

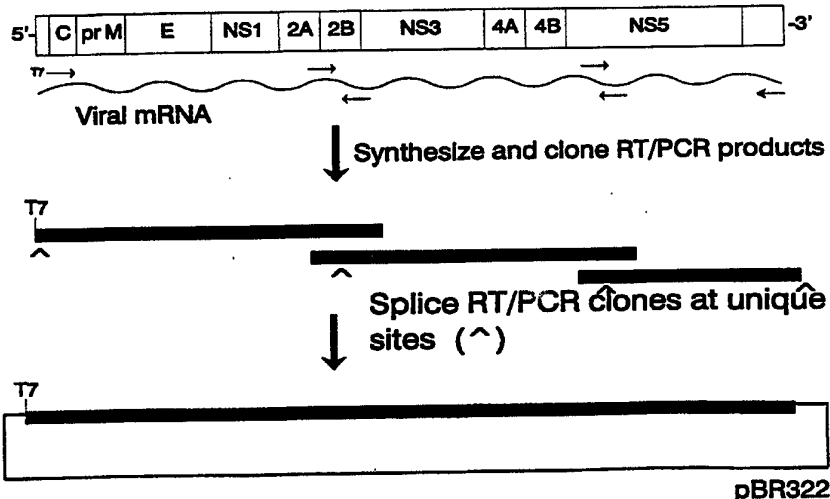


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(54) Title: INFECTIOUS DENGUE 2 VIRUS PDK-53 AS QUADRAVALENT VACCINE

Construction of DEN-2 Infectious cDNA Clone



(57) Abstract

The invention relates to infectious cDNA clones for Dengue 2 virus, strain 16681, and its live, attenuated vaccine derivative, PDK-53 (DEN-2 PDK-53). The invention also relates to infectious cDNA clones for chimeric viruses characterized as expressing structural genes of a Dengue 1, Dengue 3, or Dengue 4 attenuated virus in the context of the nonstructural genes of the Dengue 2 PDK-53 virus (DEN-2/1, DEN-2/3, DEN-2/4). The invention further relates to genetic constructs encoding these cDNAs, and host cells containing these constructs. The invention moreover relates to quadravalent vaccines providing immunity against all four serotypes of dengue virus comprising DEN-2 PDK-53 infectious clone derivative, DEN-2/1, DEN-2/3, or DEN-2/4 viruses, and related methods of immunization.

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INFECTIOUS DENGUE 2 VIRUS PDK-53 AS QUADRAVALENT VACCINE

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Field of the Invention

The invention relates to infectious cDNA clones for Dengue 2 virus, strain 16681, and its live, attenuated vaccine derivative, PDK-53 (DEN-2 PDK-53). The invention also relates to infectious cDNA clones for chimeric viruses characterized as expressing structural genes of a Dengue 1, Dengue 3, or Dengue 4 attenuated virus in the context of the nonstructural genes of the Dengue 2 PDK-53 virus (DEN-2/1, DEN-2/3, DEN-2/4). The invention further relates to genetic constructs encoding these cDNAs, and host cells containing these constructs. The invention moreover relates to quadravalent vaccines providing immunity against all four serotypes of dengue virus comprising DEN-2 PDK-53 infectious clone derivative, DEN-2/1, DEN-2/3, or DEN-2/4 viruses, and related methods of immunization.

Background of the Invention

Arthropod-borne viruses (arboviruses) are a diverse group of viruses that have been lumped together on the basis of their ecological niche, which involves cycles of transmission between vertebrate hosts and arthropod vectors such as mosquitos and ticks. The prototype arbovirus is yellow fever virus, a flavivirus, which was isolated in 1927. In the 1950s, the Rockefeller Foundation established a number of field stations in

various tropical countries for the purpose of isolating new viruses. The 1985 International Catalogue of Arboviruses Including Certain Other Viruses of Vertebrates contains registrations for 504 discrete arboviruses, 124
5 of which have caused disease in humans. Thirty-four viruses of the Flavivirus genus (family Flaviviridae) of arboviruses are human pathogens (Karabatsos, 1985). (All publications cited hereunder are incorporated herein by reference.)

10 According to a 1992 World Health Organization (WHO) press release (Press Release WHO/74, November 24, 1992), dengue hemorrhagic fever is one of the most important and increasing mosquito-transmitted infections in the world, with more than 85 countries in Asia, the Pacific Islands,
15 Africa, Central America, and South America being threatened with dengue outbreaks. Dengue fever was known in the past as "breakbone fever" due to the severe muscular and joint pain that accompanied the high fever during this infection. Dengue is an under-reported
20 disease: it is thought that millions of cases occur each year.

Dengue (DEN) viruses, which are flaviviruses, are classified antigenically into 4 serotypes (DEN-1, DEN-2, DEN-3, and DEN-4). Multiple serotypes are now endemic in
25 most countries in the tropics. DEN viruses are transmitted to humans principally by *Aedes aegypti* mosquitos throughout much of the tropical and subtropical region of the world. Viruses of all four serotypes infect humans and cause clinically inapparent infection or
30 illness ranging from dengue fever to severe and often

fatal dengue hemorrhagic fever/dengue shock syndrome (DHF/DSS). DHF/DSS has been associated epidemiologically and experimentally with immune enhancement of virus replication by preexisting, subneutralizing levels of heterotypic antibody. About 90% or more of patients with DHF/DSS are children who are 14 years old or younger (Halstead, 1970; Halstead, 1988). Case fatality rates in untreated individuals can be as high as 15-20%. Between 1956 and 1978, hospitalization of more than 350,000 dengue patients and about 12,000 deaths in Southeast Asia were reported to the WHO (Halstead, 1980). More recent dengue epidemics in Asia, the Pacific islands, the Americas, and Africa indicate that the incidence, with up to 40 million cases annually, and geographic distribution of the disease is increasing in *Aedes aegypti*-infested areas of the world (Halstead, 1984; Gubler, 1988; Brandt, 1990).

Since eradication of *Aedes aegypti* mosquitos appears to be practically infeasible, development of safe, effective vaccines against all four serotypes of DEN virus is a WHO priority (Gubler, 1988; Brandt, 1988; Brandt, 1990). Since the level of DEN virus replication in certified cell cultures yields insufficient antigenic mass to produce effective inactivated vaccines, priorities are given to developing effective live, attenuated vaccine viruses and using a variety of expression systems such as recombinant vaccinia or avipox virus (live vaccine), recombinant baculovirus (subunit vaccine), and recombinant *E. coli* (subunit vaccine) to express certain genes of the DEN viral genome (Brandt, 1988; Brandt, 1990).

Flaviviruses are enveloped RNA viruses 45 to 50 nm in diameter that contain a single-stranded, positive-sense capped RNA genome of approximately 11 kb. The RNA genome does not have a 3'-terminal poly(A) tail. Because the genetic molecule of flaviviruses is positive or messenger RNA (mRNA)-sense, naked genomic RNA injected, transfected, or electroporated into mammalian or invertebrate cells is capable of associating directly with the ribosomal protein synthetic machinery of the cell. All of the viral proteins are translated from the inserted viral genomic mRNA. These virus-specified proteins then replicate the viral genome, resulting in intracellular virus maturation and release of infectious virus from the transfected cell.

The gene organization of the flavivirus mRNA genome, illustrated below, is 5'-noncoding region (5'-NC)-capsid-premembrane/membrane (prM/M)-envelope (E)-nonstructural protein 1 (NS1)-NS2A-NS2B-NS3-NS4A-NS4B-NS5-3'-noncoding region (3'-NC). The structural proteins capsid, prM/M, and E and nonstructural proteins are translated as a large precursor polyprotein molecule from a single long open reading frame in the mRNA genome. The individual mature viral proteins are processed from the polyprotein by both cell and virus specified proteases (Westaway et al., 1985; Coia et al., 1988; Speight and Westaway, 1989; Rice et al., 1985).

Genome Organization of Dengue Virus and Other Flaviviruses

C	M	E	NS1	2A	2B	NS3	4A	4B	NS5	3'-NC
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The structural proteins are those viral proteins that are incorporated into the mature virion. The virion consists of an icosahedral capsid (C) that packages the viral genomic mRNA (nucleocapsid). The nucleocapsid is surrounded by a cell-derived lipid membrane into which the envelope (E) and mature membrane (M) proteins are imbedded. The virus-specific nonstructural genes, NS1-NS5, are expressed in the cytoplasm of the infected cell and are involved in the replication and maturation of the viral RNA genome and viral proteins.

The E glycoprotein of the virus is exposed to the environment and is involved in attachment and entry of the virus into the cell. The E protein is the primary viral immunogen against which the infected vertebrate host develops virus-specific neutralizing antibody. The E gene is the most common target for development of molecular systems to express the encoded E glycoprotein. However, immunization with various purified nonstructural genes of the virus have been shown to elicit protective immunity against challenge with wild-type virus, probably via cytotoxic T-cell mediated lysis of infected cells which express viral nonstructural proteins on the cell surface.

Vaccination can be one of the most cost effective ways to prevent dengue fever and DHF/DSS. Since 1979 the WHO has supported research on dengue vaccine development at the Mahidol University in Bangkok, Thailand (Press Release WHO/74, November 24, 1992). Investigators at Mahidol University have developed four live, attenuated candidate vaccine viruses, one for each of the four serotypes, by serial passage of the virulent parent

viruses in primary dog kidney (PDK) or fetal rhesus lung (FRhL) cell culture (Yoksan et al., 1986; Bhamarapravati et al., 1987). Phase 1 and Phase 2 clinical trials in Thailand have demonstrated that the vaccine is both safe and immunogenic in humans. The vaccines now need to be tested for efficacy in large numbers of children (Press Release WHO/74, November 24, 1992). To preclude the possible severe DHF/DSS immune enhancement phenomenon in vaccinees who might be infected naturally with a heterologous serotype of wild-type DEN virus following immunization with a single serotype of vaccine virus, it is essential that humans be vaccinated with a quadravalent vaccine to provide immunity against all four serotypes of the virus.

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Summary of the Invention

The invention provides a quadravalent vaccine providing immunity against all four serotypes of dengue virus comprising a DEN-2 PDK-53 infectious clone-derived virus.

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The invention also provides a quadravalent vaccine providing immunity against all four serotypes of dengue virus comprising a chimeric DEN-2/1 virus.

The invention further provides a quadravalent vaccine providing immunity against all four serotypes of dengue virus comprising a chimeric DEN-2/3 virus.

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The invention moreover provides a quadravalent vaccine providing immunity against all four serotypes of dengue virus comprising a chimeric DEN-2/4 virus.

The invention additionally provides a quadravalent vaccine providing immunity against all four serotypes of dengue virus comprising DEN-2 PDK-53 infectious clone-derived and chimeric DEN-2/1, DEN-2/3, and DEN-2/4 viruses.

In another aspect, the invention provides a method of immunization in which a desired immune response is produced against all four serotypes of dengue virus comprising the step of administering to a subject a quadravalent vaccine comprising DEN-2 PDK-53 infectious clone-derived and chimeric DEN-2/1, DEN-2/3, and DEN-2/4 viruses.

In yet another aspect, the invention provides a composition of matter comprising a full genome-length infectious cDNA clone for a DEN-2 virus, strain 16681.

The invention also provides a composition of matter comprising a full genome-length infectious cDNA clone for a DEN-2 virus of a strain characterized as replicating to high titer in cell culture.

The invention further provides a composition of matter comprising a full genome-length infectious cDNA clone for a DEN-2 virus, strain 16681, having the identifying characteristics of ATCC 69826.

