 Asn Ile Leu Val Ile Met Ala Val Ser Met Glu Lys Lys Leu His Asn  
 65 70 75 80  
 Ala Thr Asn Tyr Phe Leu Met Ser Leu Ala Ile Ala Asp Met Leu Val  
 85 90 95  
 Gly Leu Leu Val Met Pro Leu Ser Leu Leu Ala Ile Leu Tyr Asp Tyr  
 100 105 110  
 Val Trp Pro Leu Pro Arg Tyr Leu Cys Pro Val Trp Ile Ser Leu Asp  
 115 120 125  
 Val Leu Phe Ser Thr Ala Ser Ile Met His Leu Cys Ala Ile Ser Leu  
 130 135 140  
 Asp Arg Tyr Val Ala Ile Arg Asn Pro Ile Glu His Ser Arg Phe Asn  
 145 150 155 160  
 Ser Arg Thr Lys Ala Ile Met Lys Ile Ala Ile Val Trp Ala Ile Ser  
 165 170 175  
 Ile Gly Val Ser Val Pro Ile Pro Val Ile Gly Leu Arg Asp Glu Ser  
 180 185 190  
 Lys Val Phe Val Asn Asn Thr Thr Cys Val Leu Asn Asp Pro Asn Phe  
 195 200 205  
 Val Leu Ile Gly Ser Phe Val Ala Phe Phe Ile Pro Leu Thr Ile Met  
 210 215 220

Val Ile Thr Tyr Phe Leu Thr Ile Tyr Val Leu Arg Arg Gln Thr Leu  
 225 230 235 240  
 Met Leu Leu Arg Gly His Thr Glu Glu Glu Leu Ala Asn Met Ser Leu  
 245 250 255  
 Asn Phe Leu Asn Cys Cys Cys Lys Lys Asn Gly Gly Glu Glu Glu Asn  
 260 265 270  
 Ala Pro Asn Pro Asn Pro Asp Gln Lys Pro Arg Arg Lys Lys Lys Glu  
 275 280 285  
 Lys Arg Pro Arg Gly Thr Met Gln Ala Ile Asn Asn Glu Lys Lys Ala  
 290 295 300  
 Ser Lys Val Leu Gly Ile Val Phe Phe Val Phe Leu Ile Met Trp Cys  
 305 310 315 320  
 Pro Phe Phe Ile Thr Asn Ile Leu Ser Val Leu Cys Gly Lys Ala Cys  
 325 330 335  
 Asn Gln Lys Leu Met Glu Lys Leu Leu Asn Val Phe Val Trp Ile Gly  
 340 345 350  
 Tyr Val Cys Ser Gly Ile Asn Pro Leu Val Tyr Thr Leu Phe Asn Lys  
 355 360 365  
 Ile Tyr Arg Arg Ala Phe Ser Lys Tyr Leu Arg Cys Asp Tyr Lys Pro  
 370 375 380  
 Asp Lys Lys Pro Pro Val Arg Gln Ile Pro Arg Val Ala Ala Thr Ala  
 385 390 395 400  
 Leu Ser Gly Arg Glu Leu Asn Val Asn Ile Tyr Arg His Thr Asn Glu  
 405 410 415  
 Arg Val Ala Arg Lys Ala Asn Asp Pro Glu Pro Gly Ile Glu Met Gln  
 420 425 430  
 Val Glu Asn Leu Glu Leu Pro Val Asn Pro Ser Asn Val Val Ser Glu  
 435 440 445  
 Arg Ile Ser Ser Val  
 450

## (2) INFORMATION FOR SEQ ID NO:12:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 1764 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

- (A) NAME/KEY: repeat region
- (B) LOCATION: 436..462
- (D) OTHER INFORMATION: /note= "Sequence encoding polyaspartic acid."

(ix) FEATURE:

- (A) NAME/KEY: misc\_feature
- (B) LOCATION: 463..465
- (D) OTHER INFORMATION: /note= "Codon encoding the N-terminal amino acid of the human thrombin receptor protein."

(ix) FEATURE:

- (A) NAME/KEY: misc\_feature
- (B) LOCATION: 1630..1632
- (D) OTHER INFORMATION: /note= "Codon encoding the C-terminal amino acid of the human thrombin receptor protein. "

(ix) FEATURE:

- (A) NAME/KEY: repeat region
- (B) LOCATION: 1633..1650
- (D) OTHER INFORMATION: /note= "Sequence encoding polyhistidine."

(ix) FEATURE:

- (A) NAME/KEY: misc\_feature
- (B) LOCATION: 648..656
- (D) OTHER INFORMATION: /note= "Deleted AlwNI restriction site."

(ix) FEATURE:

- (A) NAME/KEY: misc\_feature
- (B) LOCATION: 893..898
- (D) OTHER INFORMATION: /note= "Deleted PstI restriction site."

(ix) FEATURE:

- (A) NAME/KEY: misc\_feature
- (B) LOCATION: 1301..1309
- (D) OTHER INFORMATION: /note= "Deleted AlwNI restriction site."

(ix) FEATURE:

- (A) NAME/KEY: misc\_feature
- (B) LOCATION: 1394..1402
- (D) OTHER INFORMATION: /note= "Deleted AlwNI restriction site."

- (ix) FEATURE:
  - (A) NAME/KEY: misc\_signal
  - (B) LOCATION: 374
  - (D) OTHER INFORMATION: /note= "RNA start site."
  
- (ix) FEATURE:
  - (A) NAME/KEY: mutation
  - (B) LOCATION: replace(1671, "")
  - (D) OTHER INFORMATION: /note= "C to T mutation removes AlwNI site."
  
- (ix) FEATURE:
  - (A) NAME/KEY: CDS
  - (B) LOCATION: 376..1650
  
- (ix) FEATURE:
  - (A) NAME/KEY: misc feature
  - (B) LOCATION: 376..414
  - (D) OTHER INFORMATION: /note= "Bacteriorhodopsin pre-sequence."
  
- (ix) FEATURE:
  - (A) NAME/KEY: terminator
  - (B) LOCATION: 1672..1674
  - (D) OTHER INFORMATION: /note= "Bacteriorhodopsin stop codon."
  
- (ix) FEATURE:
  - (A) NAME/KEY: terminator
  - (B) LOCATION: 1651..1653
  - (D) OTHER INFORMATION: /note= "Thrombin stop codon."

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

ATCTGCAGGA TGGGTGCAAC CGTGAAGTCC GTCACGGCTG CGTCACGACA GGAGCCGACC	60
AGCGACACCC AGAAGGTGCG AACGGTTGAG TGCCGCAACG ATCACGAGTT TTTCGTGCGC	120
TTCGAGTGGT AACACGCGTG CACGCATCGA CTTCACCGCG GGTGTTTCGA CGCCAGCCGG	180
CCGTTGAACC AGCAGGCAGC GGGCATTTCATCCGCTGT GGCCACACA CTCGGTGGGG	240
TGCGCTATTT TGGTATGGTT TGAATCCGC GTGTCGGCTC CGTGTCTGAC GGTTTCATCGG	300
TCTAAATTCC GTCACGAGCG TACCATACTG ATTGGGTCGT AGAGTTACAC ACATATCCTC	360
GTTAGGTA CT GTTGC ATG TTG GAG TTA TTG CCA ACA GCA GTG GAG GGG GTA	411
Met Leu Glu Leu Leu Pro Thr Ala Val Glu Gly Val	
1 5 10	

TCG CAG GCC CAG ATC CAG GCG CTG GAC TAC AAG GAC GAT GAT GAC GTC Ser Gln Ala Gln Ile Gln Ala Leu Asp Tyr Lys Asp Asp Asp Asp Val 15 20 25	459
GAC GCC ACC TTA GAT CCC CGG TCA TTT CTT CTC AGG AAC CCC AAT GAT Asp Ala Thr Leu Asp Pro Arg Ser Phe Leu Leu Arg Asn Pro Asn Asp 30 35 40	507
AAA TAT GAA CCA TTT TGG GAG GAT GAG GAG AAA AAT GAA AGT GGG TTA Lys Tyr Glu Pro Phe Trp Glu Asp Glu Glu Lys Asn Glu Ser Gly Leu 45 50 55 60	555
ACT GAA TAC AGA TTA GTC TCC ATC AAT AAA AGC AGT CCT CTT CAA AAA Thr Glu Tyr Arg Leu Val Ser Ile Asn Lys Ser Ser Pro Leu Gln Lys 65 70 75	603
CAA CTT CCT GCA TTC ATC TCA GAA GAT GCC TCC GGA TAT TTG ACC AGC Gln Leu Pro Ala Phe Ile Ser Glu Asp Ala Ser Gly Tyr Leu Thr Ser 80 85 90	651
TCC TGG CTG ACA CTC TTT GTC CCA TCT GTG TAC ACC GGA GTG TTT GTA Ser Trp Leu Thr Leu Phe Val Pro Ser Val Tyr Thr Gly Val Phe Val 95 100 105	699
GTC AGC CTC CCA CTA AAC ATC ATG GCC ATC GTT GTG TTC ATC CTG AAA Val Ser Leu Pro Leu Asn Ile Met Ala Ile Val Val Phe Ile Leu Lys 110 115 120	747
ATG AAG GTC AAG AAG CCG GCG GTG GTG TAC ATG CTG CAC CTG GCC ACG Met Lys Val Lys Lys Pro Ala Val Val Tyr Met Leu His Leu Ala Thr 125 130 135 140	795
GCA GAT GTG CTG TTT GTG TCT GTG CTC CCC TTT AAG ATC AGC TAT TAC Ala Asp Val Leu Phe Val Ser Val Leu Pro Phe Lys Ile Ser Tyr Tyr 145 150 155	843
TTT TCC GGC AGT GAT TGG CAG TTT GGG TCT GAA TTG TGT CGC TTC GTC Phe Ser Gly Ser Asp Trp Gln Phe Gly Ser Glu Leu Cys Arg Phe Val 160 165 170	891
ACT GCA GCA TTT TAC TGT AAC ATG TAC GCC TCT ATC TTG CTC ATG ACA Thr Ala Ala Phe Tyr Cys Asn Met Tyr Ala Ser Ile Leu Leu Met Thr 175 180 185	939
GTC ATA AGC ATT GAC CGG TTT CTG GCT GTG GTG TAT CCC ATG CAG TCC Val Ile Ser Ile Asp Arg Phe Leu Ala Val Val Tyr Pro Met Gln Ser 190 195 200	987
CTC TCC TGG CGT ACT CTG GGA AGG GCT TCC TTC ACT TGT CTG GCC ATC Leu Ser Trp Arg Thr Leu Gly Arg Ala Ser Phe Thr Cys Leu Ala Ile 205 210 215 220	1035

TGG GCT TTG GCC ATC GCA GGG GTA GTG CCT CTC GTC CTC AAG GAG CAA	1083
Trp Ala Leu Ala Ile Ala Gly Val Val Pro Leu Val Leu Lys Glu Gln	
225 230 235	
ACC ATC CAG GTG CCC GGG CTC AAC ATC ACT ACC TGT CAT GAT GTG CTC	1131
Thr Ile Gln Val Pro Gly Leu Asn Ile Thr Thr Cys His Asp Val Leu	
240 245 250	
AAT GAA ACC CTG CTC GAA GGC TAC TAT GCC TAC TAC TTC TCA GCC TTC	1179
Asn Glu Thr Leu Leu Glu Gly Tyr Tyr Ala Tyr Tyr Phe Ser Ala Phe	
255 260 265	
TCT GCT GTC TTC TTT TTT GTG CCG CTG ATC ATT TCC ACG GTC TGT TAT	1227
Ser Ala Val Phe Phe Phe Val Pro Leu Ile Ile Ser Thr Val Cys Tyr	
270 275 280	
GTG TCT ATC ATT CGA TGT CTT AGC TCT TCC GCA GTT GCC AAC CGC AGC	1275
Val Ser Ile Ile Arg Cys Leu Ser Ser Ser Ala Val Ala Asn Arg Ser	
285 290 295 300	
AAG AAG TCC CGG GCT TTG TTC CTG TCA GCT GCT GTT TTC TGC ATC TTC	1323
Lys Lys Ser Arg Ala Leu Phe Leu Ser Ala Ala Val Phe Cys Ile Phe	
305 310 315	
ATC ATT TGC TTC GGA CCC ACA AAC GTC CTC CTG ATT GCG CAT TAC TCA	1371
Ile Ile Cys Phe Gly Pro Thr Asn Val Leu Leu Ile Ala His Tyr Ser	
320 325 330	
TTC CTT TCT CAC ACT TCC ACC ACA GAG GCT GCC TAC TTT GCC TAC CTC	1419
Phe Leu Ser His Thr Ser Thr Thr Glu Ala Ala Tyr Phe Ala Tyr Leu	
335 340 345	
CTC TGT GTC TGT GTC AGC AGC ATA AGC TCG TGC ATC GAC CCC CTA ATT	1467
Leu Cys Val Cys Val Ser Ser Ile Ser Ser Cys Ile Asp Pro Leu Ile	
350 355 360	
TAC TAT TAC GCT TCC TCT GAG TGC CAG AGG TAC GTC TAC AGT ATC TTA	1515
Tyr Tyr Tyr Ala Ser Ser Glu Cys Gln Arg Tyr Val Tyr Ser Ile Leu	
365 370 375 380	
TGC TGC AAA GAA AGT TCC GAT CCC AGC AGT TAT AAC AGC AGT GGG CAG	1563
Cys Cys Lys Glu Ser Ser Asp Pro Ser Ser Tyr Asn Ser Ser Gly Gln	
385 390 395	
TTG ATG GCA AAT AAA ATG GAT ACC TGC TCT AGT AAC CTG AAT AAC AGC	1611
Leu Met Ala Ser Lys Met Asp Thr Cys Ser Ser Asn Leu Asn Asn Ser	
400 405 410	
ATA TAC AAA AAG CTG TTA ACT CAC CAC CAC CAC CAC CAC TGAGCGGCCG	1660
Ile Tyr Lys Lys Leu Leu Thr His His His His His His His	
415 420 425	

CGACCAGCGA TTGATCGCAC ACGCAGGACA GCCCCACAAC CGGCGCGGCT GTGTTCAACG 1720  
 ACACACGATG AGTCCCCCAC TCGGTCTTGT ACTCGGATCC TTTT 1764

(2) INFORMATION FOR SEQ ID NO:13:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 425 amino acids
  - (B) TYPE: amino acid
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

Met Leu Glu Leu Leu Pro Thr Ala Val Glu Gly Val Ser Gln Ala Gln  
 1 5 10 15  
 Ile Gln Ala Leu Asp Tyr Lys Asp Asp Asp Val Asp Ala Thr Leu  
 20 25 30  
 Asp Pro Arg Ser Phe Leu Leu Arg Asn Pro Asn Asp Lys Tyr Glu Pro  
 35 40 45  
 Phe Trp Glu Asp Glu Glu Lys Asn Glu Ser Gly Leu Thr Glu Tyr Arg  
 50 55 60  
 Leu Val Ser Ile Asn Lys Ser Ser Pro Leu Gln Lys Gln Leu Pro Ala  
 65 70 75 80  
 Phe Ile Ser Glu Asp Ala Ser Gly Tyr Leu Thr Ser Ser Trp Leu Thr  
 85 90 95  
 Leu Phe Val Pro Ser Val Tyr Thr Gly Val Phe Val Val Ser Leu Pro  
 100 105 110  
 Leu Asn Ile Met Ala Ile Val Val Phe Ile Leu Lys Met Lys Val Lys  
 115 120 125  
 Lys Pro Ala Val Val Tyr Met Leu His Leu Ala Thr Ala Asp Val Leu  
 130 135 140  
 Phe Val Ser Val Leu Pro Phe Lys Ile Ser Tyr Tyr Phe Ser Gly Ser  
 145 150 155 160  
 Asp Trp Gln Phe Gly Ser Glu Leu Cys Arg Phe Val Thr Ala Ala Phe  
 165 170 175  
 Tyr Cys Asn Met Tyr Ala Ser Ile Leu Leu Met Thr Val Ile Ser Ile  
 180 185 190



Asp Arg Phe Leu Ala Val Val Tyr Pro Met Gln Ser Leu Ser Trp Arg  
 195 200 205  
 Thr Leu Gly Arg Ala Ser Phe Thr Cys Leu Ala Ile Trp Ala Leu Ala  
 210 215 220  
 Ile Ala Gly Val Val Pro Leu Val Leu Lys Glu Gln Thr Ile Gln Val  
 225 230 235 240  
 Pro Gly Leu Asn Ile Thr Thr Cys His Asp Val Leu Asn Glu Thr Leu  
 245 250 255  
 Leu Glu Gly Tyr Tyr Ala Tyr Tyr Phe Ser Ala Phe Ser Ala Val Phe  
 260 265 270  
 Phe Phe Val Pro Leu Ile Ile Ser Thr Val Cys Tyr Val Ser Ile Ile  
 275 280 285  
 Arg Cys Leu Ser Ser Ser Ala Val Ala Asn Arg Ser Lys Lys Ser Arg  
 290 295 300  
 Ala Leu Phe Leu Ser Ala Ala Val Phe Cys Ile Phe Ile Ile Cys Phe  
 305 310 315 320  
 Gly Pro Thr Asn Val Leu Leu Ile Ala His Tyr Ser Phe Leu Ser His  
 325 330 335  
 Thr Ser Thr Thr Glu Ala Ala Tyr Phe Ala Tyr Leu Leu Cys Val Cys  
 340 345 350  
 Val Ser Ser Ile Ser Ser Cys Ile Asp Pro Leu Ile Tyr Tyr Tyr Ala  
 355 360 365  
 Ser Ser Glu Cys Gln Arg Tyr Val Tyr Ser Ile Leu Cys Cys Lys Glu  
 370 375 380  
 Ser Ser Asp Pro Ser Ser Tyr Asn Ser Ser Gly Gln Leu Met Ala Ser  
 385 390 395 400  
 Lys Met Asp Thr Cys Ser Ser Asn Leu Asn Asn Ser Ile Tyr Lys Lys  
 405 410 415  
 Leu Leu Thr His His His His His His  
 420 425

## (2) INFORMATION FOR SEQ ID NO:14:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 2147 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:  
(A) NAME/KEY: misc signal  
(B) LOCATION: 378..380  
(D) OTHER INFORMATION: /note= "Bacteriorhodopsin start codon."
- (ix) FEATURE:  
(A) NAME/KEY: misc feature  
(B) LOCATION: 378..416  
(D) OTHER INFORMATION: /note= "Bacteriorhodopsin pre-sequence."
- (ix) FEATURE:  
(A) NAME/KEY: CDS  
(B) LOCATION: 378..2054
- (ix) FEATURE:  
(A) NAME/KEY: misc feature  
(B) LOCATION: 417..419  
(D) OTHER INFORMATION: /note= "Codon encoding N-terminal amino acid of mature bacteriorhodopsin."
- (ix) FEATURE:  
(A) NAME/KEY: misc feature  
(B) LOCATION: 1122..1124  
(D) OTHER INFORMATION: /note= "Codon encoding amino acid number 236 of bacteriorhodopsin."
- (ix) FEATURE:  
(A) NAME/KEY: misc feature  
(B) LOCATION: 1137..1139  
(D) OTHER INFORMATION: /note= "Codon encoding amino acid number 6 of the catalytic subunit of E. coli Aspartate Transcarbamylase."
- (ix) FEATURE:  
(A) NAME/KEY: misc feature  
(B) LOCATION: 1125..1178  
(D) OTHER INFORMATION: /note= "Synthetic DNA fragment."
- (ix) FEATURE:  
(A) NAME/KEY: misc feature  
(B) LOCATION: 1125..1136  
(D) OTHER INFORMATION: /note= "Sequence encoding Factor Xa proteolytic site."
- (ix) FEATURE:  
(A) NAME/KEY: misc feature  
(B) LOCATION: 2037..2039  
(D) OTHER INFORMATION: /note= "Codon encoding amino acid number 306 of E. coli Aspartate Transcarbamylase."

(ix) FEATURE:

- (A) NAME/KEY: misc feature
- (B) LOCATION: 2040..2054
- (D) OTHER INFORMATION: /note= "Sequence encoding bacteriorhodopsin C-terminal amino acid numbers 245 through 249. "

(ix) FEATURE:

- (A) NAME/KEY: terminator
- (B) LOCATION: 2055..2057
- (D) OTHER INFORMATION: /note= "Bacteriorhodopsin stop codon."

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

TAATCTGCAG GATGGGTGCA ACCGTGAAGT CCGTCACGGC TCGTACCGA CAGGAGCCGA	60
CCAGCGACAC CCAGAAGGTG CGAACGGTTG AGTGCCGCAA CGATCACGAG TTTTTCGTGC	120
GCTTCGAGTG GTAACACGCG TGCACGCATC GACTTCACCG CGGGTGTTC GACGCCAGCC	180
GGCCGTTGAA CCAGCAGGCA GCGGGCATT CACAGCCGCT GTGGCCACA CACTCGGTGG	240
GGTGCGCTAT TTTGGTATGG TTTGGAATCC GCGTGTGGC TCCGTGTCTG ACGTTCATC	300
GGTCTAAATT CCGTCACGAG CGTACCATAC TGATTGGGTC GTAGAGTTAC ACACATATCC	360
TCGTTAGGTA CTGTTGC ATG TTG GAG TTA TTG CCA ACA GCA GTG GAG GGG	410
Met Leu Glu Leu Leu Pro Thr Ala Val Glu Gly	
1 5 10	
GTA TCG CAG GCC CAG ATC ACC GGA CGT CCG GAG TGG ATC TGG CTA GCG	458
Val Ser Gln Ala Gln Ile Thr Gly Arg Pro Glu Trp Ile Trp Leu Ala	
15 20 25	
CTC GGT ACG GCG CTA ATG GGA CTC GGG ACG CTC TAT TTC CTC GTG AAA	506
Leu Gly Thr Ala Leu Met Gly Leu Gly Thr Leu Tyr Phe Leu Val Lys	
30 35 40	
GGG ATG GGC GTC TCG GAC CCA GAT GCA AAG AAA TTC TAC GCC ATC ACG	554
Gly Met Gly Val Ser Asp Pro Asp Ala Lys Lys Phe Tyr Ala Ile Thr	
45 50 55	
ACG CTC GTC CCA GCC ATC GCG TTC ACG ATG TAC CTC TCG ATG CTG CTG	602
Thr Leu Val Pro Ala Ile Ala Phe Thr Met Tyr Leu Ser Met Leu Leu	
60 65 70 75	
GGG TAT GGC CTC ACA ATG GTA CCG TTC GGT GGG GAG CAG AAC CCC ATC	650
Gly Tyr Gly Leu Thr Met Val Pro Phe Gly Gly Glu Gln Asn Pro Ile	
80 85 90	

TAC TGG GCG CGG TAC GCT GAC TGG CTG TTC ACC ACG CCG CTG TTG TTG Tyr Trp Ala Arg Tyr Ala Asp Trp Leu Phe Thr Thr Pro Leu Leu Leu 95 100 105	698
TTA GAC CTC GCG TTG CTC GTT GAC GCG GAT CAG GGA ACG ATC CTT GCG Leu Asp Leu Ala Leu Leu Val Asp Ala Asp Gln Gly Thr Ile Leu Ala 110 115 120	746
CTC GTC GGT GCC GAC GGC ATC ATG ATC GGG ACC GGC CTG GTC GGC GCA Leu Val Gly Ala Asp Gly Ile Met Ile Gly Thr Gly Leu Val Gly Ala 125 130 135	794
CTG ACG AAG GTC TAC TCG TAC CGC TTC GTG TGG TGG GCG ATC AGC ACC Leu Thr Lys Val Tyr Ser Tyr Arg Phe Val Trp Trp Ala Ile Ser Thr 140 145 150 155	842
GCA GCG ATG CTG TAC ATC CTG TAC GTG CTG TTC TTC GGG TTC ACC TCG Ala Ala Met Leu Tyr Ile Leu Tyr Val Leu Phe Phe Gly Phe Thr Ser 160 165 170	890
AAG GCC GAA AGC ATG CGC CCC GAG GTC GCA TCC ACG TTC AAA GTA CTG Lys Ala Glu Ser Met Arg Pro Glu Val Ala Ser Thr Phe Lys Val Leu 175 180 185	938
CGT AAC GTT ACC GTT GTG TTG TGG TCC GCG TAT CCC GTC GTG TGG CTG Arg Asn Val Thr Val Val Leu Trp Ser Ala Tyr Pro Val Val Trp Leu 190 195 200	986
ATC GGC AGC GAA GGT GCG GGA ATC GTG CCG CTG AAC ATC GAG ACG CTG Ile Gly Ser Glu Gly Ala Gly Ile Val Pro Leu Asn Ile Glu Thr Leu 205 210 215	1034
CTG TTC ATG GTG CTT GAC GTG AGC GCG AAG GTC GGC TTC GGG CTC ATC Leu Phe Met Val Leu Asp Val Ser Ala Lys Val Gly Phe Gly Leu Ile 220 225 230 235	1082
CTC CTG CGC AGT CGT GCG ATC TTC GGC GAA GCC GAA GCG CCG ATC GAA Leu Leu Arg Ser Arg Ala Ile Phe Gly Glu Ala Glu Ala Pro Ile Glu 240 245 250	1130
GGT CGT CAG AAA CAT ATC ATT TCC ATA AAC GAC CTT AGT CGC GAT GAC Gly Arg Gln Lys His Ile Ile Ser Ile Asn Asp Leu Ser Arg Asp Asp 255 260 265	1178
CTT AAT CTG GTG CTG GCG ACA GCG GCG AAA CTG AAA GCA AAC CCG CAA Leu Asn Leu Val Leu Ala Thr Ala Ala Lys Leu Lys Ala Asn Pro Gln 270 275 280	1226
CCA GAG CTG TTG AAG CAC AAA GTC ATT GCC AGC TGT TTC TTC GAA GCC Pro Glu Leu Leu Lys His Lys Val Ile Ala Ser Cys Phe Phe Glu Ala 285 290 295	1274

TCT ACC CGT ACC CGC CTC TCT TTT CAA ACA TCT ATG CAC CGC CTG GGG Ser Thr Arg Thr Arg Leu Ser Phe Gln Thr Ser Met His Arg Leu Gly 300 305 310 315	1322
GCC AGC GTG GTG GGC TTC TCC GAC AGC GCC AAT ACA TCA CTG GGT AAA Ala Ser Val Val Gly Phe Ser Asp Ser Ala Asn Thr Ser Leu Gly Lys 320 325 330	1370
AAA GGC GAA ACG CTT GCC GAT ACC ATT TCA GTT ATC AGC ACT TAC GTC Lys Gly Glu Thr Leu Ala Asp Thr Ile Ser Val Ile Ser Thr Tyr Val 335 340 345	1418
GAT GCG ATA GTG ATG CGT CAT CCG CAG GAA GGT GCG GCG CGC CTG GCC Asp Ala Ile Val Met Arg His Pro Gln Glu Gly Ala Ala Arg Leu Ala 350 355 360	1466
ACC GAG TTT TCC GGC AAT GTA CCG GTA CTG AAT GCC GGT GAT GGC TCC Thr Glu Phe Ser Gly Asn Val Pro Val Leu Asn Ala Gly Asp Gly Ser 365 370 375	1514
AAC CAA CAT CCG ACG CAA ACC TTG CTG GAC TTA TTC ACT ATT CAG GAA Asn Gln His Pro Thr Gln Thr Leu Leu Asp Leu Phe Thr Ile Gln Glu 380 385 390 395	1562
ACC CAG GGG CGT CTG GAC AAT CTC CAC GTC GCA ATG GTT GGT GAC CTG Thr Gln Gly Arg Leu Asp Asn Leu His Val Ala Met Val Gly Asp Leu 400 405 410	1610
AAA TAT GGT CGC ACC GTT CAC TCC CTG ACT CAG GCG TTA GCT AAG TTC Lys Tyr Gly Arg Thr Val His Ser Leu Thr Gln Ala Leu Ala Lys Phe 415 420 425	1658
GAC GGC AAC CGT TTT TAC TTC ATC GCG CCG GAC GCG CTG GCA ATG CCG Asp Gly Asn Arg Phe Tyr Phe Ile Ala Pro Asp Ala Leu Ala Met Pro 430 435 440	1706
CAA TAC ATT CTG GAT ATG CTC GAT GAA AAA GGG ATC GCA TGG AGT CTG Gln Tyr Ile Leu Asp Met Leu Asp Glu Lys Gly Ile Ala Trp Ser Leu 445 450 455	1754
CAC AGC TCT ATT GAA GAA GTG ATG GTG GAA GTA GAC ATC CTG TAC ATG His Ser Ser Ile Glu Glu Val Met Val Glu Val Asp Ile Leu Tyr Met 460 465 470 475	1802
ACC CGC GTG CAA AAA GAG CGT CTG GAC CCG TCC GAG TAC GCC AAC GTG Thr Arg Val Gln Lys Glu Arg Leu Asp Pro Ser Glu Tyr Ala Asn Val 480 485 490	1850
AAA GCG CAG TTT GTT CTT CGC GCC AGT GAT CTC CAC AAC GCC AAA GCC Lys Ala Gln Phe Val Leu Arg Ala Ser Asp Leu His Asn Ala Lys Ala 495 500 505	1898

AAT ATG AAA GTG CTG CAT CCG TTG CCG CGT GTT GAT GAG ATT GCG ACG 1946  
 Asn Met Lys Val Leu His Pro Leu Pro Arg Val Asp Glu Ile Ala Thr  
 510 515 520

GAT GTT GAT AAA ACG CCA CAC GCC TGG TAC TTC CAG CAG GCA GGC AAC 1994  
 Asp Val Asp Lys Thr Pro His Ala Trp Tyr Phe Gln Gln Ala Gly Asn  
 525 530 535

GGG ATT TTC GCT CTG CAA GCG TTA CTG GCA CTG GTT CTG AAT CGG GCC 2042  
 Gly Ile Phe Ala Leu Gln Ala Leu Leu Ala Leu Val Leu Asn Arg Ala  
 540 545 550 555

GCG ACC AGC GAC TGATCGCACA CGCAGGACAG CCCCACAACC GCGCGGCTG 2094  
 Ala Thr Ser Asp

TGTTCAACGA CACACGATGA GTCCCCACT CGGTCTTGTA CTCGGATCCT TTT 2147

(2) INFORMATION FOR SEQ ID NO:15:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 559 amino acids
  - (B) TYPE: amino acid
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

Met Leu Glu Leu Leu Pro Thr Ala Val Glu Gly Val Ser Gln Ala Gln  
 1 5 10 15

Ile Thr Gly Arg Pro Glu Trp Ile Trp Leu Ala Leu Gly Thr Ala Leu  
 20 25 30

Met Gly Leu Gly Thr Leu Tyr Phe Leu Val Lys Gly Met Gly Val Ser  
 35 40 45

Asp Pro Asp Ala Lys Lys Phe Tyr Ala Ile Thr Thr Leu Val Pro Ala  
 50 55 60

Ile Ala Phe Thr Met Tyr Leu Ser Met Leu Leu Gly Tyr Gly Leu Thr  
 65 70 75 80

Met Val Pro Phe Gly Gly Glu Gln Asn Pro Ile Tyr Trp Ala Arg Tyr  
 85 90 95

Ala Asp Trp Leu Phe Thr Thr Pro Leu Leu Leu Leu Asp Leu Ala Leu  
 100 105 110

Leu Val Asp Ala Asp Gln Gly Thr Ile Leu Ala Leu Val Gly Ala Asp  
 115 120 125

Gly Ile Met Ile Gly Thr Gly Leu Val Gly Ala Leu Thr Lys Val Tyr  
 130 135 140  
 Ser Tyr Arg Phe Val Trp Trp Ala Ile Ser Thr Ala Ala Met Leu Tyr  
 145 150 155 160  
 Ile Leu Tyr Val Leu Phe Phe Gly Phe Thr Ser Lys Ala Glu Ser Met  
 165 170 175  
 Arg Pro Glu Val Ala Ser Thr Phe Lys Val Leu Arg Asn Val Thr Val  
 180 185 190  
 Val Leu Trp Ser Ala Tyr Pro Val Val Trp Leu Ile Gly Ser Glu Gly  
 195 200 205  
 Ala Gly Ile Val Pro Leu Asn Ile Glu Thr Leu Leu Phe Met Val Leu  
 210 215 220  
 Asp Val Ser Ala Lys Val Gly Phe Gly Leu Ile Leu Leu Arg Ser Arg  
 225 230 235 240  
 Ala Ile Phe Gly Glu Ala Glu Ala Pro Ile Glu Gly Arg Gln Lys His  
 245 250 255  
 Ile Ile Ser Ile Asn Asp Leu Ser Arg Asp Asp Leu Asn Leu Val Leu  
 260 265 270  
 Ala Thr Ala Ala Lys Leu Lys Ala Asn Pro Gln Pro Glu Leu Leu Lys  
 275 280 285  
 His Lys Val Ile Ala Ser Cys Phe Phe Glu Ala Ser Thr Arg Thr Arg  
 290 295 300  
 Leu Ser Phe Gln Thr Ser Met His Arg Leu Gly Ala Ser Val Val Gly  
 305 310 315 320  
 Phe Ser Asp Ser Ala Asn Thr Ser Leu Gly Lys Lys Gly Glu Thr Leu  
 325 330 335  
 Ala Asp Thr Ile Ser Val Ile Ser Thr Tyr Val Asp Ala Ile Val Met  
 340 345 350  
 Arg His Pro Gln Glu Gly Ala Ala Arg Leu Ala Thr Glu Phe Ser Gly  
 355 360 365  
 Asn Val Pro Val Leu Asn Ala Gly Asp Gly Ser Asn Gln His Pro Thr  
 370 375 380  
 Gln Thr Leu Leu Asp Leu Phe Thr Ile Gln Glu Thr Gln Gly Arg Leu  
 385 390 395 400  
 Asp Asn Leu His Val Ala Met Val Gly Asp Leu Lys Tyr Gly Arg Thr  
 405 410 415

Val His Ser Leu Thr Gln Ala Leu Ala Lys Phe Asp Gly Asn Arg Phe  
 420 425 430  
 Tyr Phe Ile Ala Pro Asp Ala I Ala Met Pro Gln Tyr Ile Leu Asp  
 435 445  
 Met Leu Asp Glu Lys Gly Ile Trp Ser Leu His Ser Ser Ile Glu  
 450 455 460  
 Glu Val Met Val Glu Val Asp Ile Leu Tyr Met Thr Arg Val Gln Lys  
 465 470 475 480  
 Glu Arg Leu Asp Pro Ser Glu Tyr Ala Asn Val Lys Ala Gln Phe Val  
 485 490 495  
 Leu Arg Ala Ser Asp Leu His Asn Ala Lys Ala Asn Met Lys Val Leu  
 500 505 510  
 His Pro Leu Pro Arg Val Asp Glu Ile Ala Thr Asp Val Asp Lys Thr  
 515 520 525  
 Pro His Ala Trp Tyr Phe Gln Gln Ala Gly Asn Gly Ile Phe Ala Leu  
 530 535 540  
 Gln Ala Leu Leu Ala Leu Val Leu Asn Arg Ala Ala Thr Ser Asp  
 545 550 555



THE CLAIMS DEFINING THE INVENTION ARE AS FOLLOWS:-

1. An expression vector useful for the production of heterologous polypeptide in a halobacterial host comprising:
  - a) transcription and translation regulatory DNA;
  - b) DNA encoding a heterologous polypeptide; and
  - c) DNA encoding transcription and translation stop signals;wherein said DNA of a), b) and c) is operably linked, and wherein said heterologous polypeptide is other than a halobacteria polypeptide.
2. The vector according to claim 1 further comprising DNA encoding replication and selection capability for said halobacterial host.
3. The vector according to Claim 1 further comprising additional DNA encoding the pre-sequence of the bacteriorhodopsin gene between said regulatory DNA and said DNA encoding said heterologous polypeptide for expression of a fusion polypeptide of said pre-sequence with said heterologous polypeptide.
4. The vector according to Claim 1, 2 or 3 further comprising additional DNA encoding at least a membranous domain of the bacteriorhodopsin gene and DNA operably encoding a unique protease cleavage site and a restriction site, in optional order, between said membranous domain and said DNA encoding said heterologous polypeptide, said additional DNA being 5' of said DNA encoding said heterologous polypeptide.
5. The vector according to Claim 1, 2, 3 or 4 wherein said transcription and translation regulatory sequences are from halobacteria.
6. The vector according to Claim 5 wherein said transcription and translation regulatory sequences are those of the bacteriorhodopsin gene.
7. The vector according to Claim 1, 2, 3, 4, 5, or 6 wherein said transcription and translation stop signals are from halobacteria.
8. The vector according to Claim 7 wherein said transcription and translation regulatory sequences are those of the bacteriorhodopsin gene.
9. The vector according to claim 1 wherein said heterologous polypeptide is Type HM1 human muscarinic acetylcholine receptor.
10. The vector according to Claim 1 wherein said heterologous polypeptide is the catalytic subunit of aspartate transcarbamylase from *Escherichia coli*.