In still another aspect, the invention provides a composition of matter comprising a full genome-length infectious cDNA clone for a DEN-2 virus, strain 16681, attenuated derivative, PDK-53.

The invention also provides a composition of matter comprising a full genome-length infectious cDNA clone for a DEN-2 virus attenuated derivative,

characterized as replicating to high titer in cell culture.

The invention further provides a composition of matter comprising a full genome-length infectious cDNA
5 clone for a DEN-2 virus, strain 16681, attenuated derivative, PDK-53, having the identifying characteristics of ATCC 69825.

In another aspect, the invention provides a composition of matter comprising a full genome-length
10 infectious cDNA clone of a chimeric DEN-2/1 virus, wherein the virus is characterized as expressing the prM and E genes of a DEN-1 attenuated virus in the context of the nonstructural genes of the DEN-2 PDK-53 virus. The DEN-1 attenuated virus may be DEN-1 PDK-13.

15 The invention also provides a composition of matter comprising a full genome-length infectious cDNA clone of a chimeric DEN-2 virus, wherein the virus is characterized as expressing the antigenicity of a DEN-1 attenuated virus.

20 In yet another aspect, the invention provides a composition of matter comprising a full genome-length infectious cDNA clone of a chimeric DEN-2/3 virus, wherein the virus is characterized as expressing the prM and E genes of a DEN-3 attenuated virus in the context of the
25 nonstructural genes of the DEN-2 PDK-53 virus. The DEN-3 attenuated virus may be DEN-3 PGMK30/FRhL-3.

The invention also provides a composition of matter comprising a full genome-length infectious cDNA clone of a chimeric DEN-2 virus, wherein the virus is

characterized as expressing the antigenicity of a DEN-3 attenuated virus.

In still another aspect, the invention provides a composition of matter comprising a full genome-length infectious cDNA clone of a chimeric DEN-2/4 virus, wherein the virus is characterized as expressing the prM and E genes of a DEN-4 attenuated virus in the context of the nonstructural genes of the DEN-2 PDK-53 virus. The DEN-4 attenuated virus may be DEN-4 PDK-48.

10 The invention also provides a composition of matter comprising a full genome-length infectious cDNA clone of a chimeric DEN-2 virus, wherein the virus is characterized as expressing the antigenicity of a DEN-4 attenuated virus.

15 Additionally, the invention provides a genetic construct comprising a DNA sequence operably encoding the polyprotein of DEN-2 virus, strain 16681. The polyprotein may be the polyprotein encoded by the nucleotide sequence of SEQ ID NO:1.

20 The invention also provides a genetic construct comprising a DNA sequence operably encoding at least one protein of DEN-2 virus, strain 16681. The protein may be a protein encoded by the nucleotide sequence of SEQ ID NO: 1.

25 Further, the invention provides a genetic construct comprising a DNA sequence operably encoding the polyprotein of DEN-2 virus, strain 16681, attenuated derivative, PDK-53. The polyprotein may be the polyprotein encoded by the nucleotide sequence of SEQ ID
30 NO:2.

The invention also provides a genetic construct comprising a DNA sequence operably encoding at least one protein of DEN-2 virus, strain 16681, attenuated derivative, PDK-53. The protein may be a protein encoded
5 by the nucleotide sequence of SEQ ID NO: 2.

Moreover, the invention provides a genetic construct comprising a DNA sequence operably encoding at least one structural protein of DEN-1 PDK-13. The structural protein may be a structural protein encoded by
10 the nucleotide sequence of SEQ ID NO: 124.

In another aspect, the invention provides a genetic construct comprising a DNA sequence operably encoding at least one structural protein of DEN-3 PGMK30/FRhL-3. The structural protein may be a structural
15 protein encoded by the nucleotide sequence of SEQ ID NO: 125.

In still another aspect, the invention provides a genetic construct comprising a DNA sequence operably encoding at least one structural protein of DEN-4 PDK-48. The structural protein may be a structural protein encoded
20 by the nucleotide sequence of SEQ ID NO: 126.

In yet another aspect, the invention includes a host cell comprising any of the above genetic constructs.

25

Brief Description of the Drawings

Figure 1: Strategy for construction of the full genome-length cDNA clone of DEN-2 virus. Using PCR technology, cDNA is amplified from the genomic RNA of the virus and cloned. Subclones are spliced together at
30 unique, overlapping restriction enzyme sites to construct

the full genome-length clone. Numbered arrows upstream (right arrows) and downstream (left primers used to amplify the cDNA in PCR reactions.

Figure 2: Transcription of genomic mRNA from the full-length infectious cDNA clone of DEN-2 virus. The recombinant plasmid is linearized at the unique XbaI site at the 3'-end of the genomic cDNA. Bacteriophage T7 RNA polymerase recognizes the T7 promoter engineered at the 5'-end of the cDNA and transcribes full-length viral mRNA from the cDNA template.

Figure 3: Restriction enzyme sites identified in the nucleotide sequence of the RNA genome of DEN-2 16681 virus. Locations for the sites are indicated by the genome nucleotide numbers. Restriction enzymes that cleave the DEN-2 genomic cDNA at only a single location are listed vertically at the top of the figure. The resolution of the RENZ graph is 97.5 nucleotides per dot.

Figure 4: Growth curve of DEN-2 16681 virus in C6/36 mosquito cells.

Figure 5: (A) Polaroid prints showing RT/PCR amplification of the entire mRNA genome of DEN-2 virus, strain 16681, in the form of 5 cDNA amplicons. The molecular weight marker (MW) consists of linear, double-stranded DNA markers of various base pair (bp) lengths. The top 2 gels show 5- μ l aliquots of the original RT/PCR reactions. The bottom two gels show 10% of the yield following HMC agarose gel purification of the remaining 95- μ l reaction aliquots. (B) Primers (amplimers) used in the RT/PCR reactions and the expected sizes of the resulting cDNA amplicons.

Figure 6: EcoRI restriction enzyme digests of F2, F2-Sal, Sal-F2, and F3 miniprep recombinant plasmid DNA. Plasmids from individual colonies resulting from transformation with independent ligated, recombinant plasmid molecules are numbered. The insert in the single F2-8 plasmid was too small and was discarded. The remaining recombinant plasmids contained cDNA inserts of expected size. As expected, F2-Sal cDNA contained two internal EcoRI sites; the Sal-F2 and F3 plasmids contained a single internal EcoRI site. EcoRI digestion of the recombinant plasmids regenerated linearized, wild-type 3.9-kb pCRII vector. For an undetermined reason, one of the EcoRI sites in plasmid F3-1 did not cut.

Figure 7: Schematic diagram showing the genomic locations of DEN-2 16681 virus-specific cDNA clones. Clones indicated with asterisks were spliced together at the indicated restriction enzyme sites to construct the full genome-length cDNA clone. Black horizontal bars indicate clone regions that were sequenced. Light gray regions of horizontal bars indicate clone regions that were not sequenced.

Figure 8: (A) Effect of adding Taq extender reagent to PCR reactions. The 5.2-kbp amplicon of St. Louis encephalitis virus was readily obtained by extended PCR (+) but not by standard PCR (-). (B) Agarose gel electropherogram showing DEN-2 PDK-53 F1, F2, and F3 amplicons derived by extended PCR.

Figure 9: Schematic diagram showing the genome locations of errors identified in the cDNA clones of DEN-2 16681. Errors are indicated by short vertical tick marks.

Figure 10: Schematic diagram illustrating the approximate genome locations of the nucleotide discrepancies between the data of Applicants and those of Blok *et al.* (1992) for the sequence of the genome of DEN-2 virus, strain 16681.

Figure 11: Nucleotide sequence of the genome of DEN-2 strain 16681 virus. Differences between the data determined by Blok *et al.* (1992) (DEN-2-16681.BLOK) and those obtained by Applicants (DEN-2-16681.RK). The genome nucleotide positions of the sequence differences are listed vertically. The solid squares indicate those nucleotide differences that also encode amino acid substitutions. The remaining nucleotide differences are either silent, encoding the same amino acid, or lie within the 5'-noncoding (5'-NC) or 3'-noncoding region (3'-NC).

Figure 12: Schematic diagram showing the DEN-2 PDK-53 virus-specific cDNA clones and the approximate locations of cDNA errors (vertical tick marks) identified by nucleotide sequence analyses. Clones marked with an asterisk were used in the construction of the DEN-2 PDK-53 virus-specific full-length cDNA clone. Clone #19 had a 203-bp deletion (horizontal line).

Figure 13: Schematic summary of the DEN-2 16681 vs. PDK-53 virus sequencing projects. Arrows indicate the nucleotide differences detected between the two genomes. Triangles indicate those nucleotide changes that resulted in amino acid substitutions.

Figure 14: Finalized nucleotide and amino acid sequence of the RNA genome of DEN-2 virus, strain 16681 (SEQ ID NO:1). The nucleotide and amino acid mutations

that were determined to have occurred in DEN-2 virus, strain PDK-53, are indicated at the appropriate positions (SEQ ID NO:2). The EcoRI, SstI, MuiI, and T7 promoter sites that were engineered immediately preceding the 5'-
5 terminal nucleotide of the virus-specific genomic cDNA are shown. The start positions of the viral genes and noncoding regions (5'-NC and 3'-NC) are shown. Potential sites of Asn-linked glycosylation (Asn-X-Ser or Thr, where X = any amino acid) in prM, E, and NS1 are indicated by
10 asterisks. The deduced amino acid sequence is indicated in standard single-letter abbreviation: A = Ala, C = Cys, D = Asp, E = Glu, F = Phe, G = Gly, H = His, I = Ile, K = Lys, L = Leu, M = Met, N = Asn, P = Pro, Q = Gln, R = Arg, S = Ser, T = Thr, V = Val, W = Trp, Y = Tyr.

15 **Figure 15:** Construction of intermediate clone F2 by ligating the F2-Sal SphI/HpaI fragment and Sal-F2 HpaI/KpnI fragment into pUC18. The resulting F2 clone contained a nonsilent cDNA error at genome nucleotide position 1730.

20 **Figure 16:** Correction of the intermediate F2 clone. A new PCR amplicon was cloned and sequenced. The SphI/HpaI fragment of this clone was spliced into F2 to construct F2-C having the correct nucleotide at genome position 1730.

25 **Figure 17:** Construction of the intermediate F1/3/4/5 cDNA clone for DEN-2 16681 virus. The thick solid black bars indicate DEN-2 virus-specific cDNA, illustrated with the RENZ sites of the MCS of the plasmid. The RENZ sites used in each step of the splicing strategy are indicated
30 in underlined, bold characters. The top half of the

figure shows construction of F1/3/4/5-pUC18. The bottom portion of the figure illustrates the making of F1/3/4/5-pUC19. The final step in the construction of the full genome-length cDNA clone involved the ligation of the F2-C SphI/KpnI cDNA fragment into plasmid containing cDNA F1/3/4/5 and cut with RENZs SphI/KpnI. Although F2-C cDNA could not be cloned into F1/3/4/5-pUC18, it was readily cloned into F1/3/4/5-pUC19. The pUC18 plasmid containing a small insert of cDNA made for Venezuelan equine encephalitis (VEE) virus was used simply to move F1 and F4/5 into pUC18 in a 3-molecule ligation reaction. The VEE virus-specific cDNA was spliced out during this process. Arrowheads under cDNA bars indicate orientation of mRNA-sense cDNA strand.

Figure 18: Orientation specific cloning of full genome-length cDNA of DEN-2 16681 virus into the multiple cloning site of pUC19. Although the full-length cDNA was readily cloned in pUC19, multiple attempts to insert the cDNA into pUC18 failed. Presumably, interaction of the cDNA with pUC18-specific gene transcripts, translation of a toxic DEN-2 polypeptide, or translation of a toxic pUC18/DEN-2 fusion polypeptide produced deleterious effects in *E. coli*. Large arrows indicate orientation of mRNA-sense cDNA strands in the pUC plasmid backbone. Smaller arrows indicate orientations of the *lac Z* and ampicillin genes as well as the origin of replication. DEN-2 insert is indicated by a thick solid black line.

Figure 19: Insertion of the MCS of plasmid pUC19 into pBR322 in both orientations to construct pBRUC-138 and pBRUC-139. The pUC18 HindIII (blunt-ended = BL)/EcoRI

MCS fragment was ligated into pBR322 cut with *Ava*I (BL)/*Eco*RI to construct pBRUC-138. The pUC18 *Eco*RI (BL)/*Hind*III MCS fragment was ligated into pBR322 cut with *Ava*I (BL)/*Hind*III to make pBRUC-139. In both cases, the tetracycline gene of pBR322 was removed. pBRUC-138 = 2992-bp (61-bp MCS + 2931-bp pBR322 deletion vector). pBRUC-139 = 3022-bp (61-bp MCS + 2961-bp pBR322 deletion vector). Orientations of ORI, ROP, and the Amp gene are indicated.

10 **Figure 20:** Construction of pD2/IC-30P, the full genome-length cDNA clone of DEN-2 16681 virus, in plasmid pBR322 (pBRUC-139 (*Sph*I-) derivative). The F3/4/5 clone cDNA was ligated into pBRUC-139 first (Top of Figure), followed by F1-E and F2-C. Viable, infectious DEN-2 virus was successfully obtained from viral mRNA transcribed from this clone.

Figure 21: Construction of pD2/IC-130V, the full genome-length cDNA clone of DEN-2 PDK-53 virus. A nonsilent error in cDNA clone F3-3C was corrected by splicing in a correct *Bst*BI/*Nhe*I fragment from clone F3.5-6 (Top). The resulting corrected clone F3-3CC was spliced into the 16681 F345-F clone in pBRUC-139. cDNA fragments F1-79B, F2-16B, and the recombinant F3/4/5 vector DNA were spliced together in a single ligation reaction to produce pD2/IC-130V. The *Nhe*I site occurs at genome nucleotide position 6646. Therefore, the PDK-53 virus-specific full-length cDNA clone contains the parental 16681 virus-specific nucleotide at position 8571. This nucleotide difference is silent; it does not encode an amino acid change. Other than the 8571 position, DEN-2 16681 and

PDK-53 viruses are identical in nucleotide sequence from nucleotide position 6646 to the 3' terminus of the genome.

Figure 22: Agarose gel electropherogram of viral genomic mRNA extracted from gradient-purified, wild-type DEN-2 16681 virus and Venezuelan equine encephalitis (VEE) virus. The quantity of RNA loaded onto the gel ranged from 22 ng to 383 ng. The stock RNA was quantitated spectrophotometrically at 260 nm. The genome-length RNA band is clearly visible between the 4153-bp and 6788-bp MW marker bands. Bands were visualized by incorporating 200 ng/ml of ethidium bromide stain in the gel and electrophoresis buffer.

Figure 23: Transcription of RNA from pVE/IC-92 (VEE virus clone) and pD2/IC-20 (DEN-2 16681 virus clone). Transcription reaction conditions (100 ng linearized DNA template, 12.5 mM DTT, 2.7 u/ μ l RNasin, 0.15 mM NTPs, 3.3 U/ μ l T7 RNA polymerase (Stratagene) in commercial buffer (Stratagene)) yielded high quantity and quality of infectious mRNA transcripts from the pVE/IC-92 clone and 3'-end truncation products of that clone. However, these reaction conditions failed to permit transcription of RNA from the pD2/IC-20 clone or two of its 3'-end transcription products (clone linearized at the NsiI or MroI site instead of at the 3'-terminal XbaI site). pVE/IC-92 plasmid linearized at the MluI (3'-terminal), SphI, Tth111I, HindIII, SalI, and StuI sites in the cDNA clone yielded RNA transcripts of 11447, 11377, 7541, 2407, 1620, and 674 base length, respectively (the more intense, prominent bands in these gel lanes).

Figure 24: Transcription of RNA from the DEN-2 16681 cDNA clone pD2/IC-20. (A) Transcription of RNA using different quantities of linearized plasmid template (a,b). The cap analog m7G(5')ppp(5')A was not included in the reaction. (B) Transcription of 5'-capped RNA with inclusion of cap analog in the reaction. Transcription was accomplished with the Ampliscribe transcription kit from Epicentre Technologies. T7 pol = bacteriophage T7 RNA polymerase.

Figure 25: Transcription of full genome-length, infectious viral mRNA from XbaI-linearized DEN-2 16681 plasmid pD2/IC-30P (A and D replicate clones resulting from independent bacterial colonies transformed with the recombinant pBRUC/DEN-2 plasmid) and PDK-53 plasmid pD2/IC-130V (F and J replicates). Genomic "viral RNA" extracted from gradient-purified wild-type DEN-2 16681 virus was electrophoresed in lanes 2 and 10. Aliquots of transcription reactions sampled before (T7 RNA polymerase "-") and after (T7 Pol "+") addition of T7 RNA polymerase are shown. Only the linearized plasmid DNA template is observed in the absence of the polymerase.

Figure 26: Transcription of RNA from pD2/IC-20, pD2/IC-30P, and pD2/IC-130V in the presence or absence of T7 RNA polymerase or cap analog in the transcription reaction. All lanes shown are on a single gel. Transcription was performed with the Ampliscribe transcription kit.

Figure 27: Derivation tree for the construction of the DEN-2 16681 and PDK-53 virus-specific full genome-length cDNA clones pD2/IC-30P and pD2/IC-130V,

respectively, and chimeric 16681/PDK-53 clones derived from the two prototype clones.

Figure 28: Genotype maps of DEN-2 16681 and PDK-53 virus-specific full genome-length cDNAs and their chimeric derivatives. The scale at the top indicates relative genome nucleotide position in thousands. The graph resolution is 119.1444 bp/dot. cDNA regions contributed by the parental DEN-2 16681 virus are indicated by solid black bars. Regions derived from the DEN-2 PDK-53 vaccine virus are indicated by stippled bars. The 8 mutations identified by sequence analyses of the genomes of the 16681 and PDK-53 viruses are indicated. The virus-specific 5-noncoding nucleotides are indicated in lower case characters. The amino acids encoded by the virus-specific nucleotide mutations in the protein coding region of the genome are indicated in upper case, single-letter amino acid abbreviation.

Figure 29: Results of spot-sequencing PCR amplicons amplified from seed stocks of viruses derived from full genome-length cDNA clones. Dots indicate nucleotide sequence identity to the DEN-2 16681 virus. The expected virus-specific nucleotides for the genotype of each virus are shown. Those nucleotide positions that have actually been confirmed by sequence analysis are indicated by underlined nucleotide base characters. The actual genome nucleotide positions are indicated at the bottom of the Figure.

Figure 30: Recombinant full-length pD2/IC-30P-A and pD2/IC-130V-F plasmids extracted from 1-ml aliquots of *E. coli* TB-1 cultures submitted to ATCC.

Figure 31: Partial nucleotide sequences of candidate vaccine viruses:

DEN-1 16007 PDK-13 (D1.VAC) (SEQ ID NO: 124)
 DEN-2 16681 PDK-53 (D2.VAC) (see SEQ ID NO: 2)
 5 DEN-3 16562 PGMK-30/FRhL-3 (D3.VAC) (SEQ ID NO: 125)
 DEN-4 1036 PDK-48 (D4.VAC) (SEQ ID NO: 126)

aligned with the nucleotide and deduced amino acid sequences of DEN-2 16681 virus (see SEQ ID NO:1). Dots in the DEN-1, DEN-3, and DEN-4 sequences signify identity
 10 with the DEN-2 sequence.

Figure 32: Partial amino acid sequences of candidate vaccine viruses:

DEN-1 16007 PDK-13 (D1.VAC) (SEQ ID NO: 124)
 DEN-2 16681 PDK-53 (D2.VAC) (see SEQ ID NO: 2)
 15 DEN-3 16562 PGMK-30/FRhL-3 (D3.VAC) (SEQ ID NO: 125)
 DEN-4 1036 PDK-48 (D4.VAC) (SEQ ID NO: 126)

aligned with the deduced amino acid sequence of DEN-2 16681 virus (see SEQ ID NO:1). Dots in the DEN-1, DEN-3, and DEN-4 sequences signify identity with the DEN-2
 20 sequence.

Figure 33: Mutagenesis analysis of the 5' end of the prM gene. The 447-452 sequence ("AACCAC" in DEN-2) can be mutated to "CTCGAG" in all four DEN viruses to create a XhoI site for cassette splicing. This modification
 25 results in conservative Thr-Thr to Ser-Ser substitutions at amino acid positions prM 4-5 in DEN-2 virus. By creating this XhoI site, all four viruses will contain the sequence FHLSSR at amino acid positions prM 1-6 (see Figure 32). Nucleotide mutations that are necessary to
 30 create the XhoI site are indicated by bold, underlined

characters in the nucleotide sequences of D2.VAC, D1.VAC, D3.VAC, and D4.VAC and their respective primers designed for amplification in PCR.

Figure 34: Mutagenesis analysis of the 3' end of the E gene. The 2344-2349 sequence ("TCACGC" in DEN-2) can be mutated to "TCTAGA" in all four DEN viruses to create a XbaI site for cassette splicing. This modification results in no amino acid change in DEN-2 at this site, but substitutions do occur in the other three viruses. By creating this XhoI site, all four viruses will contain the sequence SRS at amino acid positions E 470-472 (see Figure 32). Nucleotide mutations that are necessary to create the XbaI site are indicated by bold, underlined characters in the nucleotide sequences of D2.VAC, D1.VAC, D3.VAC, and D4.VAC and their respective primers designed for amplification in PCR.

Figure 35: Construction of DEN-2 PDK-53 cassette plasmids pF1-Xho and pF2-Xba. (A) pF1-Xho: Clone PCR cDNA amplicons F1-prM5' and F1-prM3' into TA-vector. Sequence and splice correct clones together at the SphI site in the TA-vector to construct pF1-prM53 (not shown). Subclone the prM53 cDNA into SstI/SphI-cut pF1-E (see Figure 20) to construct pF1-Xho. (B) pF2-Xba: Clone PCR cDNA amplicons F2-E5' and F2-E3' into TA-vector. Splice correct clones together at the XbaI site in the TA-vector to construct pF2-E53 (not shown). Subclone the SphI/HpaI E53 cDNA fragment into pF2-16B (see Figure 21), which itself is subcloned into pBRUC-139 between the SphI/KpnI sites (not shown), to construct pF2-Xho. PCR amplicon designations are underlined. Solid black bars indicate newly

synthesized and sequence-characterized cDNA. Stippled bar indicates previously synthesized cDNA. Graph resolution = 64.1857 nucleotides/dot.