11. A halobacterial host transformed with a vector according to claim 1, 2, 3, 4, 5, 6, 7 or 8.

12. A method for producing a heterologous polypeptide in a halobacterial host comprising:  
a) transforming a halobacterial host with an expression vector, said expression vector comprising:

- i) transcription and translation regulatory DNA;
- ii) DNA encoding a heterologous polypeptide; and
- iii) DNA encoding transcriptional and translation stop signals;
- iv) wherein said DNA of i), ii), and iii) is operably linked;

b) transforming a halobacterial host with said expression vector; and

c) causing expression of said DNA encoding the heterologous polypeptide;

wherein said heterologous polypeptide is other than a halobacteria polypeptide.

13. The method according to claim 12 wherein said expression vector further comprises additional DNA encoding at least a membranous domain of the bacteriorhodopsin gene and DNA operably encoding a unique protease cleavage site and a restriction site, in optional order, between said membranous domain and said DNA encoding said heterologous polypeptide, said additional DNA being 5' of said DNA encoding said heterologous polypeptide.

14. The method according to Claim 12 or 13 wherein said transcription and translation regulatory sequences are from halobacteria.

15. The method according to Claim 14 wherein said transcription and translation regulatory sequences are those of the bacteriorhodopsin gene.

16. The method according to Claim 12, 13, 14 or 15 wherein said transcription and translation stop signals are from halobacteria.

17. The method according to Claim 16 wherein said transcription and translation regulatory sequences are those of the bacteriorhodopsin gene.

18. A method for producing a heterologous polypeptide in a halobacterial host comprising causing expression of DNA encoding said heterologous polypeptide within an operable expression vector transformed into said halobacterial host, wherein said heterologous polypeptide is other than a halobacteria polypeptide.

19. A method for producing a heterologous polypeptide in a halobacterial host comprising:  
a) transforming said halobacterial host with an expression vector encoding a fusion polypeptide, said expression vector comprising:

- i) transcription and translation regulatory DNA;
- ii) DNA encoding a unique protease cleavage site;



- iii) DNA encoding a heterologous polypeptide;
  - iv) DNA encoding a membranous domain of bacteriorhodopsin;
  - v) wherein said DNA of i), ii), iii) and iv) is operably linked;
  - b) causing expression of said fusion polypeptide such that the fusion polypeptide is localized in the membranes; and
  - c) incubating said membrane-bound fusion polypeptide with a protease such that said protease cleaves at said unique protease cleavage site to release said heterologous polypeptide from said membranes;
- wherein said heterologous polypeptide is other than a halobacteria polypeptide.

20. The method according to Claim 19 wherein said transcription and translation regulatory sequences are from halobacteria.

21. The method according to Claim 20 wherein said transcription and translation regulatory sequences are those of the bacteriorhodopsin gene.

22. The method according to Claim 19 or 20 wherein said transcription and translation stop signals are from halobacteria.

23. The method according to Claim 22 wherein said transcription and translation regulatory sequences are those of the bacteriorhodopsin gene.

Dated this 17 day of April 1997

THE REGENTS OF THE UNIVERSITY OF CALIFORNIA  
Patent Attorneys for the Applicant  
PETER MAXWELL & ASSOCIATES



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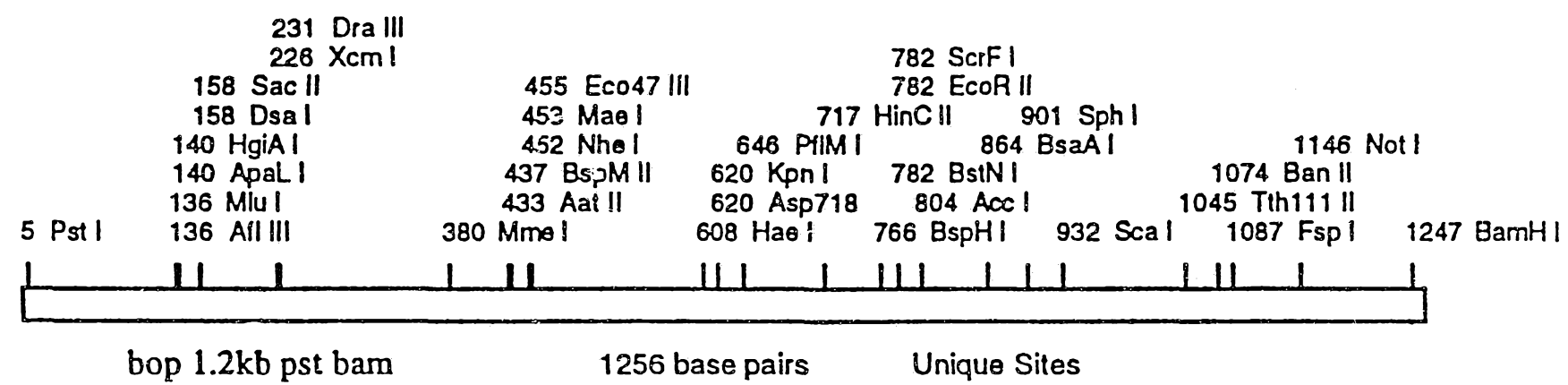


FIG. 1

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ATCTGCAGGA TGGGTGCAAC CGTGAAGTCC GTCACGGCTG CGTCACGACA GGAGCCGACC	60
AGCGACACCC AGAAGGTGCG AACGGTTGAG TGCCGCAACG ATCACGAGTT TTTCGTGCGC	120
TTCGAGTGGT AACACGCGTG CACGCATCGA CTTCACCGCG GGTGTTTCGA CGCCAGCCGG	180
CCGTTGAACC AGCAGGCAGC GGGCATTTC AAGCCGCTGT GGCCACACA CTCGGTGGGG	240
TGCGCTATTT TGGTATGGTT TGAATCCGC GTGTCGGCTC CGTGTCTGAC GGTTTCATCGG	300
TCTAAATTCC GTCACGAGCG TACCATACTG ATTGGGTCGT AGAGTTACAC ACATATCCTC	360
GTTAGGTACT GTTGC ATG TTG GAG TTA TTG CCA ACA GCA GTG GAG GGG GTA	411
Met Leu Glu Leu Leu Pro Thr Ala Val Glu Gly Val	
1 5 10	
TCG CAG GCC CAG ATC ACC GGA CGT CCG GAG TGG ATC TGG CTA GCG CTC	459
Ser Gln Ala Gln Ile Thr Gly Arg Pro Glu Trp Ile Trp Leu Ala Leu	
15 20 25	
GGT ACG GCG CTA ATG GGA CTC GGG ACG CTC TAT TTC CTC GTG AAA GGG	507
Gly Thr Ala Leu Met Gly Leu Gly Thr Leu Tyr Phe Leu Val Lys Gly	
30 35 40	

FIG. 2(A)

ATG	GGC	GTC	TCG	GAC	CCA	GAT	GCA	AAG	AAA	TTC	TAC	GCC	ATC	ACG	ACG	555
Met	Gly	Val	Ser	Asp	Pro	Asp	Ala	Lys	Lys	Phe	Tyr	Ala	Ile	Thr	Thr	
45					50					55					60	

CTC	GTC	CCA	GCC	ATC	GCG	TTC	ACG	ATG	TAC	CTC	TCG	ATG	CTG	CTG	GGG	603
Leu	Val	Pro	Ala	Ile	Ala	Phe	Thr	Met	Tyr	Leu	Ser	Met	Leu	Leu	Gly	
				65					70						75	

TAT	GGC	CTC	ACA	ATG	GTA	CCG	TTC	GGT	GGG	GAG	CAG	AAC	CCC	ATC	TAC	651
Tyr	Gly	Leu	Thr	Met	Val	Pro	Phe	Gly	Gly	Glu	Gln	Asn	Pro	Ile	Tyr	
			80					85					90			

TGG	GCG	CGG	TAC	GCT	GAC	TGG	CTG	TTC	ACC	ACG	CCG	CTG	TTG	TTG	TTA	609
Trp	Ala	Arg	Tyr	Ala	Asp	Trp	Leu	Phe	Thr	Thr	Pro	Leu	Leu	Leu	Leu	
		95					100					105				

GAC	CTC	GCG	TTG	CTC	GTT	GAC	GCG	GAT	CAG	GGA	ACG	ATC	CTT	GCG	CTC	747
Asp	Leu	Ala	Leu	Leu	Val	Asp	Ala	Asp	Gln	Gly	Thr	Ile	Leu	Ala	Leu	
	110					115					120					

GTC	GGT	GCC	GAC	GGC	ATC	ATG	ATC	GGG	ACC	GGC	CTG	GTC	GCC	GCA	CTG	795
Val	Gly	Ala	Asp	Gly	Ile	Met	Ile	Gly	Thr	Gly	Leu	Val	Gly	Ala	Leu	
125					130					135					140	

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FIG. 2(B)

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ACG	AAG	GTC	TAC	TCG	TAC	CGC	TTC	GTG	TGG	TGG	GCG	ATC	AGC	ACC	GCA	843
Thr	Lys	Val	Tyr	Ser	Tyr	Arg	Phe	Val	Trp	Trp	Ala	Ile	Ser	Thr	Ala	
				145					150					155		
GCG	ATG	CTG	TAC	ATC	CTG	TAC	GTG	CTG	TTC	TTC	GGG	TTC	ACC	TCG	AAG	891
Ala	Met	Leu	Tyr	Ile	Leu	Tyr	Val	Leu	Phe	Phe	Gly	Phe	Thr	Ser	Lys	
			160					165					170			
GCC	GAA	AGC	ATG	CGC	CCC	GAG	GTC	GCA	TCC	ACG	TTC	AAA	GTA	CTG	CGT	939
Ala	Glu	Ser	Met	Arg	Pro	Glu	Val	Ala	Ser	Thr	Phe	Lys	Val	Leu	Arg	
		175					180					185				
AAC	GTT	ACC	GTT	GTG	TTG	TGG	TCC	GCG	TAT	CCC	GTC	GTG	TGG	CTG	ATC	987
Asn	Val	Thr	Val	Val	Leu	Trp	Ser	Ala	Tyr	Pro	Val	Val	Trp	Leu	Ile	
	190					195					200					
GGC	AGC	GAA	GGT	GCG	GGA	ATC	GTG	CCG	CTG	AAC	ATC	GAG	ACG	CTG	CTG	1035
Gly	Ser	Glu	Gly	Ala	Gly	Ile	Val	Pro	Leu	Asn	Ile	Glu	Thr	Leu	Leu	
205					210					215				220		
TTC	ATG	GTG	CTT	GAC	GTG	AGC	GCG	AAG	GTC	GGC	TTC	GGG	CTC	ATC	CTC	1083
Phe	Met	Val	Leu	Asp	Val	Ser	Ala	Lys	Val	Gly	Phe	Gly	Leu	Ile	Leu	
				225					230					235		

FIG. 2(C)

CTG CGC AGT CGT GCG ATC TTC GGC GAA GCC GAA GCG CCG GAG CCG TCC	1131
Leu Arg Ser Arg Ala Ile Phe Gly Glu Ala Glu Ala Pro Glu Pro Ser	
240                                245                                250	

GCC GGC GAC GGC GCG GCC GCG ACC AGC GAC TGATCGCACA CGCAGGACAG	1181
Ala Gly Asp Gly Ala Ala Ala Thr Ser Asp	
255                                260	

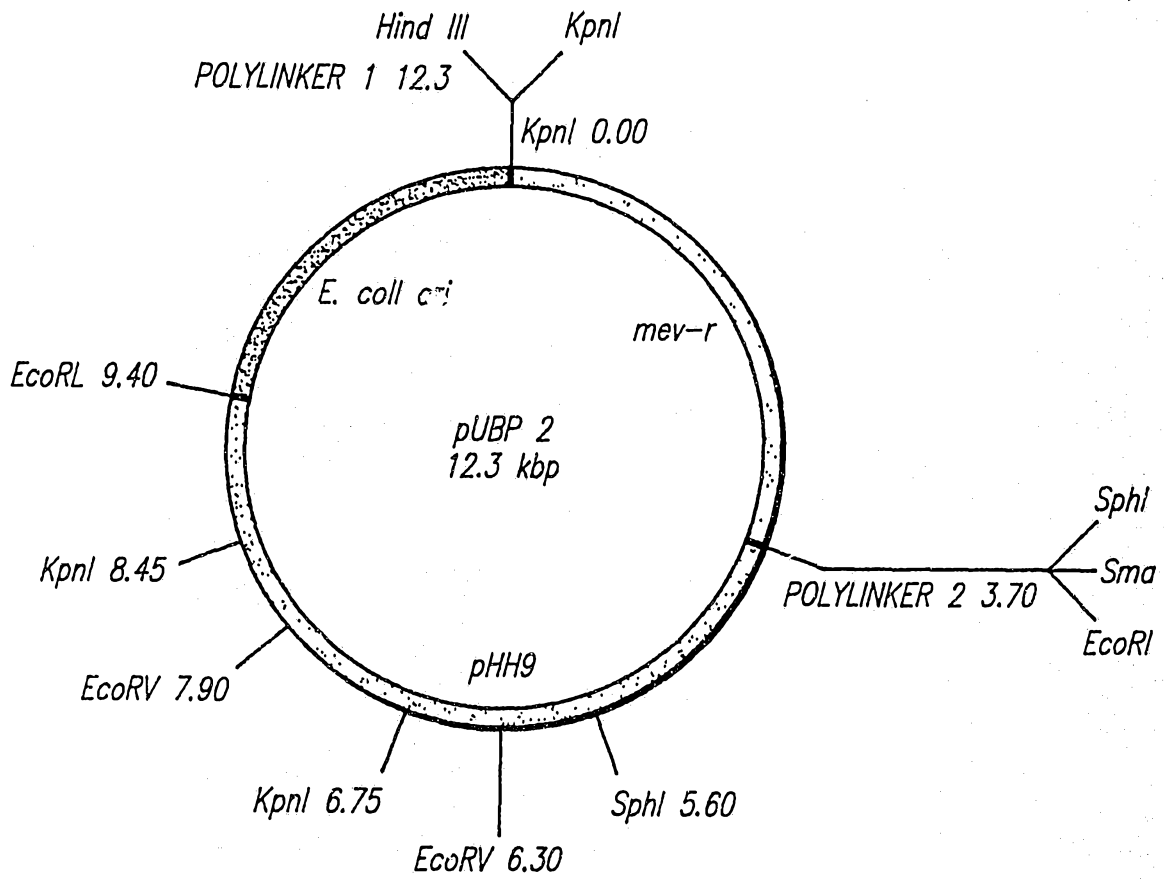
CCCCACAACC GCGCGGCTG TGTTCAACGA CACACGATGA GTCCCCACT CGGTCTTGTA 1241

CTCGGATCCT TTT 1254

FIG. 2(D)

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POLYLINKER 1 : 12.3/HindIII.SphI.MluI.XhoI.PstI.SalI.XbaI.BamHI.HindIII.XbaI.KpnI.  
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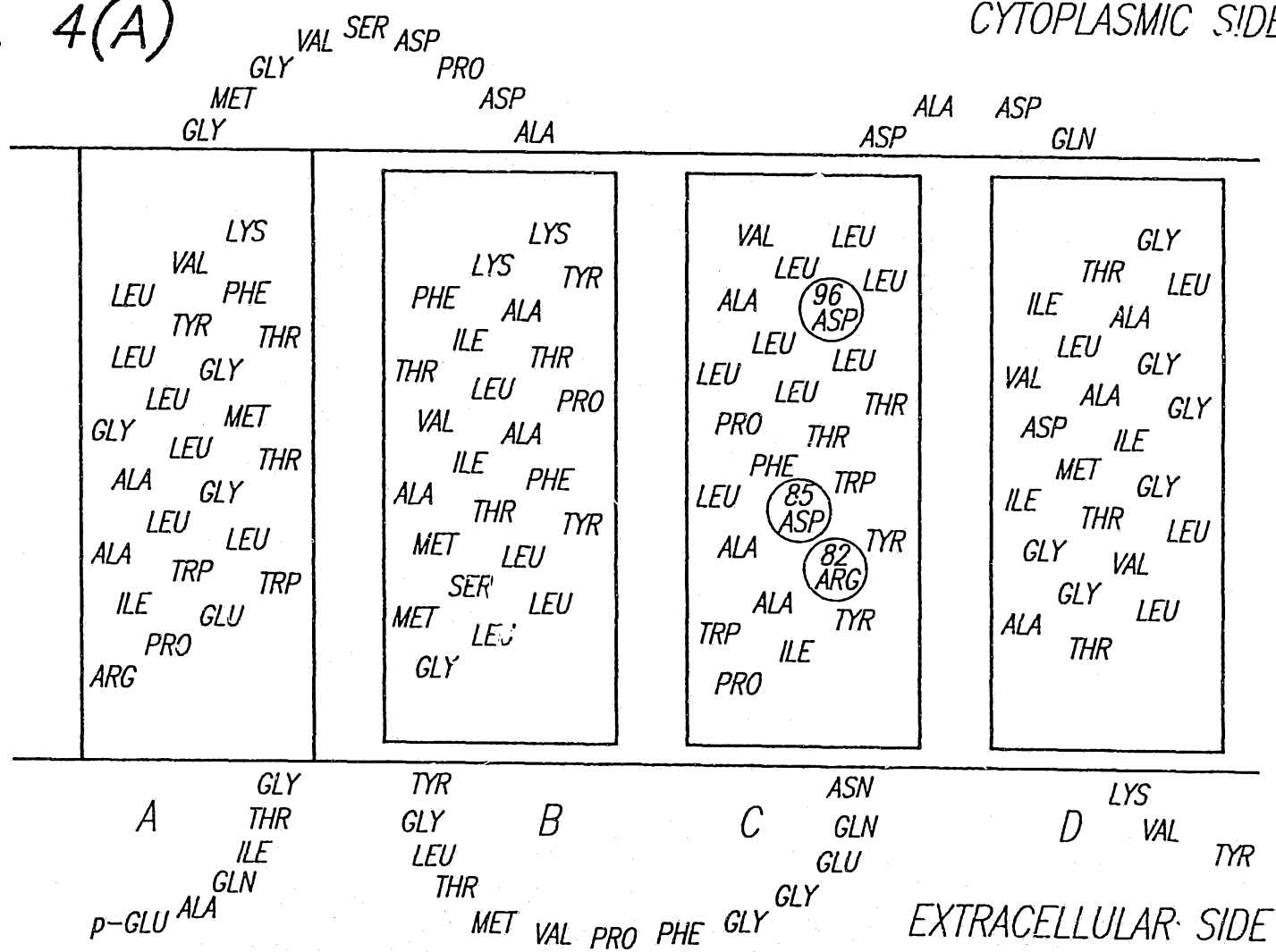
FIG. 3