Figure 36: Construction of chimeric plasmids

5 containing the prM and E genes (XhoI-XbaI cDNA fragment) of DEN-1, DEN-3, or DEN-4 candidate vaccine virus within the genetic background of DEN-2 PDK-53 virus. pD2V-CAS12 was constructed by ligating the SstI/SphI fragment of pF1-Xho and SphI/KpnI fragment of pF2-Xba (see Figure 33) into

10 a truncated form of pD2/IC-130V (see Figure 21). pD2/IC-130V was truncated by restricting the full-length clone at the NsiI-4696 and 3'-end XbaI sites, blunt-ending with T4 DNA polymerase, and religating. This procedure removed genome nucleotides 4696-10723, thereby removing the XhoI-

15 5426 and 3'-end XbaI sites, which would otherwise interfere with construction of chimeric plasmid cassettes using XhoI and XbaI sites. The cassette strategy employs PCR amplification of DEN-1, DEN-3, and DEN-4 cDNAs containing the prM and E genes; cutting the amplicons with

20 XhoI/XbaI; cloning resulting fragments into pD2V-CAS12 to construct pD1V-CAS12, pD3V-CAS12, and pD4V-CAS12 chimeric cassettes; confirming the chimeric XhoI/XbaI insert by nucleotide sequence analysis; and then subcloning the SstI/KpnI fragment of the chimeric cassette into pD2/IC-

25 130V to construct the chimeric full genome-length cDNA clones from which chimeric DEN-2/1, -2/3, and -2/4 viruses are derived. The genetic background of DEN-2 PDK-53 virus is illustrated by the solid black bars. The heterologous DEN-1, DEN-3, and DEN-4 cDNA inserts are indicated by the

30 stippled bars. The pBRUC-139 plasmid backbone is not

illustrated for pD1V-CAS12, pD3V-CAS12, or pD4V-CAS12 chimeric plasmid. Resolution = 110.5464 bp/dot.

Detailed Description of the Invention

5 We developed a quadravalent vaccine by initially constructing a full genome-length infectious cDNA clone for DEN-2 virus. We chose serotype 2 of DEN virus because virus strains of this serotype generally replicate to high titer in cell culture. We chose to develop an infectious
10 clone for the 16681 strain of DEN-2 virus because the candidate vaccine viruses developed by Mahidol University are currently the best live, attenuated vaccine virus candidates in terms of immunogenic efficacy and lack of reactogenicity in vaccinees. We developed an infectious
15 cDNA clone of the 16681 strain, which is the parent to the DEN-2 PDK-53 candidate vaccine virus developed at Mahidol University, to permit engineering of second and later generation live, attenuated DEN vaccine viruses.

The infectious clone strategy was initiated with the
20 virulent parental 16681 strain obtained from the Division of Vector-Borne Infectious Diseases (DVBID) of the Centers for Disease Control and Prevention (CDC) virus collection. We synthesized cDNA from the DEN-2 16681 viral RNA. The immediate objective was to obtain an accurate full genome-
25 length infectious cDNA clone of the 16681 strain of DEN-2 virus, since it was essential to develop a reliable experimental system to permit routine genetic engineering of the cDNA and recovery of virus. Our approach involved using polymerase chain reaction (PCR) technology to create
30 cDNA clones that could be spliced together to construct a

single full genome-length clone (Figure 1) from which full-length, infectious DEN-2 genomic mRNA could be transcribed (Figure 2).

The first full-length sequence-characterized cDNA clone, designated pD2/IC-20, was constructed in the high copy number pUC19 plasmid vector. Successful transcription of genome-length DEN-2 16681 viral RNA from pD2/IC-20 was clearly demonstrated by agarose gel electrophoresis of the transcription reaction product.

10 However, RNA transcribed from this particular clone failed to yield infectious virus. It was determined that cDNA errors had occurred during the clone manipulations. We then decided to reconstruct the full-length clone in the low copy number pBR322 plasmid. The full-length cDNA of

15 DEN-2 16681 virus was successfully moved into pBR322 to construct pD2/IC-30P. Full-length, infectious DEN-2 16681 genomic RNA was subsequently transcribed from pD2/IC-30P.

The DEN-1 PDK-13, DEN-2 PDK-53, DEN-3 PGMK-30/FRhL-3, and DEN-4 PDK-48 vaccine viruses were obtained from

20 Mahidol University. Our goal involved replacement of the entire genomic cDNA backbone of the DEN-2 16681 full-length clone with the cognate cDNA cloned from the genome of the DEN-2 PDK-53 candidate vaccine virus. The prM and E genes of the DEN-2 PDK-53 virus are then replaced with

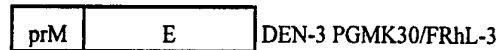
25 the prM and E genes of the DEN-1 PDK-13, DEN-3 PGMK30/FRhL-3, and DEN-4 PDK-48 candidate vaccine viruses to construct chimeric DEN-2/1, DEN-2/3, and DEN-2/4 viruses containing the nonstructural genes of the DEN-2 PDK-53 virus and the prM and E genes of the heterologous

30 DEN viruses.

DEN-2 PDK-53 Infectious cDNA Clone Backbone

	C	M	E	NS1	2A	2B	NS3	4A	4B	NS5	3'-NC
--	---	---	---	-----	----	----	-----	----	----	-----	-------

5



10

It is contemplated that chimeric, infectious clone-derived DEN-2/1, DEN-2/3, and DEN-2/4 viruses will result in immediate improvement in the efficacy of a quadravalent vaccine. Our preliminary data from Mahidol University indicate that very small amounts of the DEN-2 PDK-53 vaccine virus were required to infect and immunize humans. However, the DEN-1, DEN-3, and DEN-4 vaccine virus candidates had approximately 30-fold to 2000-fold lower infectivity for humans. The low infective efficacies of the DEN-1, DEN-3, and DEN-4 viruses create significant problems in terms of vaccine efficacy in eliciting seroconversion in vaccinees, as well as problems of vaccine production for mass vaccination programs, since a large volume, up to 1 ml, of undiluted cell culture-derived vaccine virus must be administered to achieve even minimal levels of infectivity for these viruses. Since the increased infectivity of the DEN-2 PDK-53 vaccine virus is likely due to more efficient virus replication, and since this replicative efficacy is controlled by the nonstructural proteins of the virus, then chimeric vaccine viruses that express the relevant immunogenic structural proteins of DEN-1, DEN-3, or DEN-4 virus in the context of replication control by the nonstructural gene products of

30

the DEN-2 PDK-53 virus should replicate better and be more infective and immunogenic in human vaccinees than the original DEN-1, DEN-3, and DEN-4 vaccine viruses containing nonchimeric genotypes.

5

A quadravalent vaccine is obtained upon completion of the following steps:

- 10 (1) A full genome-length infectious cDNA clone for a DEN-2 virus, strain 16681, is constructed.
- 15 (2) A full genome-length infectious cDNA clone for a DEN2-16681 attenuated derivative, PDK-53, is constructed, preferably by substituting the genomic cDNA backbone of the DEN2-16681 full length clone with the corresponding cDNA cloned from the genome of the DEN-2 PDK-53 candidate vaccine virus.
- 20 (3) The candidate DEN-1, DEN-3, and DEN-4 vaccine viruses are subjected to PCR amplification of cDNA from extracted genomic RNA, and chimeric infectious cDNA clones expressing the prM and E genes of DEN-1, DEN-3, and DEN-4 viruses,
25 respectively, in the context of the nonstructural genes of the DEN-2 PDK-53 virus are constructed.

- (4) The infectious clone-derived chimeric DEN-2/1, DEN-2/3, and DEN-2/4 vaccine viruses are tested to ensure that they:
- 5 (a) Are viable;
- (b) Express appropriate virus-specific immunogens;
- (c) Replicate to sufficient titer in cell culture;
- 10 (d) Are infectious and immunogenic for humans; and
- (e) Retain phenotypic markers of attenuation.

There is no good animal model for investigating dengue pathogenesis. DEN viruses are naturally transmitted between mosquitos and humans. Although lower primates can be infected with these viruses, they do not develop the clinical profiles that occur in humans. Infectious clone-derived viruses can be compared to their more virulent parental strains using certain *in vitro* and *in vivo* markers:

15
20

In Vitro Markers:

- 25 Plaque size in cell culture;
- Temperature sensitivity;
- Cytopathic effects (CPE) in LLC-MK₂ cells; and
- Replication in macrophages.

In Vivo Markers:

- Virulence by intracranial route in mice;
Viremia in monkeys;
5 Virulence by intracranial route in monkeys; and
Elicitation of neutralizing antibodies in
animals.

Infectious cDNA clones are expressed, the resulting
10 RNA transcripts are transfected into permissible cells,
and the live, attenuated viruses are formulated into
vaccines.

Additionally, the DEN-2 PDK-53 and chimeric DEN-
2/1, DEN-2/3, and DEN-2/4 infectious cDNA clones can by
15 themselves confer immunity by DNA immunization, a form of
gene therapy involving the direct inoculation of naked DNA
into the host such that its expression produces an immune
response (e.g., Ulmer et al., 1993 (DNA immunization
protected against influenza); Cox et al., 1993 (DNA
20 immunization protected against herpesvirus); Xiang et al.,
1994 (DNA immunization protected against rabies); Sedegah
et al., 1994 (DNA immunization protected against
malaria)).

Moreover, infectious cDNA clones are exquisite tools
25 for studying the molecular biology of virus structure,
function, and replication. This has been amply
demonstrated for many RNA viruses in the literature,
including Venezuelan equine encephalitis virus as reported
by Kinney et al. (1989). A successful infectious cDNA
30 clone of DEN-2 virus permits important investigations of

dengue virus replication, pathogenesis, and antigenic structure. Infectious clone cDNA templates permit the directed engineering of virus vaccines. Directed site-specific, nonrandom mutations can readily be made in
5 infectious cDNA clones, and therefore in clone-derived viruses, using a wide variety of DNA modification enzymes, restriction endonucleases, and *in vitro* mutagenesis methods. DNA is easier to manipulate than RNA, and the 10^{-9} error rate of DNA replication is much lower than the 10^{-3}
10 - 10^{-4} error rate produced by RNA polymerases. Infectious cDNA clones permit direct analyses of the phenotypic effects of individual and cumulative mutations in the viral genome. An infectious cDNA clone provides a "gold standard" reference sequence for a vaccine.

15

Particular aspects of the invention may be more readily understood by reference to the following examples, which are intended to exemplify the invention, without limiting its scope to the particular exemplified
20 embodiments.

EXAMPLESInformation:

5

Most of the background, protocols, and recipes used in recombinant DNA work can be found in *Molecular Cloning: A Laboratory Manual* (Sambrook et al., 1989), and *Current Protocols in Molecular Biology* (Ausubel et al., 1989).

10

Viruses:

The virulent parental DEN-2 16681 strain was immediately available in the DVBID collection of viruses. We received the DEN-1 PDK-13, DEN-2 PDK-53, DEN-3 PGMK-30/FRhL-3, and DEN-4 PDK-48 vaccine viruses from Mahidol University. The DEN vaccine viruses were passaged in primary dog kidney (PDK) cells because this cell culture is included among those cell types that are certified for human use by the Bureau of Biologics, US Food and Drug Administration (Yoksan et al., 1986). The virus strain designations are shown below:

			Vaccine
		Parent	Derivative
	<u>Virus</u>	<u>Strain</u>	<u>Strain</u>
25	DEN-1	16007	PDK-13
	DEN-2	16681	PDK-53
30	DEN-3	16562	PGMK-30/FRhL-3

DEN-4 1036 PDK-48

PDK = primary dog kidney cells

FRhL = fetal rhesus lung cells

5 PGMK = primary green monkey kidney cells

DEN-1 16007 Parent

- ▶ Recovered from serum of a patient with hemorrhagic fever and shock in Thailand in 1964
- 10 ▶ Passaged 3X in BS-C-1 cells, 1X in LLC-MK₂ cells
- ▶ Passaged 2X in *Toxorhynchites amboinensis* mosquitos
- ▶ PDK-1

↓

PDK-43 Vaccine

15

DEN-2 16681 Parent

- ▶ Recovered from serum of a patient with hemorrhagic fever and shock in Thailand in 1964
- ▶ Passaged 3X in BS-C-1 cells, 1X in LLC-MK₂ cells
- 20 ▶ Passaged 2X in *Toxorhynchites amboinensis* mosquitos
- ▶ PDK-1

↓

PDK-53 Vaccine

25 DEN-3 16562 Parent

- ▶ Recovered from serum of a patient with hemorrhagic fever and shock in the Philippines in 1964
- ▶ Passaged 3X in BS-C-1 cells, 1X in LLC-MK₂ cells
- ▶ Passaged 2X in *Toxorhynchites amboinensis* mosquitos
- 30 ▶ PGMK-1

- ↓
PGMK-30 DEN-3 virus grown in PGMK cells
↓ replicated to very low titer in
PDK FRhL-3 Vaccine cells (Yoksan et al., 1986)
- 5
- DEN-4 1036 Parent
- ▶ Recovered from serum of a patient with dengue fever in Indonesia in 1976
 - ▶ Passed 4X in *Aedes aegypti* mosquitos
- 10 ▶ PDK-1
- ↓
PDK-48

The DEN-2 full-length cDNA clone was derived from the
15 DVVID seed of DEN-2 16681 virus, which had the passage history:

- Human
- 3X BS-C-1 cells
- 20 2X LLC-MK₂ cells
- 2X *T. amboinensis* mosquitos
- 4X C6/36 cells (*Aedes albopictus*)

Complementary DNA (cDNA) was amplified by RT/PCR
25 directly, without further cell culture passage, from virus present in vaccine vials of the DEN-1 PDK-43, DEN-2 PDK-53, DEN-3 PGMK-30/FRhL-3, and DEN-4 PDK-48 viruses.

Stock virus seed was prepared from virus-infected cells grown in 75 or 150 cm² plastic tissue culture flasks.
30 The culture medium was clarified by centrifugation for 30

min at 10,000 rpm in a Sorvall GSA rotor, bringing the final concentration of fetal bovine serum (FBS) to 10% (v/v), and then freezing the clarified virus suspension in aliquots of 0.5 - 1.0 ml at -70°C. Gradient purified DEN-
5 2 16681 virus was prepared according to the method of Obijeski et al. (1976) as reported by Kinney et al. (1983).

Cell Lines:

10

Infectious virus was derived from the infectious cDNA clones by electroporation of BHK-21-15 (baby hamster kidney-21, clone 15) cells with transcribed viral RNA. Viruses were also grown in LLC-MK₂ monkey kidney cells,
15 Vero African green monkey kidney cells, and C6/36 mosquito cells (*Aedes albopictus* C6 cells, clone 36, Igarashi (1978)). All four cell lines were grown in Eagle's minimal essential medium (MEM) supplemented with 10% (v/v) heat-inactivated (56°C for 30 min) FBS, 1.25 g/L of sodium
20 bicarbonate, 100 units/ml of penicillin G, and 100 µg/ml of streptomycin sulfate. Confluent cell monolayers grown in plastic tissue culture flasks were infected by decanting the growth medium, permitting the virus inoculum to adsorb for 1.5 h at 37°C, and then adding MEM
25 containing 5% FBS. For plaque titration of viruses, confluent cell monolayers in plastic 6-well trays were inoculated with 200 µl of the appropriate dilution of virus. Virus was adsorbed to the cell monolayer for 1.5 h at 37 °C. The cells were then overlaid with 3 ml of 1%
30 (w/v) Noble agar (maintained at 40°C) in MEM lacking

phenol red pH indicator and containing 2% FBS and 0.01% (w/v) DEAE-dextran. Following incubation for 6 days at 37 °C in a 5% CO₂ atmosphere, a second 1-ml agar overlay containing 50 µg/ml of neutral red vital stain was added.

5 Viral plaques were counted 2-5 days later.

E. coli:

The *E. coli* K-12 strains used in this project

10 included XL1-Blue, MC-1061, SURE, JM101, and TB-1. Recombinant plasmid containing full genome-length cDNA of DEN-2 virus was successfully replicated in *E. coli* XL1-Blue, MC-1061, and TB-1. Flavivirus cDNA, particularly the gene region encoding the envelope glycoprotein, is

15 troublesome in *E. coli*. Bacteria hosting the recombinant plasmid containing the full-length cDNA clone grew slowly and were often difficult to streak for isolation on agar plates containing selective antibiotic. Transformation efficiencies were sometimes improved somewhat by

20 incubation of agar plates at 30°C or ambient temperature rather than at 37°C. Bacterial stocks were stored frozen at -70°C in 10% (v/v) glycerol.

Precautions for Working with RNA:

25

RNA is a fragile molecule that is very readily degraded by the many ubiquitous RNases present in the environment. Many of these RNases are resistant to treatment with detergents and heat, including autoclaving.

30 All reagents and materials that contacted the viral RNA in

this project were RNase-free to avoid degradation of the viral RNA by these ubiquitous, very stable enzymes. The investigator wore tight-fitting gloves, maintained all reagents on ice, used a plastic tool to open the lids of
5 microtubes, used individually packaged pipets, preferably plastic for aqueous solutions, disposable plasticware which is generally RNase-free before opening, and used "For RNA Only" microtubes, Gilson micropipetors (P-10, P-20, P-100, P-200, P-1000) and tips with aerosol barriers.
10 Use of recycled glassware was avoided. Weigh boats, magnetic stirrers, and pH meters were not used. Chemicals were weighed in sterile, RNase-free disposable plastic 50-ml centrifuge tubes, and solutions were adjusted to the appropriate pH by aliquoting a small volume of the
15 solution onto pH paper. Whenever possible, commercially prepared, guaranteed RNase-free reagents were purchased. Otherwise, newly-opened chemicals were reserved "For RNA Only". Water and stock salt solutions, except for those containing Tris, were treated overnight with 0.1% (v/v)
20 diethylpyrocarbonate (DEPC) to inactivate RNases via alkylation and then autoclaved for 20 min. It is advisable to use the best sterile technique when working with RNA.

25 Extraction of Viral Genomic RNA from Virus Seed:

Virus seeds containing at least 10^6 PFU (plaque forming units)/ml of virus are ideal for providing appropriate yields of RNA. Seed with virus titer of 10^4 or
30 lower can be problematic in terms of yielding sufficient

RNA. For these low-titer seeds it is best to pool the yields of several extracted seed aliquots.

RNA extraction involved the addition of 200 μ l of cold RNA lysis buffer (4 M guanidine isothiocyanate, 25 mM sodium citrate, pH 7.0, 0.5% (w/v) sarkosyl, and 100 mM beta-mercaptoethanol), and 30 μ l of 3 M sodium acetate, pH 5.2, to an empty RNase-free 1.5-ml microtube on ice. In a biosafety cabinet, 200 μ l of DEN virus seed was added to the microtube and mixed vigorously for 30 sec with a mechanical mixer. The tube was centrifuged briefly to pellet the liquid; then 400 μ l of cold phenol (commercially supplied by AMRESCO) equilibrated to pH 4.5 and 80 μ l of cold chloroform were added. The tube was mixed vigorously for 30 sec, placed on ice for 15 min, mixed again, then centrifuged for 1 min at maximum speed in a refrigerated microcentrifuge to separate the aqueous and organic phases. The top aqueous phase containing the extracted RNA was transferred to a fresh 1.5-ml microtube on ice, 400 μ l of cold isopropanol was added, and the tube was incubated for at least 1 h or overnight at -20°C . The RNA was precipitated by centrifugation for 10 min at maximum speed at 4°C . The supernatant was removed with a pipet rather than by decantation and rinsed with 500 μ l of 75% (v/v) ethanol. After spinning again for 10 min, the ethanol was removed with a pipet. The tube was centrifuged again briefly and the residual liquid was removed with a micropipet. The RNA pellet was air dried briefly, resuspended in 50 μ l of cold RNase-free dH_2O , and stored frozen. For seeds containing low virus titer, the

RNA pellets in 3-6 microtubes were pooled in a total volume of 50 μ l.

RT/PCR Synthesis of Dengue Virus-Specific cDNA Fragments

5

Full-length genomic mRNA was extracted directly from 200 μ l of DEN virus seed. The standard reverse transcriptase/polymerase chain reaction (RT/PCR) was performed in a 100- μ l reaction solution containing 5-18 μ l
10 of the extracted viral RNA, 1 μ l each of 100 μ M stock solutions (stored frozen in dH₂O) of the upstream mRNA-sense primer-amplimer and downstream complementary-sense primer-amplimer, 10 μ l of 10X standard PCR buffer (500 mM KCl, 100 mM Tris-HCl, pH 8.5, 15 mM MgCl₂ and 0.1% (w/v)
15 gelatin), 8.0 μ l of 2.5 mM dNTPS (2.5 mM each of dATP, dCTP, dGTP, and dTTP; Pharmacia-LKB), 0.5 μ l of 1 M dithiothreitol (DTT), 0.5 μ l of RNase inhibitor (RNasin, 40 U/ μ l, Boehringer-Mannheim), 0.5 μ l of Taq DNA
polymerase (5 U/ μ l, Perkin-Elmer), and 0.5 μ l of RAV-2
20 reverse transcriptase (18 U/ μ l, Takara). The reaction solution was made as two components:

38

5	▶ PCR Reaction Mix:	10.0 μ l 8.0 μ l 0.5 μ l 0.5 μ l 0.5 μ l	10X Standard PCR Buffer 2.5 mM dNTPs 1 M DTT RNasin (40 U/ μ l) Taq DNA Polymerase (5 U/ μ l)
10		0.5 μ l <u>60.0 μl</u> 80.0 μ l	RAV-2 RT (18 U/ μ l) <u>RNase-Free dH₂O</u> Reaction Mix for 1 reaction. Make more than needed for all reaction tubes. Store excess at -70°C for reuse.
15	▶ Template/Primer Mix:	18.0 μ l 1.0 μ l <u>1.0 μl</u> 20.0 μ l	DEN-2 RNA Template 100 μ M Up-Amplimer <u>100 μM Down-Amplimer</u>
20	▶ Reaction Solution:	80.0 μ l <u>20.0 μl</u> 100.0 μ l	PCR Reaction Mix <u>Template/Primer Mix</u> In a thin-walled, 200- μ l microtube.

25

The RT/PCR reactions in thin-wall 200- μ l microtubes (Phenix Research Products) were incubated without oil overlay in a Perkin-Elmer Model 9600 thermocycler according to the following program:

30

35

40

50 °C for 60 min = First strand cDNA synthesis
by reverse transcriptase

94°C for 4 min

50°C for 1 min

72°C for 5 min

94°C for 30 sec

55°C for 30 sec

72°C for 5 min

Delta +10 sec/cycle

30 Cycles

Following completion of the RT/PCR reactions, 5- μ l aliquots of each of the 100- μ l reactions were analyzed by agarose gel electrophoresis. The DNA bands in the agarose gel were stained in ethidium bromide (500 ng/ml) solution and visualized on an ultraviolet light box. Since extraneous non-target cDNA bands are often amplified in addition to the target cDNA molecules, the remaining 95 μ l of each RT/PCR reaction was electrophoresed in a larger, preparative agarose gel, and the target cDNA was stained briefly, excised with a razor blade, and physically extracted from the agarose slice.