FIG. 4(A)

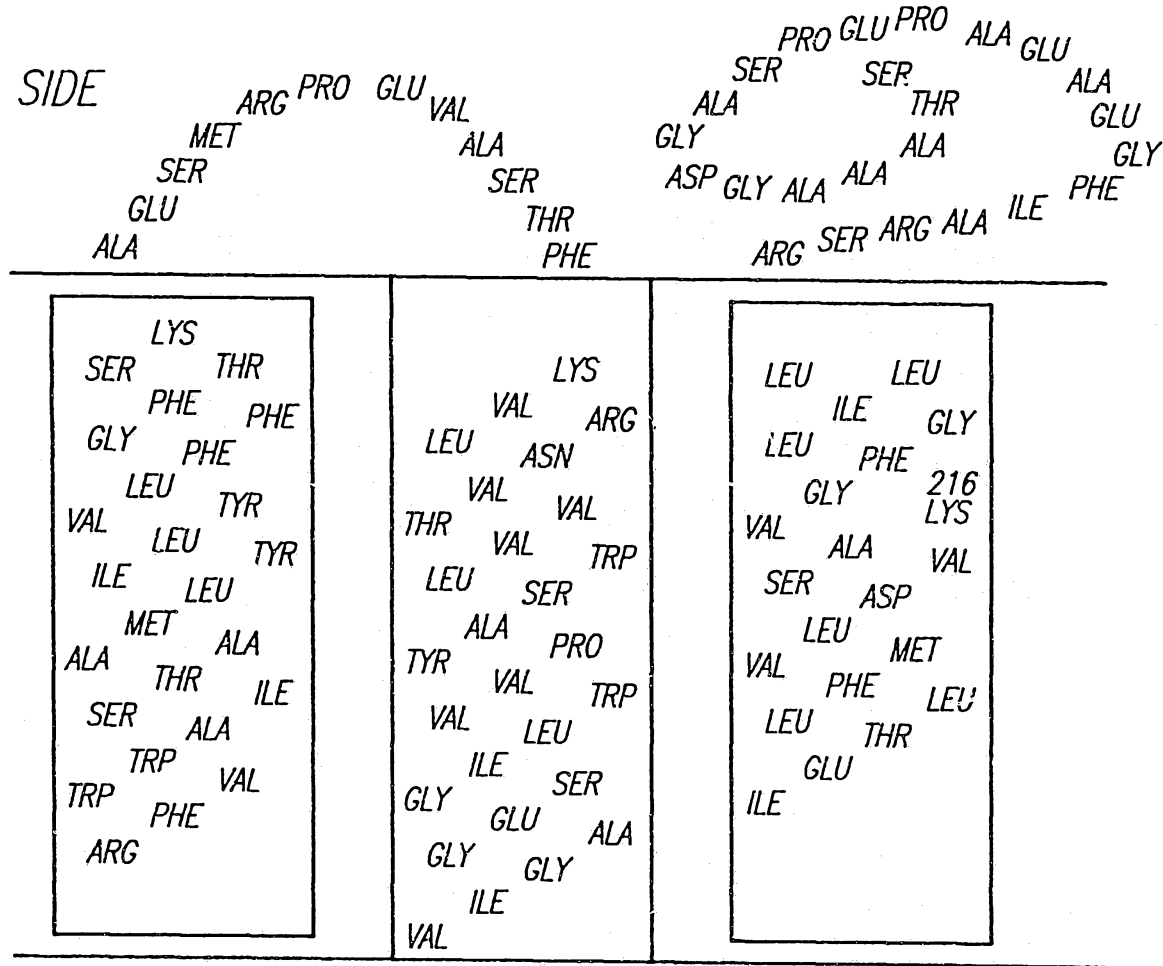
CYTOPLASMIC SIDE

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CYTOPLASMIC SIDE



TYR  
 SER E      F      PRO      LEU      ASN      G

EXTRACELLULAR SIDE

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FIG. 4(B)

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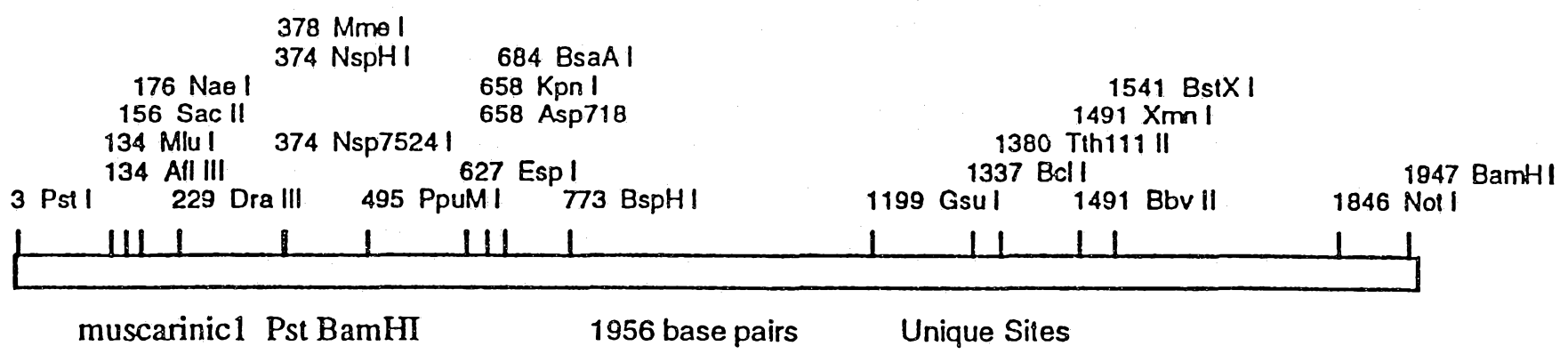


FIG. 5

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ATCTGCAGGA	TGGGTGCAAC	CGTGAAGTCC	GTCACGGCTG	CGTCACGACA	GGAGCCGACC													60
AGCGACACCC	AGAAGGTGCG	AACGGTTGAG	TGCCGCAACG	ATCACGAGTT	TTTCGTGCGC													120
TTCGAGTGGT	AACACGCGTG	CACGCATCGA	CTTCACCGCG	GGTGTTCGA	CGCCAGCCGG													180
CCGTTGAACC	AGCAGGCAGC	GGGCATTTCA	CATCCGCTGT	GGCCACACA	CTCGGTGGGG													240
TGCGCTATTT	TGGTATGGTT	TGGAATCCGC	GTGTCGGCTC	CGTGTCTGAC	GGTTCATCGG													300
TCTAAATTCC	GTCACGAGCG	TACCATACTG	ATTGGGTCGT	AGAGTTACAC	ACATATCCTC													360
GTTAGGTACT	GTTGC	ATG	TTG	GAG	TTA	TTG	CCA	ACA	GCA	GTG	GAG	GGG	GTA					411
		Met	Leu	Glu	Leu	Leu	Pro	Thr	Ala	Val	Glu	Gly	Val					
		1				5					10							
TCG	CAG	GCC	CAG	ATC	CAG	GCG	CTG	ATG	AAC	ACT	TCA	GCC	CCA	CCT	GCT			459
Ser	Gln	Ala	Gln	Ile	Gln	Ala	Leu	Met	Asn	Thr	Ser	Ala	Pro	Pro	Ala			
		15					20					25						
GTC	AGC	CCC	AAC	ATC	ACC	GTC	CTG	GCA	CCA	GGA	AAG	GGT	CCC	TGG	CAA			507
Val	Ser	Pro	Asn	Ile	Thr	Val	Leu	Ala	Pro	Gly	Lys	Gly	Pro	Trp	Gln			
		30					35				40							

FIG. 6(A)

GTG GCC TTC ATT GGG ATC ACC ACG GGC CTC CTG TCG CTA GCC ACA GTG 555  
 Val Ala Phe Ile Gly Ile Thr Thr Gly Leu Leu Ser Leu Ala Thr Val  
 45 50 55 60

ACA GGC AAC CTG CTG GTA CTC ATC TCT TTC AAG GTC AAC ACG GAG CTC 603  
 Thr Gly Asn Leu Leu Val Leu Ile Ser Phe Lys Val Asn Thr Glu Leu  
 65 70 75

AAG ACA GTC AAT AAC TAC TTC CTG CTG AGC CTG GCC TGT GCT GAC CTC 651  
 Lys Thr Val Asn Asn Tyr Phe Leu Leu Ser Leu Ala Cys Ala Asp Leu  
 80 85 90

ATC ATC GGT ACC TTC TCC ATG AAC CTC TAT ACC ACG TAC CTG CTC ATG 699  
 Ile Ile Gly Thr Phe Ser Met Asn Leu Tyr Thr Thr Tyr Leu Leu Met  
 95 100 105

GGC CAC TGG GCT CTG GGC ACG CTG GCT TGT GAC CTC TGG CTG GCC CTG 747  
 Gly His Trp Ala Leu Gly Thr Leu Ala Cys Asp Leu Trp Leu Ala Leu  
 110 115 120

GAC TAT GTG GCC AGC AAT GCC TCC GTC ATG AAT CTG CTG CTC ATC AGC 795  
 Asp Tyr Val Ala Ser Asn Ala Ser Val Met Asn Leu Leu Leu Ile Ser  
 125 130 135 140

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FIG. 6(B)

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TTT	GAC	CGC	TAC	TTC	TCC	GTG	ACT	CGG	CCC	CTG	AGC	TAC	CGT	GCC	AAG	843
Phe	Asp	Arg	Tyr	Phe	Ser	Val	Thr	Arg	Pro	Leu	Ser	Tyr	Arg	Ala	Lys	
				145					150					155		
CGC	ACA	CCC	CGC	CGC	GCA	GCT	CTG	ATG	ATC	GGC	CTG	GCC	TGG	CTG	GTT	891
Arg	Thr	Pro	Arg	Arg	Ala	Ala	Leu	Met	Ile	Gly	Leu	Ala	Trp	Leu	Val	
			160					165					170			
TCC	TTT	GTG	CTC	TGG	GCC	CCA	GCC	ATC	CTC	TTC	TGG	CAA	TAC	CTG	GTA	939
Ser	Phe	Val	Leu	Trp	Ala	Pro	Ala	Ile	Leu	Phe	Trp	Gln	Tyr	Leu	Val	
		175					180					185				
GGG	GAG	CGG	ACG	ATG	CTA	GCT	GGG	CAG	TGC	TAC	ATC	CAG	TTC	CTC	TCC	987
Gly	Glu	Arg	Thr	Met	Leu	Ala	Gly	Gln	Cys	Tyr	Ile	Gln	Phe	Leu	Ser	
	190					195				200						
CAG	CCC	ATC	ATC	ACC	TTT	GGC	ACA	GCC	ATG	GCT	GCC	TTC	TAC	CTC	CCT	1035
Gln	Pro	Ile	Ile	Thr	Phe	Gly	Thr	Ala	Met	Ala	Ala	Phe	Tyr	Leu	Pro	
205					210				215						220	
GTC	ACA	GTC	ATG	TGC	ACG	CTC	TAC	TGG	CGC	ATC	TAC	CGG	GAG	ACA	GAG	1083
Val	Thr	Val	Met	Cys	Thr	Leu	Tyr	Trp	Arg	Ile	Tyr	Arg	Glu	Thr	Glu	
				225					230					235		

FIG. 6(C)

SUBSTITUTE SHEET (RULE 26)

AAC	CGA	GCA	CGG	GAG	CTG	GCA	GCC	CTT	CAG	GGC	TCC	GAG	ACG	CCA	GGC	1131
Asn	Arg	Ala	Arg	Glu	Leu	Ala	Ala	Leu	Gln	Gly	Ser	Glu	Thr	Pro	Gly	
			240					245					250			
AAA	GGG	GGT	GGC	AGC	AGC	AGC	AGC	TCA	GAG	AGG	TCT	CAG	CCA	GGG	GCA	1179
Lys	Gly	Gly	Gly	Ser	Ser	Ser	Ser	Ser	Glu	Arg	Ser	Gln	Pro	Gly	Ala	
			255					260				265				
GAG	GGC	TCA	CCA	GAG	ACT	CCT	CCA	GGC	CGC	TGC	TGT	CGC	TGC	TGC	CGG	1227
Glu	Gly	Ser	Pro	Glu	Thr	Pro	Pro	Gly	Arg	Cys	Cys	Arg	Cys	Cys	Arg	
			270					275				280				
GCC	CCA	AGG	CTG	CTG	CAA	GCC	TAC	AGC	TGG	AAG	GAA	GAA	GAG	GAA	GAG	1275
Ala	Pro	Arg	Leu	Leu	Gln	Ala	Tyr	Ser	Trp	Lys	Glu	Glu	Glu	Glu	Glu	
					290					295					300	
GAC	GAA	GGC	TCC	ATG	GAG	TCC	CTC	ACA	TCC	TCA	GAG	GGA	GAG	GAG	CCT	1323
Asp	Glu	Gly	Ser	Met	Glu	Ser	Leu	Thr	Ser	Ser	Glu	Gly	Glu	Glu	Pro	
				305					310					315		
GGC	TCC	GAA	GTG	GTG	ATC	AAG	ATG	CCA	ATG	GTG	GAC	CCC	GAG	GCA	CAG	1371
Gly	Ser	Glu	Val	Val	Ile	Lys	Met	Pro	Met	Val	Asp	Pro	Glu	Ala	Gln	
			320					325					330			

FIG. 6(D)



GCC	CCC	ACC	AAG	CAG	CCC	CCA	CGG	AGC	TCC	CCA	AAT	ACA	GTC	AAG	AGG	1419
Ala	Pro	Thr	Lys	Gln	Pro	Pro	Arg	Ser	Ser	Pro	Asn	Thr	Val	Lys	Arg	
		335					340					345				
CCG	ACT	AAG	AAA	GGG	CGT	GAT	CGA	GCT	GGC	AAG	GGC	CAG	AAG	CCC	CGT	1467
Pro	Thr	Lys	Lys	Gly	Arg	Asp	Arg	Ala	Gly	Lys	Gly	Gln	Lys	Pro	Arg	
		350				355					360					
GGA	AAG	GAG	CAG	CTG	GCC	AAG	CGG	AAG	ACC	TTC	TCG	CTG	GTC	AAG	GAG	1515
Gly	Lys	Glu	Gln	Leu	Ala	Lys	Arg	Lys	Thr	Phe	Ser	Leu	Val	Lys	Glu	
		365			370					375					380	
AAG	AAG	GCG	GCT	CGG	ACC	CTG	AGT	GCC	ATC	CTC	CTG	GCC	TTC	ATC	CTC	1563
Lys	Lys	Ala	Ala	Arg	Thr	Leu	Ser	Ala	Ile	Leu	Leu	Ala	Phe	Ile	Leu	
				385					390					395		
ACC	TGG	ACA	CCG	TAC	AAC	ATC	ATG	GTG	CTG	GTG	TCC	ACC	TTC	TGC	AAG	1611
Thr	Trp	Thr	Pro	Tyr	Asn	Ile	Met	Val	Leu	Val	Ser	Thr	Phe	Cys	Lys	
			400					405					410			
GAC	TGT	GTT	CCC	GAG	ACC	CTG	TGG	GAG	CTG	GGC	TAC	TGG	CTG	TGC	TAC	1659
Asp	Cys	Val	Pro	Glu	Thr	Leu	Trp	Glu	Leu	Gly	Tyr	Trp	Leu	Cys	Tyr	
		415					420					425				

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FIG. 6(E)

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GTC AAC AGC ACC ATC AAC CCC ATG TGC TAC GCA CTC TGC AAC AAA GCC	1707
Val Asn Ser Thr Ile Asn Pro Met Cys Tyr Ala Leu Cys Asn Lys Ala	
430 435 440	

TTC CGG GAC ACC TTT CGC CTG CTG CTT TGC CGC TGG GAC AAG AGA CGC	1755
Phe Arg Asp Thr Phe Arg Leu Leu Leu Cys Arg Trp Asp Lys Arg Arg	
445 450 455 460	

TGG CGC AAG ATC CCC AAG CGC CCT GGC TCC GTG CAC CGC ACT CCC TCC	1803
Trp Arg Lys Ile Pro Lys Arg Pro Gly Ser Val His Arg Thr Pro Ser	
465 470 475	

CGC CAA TGC TGATAGTCCC CTCTCCTGCA TCCCTCCACC CCAGCGGCCG	1852
Arg Gln Cys	

CGACCAGCGA TTGATCGCAC ACGCAGGACA GCCCCACAAC CGGCGCGGCT GTGTTCAACG	1912
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ACACACGATG AGTCCCCCAG TCGGTCTTGT ACTCGGATCC TTTT	1956
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FIG. 6(F)

SUBSTITUTE SHEET (RULE 26)