High-Melt-Crush (HMC) Extraction of DNA from Agarose:

An agarose gel slice containing DNA was placed in a 1.5-ml microtube and crushed thoroughly with a spatula or pestle. The volume of the crushed agarose was brought to 400-500 μ l with TE buffer (10 mM Tris-HCl, pH 7.5, 1 mM disodium EDTA) and 400 μ l of phenol (supplied by AMRESCO), pH 8, was added. The agarose suspension was mixed vigorously using a mechanical mixer, frozen, thawed and mixed, frozen, thawed and mixed, and then centrifuged for 10 min at maximum speed at 4°C. The top aqueous phase was transferred to a fresh microtube, extracted with 400 μ l of phenol:chloroform:isoamyl alcohol (25:24:1) and centrifuged for 2 min. The top aqueous phase was transferred to a fresh tube and extracted with 700 μ l of diethyl ether or chloroform. If chloroform was used, the top phase was again transferred to a fresh tube after a brief spin to separate phases. The DNA was precipitated

for at least 30 min at -70°C or overnight at -20°C following addition of 2.5 volumes (essentially filling the microtube) of 95% ethanol containing 300 mM ammonium acetate and 10 mM MgCl₂. The DNA was pelleted at 4°C by
5 centrifugation for 20 min at maximum speed. The liquid was decanted, and the DNA pellet was rinsed with 500 µl of 75% ethanol, air-dried briefly, dissolved in 30 µl of TE buffer, and stored frozen or in the refrigerator. A 3-µl
10 aliquot of the extracted DNA was analyzed for purity and quantity by agarose gel electrophoresis. Generally, 20-80% of the DNA loaded onto a gel can be recovered from the gel by this method.

Agarose Gels:

15

DNA was analyzed by electrophoresis in 1% (w/v) agarose gels run in TBE buffer (100 mM Tris-HCl, pH 8, 91 mM boric acid, and 20 mM disodium EDTA). DNA bands were visualized by staining the gel in water containing 500
20 ng/ml of ethidium bromide and exposure to ultraviolet light. Gels used for analyzing RNA transcripts were made with RNase-free reagents. Ethidium bromide stain was incorporated in the gel and running buffer so that the RNA bands could be visualized immediately. To obtain gel-
25 purified DNA fragments, DNA was electrophoresed in 0.7% (w/v) agarose gels made with genetic technology grade Seakem agarose (FMC) or with biotechnology grade agarose (3:1 high resolution blend, AMRESCO).

Cloning of Dengue Virus-Specific cDNA Fragments:

Some DNA polymerases add an extra "A" nucleotide
5 overhang at the 3'-end of synthesized DNA strands. The
Taq DNA polymerase does this. To enable the cloning of
DNA molecules synthesized using Taq DNA polymerase, TA-
cloning vectors have been engineered (Marchuk *et al.*,
1991). These vectors generally have a single "T" overhang
10 engineered at the 3'-terminus of EcoRV-cut, blunt-ended,
linearized plasmid vector. The EcoRV site occurs within
the multiple cloning site (MCS) of the plasmid. The MCS
is a series of contiguous, unique restriction enzyme
(REnz) sites engineered into a vector plasmid to permit
15 subcloning of exogenous DNA fragments following
restriction with a variety of RENzS. The HMC-purified DEN
cDNA amplicons were cloned into the 3900-bp pCRII
(Invitrogen), the 2887-bp pT7Blue(R) (pT7Blue, Novagen),
or the 3003-bp pGEM-5Zf (Promega) TA-vector plasmid. The
20 RENz sites available in the MCS region of these TA-
vectors, as well as the RENz sites of the MCS of the
general purpose cloning plasmids, pUC18 and pUC19, used in
this project are shown below.

RENZ Sites Present in the MCS of Several Cloning Vectors

	<u>pUC18</u>	<u>pUC19</u>	<u>pT7Blue</u>	<u>pCRII</u>	<u>pGEM-5Zf</u>
5			T7	SP6	T7
	EcoRI	HindIII	HindIII	NsiI	ApaI
	SstI	SphI	BspMI	HindIII	AatII
	KpnI	PstI	SphI	KpnI	SphI
10	SmaI	SalI	PstI	SstI	NcoI
	BamHI	XbaI	Sse8387I	BamHI	SstII
	XbaI	BamHI	SalI	SpeI	<u>EcoRV</u>
	SalI	SmaI	AccI	BstXI	SpeI
	PstI	KpnI	HincII	EcoRI	NotI
15	SphI	SstI	XbaI	<u>EcoRV</u>	PstI
	HindIII	EcoRI	SpeI	EcoRI	SalI
			NdeI	PstI	NdeI
			<u>EcoRV</u>	BstXI	SacI
			BamHI	NotI	BstXI
20			AvaI	AvaI	NsiI
			SmaI	SphI	SP6
			KpnI	NsiI	
			SacI	XbaI	
			BanII	ApaI	
25			EcoRI	T7	

The pUC18/19 plasmids possess identical MCS sites in reverse orientation in the plasmid backbone. Their purpose is to permit cloning of DNA in either orientation into the plasmid using the same pair of RENZs - this

reversibility was exploited in this project. The TA-vectors used here all possessed T7 and/or SP6 bacteriophage RNA promoters to enable RNA transcription from cloned DNA. These promoters were not used in this project. All of the plasmids contain the gene for ampicillin resistance. They also contained the *lac Z* portion of the *E. coli lac* operon. This permits color discrimination between bacterial colonies that receive a recombinant or a wild-type plasmid. In the presence of IPTG and X-gal, bacterial colonies that are transformed with a wild-type plasmid lacking a cDNA insert develop a blue color, whereas cells that receive a recombinant plasmid with cDNA cloned into the MCS of the plasmid are white. Agar plates contained 800 μ g of IPTG and 800 μ g of X-gal.

Fifty to 100 ng of HMC-purified amplicon was ligated to 50 ng of the pCRII vector using the TA-vector cloning kit supplied by Invitrogen exactly as specified by the instructions supplied with the kit. Frozen, transformation competent *E. coli* INV α F' cells, supplied with the Invitrogen kit and stored at -70°C, were transformed with the ligated DNA as described in the kit instructions. The transformed cells were plated on YTA₅₀ agar plates (8 g of DIFCO tryptone, 5 g of DIFCO yeast extract, 5 g of NaCl, and 15 g of BACTO agar per liter of dH₂O) containing 50 μ g/ml of ampicillin. Only bacterial cells transformed with the pCRII plasmid, which contains an ampicillin resistance gene, grow on this medium. The agar plates were incubated at 37°C overnight.

Similarly, cDNA was ligated to the other TA-vectors or to pUC18/19 cut with the appropriate RENZ(s). Ligations were performed at room temperature or at 12°C. *E. coli* XL1-Blue, SURE, TB-1, or MC-1061 cells were transformed by electroporation and plated on YTA₅₀ plates. Electroporation was performed according to Dower et al. (1988) using cuvettes with a 2-cm electrode gap in a Bio-Rad Gene Pulser set at 2.5 kV voltage, 25 μF capacitance, and 200 ohms resistance. Electroporation-competent cells were prepared by growing a fresh bacterial culture to an optical density of 0.5-0.7 at 600 nm. The cells from 1.5 - 3 L of culture were pelleted by centrifugation for 10 min at 4°C and 5000 rmp in a Sorvall GSA rotor, pooled, washed twice in 1 mM Hepes buffer, and resuspended in 2 ml of 10% (v/v) sterile glycerol per L of original culture. The concentrated cells in glycerol were stored at -70°C.

Bacterial colonies were transferred to 2 ml of 2XYT-Amp₅₀ broth (16 g of tryptone, 20 g of yeast extract, and 5 g of NaCl per liter of dH₂O) and incubated overnight with shaking at 300 rpm at 37°C in a floor model incubator - shaker (model Innova 4300, New Brunswick). Recombinant plasmid was extracted from these 2-ml minicultures and analyzed by agarose gel electrophoresis for the presence of cDNA insert. Recombinant plasmids are larger than wild type vector plasmid because of the cDNA insert, and they migrate more slowly than wild type plasmid in agarose gels.

All of the DEN-2 16681 virus-specific cDNA amplicons were cloned into the pCRII TA-vector. Aliquots of insert-positive miniprep plasmids were digested with the

restriction enzyme EcoRI. Since the pCRII MCS contains two EcoRI recognition sites (palindromic hexameric sequence GAATTC) on either side of the EcoRV cDNA cloning site, this RENZ cleaved the cDNA insert from the plasmid vector and cleaved any EcoRI sites that were present within the cDNA itself. The EcoRI-restricted DNA was analyzed by agarose gel electrophoresis to determine that the cloned cDNA was of appropriate size. In our experience, cloning of PCR-derived cDNA amplicons 2000 bp or smaller in size into the TA-vector is efficient. Cloning amplicons larger than 3500 bp into the TA-vector can be very difficult.

After screening, certain of the miniprep plasmids were selected for further analysis. Their corresponding bacterial minicultures were streaked for isolation on YTA₅₀ plates, and an isolated colony was inoculated into 50-200 ml of YTA₅₀ broth to grow up a preparative amount of recombinant plasmid. The preparative scale for the extraction of the plasmid was essentially identical to that for minipreps except for scaled up volumes.

Extraction of Plasmid DNA from Minicultures of *E. coli*:

White colonies containing recombinant plasmid were picked with a sterile toothpick and shaken overnight at 300 rpm in 2 ml of 2X-YTA₅₀ broth. Each miniculture was decanted into a 1.5-ml microtube, and the cells were pelleted by centrifugation at 6000 rpm for 2 min. The supernatant was aspirated, and the cell pellet was resuspended gently by up/down micropipeting in 200 μ l of

GTE buffer (50 mM glucose, 25 mM Tris-HCl, pH 8.0, and 25 mM disodium EDTA) and then mixed with 300 μ l of lysis buffer (0.2 N NaOH, 1% (w/v) sodium dodecylsulfate (SDS)). After incubation on ice for 5 min, 300 μ l of cold

5 potassium acetate solution (3 M potassium acetate, 7 M acetic acid, pH 4.8) was added, and the solution was chilled for 5 min on ice and then centrifuged at maximum speed for 10 min at 4°C. The supernatant was poured into a fresh microtube, RNase A was added to 20 μ g/ml, and the

10 mixture was incubated at 37°C for 30 min. The sample was extracted twice with 600 μ l of chloroform and centrifuged for 1 min at maximum speed at room temperature. The DNA pellet was dissolved in 32 μ l of dH₂O. Eight μ l of 4M NaCl and 40 μ l of 13% (w/v) PEG-8000 was added, and the mixed

15 solution was incubated for 5 min on ice. The sample was centrifuged for 15 min at maximum speed at 4°C, the liquid was aspirated with a micropipet , and the pellet was rinsed with 500 μ l of 75% ethanol. The air dried pellet was dissolved in 30 μ l of dH₂O and stored frozen until

20 used.

Extraction of Plasmid DNA from Large Cultures of *E. coli*:

Preparative-scale plasmid extraction was performed by

25 inoculating 100 ml of 2X-YTA₅₀ broth with 2 ml of an overnight culture of *E. coli*. The culture was shaken overnight at 300 rpm and 37°C. The cells were pelleted by centrifugation for 10 min at 5000 rpm in a Sorvall GSA rotor and resuspended in 6 ml of cold GTE buffer. Nine ml

30 of a freshly made solution of 0.2 N NaOH and 1% (w/v) SDS

was added. The sample was incubated for 5 min on ice, then 9 ml of cold 3 M potassium acetate solution was added. After another 5-min incubation on ice, the tube was centrifuged for 20 min at 10,000 rpm at room temperature and the supernatant was transferred to a fresh 30-ml glass tube. RNase A was added to 20 $\mu\text{g}/\text{ml}$, and the sample was incubated for 30 min at 37°C and then extracted twice with 6 ml of chloroform. Twelve ml of room-temperature isopropanol was added and the tube was centrifuged immediately for 20 min at 10,000 rpm at room temperature. The supernatant was decanted, and the DNA pellet was rinsed with 1 ml of 75% ethanol, air dried briefly, and resuspended in 480 μl of dH_2O . The DNA was precipitated by addition of 120 μl of 4 M NaCl and 600 μl of 13% PEG-8000, incubation for 5 min on ice, and centrifugation for 15 min at maximum speed at 4°C. The DNA pellet was rinsed with 500 μl of 75% ethanol, air dried briefly, rehydrated in TE buffer, and stored frozen.

20 **Nucleotide Sequence Analysis of the Dengue cDNA Clones:**

Nucleotide sequence analyses of DEN-2 16681 cDNA clones #1-#15 were performed by cloning EcoRI restriction fragments of each clone into the single-stranded bacteriophage M13mp18 or M13mp19. Since this is not the current method of choice for sequencing, the method will be described only briefly here. The procedure used for the extraction of plasmid DNA from bacterial cells was also used to extract the intracellular double-stranded replicative form (RF) DNA of M13 from bacteriophage-

infected *E. coli* JM101 cells. The RF DNA was linearized at the EcoRI site of the MCS and ligated to the DEN-2 HMC-purified EcoRI cDNA restriction fragments.

Electroporation-competent *E. coli* JM101 cells were
5 transformed by electroporation and plated onto H-agar plates (10 g of DIFCO tryptone, 5 g of NaCl, 15 g of BACTO agar, and 1% (w/v) thiamine per liter of dH₂O) containing 800 µg each of isopropyl-β-D-galactopyranoside (IPTG) and 5-bromo-4-chloro-3-indolyl-β-D-galactopyranoside (BCIG or
10 X-gal). The electroporated cells were mixed with 300 µl of a fresh logarithmic culture of JM101 cells and 3 ml of warm (51°C) top H-agar containing 9 g/L of agar and then poured onto the H-agar plates. Cells that were transfected with recombinant DNA supported replication of
15 recombinant M13 virus, resulting in the formation of bacteriophage plaques in the JM101 cell lawn on the agar plate. The IPTG/BCIG histochemistry of the system permitted identification of white plaques containing recombinant bacteriophage into which cDNA had been ligated
20 into the EcoRI site of the MCS, whereas wild-type nonrecombinant M13 bacteriophage produced blue plaques. Isolated plaques were picked, inoculated into 3 ml of a fresh, pre-logarithmic phase culture of JM101, and shaken at 37°C for 8-16 h. The minicultures were clarified by
25 centrifugation in 1.5-ml microtubes, the bacteriophage particles were precipitated with PEG-8000, and the single-stranded, circular bacteriophage DNA was isolated from the virions by phenol extraction. The recombinant, circular, single-stranded bacteriophage DNA was sequenced by the
30 dideoxynucleotide termination method. Sequencing kits can

be purchased from various commercial vendors. Radioactive ^{32}P -dCTP or ^{35}S -dCTP was incorporated into the strands synthesized in the sequencing reactions. Sequencing was accomplished with many DEN-2 virus-specific primers
5 designed to sequence the entire genome. The sequence reactions were electro-phoresed in 6% (w/v) polyacrylamide gels, which were dried onto filter paper and overlaid with X-ray film. The DNA bands of the autoradiographs were read by the investigator, and the data was entered into a
10 sequence project data spreadsheet. This sequencing method has been used extensively in the past (e.g., Kinney et al., 1986; Johnson et al., 1986; Deubel et al., 1986; Deubel et al., 1988; Kinney et al., 1989; Trent et al., 1987).

15 Nucleotide sequencing was also performed by the current method of direct sequencing of double-stranded plasmid DNA by the dideoxynucleotide termination method using the Applied Biosystems Taq DyeDeoxy Terminator Cycle Sequencing Kit, cycle sequencing in the Model 9600
20 thermocycler according to the instruction manual supplied with the kit, and analyzing the DNA sequence on an ABI Model 373A DNA sequencing apparatus. Sequencing reactions in 200- μl thin-walled microtubes contained 9.5 μl of reaction mix (buffer, the four dideoxynucleotides, and Taq
25 polymerase supplied in the kit), 7.0 μl of double or single-stranded template DNA (150 pg/bp), and 3.2 μl of 10 μM sequencing primer (32 pmol). After mixing, the reactions were placed in a Perkin-Elmer Model 9600 thermocycler, and programmed cycle sequencing was
30 performed for 25 cycles of incubation at 96°C for 15 sec,

50°C for 15 sec, and 60°C for 4 min. Strand extension was performed at 60°C rather than 72°C because the fluorescent dye-labeled dideoxynucleotide terminators are heat sensitive. The reaction was then applied to a Centrisep gel column (Princeton Separations) to remove unincorporated dye-labeled dideoxynucleotides according to the instructions supplied with the columns. The eluted DNA was vacuum dried for 1 h using a Savant Speed Vac Concentrator and stored at -70°C. The DNA was hydrated with 5 μ l of deionized formamide and 1 μ l of 50 mM disodium EDTA, then heated in an aluminum block for 2 min at 90°C. A 3- μ l aliquot of the denatured DNA sample was applied to one of 24 wells of a polyacrylamide-urea gel in an Applied Biosystems 373A DNA sequencer. The color-coded sequence chromatograph was read by visual inspection, and the resulting nucleotide sequence was entered into a computer-maintained sequence data spreadsheet. The sequencing kit incorporates dideoxynucleotide terminators that are each labeled with a unique fluorescent dye that permits laser detection of all four terminators in a single polyacrylamide gel lane in the Model 373 sequencer. The data was recorded in the form of colored chromatograms that are easily read by the investigator. Single-stranded recombinant M13 DNA can also be sequenced in this manner.

25

Extraction of M13 Single-Stranded DNA for Sequencing:

White bacteriophage plaques containing recombinant M13 DNA were picked with sterile toothpicks and placed into 2-ml slightly turbid (less than 0.15 A_{600}) cultures of

30

E. coli JM101. The cultures were shaken at 300 rpm and 37°C overnight and then clarified by centrifugation in microtubes at maximum speed for 10 min at room temperature. One ml of the supernatant was transferred to a fresh 1.5-ml microtube containing 200 µl of sterile 20% (w/v) PEG-8000 in 250 mM NaCl. The tubes were mixed by inversion, incubated for 15 min at room temperature, and centrifuged at maximum speed for 5 min at room temperature. The PEG supernatant was removed completely, and the DNA pellet was resuspended in 300 µl of TE buffer. An equal volume of pH 8-buffered phenol was added, and the solution was mixed vigorously several times during a period of 20 min at room-temperature. The tube was centrifuged for 5 min at room temperature, and the top aqueous phase was transferred to a fresh 1.5-ml microtube. After sequential extraction with phenol:chloroform:isoamyl alcohol and chloroform, the DNA was precipitated by adding 2.5 volumes of 95% ethanol containing 300 mM ammonium acetate and 10 mM MgCl₂, and incubating at -20°C overnight. The tube was centrifuged at maximum speed for 15 min at 4°C, and the supernatant was decanted. Following a rinse with 500 µl of 75% ethanol, the DNA was air dried briefly, resuspended in 60 µl of TE buffer, and stored at 4°C.

25 **Primers:**

Primer design was based on the sequence of DEN-2 virus, strain 16681, published by Blok et al. (1992), and DEN-2 virus, Jamaican strain 1409, as reported by Deubel et al. (1986) and Deubel et al. (1988).

Primers were synthesized by the Biotechnology Core Facility at the CDC in Atlanta, Georgia. We received the dried primers via mail and adjusted them to a concentration of 100 μ M in dH₂O. The designations and sequences of all of the primers-amplimers used in this project are listed in Appendix A.

To amplify the 3'-end of the DEN-2 virus genome, a downstream amplimer was designed that was complementary to the published sequence of the 3' terminus of the genome. A unique XbaI restriction enzyme site was incorporated at the 5' end of this amplimer to provide a unique site to permit linearization of the recombinant plasmid containing the full-length cDNA clone at the 3' terminus of the cloned genomic cDNA. This linearization was necessary to obtain appropriately terminated DEN virus-specific run-off RNA transcripts from the cDNA clone in transcription reactions with bacteriophage T7 RNA polymerase. Linearization at this 3'-terminal XbaI site resulted in the incorporation of a 5-nucleotide TCTAG extension to the 3' terminus of the genomic mRNA transcribed from the full-length cDNA clone of DEN-2 16681 virus, and a 4-nucleotide CTAG extension to the 3' terminus of RNA transcribed from the DEN-2 PDK-53 cDNA clone. The difference between the two cDNA clones in the length of the extraneous 3'-terminal extension was due to the differently designed 3'-terminal amplimers used to obtain the 3' end genomic cDNA amplicon. Amplimer cD2-10687.XBA or cD2-10687.X2 was used to amplify and clone the 3'-terminal portion of DEN-2 16681 or PDK-53 virus, respectively.

The promoter for the bacteriophage T7 RNA polymerase was engineered at the 5' terminus of the cloned genomic cDNA by incorporating the recognition sequence of the T7 RNA polymerase into the sequence of the 5'-terminal
5 upstream, mRNA-sense amplicon D2-SMT71 immediately preceding the 5'-terminal nucleotide of the DEN-2 viral genome. This design ensured that the T7 RNA polymerase initiated RNA transcription at the 5'-terminal nucleotide of the DEN-2 virus-specific cDNA (Milligan et al., 1987).

10 Amplimers for PCR reactions were designed to take advantage of RENZ sites identified within the nucleotide sequence of the genome of DEN-2 16681 virus. cDNA molecules were amplified to permit ligation or splicing together of overlapping contiguous cDNA clones at shared,
15 overlapping, unique RENZ sites (Figure 3).

Transcription of Genomic mRNA from DEN Virus-Specific Full-Length cDNA Clones:

The recombinant plasmid containing the full-length
20 cDNA clone was prepared for RNA transcription by linearization at the unique XbaI site located at the 3' terminus of the cloned genomic cDNA. The restriction reaction containing the XbaI-restricted plasmid was extracted sequentially with phenol:chloroform:isoamyl
25 alcohol and chloroform and then precipitated. The DNA was redissolved in 50 μ l of TE buffer and digested with proteinase K at a concentration of 1 mg/ml for 1 h at 37°C to hydrolyze contaminating RNases. The sample was then extracted twice with "For RNA Only"
30 phenol:chloroform:isoamyl alcohol buffered to pH 8,

extracted twice with chloroform to remove traces of phenol, and precipitated by adding one-tenth volume of RNase-free 3 M sodium acetate, pH 5.2, and 2.5 volumes of ethanol and incubating for at least 1 h at -70°C or
5 overnight at -20°C.

DEN-2 virus-specific genomic RNA was transcribed from the linearized cDNA template using a commercial T7 transcription kit (Ampliscribe T7 transcription kit, Epicentre Technologies). Transcription reactions were
10 performed for 2 h at 37°C in RNase-free 1.5-ml microtubes in 20- μ l reactions containing 100-1000 ng of linearized DNA template, 7.5 mM each of CTP, GTP, and UTP, 0.75 mM ATP, 2.7 mM m⁷GpppA cap analog, 6.7 mM DTT, 2.0 μ l of a 10X concentration of a proprietary buffer supplied with
15 the commercial kit, and 2.0 μ l of the proprietary Ampliscribe enzyme solution supplied with the kit. Reaction solutions were used directly and without further treatment to transfect BHK-21 cells.