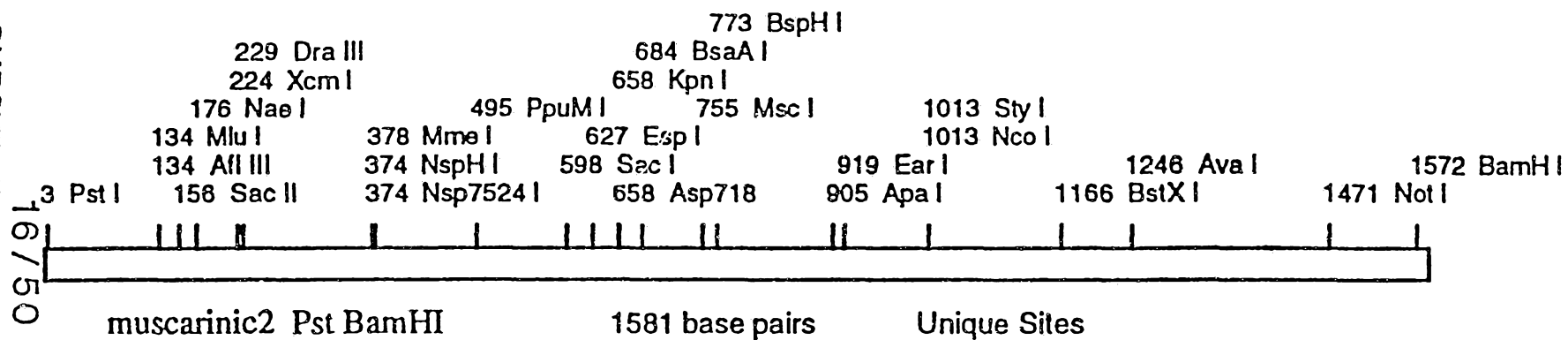


FIG. 7

SUBSTITUTE SHEET (RULE 26)  
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ATCTCCAGGA	TGGGTGCAAC	CGTGAAGTCC	GTCACGGCTG	CGTCACGACA	GGAGCCGACC		60									
AGCGACACCC	AGAAGGTGCG	AACGGTTGAG	TGCCGCAACG	ATCACGAGTT	TTTCGTGCGC		120									
TTCGAGTGGT	AACACGCGTG	CACGCATCGA	CTTCACCGCG	GGTGTTCGA	CGCCAGCCGG		180									
CCGTTGAACC	AGCAGGCAGC	GGGCATTTCA	CATCCGCTGT	GGCCACACA	CTCGGTGGGG		240									
TGCGCTATTT	TGGTATGGTT	TGGAATCCGC	GTGTCGGCTC	CGTGTCTGAC	GGTTCATCGG		300									
TCTAAATTCC	GTCACGAGCG	TACCATACTG	ATTGGGTCGT	AGAGTTACAC	ACATATCCTC		360									
GTTAGGTACT	GTTGC	ATG	TTG	GAG	TTA	TTG	CCA	ACA	GCA	GTG	GAG	GGG	GTA	411		
		Met	Leu	Glu	Leu	Leu	Pro	Thr	Ala	Val	Glu	Gly	Val			
		1				5					10					
TCG	CAG	GCC	CAG	ATC	CAG	GCG	CTG	ATG	AAC	ACT	TCA	GCC	CCA	CCT	GCT	459
Ser	Gln	Ala	Gln	Ile	Gln	Ala	Leu	Met	Asn	Thr	Ser	Ala	Pro	Pro	Ala	
		15				20					25					
GTC	AGC	CCC	AAC	ATC	ACC	GTC	CTG	GCA	CCA	GGA	AAG	GGT	CCC	TGG	CAA	507
Val	Ser	Pro	Asn	Ile	Thr	Val	Leu	Ala	Pro	Gly	Lys	Gly	Pro	Trp	Gln	
	30					35				40						

FIG. 8(A)

GTG	GCC	TTC	ATT	GGG	ATC	ACC	ACG	GGC	CTC	CTG	TCG	CTA	GCC	ACA	GTG	555
Val	Ala	Phe	Ile	Gly	Ile	Thr	Thr	Gly	Leu	Leu	Ser	Leu	Ala	Thr	Val	
45					50					55					60	
ACA	GCC	AAC	CTG	CTG	GTA	CTC	ATC	TCT	TTC	AAG	GTC	AAC	ACG	GAG	CTC	603
Thr	Gly	Asn	Leu	Leu	Val	Leu	Ile	Ser	Phe	Lys	Val	Asn	Thr	Glu	Leu	
				65					70					75		
AAG	ACA	GTC	AAT	AAC	TAC	TTC	CTG	CTG	AGC	CTG	GCC	TGT	GCT	GAC	CTC	651
Lys	Thr	Val	Asn	Asn	Tyr	Phe	Leu	Leu	Ser	Leu	Ala	Cys	Ala	Asp	Leu	
			80					85					90			
ATC	ATC	GGT	ACC	TTC	TCC	ATG	AAC	CTC	TAT	ACC	ACG	TAC	CTG	CTC	ATG	699
Ile	Ile	Gly	Thr	Phe	Ser	Met	Asn	Leu	Tyr	Thr	Thr	Tyr	Leu	Leu	Met	
		95					100					105				
GGC	CAC	TGG	GCT	CTG	GGC	ACG	CTG	GCT	TGT	GAC	CTC	TGG	CTG	GCC	CTG	747
Gly	His	Trp	Ala	Leu	Gly	Thr	Leu	Ala	Cys	Asp	Leu	Trp	Leu	Ala	Leu	
	110					115					120					
GAC	TAT	GTG	GCC	AGC	AAT	GCC	TCC	GTC	ATG	AAT	CTG	CTG	CTC	ATC	AGC	795
Asp	Tyr	Val	Ala	Ser	Asn	Ala	Ser	Val	Met	Asn	Leu	Leu	Leu	Ile	Ser	
125					130					135					140	

FIG. 8(B)

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TTT	GAC	CGC	TAC	TTC	TCC	GTG	ACT	CGG	CCC	CTG	AGC	TAC	CGT	GCC	AAG	843
Phe	Asp	Arg	Tyr	Phe	Ser	Val	Thr	Arg	Pro	Leu	Ser	Tyr	Arg	Ala	Lys	
				145					150					155		
CGC	ACA	CCC	CGC	CGC	GCA	GCT	CTG	ATG	ATC	GGC	CTG	GCC	TGG	CTG	GTT	891
Arg	Thr	Pro	Arg	Arg	Ala	Ala	Leu	Met	Ile	Gly	Leu	Ala	Trp	Leu	Val	
			160					165					170			
TCC	TTT	GTG	CTC	TGG	GCC	CCA	GCC	ATC	CTC	TTC	TGG	CAA	TAC	CTG	GTA	939
Ser	Phe	Val	Leu	Trp	Ala	Pro	Ala	Ile	Leu	Phe	Trp	Gln	Tyr	Leu	Val	
		175					180					185				
GGG	GAG	CGG	ACG	ATG	CTA	GCT	GGG	CAG	TGC	TAC	ATC	CAG	TTC	CTC	TCC	987
Gly	Glu	Arg	Thr	Met	Leu	Ala	Gly	Gln	Cys	Tyr	Ile	Gln	Phe	Leu	Ser	
	190					195				200						
CAG	CCC	ATC	ATC	ACC	TTT	GGC	ACA	GCC	ATG	GCT	GCC	TTC	TAC	CTC	CCT	1035
Gln	Pro	Ile	Ile	Thr	Phe	Gly	Thr	Ala	Met	Ala	Ala	Phe	Tyr	Leu	Pro	
205					210					215					220	
GTC	ACA	GTC	ATG	TGC	ACG	CTC	TAC	TGG	CGC	ATC	TAC	CGG	GAG	ACA	GAG	1083
Val	Thr	Val	Met	Cys	Thr	Leu	Tyr	Trp	Arg	Ile	Tyr	Arg	Glu	Thr	Glu	
				225					230					235		

FIG. 8(C)

SUBSTITUTE SHEET (RULE 26)  
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AAC	CGA	GCA	CGG	GAG	CTG	GCA	GCC	CTT	CAG	GGC	TCC	GAG	ACG	CCA	GGC	1131
Asn	Arg	Ala	Arg	Glu	Leu	Ala	Ala	Leu	Gln	Gly	Ser	Glu	Thr	Pro	Gly	
			240					245					250			
AAA	AAG	GAG	AAG	AAG	GCG	GCT	CGG	ACC	CTG	AGT	GCC	ATC	CTC	CTG	GCC	1179
Lys	Lys	Glu	Lys	Lys	Ala	Ala	Arg	Thr	Leu	Ser	Ala	Ile	Leu	Leu	Ala	
		255					260				265					
TTC	ATC	CTC	ACC	TGG	ACA	CCG	TAC	AAC	ATC	ATG	GTG	CTG	GTG	TCC	ACC	1227
Phe	Ile	Leu	Thr	Trp	Thr	Pro	Tyr	Asn	Ile	Met	Val	Leu	Val	Ser	Thr	
	270					275				280						
TTC	TGC	AAG	GAC	TGT	GTT	CCC	GAG	ACC	CTG	TGG	GAG	CTG	GGC	TAC	TGG	1275
Phe	Cys	Lys	Asp	Cys	Val	Pro	Glu	Thr	Leu	Trp	Glu	Leu	Gly	Tyr	Trp	
285				290						295					300	
CTG	TGC	TAC	GTC	AAC	AGC	ACC	ATC	AAC	CCC	ATG	TGC	TAC	GCA	CTC	TGC	1323
Leu	Cys	Tyr	Val	Asn	Ser	Thr	Ile	Asn	Pro	Met	Cys	Tyr	Ala	Leu	Cys	
				305					310					315		
AAC	AAA	GCC	TTC	CGG	GAC	ACC	TTT	CGC	CTG	CTG	CTT	TGC	CGC	TGG	GAC	1371
Asn	Lys	Ala	Phe	Arg	Asp	Thr	Phe	Arg	Leu	Leu	Leu	Cys	Arg	Trp	Asp	
		320						325					330			

FIG. 8(D)

AAG AGA CGC TGG CGC AAG ATC CCC AAG CGC CCT GGC TCC GTG CAC CGC	1419
Lys Arg Arg Trp Arg Lys Ile Pro Lys Arg Pro Gly Ser Val His Arg	
335 340 345	
ACT CCC TCC CGC CAA TGC TGATAGTCCC CTCTCCTGCA TCCCTCCACC	1467
Thr Pro Ser Arg Gln Cys	
350	
CCAGCGGCCG CGACCAGCGA TTGATCGCAC ACGCAGGACA GCCCCACAAC CGGCGCGGCT	1527
GTGTTCAACG ACACACGATG AGTCCCCCAC TCGGTCTTGT ACTCGGATCC TTTT	1581

FIG. 8(E)



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SUBSTITUTE SHEET (RULE 26)

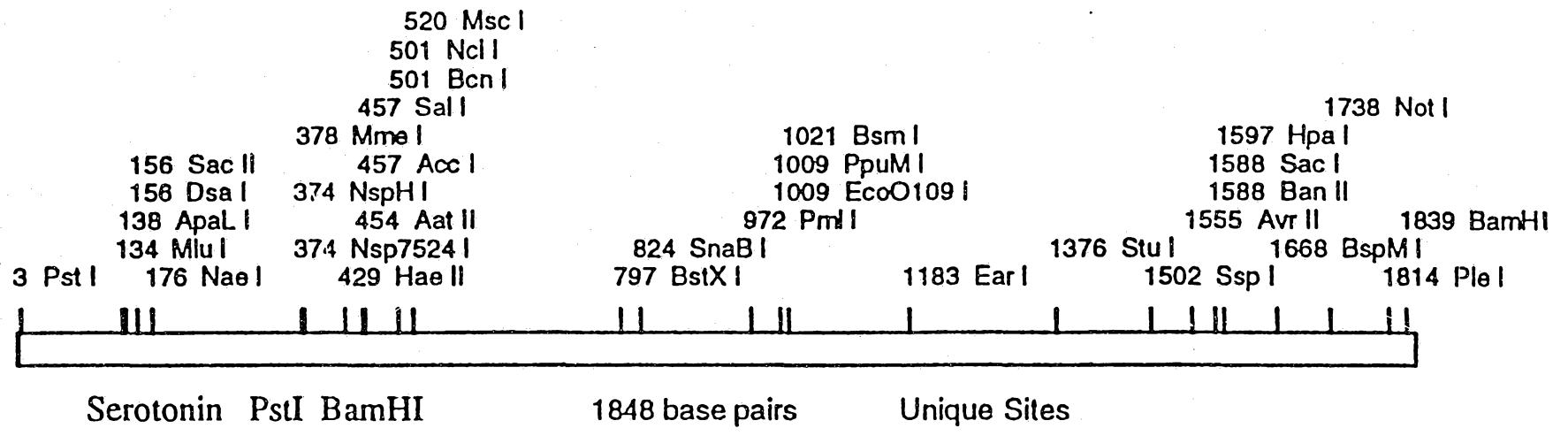


FIG. 9

SUBSTITUTE SHEET (RULE 26)  
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ATCTGCAGGA TGGGTGCAAC CGTGAAGTCC GTCACGGCTG CGTCACGACA GGAGCCGACC	60
AGCGACACCC AGAAGGTGCG AACGGTTGAG TGCCGCAACG ATCACGAGTT TTTCGTGCGC	120
TTCGAGTGGT AACACGCGTG CACGCATCGA C TTCACCGCG GGTGTTTCGA CGCCAGCCGG	180
CCGTTGAACC AGCAGGCAGC GGGCATTTC A CATCCGCTGT GGCCACACA CTCGGTGGGG	240
TGCGCTATTT TGGTATGGTT TGAATCCGC GTGTCGGCTC CGTGTCTGAC GGTTTCATCGG	300
TCTAAATTCC GTCACGAGCG TACCATACTG ATTGGGTCGT AGAGTTACAC ACATATCCTC	360
GTTAGGTACT GTTGC ATG TTG GAG TTA TTG CCA ACA GCA GTG GAG GGG GTA	411
Met Leu Glu Leu Leu Pro Thr Ala Val Glu Gly Val	
1 5 10	
TCG CAG GCC CAG ATC CAG GCG CTG GAC TAC AAG GAC GAT GAT GAC GTC	459
Ser Gln Ala Gln Ile Gln Ala Leu Asp Tyr Lys Asp Asp Asp Asp Val	
15 20 25	
GAC ACT TTT AAT TCC TCC GAT GGT GGA CGC TTG TTT CAA TTC CCG GAC	507
Asp Thr Phe Asn Ser Ser Asp Gly Gly Arg Leu Phe Gln Phe Pro Asp	
30 35 40	

FIG. 10(A)

SUBSTITUTE SHEET (RULE 26)  
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GGG	GTA	CAA	AAC	TGG	CCA	GCA	CTT	TCA	ATC	GTC	GTG	ATT	ATA	ATC	ATG	555
Gly	Val	Gln	Asn	Trp	Pro	Ala	Leu	Ser	Ile	Val	Val	Ile	Ile	Ile	Met	
45					50					55					60	
ACA	ATA	GGG	GGC	AAC	ATT	CTT	GTT	ATC	ATG	GCA	GTA	AGC	ATG	GAG	AAG	603
Thr	Ile	Gly	Gly	Asn	Ile	Leu	Val	Ile	Met	Ala	Val	Ser	Met	Glu	Lys	
				65					70					75		
AAA	CTG	CAC	AAT	GCA	ACC	AAT	TAC	TTC	TTA	ATG	TCC	CTA	GCC	ATT	GCT	651
Lys	Leu	His	Asn	Ala	Thr	Asn	Tyr	Phe	Leu	Met	Ser	Leu	Ala	Ile	Ala	
			80					85					90			
GAT	ATG	CTG	GTG	GGA	CTA	CTT	GTC	ATG	CCC	CTG	TCC	CTG	CTT	GCT	ATT	699
Asp	Met	Leu	Val	Gly	Leu	Leu	Val	Met	Pro	Leu	Ser	Leu	Leu	Ala	Ile	
		95					100					105				
CTT	TAT	GAT	TAT	GTC	TGG	CCT	TTA	CCT	AGA	TAT	TTG	TGC	CCC	GTC	TGG	747
Leu	Tyr	Asp	Tyr	Val	Trp	Pro	Leu	Pro	Arg	Tyr	Leu	Cys	Pro	Val	Trp	
	110					115					120					
ATT	TCA	CTA	GAT	GTG	CTA	TTT	TCA	ACT	GCG	TCC	ATC	ATG	CAC	CTC	TGC	795
Ile	Ser	Leu	Asp	Val	Leu	Phe	Ser	Thr	Ala	Ser	Ile	Met	His	Leu	Cys	
125					130					135					140	

FIG. 10(B)

GCC	ATA	TCG	CTG	GAC	CGG	TAT	GTA	GCA	ATA	CGT	AAT	CCT	ATT	GAG	CAT	843
Ala	Ile	Ser	Leu	Asp	Arg	Tyr	Val	Ala	Ile	Arg	Asn	Pro	Ile	Glu	His	
				145						150					155	
AGC	CGG	TTC	AAT	TCG	CGG	ACT	AAG	GCC	ATC	ATG	AAG	ATT	GCC	ATC	GTT	891
Ser	Arg	Phe	Asn	Ser	Arg	Thr	Lys	Ala	Ile	Met	Lys	Ile	Ala	Ile	Val	
			160							165					170	
TGG	GCA	ATA	TCA	ATA	GGA	GTT	TCA	GTT	CCT	ATC	CCT	GTG	ATT	GGA	CTG	939
Trp	Ala	Ile	Ser	Ile	Gly	Val	Ser	Val	Pro	Ile	Pro	Val	Ile	Gly	Leu	
			175							180					185	
AGG	GAC	GAA	AGC	AAA	GTG	TTC	GTG	AAT	AAC	ACC	ACG	TGC	GTG	CTC	AAT	987
Arg	Asp	Glu	Ser	Lys	Val	Phe	Val	Asn	Asn	Thr	Thr	Cys	Val	Leu	Asn	
	190							195				200				
GAC	CCC	AAC	TTC	GTT	CTC	ATC	GGG	TCC	TTC	GTG	GCA	TTC	TTC	ATC	CCG	1035
Asp	Pro	Asn	Phe	Val	Leu	Ile	Gly	Ser	Phe	Val	Ala	Phe	Phe	Ile	Pro	
205					210						215				220	
TTG	ACG	ATT	ATG	GTG	ATC	ACC	TAC	TTC	TTA	ACG	ATC	TAC	GTC	CTG	CGC	1083
Leu	Thr	Ile	Met	Val	Ile	Thr	Tyr	Phe	Leu	Thr	Ile	Tyr	Val	Leu	Arg	
				225						230					235	

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FIG. 10(C)

CGT	CAA	ACT	CTG	ATG	TTA	CTT	CGA	GGT	CAC	ACC	GAG	GAG	GAA	CTG	GCT	1131
Arg	Gln	Thr	Leu	Met	Leu	Leu	Arg	Gly	His	Thr	Glu	Glu	Glu	Leu	Ala	
			240					245					250			
AAT	ATG	AGC	CTG	AAC	TTT	CTG	AAC	TGC	TGC	TGC	AAG	AAG	AAT	GGT	GGT	1179
Asn	Met	Ser	Leu	Asn	Phe	Leu	Asn	Cys	Cys	Cys	Lys	Lys	Asn	Gly	Gly	
		255					260						265			
GAG	GAA	GAG	AAC	GCT	CCG	AAC	CCT	AAT	CCA	GAT	CAG	AAA	CCA	CGT	CGA	1227
Glu	Glu	Glu	Asn	Ala	Pro	Asn	Pro	Asn	Pro	Asp	Gln	Lys	Pro	Arg	Arg	
	270						275				280					
AAG	AAG	AAA	GAA	AAG	CGT	CCC	AGA	GGC	ACC	ATG	CAA	GCT	ATC	AAC	AAC	1275
Lys	Lys	Lys	Glu	Lys	Arg	Pro	Arg	Gly	Thr	Met	Gln	Ala	Ile	Asn	Asn	
285					290					295					300	
GAA	AAG	AAA	GCT	TCC	AAA	GTC	CTT	GGC	ATT	GTA	TTC	TTT	GTG	TTT	CTG	1323
Glu	Lys	Lys	Ala	Ser	Lys	Val	Leu	Gly	Ile	Val	Phe	Phe	Val	Phe	Leu	
				305					310					315		
ATC	ATG	TGG	TGC	CCG	TTT	TTC	ATC	ACC	AAT	ATC	CTG	TCG	GTT	CTT	TGT	1371
Ile	Met	Trp	Cys	Pro	Phe	Phe	Ile	Thr	Asn	Ile	Leu	Ser	Val	Leu	Cys	
			320					325					330			

FIG. 10(D)

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SUBSTITUTE SHEET (RULE 26)

GGG AAG GCC TGT AAC CAA AAG CTA ATG GAG AAG CTT CTC AAT GTG TTT 1419  
 Gly Lys Ala Cys Asn Gln Lys Leu Met Glu Lys Leu Leu Asn Val Phe  
 335 340 345

GTG TGG ATT GGC TAT GTG TGT TCA GGC ATC AAT CCT CTG GTG TAC ACT 1467  
 Val Trp Ile Gly Tyr Val Cys Ser Gly Ile Asn Pro Leu Val Tyr Thr  
 350 355 360

CTC TTT AAT AAA ATT TAC CGA AGG GCT TTC TCT AAA TAT TTG CGC TGC 1515  
 Leu Phe Asn Lys Ile Tyr Arg Arg Ala Phe Ser Lys Tyr Leu Arg Cys  
 365 370 375 380

GAT TAT AAG CCA GAC AAA AAG CCT CCT GTT CGA CAG ATT CCT AGG GTT 1563  
 Asp Tyr Lys Pro Asp Lys Lys Pro Pro Val Arg Gln Ile Pro Arg Val  
 385 390 395

GCT GCC ACT GCT TTG TCT GGG AGG GAG CTC AAT GTT AAC ATT TAT CGG 1611  
 Ala Ala Thr Ala Leu Ser Gly Arg Glu Leu Asn Val Asn Ile Tyr Arg  
 400 405 410

CAT ACC AAT GAA CGT GTG GCT AGG AAA GCT AAT GAC CCT GAG CCT GGC 1659  
 His Thr Asn Glu Arg Val Ala Arg Lys Ala Asn Asp Pro Glu Pro Gly  
 415 420 425

FIG. 10(E)

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ATA GAG ATG CAG GTG GAG AAC TTA GAG CTG CCA GTC AAC CCC TCT AAT	1707
Ile Glu Met Gln Val Glu Asn Leu Glu Leu Pro Val Asn Pro Ser Asn	
430 435 440	

GTG GTC AGC GAG AGG ATT AGT AGT GTG TGAGCGGCCG CGACCAGCGA	1754
Val Val Ser Glu Arg Ile Ser Ser Val	
445 450	

TTGATCGCAC ACGCAGGACA GCCCACAAC CGGCGGGCT GTGTTCAACG ACACACGATG	1814
---	------

AGTCCCCAC TCGGTCTTGT ACTCGGATCC TTTT	1848
--------------------------------------	------

FIG. 10(F)

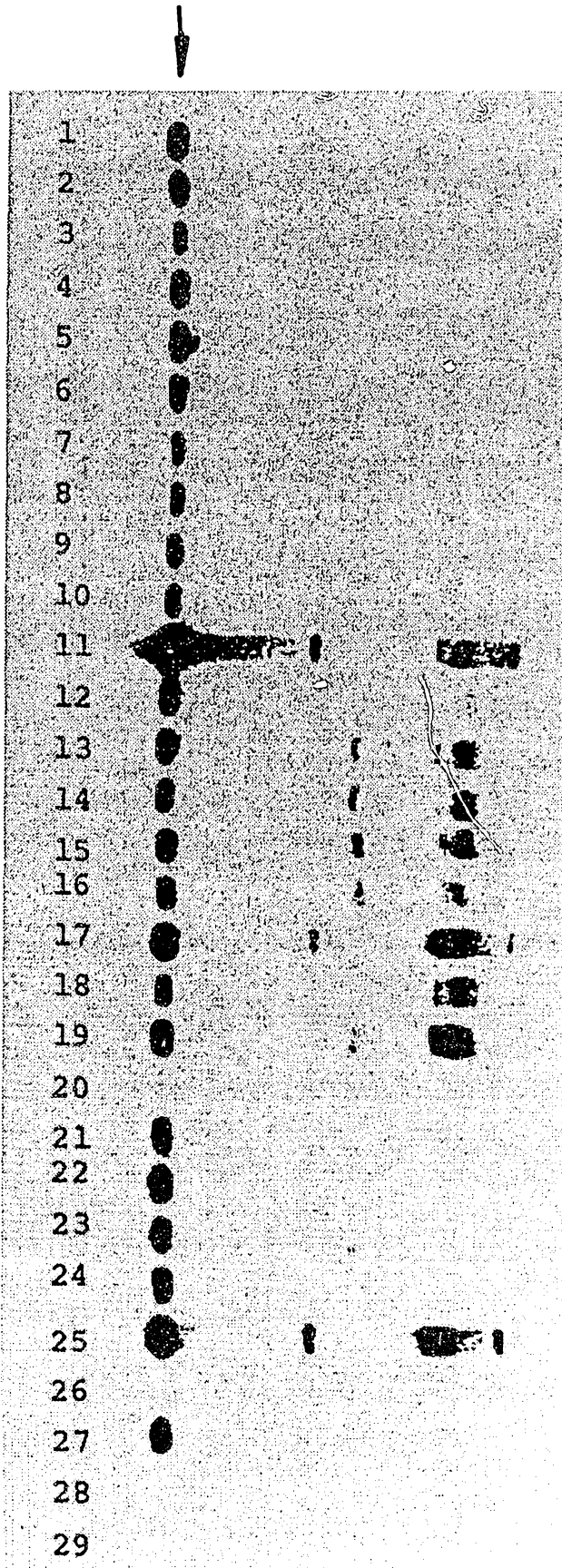


FIG. 11



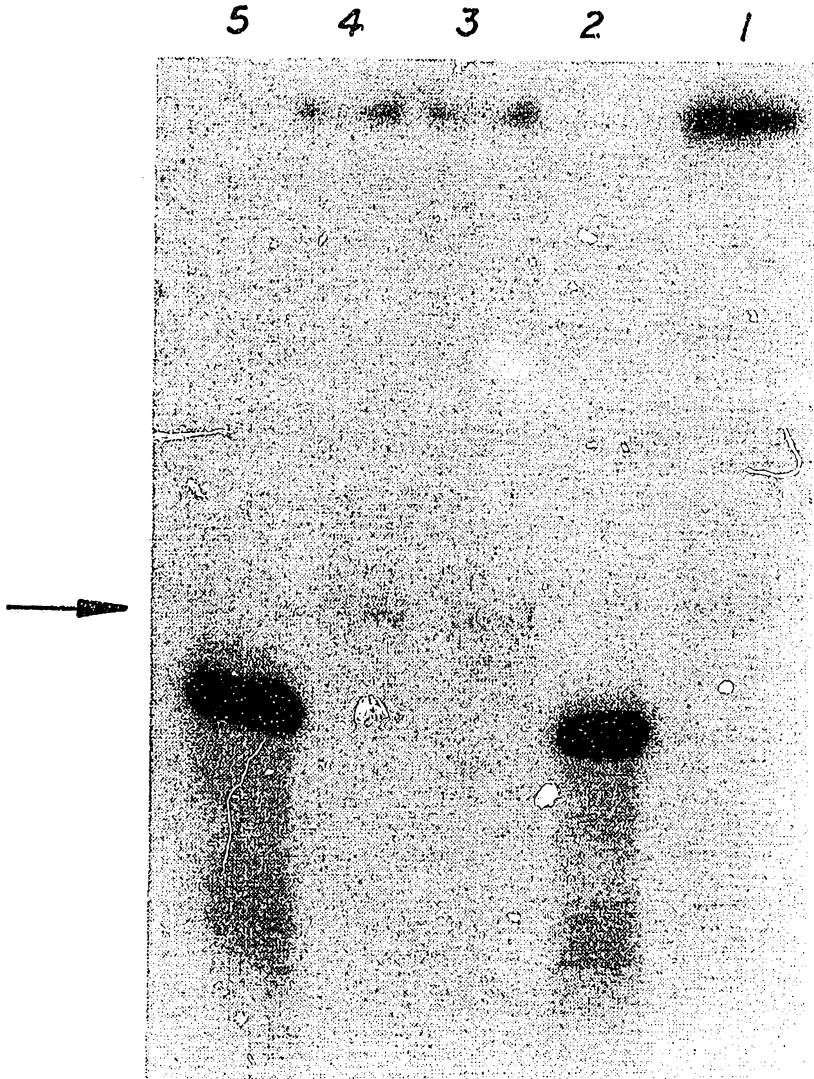


FIG. 12

SUBSTITUTE SHEET (RULE 26)

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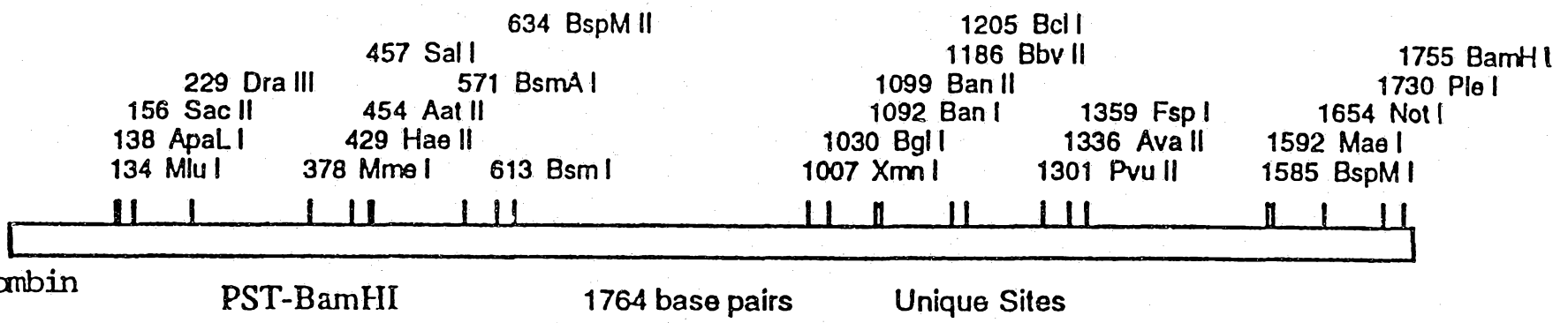


FIG. 13

SUBSTITUTE SHEET (RULE 26)  
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ATCTGCAGGA TGGGTGCAAC CGTGAAGTCC GTCACGGCTG CGTCACGACA GGAGCCGACC	60
AGCGACACCC AGAAGGTGCG AACGGTTGAG TGCCGCAACG ATCACGAGTT TTTCGTGCGC	120
TTCGAGTGGT AACACGCGTG CACGCATCGA CTTACCGCG GGTGTTTCGA CGCCAGCCGG	180
CCGTTGAACC AGCAGGCAGC GGGCATTTC AATCCGCTGT GGCCACACA CTCGGTGGGG	240
TGCGCTATTT TGGTATGGTT TGAATCCGC GTGTCGGCTC CGTGTCTGAC GGTTTCATCGG	300
TCTAAATTCC GTCACGAGCG TACCATACTG ATTGGGTCGT AGAGTTACAC ACATATCCTC	360
GTTAGGTACT GTTGC ATG TTG GAG TTA TTG CCA ACA GCA GTG GAG GGG GTA	411
Met Leu Glu Leu Leu Pro Thr Ala Val Glu Gly Val	
1 5 10	
TCG CAG GCC CAG ATC CAG GCG CTG GAC TAC AAG GAC GAT GAT GAC GTC	459
Ser Gln Ala Gln Ile Gln Ala Leu Asp Tyr Lys Asp Asp Asp Asp Val	
15 20 25	
GAC GCC ACC TTA GAT CCC CGG TCA TTT CTT CTC AGG AAC CCC AAT GAT	507
Asp Ala Thr Leu Asp Pro Arg Ser Phe Leu Leu Arg Asn Pro Asn Asp	
30 35 40	

FIG. 14(A)

AAA	TAI	GAA	CCA	TTT	TGG	GAG	GAT	GAG	GAG	AAA	AAT	GAA	AGT	GGG	TTA	555
Lys	Tyr	Glu	Pro	Phe	Trp	Glu	Asp	Glu	Glu	Lys	Asn	Glu	Ser	Gly	Leu	
45					50					55					60	

ACT	GAA	TAC	AGA	TTA	GTC	TCC	ATC	AAT	AAA	AGC	AGT	CCT	CTT	CAA	AAA	603
Thr	Glu	Tyr	Arg	Leu	Val	Ser	Ile	Asn	Lys	Ser	Ser	Pro	Leu	Gln	Lys	
				65					70					75		

CAA	CTT	CCT	GCA	TTC	ATC	TCA	GAA	GAT	GCC	TCC	GGA	TAT	TTG	ACC	AGC	651
Gln	Leu	Pro	Ala	Phe	Ile	Ser	Glu	Asp	Ala	Ser	Gly	Tyr	Leu	Thr	Ser	
			80					85					90			

TCC	TGG	CTG	ACA	CTC	TTT	GTC	CCA	TCT	GTG	TAC	ACC	GGA	GTG	TTT	GTA	699
Ser	Trp	Leu	Thr	Leu	Phe	Val	Pro	Ser	Val	Tyr	Thr	Gly	Val	Phe	Val	
		95					100					105				

GTC	AGC	CTC	CCA	CTA	AAC	ATC	ATG	GCC	ATC	GTT	GTG	TTC	ATC	CTG	AAA	747
Val	Ser	Leu	Pro	Leu	Asn	Ile	Met	Ala	Ile	Val	Val	Phe	Ile	Leu	Lys	
	110					115				120						

ATG	AAG	GTC	AAG	AAG	CCG	GCG	GTG	GTG	TAC	ATG	CTG	CAC	CTG	GCC	ACG	795
Met	Lys	Val	Lys	Lys	Pro	Ala	Val	Val	Tyr	Met	Leu	His	Leu	Ala	Thr	
125					130					135					140	

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FIG. 14(B)

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 SUBSTITUTE SHEET (RULE 26)

GCA	GAT	GTG	CTG	TTT	GTG	TCT	GTG	CTC	CCC	TTT	AAG	ATC	AGC	TAT	TAC	843
Ala	Asp	Val	Leu	Phe	Val	Ser	Val	Leu	Pro	Phe	Lys	Ile	Ser	Tyr	Tyr	
				145					150					155		
TTT	TCC	GGC	AGT	GAT	TGG	CAG	TTT	GGG	TCT	GAA	TTG	TGT	CGC	TTC	GTC	891
Phe	Ser	Gly	Ser	Asp	Trp	Gln	Phe	Gly	Ser	Glu	Leu	Cys	Arg	Phe	Val	
			160					165					170			
ACT	GCA	GCA	TTT	TAC	TGT	AAC	ATG	TAC	GCC	TCT	ATC	TTG	CTC	ATG	ACA	939
Thr	Ala	Ala	Phe	Tyr	Cys	Asn	Met	Tyr	Ala	Ser	Ile	Leu	Leu	Met	Thr	
		175					180					185				
GTC	ATA	AGC	ATT	GAC	CGG	TTT	CTG	GCT	GTG	GTG	TAT	CCC	ATG	CAG	TCC	987
Val	Ile	Ser	Ile	Asp	Arg	Phe	Leu	Ala	Val	Val	Tyr	Pro	Met	Gln	Ser	
	190					195					200					
CTC	TCC	TGG	CGT	ACT	CTG	GGA	AGG	GCT	TCC	TTC	ACT	TGT	CTG	GCC	ATC	1035
Leu	Ser	Trp	Arg	Thr	Leu	Gly	Arg	Ala	Ser	Phe	Thr	Cys	Leu	Ala	Ile	
205					210				215						220	
TGG	GCT	TTG	GCC	ATC	GCA	GGG	GTA	GTG	CCT	CTC	GTC	CTC	AAG	GAG	CAA	1083
Trp	Ala	Leu	Ala	Ile	Ala	Gly	Val	Val	Pro	Leu	Val	Leu	Lys	Glu	Gln	
				225					230					235		

FIG. 14(C)

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ACC	ATC	CAG	GTG	CCC	GGG	CTC	AAC	ATC	ACT	ACC	TGT	CAT	GAT	GTG	CTC	1131
Thr	Ile	Gln	Val	Pro	Gly	Leu	Asn	Ile	Thr	Thr	Cys	His	Asp	Val	Leu	
			240					245					250			

AAT	GAA	ACC	CTG	CTC	GAA	GGC	TAC	TAT	GCC	TAC	TAC	TTC	TCA	GCC	TTC	1179
Asn	Glu	Thr	Leu	Leu	Glu	Gly	Tyr	Tyr	Ala	Tyr	Tyr	Phe	Ser	Ala	Phe	
		255					260					265				

TCT	GCT	GTC	TTC	TTT	TTT	GTG	CCG	CTG	ATC	ATT	TCC	ACG	GTC	TGT	TAT	1227
Ser	Ala	Val	Phe	Phe	Phe	Val	Pro	Leu	Ile	Ile	Ser	Thr	Val	Cys	Tyr	
	270					275					280					

GTG	TCT	ATC	ATT	CGA	TGT	CTT	AGC	TCT	TCC	GCA	GTT	GCC	AAC	CGC	AGC	1275
Val	Ser	Ile	Ile	Arg	Cys	Leu	Ser	Ser	Ser	Ala	Val	Ala	Asn	Arg	Ser	
285				290						295					300	

AAG	AAG	TCC	CGG	GCT	TTG	TTC	CTG	TCA	GCT	GCT	GTT	TTC	TGC	ATC	TTC	1323
Lys	Lys	Ser	Arg	Ala	Leu	Phe	Leu	Ser	Ala	Ala	Val	Phe	Cys	Ile	Phe	
				305					310					315		

ATC	ATT	TGC	TTC	GGA	CCC	ACA	AAC	GTC	CTC	CTG	ATT	GCG	CAT	TAC	TCA	1371
Ile	Ile	Cys	Phe	Gly	Pro	Thr	Asn	Val	Leu	Leu	Ile	Ala	His	Tyr	Ser	
			320					325					330			

FIG. 14(D)

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TTC	CTT	TCT	CAC	ACT	TCC	ACC	ACA	GAG	GCT	GCC	TAC	TTT	GCC	TAC	CTC	1419
Phe	Leu	Ser	His	Thr	Ser	Thr	Thr	Glu	Ala	Ala	Tyr	Phe	Ala	Tyr	Leu	
		335						340					345			
CTC	TGT	GTC	TGT	GTC	AGC	AGC	ATA	AGC	TCG	TGC	ATC	GAC	CCC	CTA	ATT	1467
Leu	Cys	Val	Cys	Val	Ser	Ser	Ile	Ser	Ser	Cys	Ile	Asp	Pro	Leu	Ile	
	350						355				360					
TAC	TAT	TAC	GCT	TCC	TCT	GAG	TGC	CAG	AGG	TAC	GTC	TAC	AGT	ATC	TTA	1515
Tyr	Tyr	Tyr	Ala	Ser	Ser	Glu	Cys	Gln	Arg	Tyr	Val	Tyr	Ser	Ile	Leu	
365					370					375					380	
TGC	TGC	AAA	GAA	AGT	TCC	GAT	CCC	AGC	AGT	TAT	AAC	AGC	AGT	GGG	CAG	1563
Cys	Cys	Lys	Glu	Ser	Ser	Asp	Pro	Ser	Ser	Tyr	Asn	Ser	Ser	Gly	Gln	
				385					390					395		
TTG	ATG	GCA	AGT	AAA	ATG	GAT	ACC	TGC	TCT	AGT	AAC	CTG	AAT	AAC	AGC	1611
Leu	Met	Ala	Ser	Lys	Met	Asp	Thr	Cys	Ser	Ser	Asn	Leu	Asn	Asn	Ser	
			400					405					410			

FIG. 14(E)

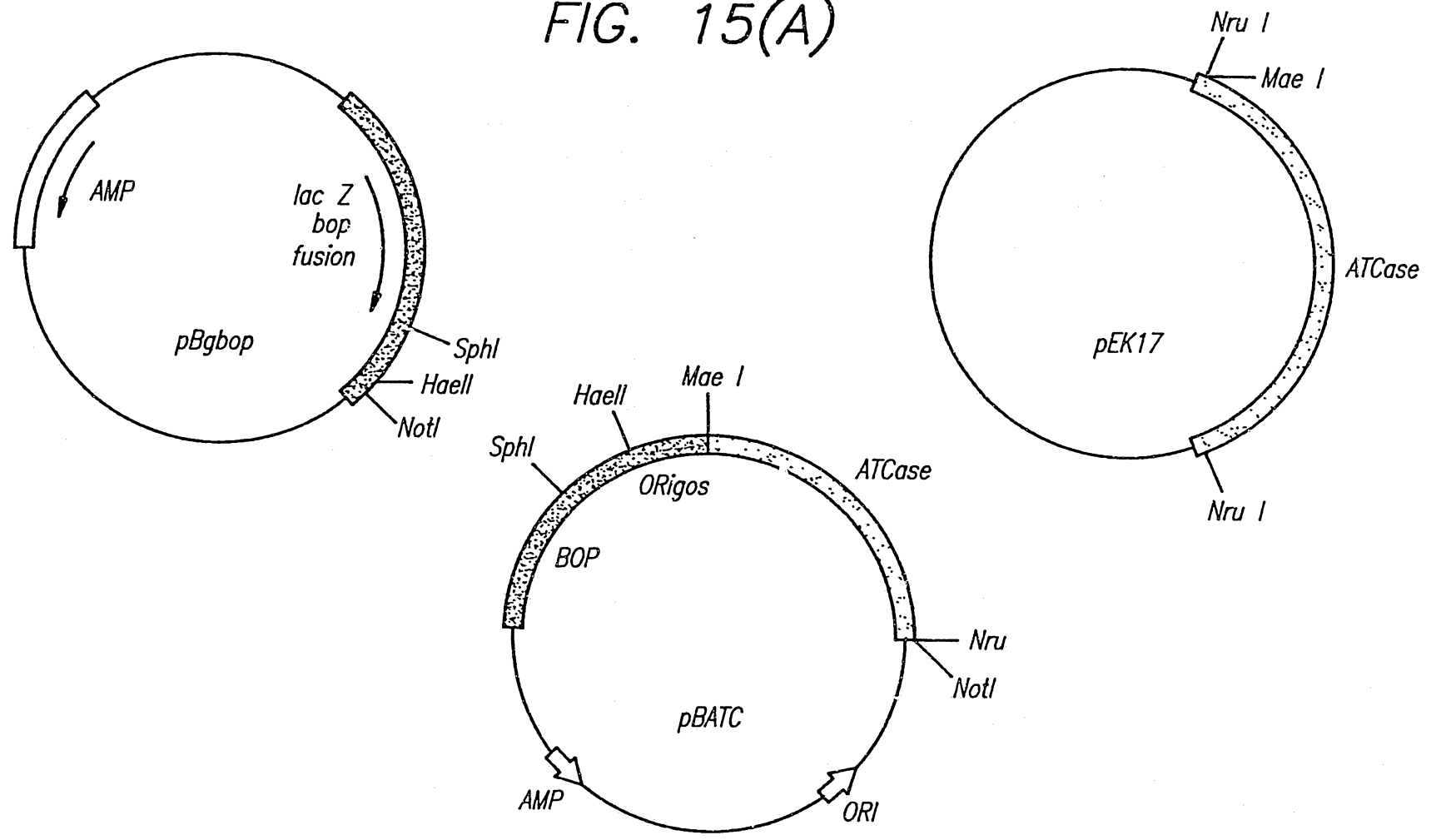
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ATA TAC AAA AAG CTG TTA ACT CAC CAC CAC CAC CAC CAC TGAGCGGCCG	1660
Ile Tyr Lys Lys Leu Leu Thr His His His His His His	
415 420 425	
CGACCAGCGA TTGATCGCAC ACGCAGGACA GCCCCACAAC CGGCGCGGCT GTGTTCAACG	1720
ACACACGATG AGTCCCCCAC TCGGTCTTGT ACTCGGATCC TTTT	1764

FIG. 14(F)



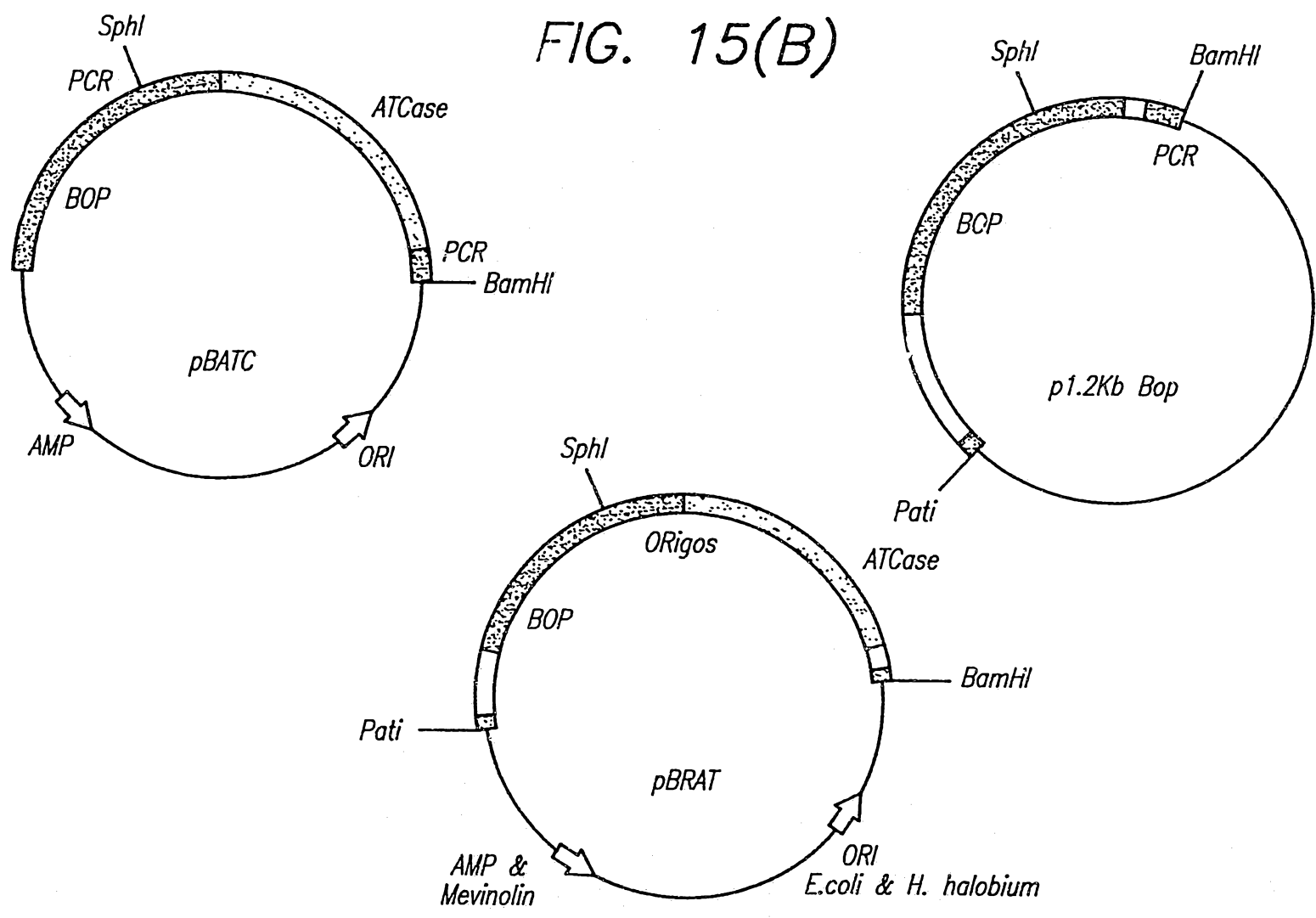
FIG. 15(A)



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FIG. 15(B)



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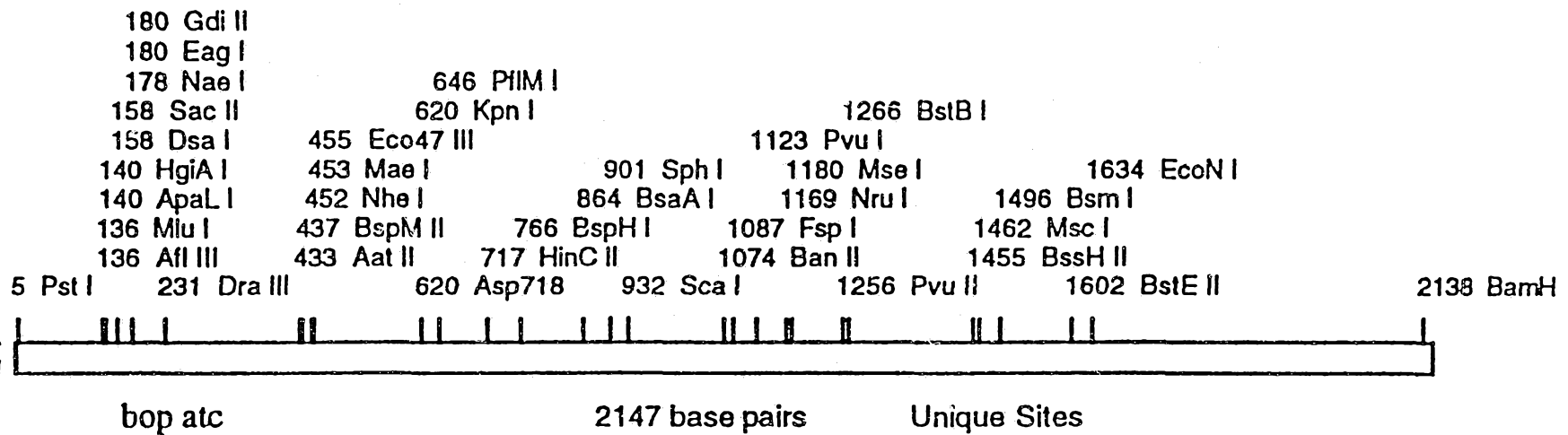


FIG. 16

SUBSTITUTE SHEET (RULE 26)  
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TAATCTGCAG GATGGGTGCA ACCGTGAAGT CCGTCACGGC TCGGTCACGA CAGGAGCCGA	60
CCAGCGACAC CCAGAAGGTG CGAACGGTTG AGTGCCGCAA CGATCACGAG TTTTTCGTGC	120
GCTTCGAGTG GTAACACGCG TGCACGCATC GACTTCACCG CGGGTGTTC GACGCCAGCC	180
GGCCGTTGAA CCAGCAGGCA GCGGGCATT CACAGCCGCT GTGGCCACA CACTCGGTGG	240
GGTGCGCTAT TTTGGTATGG TTTGGAATCC GCGTGTCGGC TCCGTGTCTG ACGGTTTCATC	300
GGTCTAAATT CCGTCACGAG CGTACCATAC TGATTGGGTC GTAGAGTTAC ACACATATCC	360
TCGTTAGGTA CTGTTGC ATG TTG GAG TTA TTG CCA ACA GCA GTG GAG GGG	410
Met Leu Glu Leu Leu Pro Thr Ala Val Glu Gly	
1 5 10	
GTA TCG CAG GCC CAG ATC ACC GGA CGT CCG GAG TGG ATC TGG CTA GCG	458
Val Ser Gln Ala Gln Ile Thr Gly Arg Pro Glu Trp Ile Trp Leu Ala	
15 20 25	
CTC GGT ACG GCG CTA ATG GGA CTC GGG ACG CTC TAT TTC CTC GTG AAA	506
Leu Gly Thr Ala Leu Met Gly Leu Gly Thr Leu Tyr Phe Leu Val Lys	
30 35 40	

FIG. 17(A)

GGG	ATG	GGC	GTC	TCG	GAC	CCA	GAT	GCA	AAG	AAA	TTC	TAC	GCC	ATC	ACG	554
Gly	Met	Gly	Val	Ser	Asp	Pro	Asp	Ala	Lys	Lys	Phe	Tyr	Ala	Ile	Thr	
	45					50					55					
ACG	CTC	GTC	CCA	GCC	ATC	GCG	TTC	ACG	ATG	TAC	CTC	TCG	ATG	CTG	CTG	602
Thr	Leu	Val	Pro	Ala	Ile	Ala	Phe	Thr	Met	Tyr	Leu	Ser	Met	Leu	Leu	
	60				65				70						75	
GGG	TAT	GGC	CTC	ACA	ATG	GTA	CCG	TTC	GGT	GGG	GAG	CAG	AAC	CCC	ATC	650
Gly	Tyr	Gly	Leu	Thr	Met	Val	Pro	Phe	Gly	Gly	Glu	Gln	Asn	Pro	Ile	
				80					85					90		
TAC	TGG	GCG	CGG	TAC	GCT	GAC	TGG	CTG	TTC	ACC	ACG	CCG	CTG	TTG	TTG	698
Tyr	Trp	Ala	Arg	Tyr	Ala	Asp	Trp	Leu	Phe	Thr	Thr	Pro	Leu	Leu	Leu	
			95					100					105			
TTA	GAC	CTC	GCG	TTG	CTC	GTT	GAC	GCG	GAT	CAG	GGA	ACG	ATC	CTT	GCG	746
Leu	Asp	Leu	Ala	Leu	Leu	Val	Asp	Ala	Asp	Gln	Gly	Thr	Ile	Leu	Ala	
		110					115					120				
CTC	GTC	GGT	GCC	GAC	GGC	ATC	ATG	ATC	GGG	ACC	GGC	CTG	GTC	GGC	GCA	794
Leu	Val	Gly	Ala	Asp	Gly	Ile	Met	Ile	Gly	Thr	Gly	Leu	Val	Gly	Ala	
	125					130					135					

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FIG. 17(B)

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CTG	ACG	AAG	GTC	TAC	TCG	TAC	CGC	TTC	GTG	TGG	TGG	GCG	ATC	AGC	ACC	842
Leu	Thr	Lys	Val	Tyr	Ser	Tyr	Arg	Phe	Val	Trp	Trp	Ala	Ile	Ser	Thr	
140							145				150				155	

GCA	GCG	ATG	CTG	TAC	ATC	CTG	TAC	GTG	CTG	TTC	TTC	GGG	TTC	ACC	TCG	890
Ala	Ala	Met	Leu	Tyr	Ile	Leu	Tyr	Val	Leu	Phe	Phe	Gly	Phe	Thr	Ser	
				160						165					170	

AAG	GCC	GAA	AGC	ATG	CGC	CCC	GAG	GTC	GCA	TCC	ACG	TTC	AAA	GTA	CTG	938
Lys	Ala	Glu	Ser	Met	Arg	Pro	Glu	Val	Ala	Ser	Thr	Phe	Lys	Val	Leu	
			175							180					185	

CGT	AAC	GTT	ACC	GTT	GTG	TTG	TGG	TCC	GCG	TAT	CCC	GTC	GTG	TGG	CTG	986
Arg	Asn	Val	Thr	Val	Val	Leu	Trp	Ser	Ala	Tyr	Pro	Val	Val	Trp	Leu	
		190						195						200		

ATC	GGC	AGC	GAA	GGT	GCG	GGA	ATC	GTG	CCG	CTG	AAC	ATC	GAG	ACG	CTG	1034
Ile	Gly	Ser	Glu	Gly	Ala	Gly	Ile	Val	Pro	Leu	Asn	Ile	Glu	Thr	Leu	
	205						210					215				

CTG	TTC	ATG	GTG	CTT	GAC	GTG	AGC	GCG	AAG	GTC	GCC	TTC	GGG	CTC	ATC	1082
Leu	Phe	Met	Val	Leu	Asp	Val	Ser	Ala	Lys	Val	Gly	Phe	Gly	Leu	Ile	
220					225					230					235	

FIG. 17(C)

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CTC	CTG	CGC	AGT	CGT	GCG	ATC	TTC	GGC	GAA	GCC	GAA	GCG	CCG	ATC	GAA	1130
Leu	Leu	Arg	Ser	Arg	Ala	Ile	Phe	Gly	Glu	Ala	Glu	Ala	Pro	Ile	Glu	
				240					245					250		
GGT	CGT	CAG	AAA	CAT	ATC	ATT	TCC	ATA	AAC	GAC	CTT	AGT	CGC	GAT	GAC	1178
Gly	Arg	Gln	Lys	His	Ile	Ile	Ser	Ile	Asn	Asp	Leu	Ser	Arg	Asp	Asp	
			255					260						265		
CTT	AAT	CTG	GTG	CTG	GCG	ACA	GCG	GCG	AAA	CTG	AAA	GCA	AAC	CCG	CAA	1226
Leu	Asn	Leu	Val	Leu	Ala	Thr	Ala	Ala	Lys	Leu	Lys	Ala	Asn	Pro	Gln	
		270					275					280				
CCA	GAG	CTG	TTG	AAG	CAC	AAA	GTC	ATT	GCC	AGC	TGT	TTC	TTC	GAA	GCC	1274
Pro	Glu	Leu	Leu	Lys	His	Lys	Val	Ile	Ala	Ser	Cys	Phe	Phe	Glu	Ala	
	285					290					295					
TCT	ACC	CGT	ACC	CGC	CTC	TCT	TTT	CAA	ACA	TCT	ATG	CAC	CGC	CTG	GGG	1322
Ser	Thr	Arg	Thr	Arg	Leu	Ser	Phe	Gln	Thr	Ser	Met	His	Arg	Leu	Gly	
300					305					310					315	
GCC	AGC	GTG	GTG	GGC	TTC	TCC	GAC	AGC	GCC	AAT	ACA	TCA	CTG	GGT	AAA	1370
Ala	Ser	Val	Val	Gly	Phe	Ser	Asp	Ser	Ala	Asn	Thr	Ser	Leu	Gly	Lys	
				320					325					330		

FIG. 17(D)

AAA	GGC	GAA	ACG	CTT	GCC	GAT	ACC	ATT	TCA	GTT	ATC	AGC	ACT	TAC	GTC	1418
Lys	Gly	Glu	Thr	Leu	Ala	Asp	Thr	Ile	Ser	Val	Ile	Ser	Thr	Tyr	Val	
			335					340					345			

GAT	GCG	ATA	GTG	ATG	CGT	CAT	CCG	CAG	GAA	GGT	GCG	GCG	CGC	CTG	GCC	1466
Asp	Ala	Ile	Val	Met	Arg	His	Pro	Gln	Glu	Gly	Ala	Ala	Arg	Leu	Ala	
		350					355					360				

ACC	GAG	TTT	TCC	GGC	AAT	GTA	CCG	GTA	CTG	AAT	GCC	GGT	GAT	GGC	TCC	1514
Thr	Glu	Phe	Ser	Gly	Asn	Val	Pro	Val	Leu	Asn	Ala	Gly	Asp	Gly	Ser	
	365					370					375					

AAC	CAA	CAT	CCG	ACG	CAA	ACC	TTG	CTG	GAC	TTA	TTC	ACT	ATT	CAG	GAA	1562
Asn	Gln	His	Pro	Thr	Gln	Thr	Leu	Leu	Asp	Leu	Phe	Thr	Ile	Gln	Glu	
380					385				390						395	

ACC	CAG	GGG	CGT	CTG	GAC	AAT	CTC	CAC	GTC	GCA	ATG	GTT	GGT	GAC	CTG	1610
Thr	Gln	Gly	Arg	Leu	Asp	Asn	Leu	His	Val	Ala	Met	Val	Gly	Asp	Leu	
			400						405					410		

AAA	TAT	GGT	CGC	ACC	GTT	CAC	TCC	CTG	ACT	CAG	GCG	TTA	GCT	AAG	TTC	1658
Lys	Tyr	Gly	Arg	Thr	Val	His	Ser	Leu	Thr	Gln	Ala	Leu	Ala	Lys	Phe	
			415					420					425			

FIG. 17(E)

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GAC	GGC	AAC	CGT	TTT	TAC	TTC	ATC	GCG	CCG	GAC	GCG	CTG	GCA	ATG	CCG	1706
Asp	Gly	Asn	Arg	Phe	Tyr	Phe	Ile	Ala	Pro	Asp	Ala	Leu	Ala	Met	Pro	
		430					435					440				

CAA	TAC	ATT	CTG	GAT	ATG	CTC	GAT	GAA	AAA	GGG	ATC	GCA	TGG	AGT	CTG	1754
Gln	Tyr	Ile	Leu	Asp	Met	Leu	Asp	Glu	Lys	Gly	Ile	Ala	Trp	Ser	Leu	
		445				450					455					

CAC	AGC	TCT	ATT	GAA	GAA	GTG	ATG	GTG	GAA	GTA	GAC	ATC	CTG	TAC	ATG	1802
His	Ser	Ser	Ile	Glu	Glu	Val	Met	Val	Glu	Val	Asp	Ile	Leu	Tyr	Met	
					465					470					475	

ACC	CGC	GTG	CAA	AAA	GAG	CGT	CTG	GAC	CCG	TCC	GAG	TAC	GCC	AAC	GTG	1850
Thr	Arg	Val	Gln	Lys	Glu	Arg	Leu	Asp	Pro	Ser	Glu	Tyr	Ala	Asn	Val	
				480					485					490		

AAA	GCG	CAG	TTT	GTT	CTT	CGC	GCC	AGT	GAT	CTC	CAC	AAC	GCC	AAA	GCC	1898
Lys	Ala	Gln	Phe	Val	Leu	Arg	Ala	Ser	Asp	Leu	His	Asn	Ala	Lys	Ala	
			495					500					505			

AAT	ATG	AAA	GTG	CTG	CAT	CCG	TTG	CCG	CGT	GTT	GAT	GAG	ATT	GCG	ACG	1946
Asn	Met	Lys	Val	Leu	His	Pro	Leu	Pro	Arg	Val	Asp	Glu	Ile	Ala	Thr	
		510					515					520				

FIG. 17(F)

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GAT	GTT	GAT	AAA	ACG	CCA	CAC	GCC	TGG	TAC	TTC	CAG	CAG	GCA	GGC	AAC	1994
Asp	Val	Asp	Lys	Thr	Pro	His	Ala	Trp	Tyr	Phe	Gln	Gln	Ala	Gly	Asn	
	525					530					535					
GGG	ATT	TTC	GCT	CTG	CAA	GCG	TTA	CTG	GCA	CTG	GTT	CTG	AAT	CGG	GCC	2042
Gly	Ile	Phe	Ala	Leu	Gln	Ala	Leu	Leu	Ala	Leu	Val	Leu	Asn	Arg	Ala	
540					545					550					555	
GCG	ACC	AGC	GAC	TGATCGCACA	CGCAGGACAG	CCCCACAACC	GGCGCGGCTG									2094
Ala	Thr	Ser	Asp													
TGTTCAACGA	CACACGATGA	GTCCCCACT	CGGTCTTGTA	CTCGGATCCT	TTT											2147

FIG. 17(G)

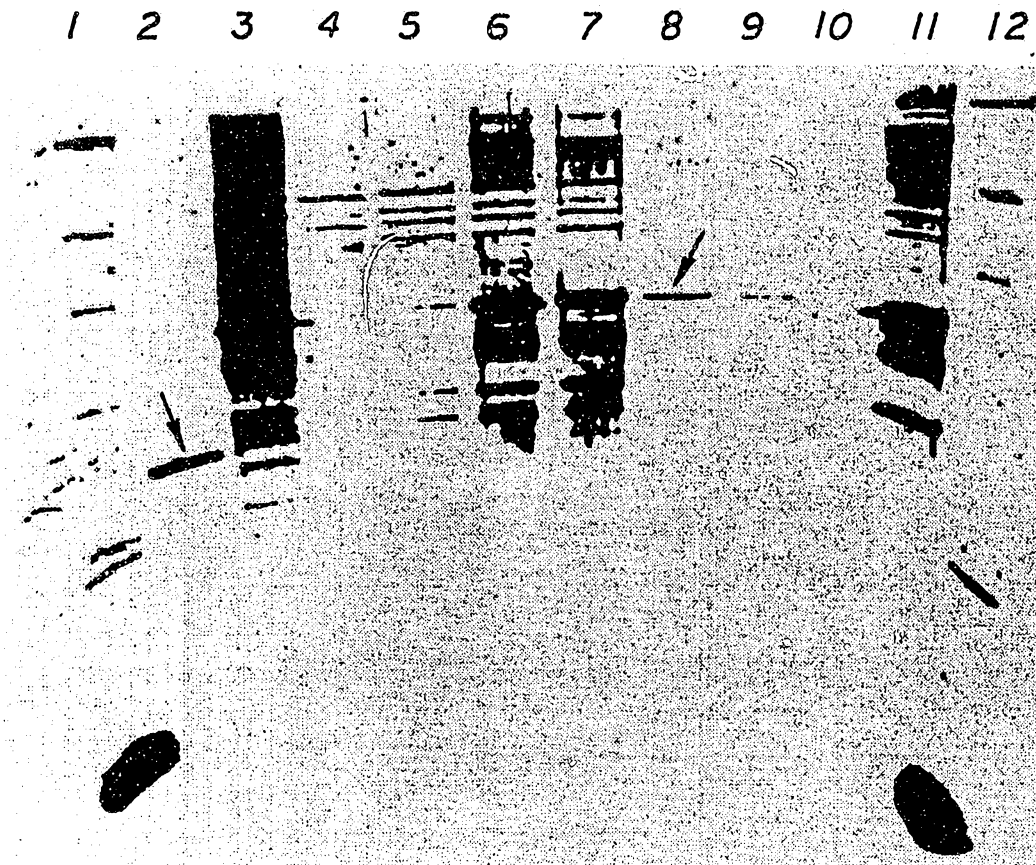


FIG. 18

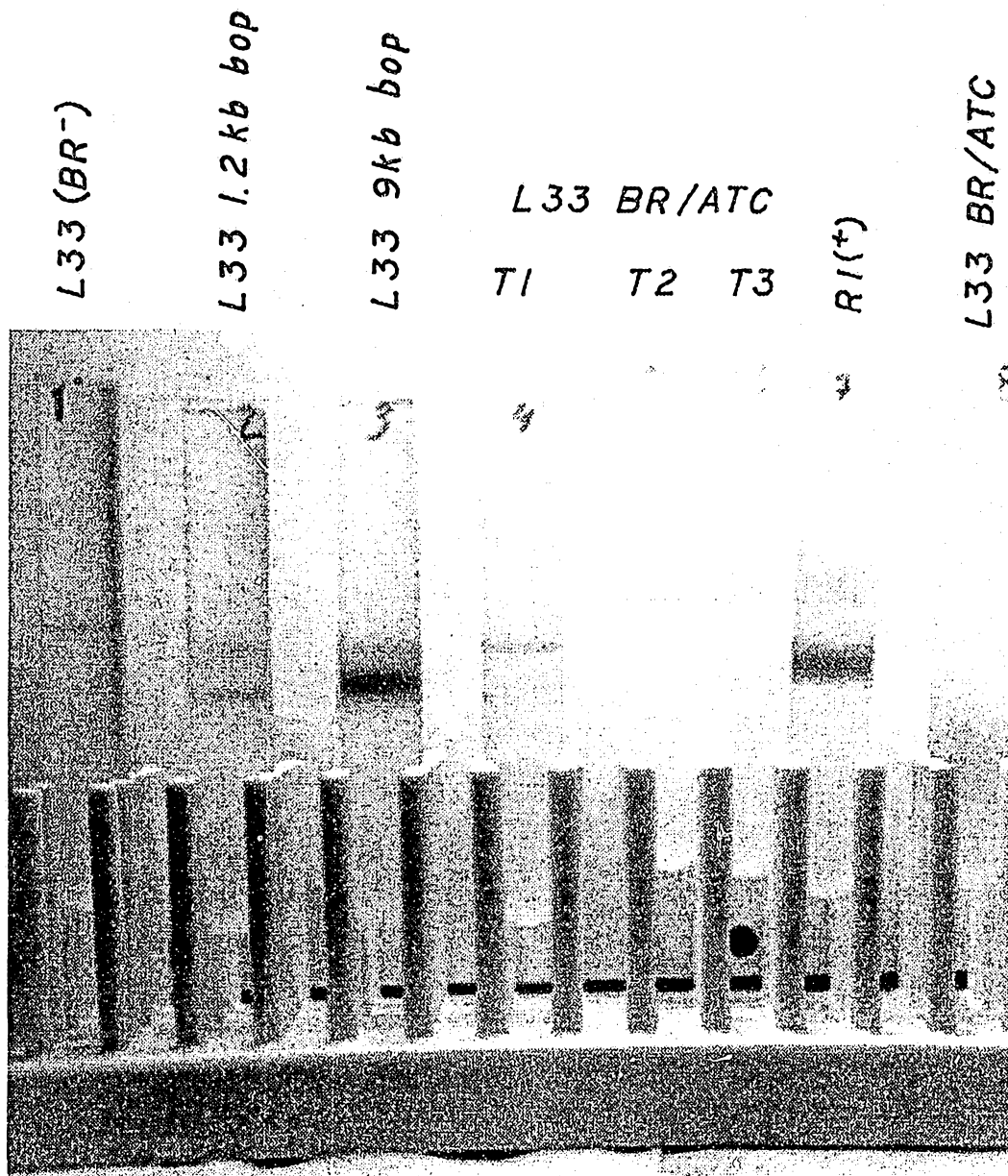
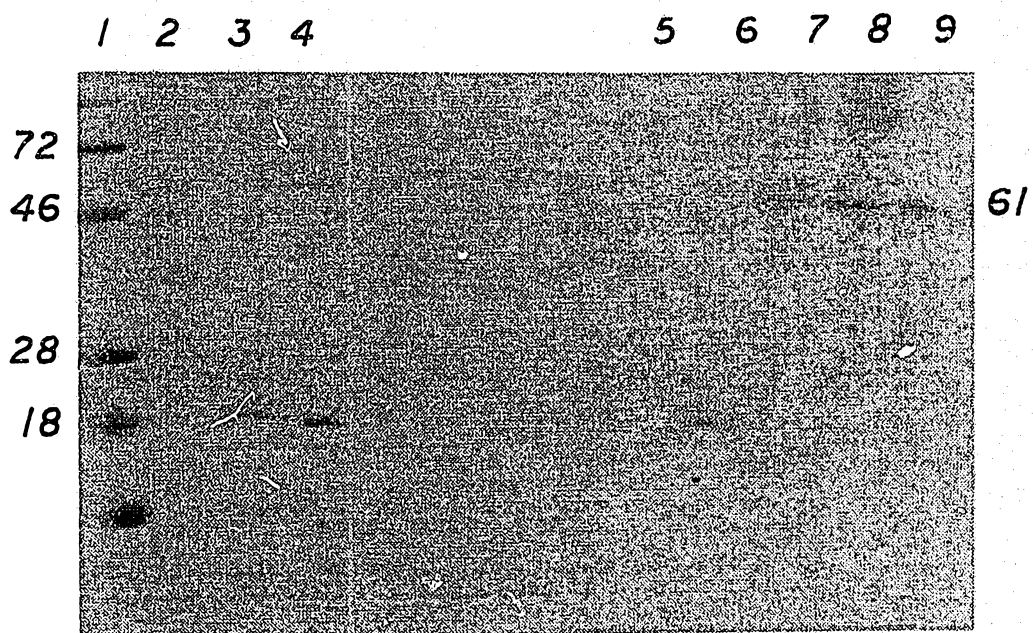


FIG. 19(A)



*FIG. 19(B)*

## INTERNATIONAL SEARCH REPORT

 International Application No  
 PCT/US 94/02388

## A. CLASSIFICATION OF SUBJECT MATTER

C 12 N 15/11, C 12 N 1/21, C 12 N 15/00, C 12 P 21/00

According to International Patent Classification (IPC) or to both national classification and IPC<sup>5</sup>

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

C 12 N, C 12 P

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF THE UNITED STATES OF AMERICA, vol. 87, issued 1990, September U. BLASEIO et al. "Transformation of Halobacterium halobium: Development of vectors and investigation of gas vesicle synthesis", pages 6772-6776, the whole article.	1, 2, 10
A	PATENT ABSTRACTS OF JAPAN, unexamined applications, c field, vol. 8, no. 126, issued 1994, June 13 THE PATENT OFFICE JAPANESE GOVERNMENT,	1, 2, 10

 Further documents are listed in the continuation of box C.

 Patent family members are listed in annex.

## \* Special categories of cited documents:

- \*A\* document defining the general state of the art which is not considered to be of particular relevance
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Date of the actual completion of the international search

09 August 1994

Date of mailing of the international search report

26-08-1994

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INTERNATIONAL SEARCH REPORT

-2-

International Application No  
PCT/US 94/02388

C(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
	<p>page 30 C 228; &amp; JP,A,59-36 700 (MITSUBISHI KASEI KOGYO). -----</p>	