20 **Transfection of BHK-21 Cells with Genomic RNA Transcripts:**

BHK-21 clone 15 cells were transfected with RNA transcripts by electroporation (Liljeström et al., 1991). Fresh cultures of BHK-21 cells were grown to 90%
25 confluency, rinsed twice with cold RNase-free phosphate buffered saline (PBS), and released from the plastic by incubation with 3 ml of commercial trypsin-EDTA solution (GIBCO-BRL). The cells were pelleted by low-speed centrifugation at 1200 rpm for 5 min in a Beckman GPKR
30 centrifuge. The cells were washed twice with cold PBS,

resuspended in cold PBS and kept on ice. The cells were counted using a hemacytometer and microscope, and the cell concentration was adjusted to 10^7 cells/ml. One-half ml of the washed, adjusted cells were mixed with each

5 transcription reaction solution in 1.5-ml microtubes on ice. The mixture was transferred to a cold electroporation cuvette with 0.2-cm electrode gap, which was placed in the cuvette holder of the Bio-Rad Gene Pulser. The cells were shocked twice using settings of

10 1.5 kV voltage, 25 μ FD of capacitance, and resistance set to infinity. The shocked cells were incubated for 10 min at room temperature and then added to 75 cm² tissue flasks containing 20 ml of MEM containing 10% FBS. Transfected cell cultures were incubated at 37°C for 5-8 days until

15 CPE was evident in the cell monolayer and/or expression of DEN virus-specific antigens was identified in an aliquot of the cell monolayer scraped from the flask using DEN virus-specific mouse hyperimmune ascitic fluid or monoclonal antibodies in indirect immunofluorescence

20 tests.

RESULTS

Replication of DEN-2 16681 Virus:

25 DEN-2 16681 virus replicates to high titer in cell culture. The CDC virus seed used in this study contained 2.0×10^7 plaque forming units (PFU)/ml. This titer was determined by plaque titration of the seed virus in monolayer cultures of Vero cells. This seed titered $1.3 \times$

30 10^4 PFU/ml in LLC-MK₂ cells. A growth curve for this virus

was determined in C6/36 *Aedes albopictus* cell culture (Figure 4). This level of replication is quite high for a flavivirus. The DEN-2 16681 virus is eminently suitable to serve as the parent to an infectious cDNA clone of DEN
5 virus.

The DEN-2 PDK-53 vaccine virus, taken directly from a vaccine vial obtained from Mahidol University, contained 3.4×10^4 PFU/ml of virus, as titrated in Vero cell monolayers, and 1.5×10^4 PFU/ml as titrated in LLC-MK₂
10 cell monolayers.

RT/PCR Amplification and Cloning of DEN-2 16681 cDNA:

The entire genome of DEN-2 virus, parental strain
15 16681, was amplified from genomic RNA in the form of 5 cDNA clones of various sizes (T7-F1, F2, F3, F4, and F5). PCR amplification with 5 sets of upstream and downstream amplimers yielded the predicted amplicon sizes in PCR reactions. Figure 5 shows the migration of these cDNA
20 fragments in agarose gels.

Recombinant plasmids, obtained by ligating the cDNA amplicons into the pCRII TA-vector, were extracted from minicultures derived from transformed *E. coli* XL1-Blue colonies. Uncut plasmids were screened for the presence
25 of cDNA insert by comparing their mobility in agarose gels with the mobility of uncut wild-type pCRII vector plasmid. Selected plasmids were then restricted with the restriction enzyme EcoRI to confirm the size of the inserted cDNA fragment. EcoRI digests of F2-Sal, Sal-F2,

and F3 plasmids derived from independent transformed bacterial colonies are shown in Figure 6.

The following 15 DEN-2 16681 virus-specific cDNA clones, shown schematically in Figure 7, were selected for nucleotide sequence analysis:

	<u>Clone</u>	<u>RT/PCR Amplicon</u>	
	1	F1	- A8
	2	F1	- A21
10	3	F1	- A25
	4	F1	- A26
	5	F2-Sal	- AA2-4
	6	F2-Sal	- AA2-8
	7	Sal-F2	- AA3-3
15	8	Sal-F2	- AA3-4
	9	F3	- AA4-4
	10	F3	- AA4-6
	11	F4	- 10
	12	F4	- 12
20	13	F5	- AA6-1
	14	F5	- AA6-2
	15	F5	- AA6-4

25

RT/PCR Amplification and Cloning of DEN-2 PDK-53 cDNA:

The entire genome of DEN-2 virus, vaccine strain PDK-53, was amplified from genomic RNA in the form of 23 cDNA clones of various sizes. Even though the PDK-53 vaccine contained only about 10^4 PFU/ml of virus, we were able to routinely amplify cDNA from RNA that was extracted directly from this seed virus. To accomplish this, we routinely use the "extended PCR method", incorporating the Taq extender reagent (Stratagene) in the PCR reactions. We had previously shown that the Taq extender

significantly enhanced yields of large molecular weight amplicons in the PCR amplification of the nonstructural genes of the flavivirus, St. Louis encephalitis virus (Figure 8A). For extended PCR reactions, reaction

5 mixtures were made as for standard PCR reactions, but the standard PCR buffer was replaced with the Taq extender buffer and 1 unit of AmpliTaq DNA polymerase (Perkin-Elmer) and 1 unit of the Taq extender enzyme per kbp of expected amplicon size was included in the reaction.

10 Figure 8B shows the correct agarose gel migration of large cDNA amplicons F1 (containing the T7 RNA polymerase promoter at the 5' end of the mRNA-sense strand of the amplicon), F2, and F3 obtained by PCR amplification using DEN-2 PDK-53 viral genomic RNA as template. The standard

15 PCR reaction also worked for a number of DEN-2 PDK-53 amplifications.

The PDK-53 PCR products were cloned into the pGEM-5Zf TA-vector (Promega) or the pT7Blue(R) TA-vector (Novagen). Although we seemed to have the best cloning efficiency of

20 PCR amplicons in the pCRII TA-vector, the other vector kits were less expensive and worked well. The cloning efficiency of PCR products into the TA-vector decreased rapidly as amplicon size increased beyond 2000 bp.

25 The following 23 DEN-2 PDK-53 virus-specific cDNA clones were selected for nucleotide sequence analysis:

	<u>CLONE</u>	<u>RT/PCR AMPLICON</u>	<u>Expected Amplicon Length</u>	<u>Up-Amplimer</u>	<u>Down-Amplimer</u>
5	1	F-5	1552-bp	D2-SMT71	cD2-1510
	2	F1-7	"		"
	3	F1-9	"		"
	4	F1-75A	"		"
	5	F1-79B	"		"
10	6	F2-14	3355-bp	D2-1261	cD2-4615
	7	F2-16B	"		"
	8	F3-33	2676-bp	D2-4257	cD2-6932
	19	F3-3C	"		"
	10	F4-9	2373-bp	D2-6493	cD2-8865
15	11	F4.9-22	2937-bp	D2-6493	cD2-9429
	12	F4.9-53	"		"
	13	F4.5-1	1897-bp	D2-8440	cD2-10337
	14	F4.5-2	"		"
	15	F4.5-6	"		"
20	16	F4.5-7	"		"
	17	F5-72	1914-bp	D2-8773	cD2-10687.X2
	18	F5-77	"		"
	19	F5-78	"		"
	20	F3.5-4	1375-bp	D2-6046	cD2-7420
25	21	F3.5-6	"		"
	22	F3.5-19	"		"
	23	F3-3K	2676-bp	D2-4257	cD2-6932

30 Nucleotide Sequence Analyses of DEN-2 16681 cDNA Clones:

EcoRI fragments of the 15 DEN-2 16681 virus-specific cDNA clones were subcloned into the single-stranded bacteriophage M13mp18 or M13mp19 for sequencing.

35 Sequencing of the entire viral genome was performed manually using radioisotopic labeling and exposure,

development, and reading of autoradiographs. The data was read from the films and entered by hand into a sequence data spreadsheet.

The locations of observed cDNA artifacts or "errors" dictated the splicing strategy of subclones to construct the full genome-length clone. If the nucleotide at a particular position of one cDNA clone differed from the nucleotides at that same position in 2 or more independent clones, then the nucleotide in the first clone was deemed to be an error. If only 2 cDNA clones were sequenced for a given region of the genome and they differed in sequence at a particular position, then if one of the cDNA clones agreed with the sequence data of Blok *et al.* (1992), then the clone containing the nucleotide that was in agreement with the latter investigators was deemed to be correct. The approximate locations of the cDNA errors identified in the 16681 clones are illustrated in Figure 9.

The full genome-length cDNA clone of DEN-2 16681 virus was first constructed in pUC19. Unfortunately, RNA transcribed from this clone was not infectious. When over 90% of the full-length cDNA in the clone was resequenced, it was determined that several mutations had occurred during splicing and cloning manipulations of the subclones in *E. coli*. One of these mutations was a base deletion in the NS4B gene. This deletion would cause a frameshift of the amino acid sequence, resulting in ribosomal translation of a nonsense polypeptide downstream of the mutation point. This fatal deletion, by itself, would explain the noninfectious nature of the RNA transcribed from the first full-length clone in pUC19.

The final, correct cDNA subclones (F1-E, F2-E, F3/4/5-F) that were incorporated into the full-length, successfully-infectious clone of 16681 virus were reanalyzed by direct sequencing of the double-stranded plasmid DNA via the thermocycling method using the Taq DyeDeoxy Terminator Cycle Sequencing Kit. Sequence analysis was performed using the automated 373A DNA sequencing machine. The color-coded sequence chromatograms were read by the investigator and the data was entered manually into a computer-based spreadsheet.

We independently confirmed the sequence of the 5'-terminal 32 nucleotides of the DEN-2 16681 viral genome. A 5'-end RNA-cDNA hybrid molecule, made with primer cD2-996 and reverse transcriptase, was 3'-tailed with dCTP and annealed to dGTP-tailed, PstI-cut M13mp19 RF DNA. One of the resulting M13 clones had a cDNA run-off product containing the 5'-terminal end of the genome. The 5'-end sequence was identical to that published for DEN-2 1409 (Deubel et al., 1988) and DEN-2 16681 (Blok et al., 1992). We have not independently confirmed the sequence of the 3'-terminal 36 nucleotides of DEN-2 16681 virus or the 5'- or 3'-terminal nucleotides of DEN-2 PDK-53 virus.

We sequenced uncloned, PCR-derived amplicon cDNA fragments directly for the following regions of the DEN-2 16681 viral genome: nucleotides 70-260, 330-870, 890-1690, 1890-3720, 3770-4050, 4080-4320, and the 3'-terminal 9990-10686. Unlike the sequencing of cloned DNA, direct analysis of PCR amplicons provides sequence information for the majority population of amplified cDNA molecules,

and therefore for the majority population of template RNA molecules.

We observed very early in the project that the nucleotide sequence of DEN-2 16681 virus that we
5 determined at the CDC laboratory differed significantly from the sequence of DEN-2 16681 virus as published by Blok et al. (1992). Our nucleotide sequence differed from that published by Blok et al. (1992) at 60 nucleotide positions, which were located throughout the genome.
10 Amino acid substitutions were encoded by 26 of these nucleotide differences. The approximate genomic locations of the nucleotide differences are illustrated in the schematic diagram in Figure 10. The exact nucleotide positions of the discrepancies are shown in Figure 11.

15

Nucleotide Sequence Analyses of DEN-2 PDK-53 cDNA Clones:

The DEN-2 PDK-53 virus-specific cDNA clones were analyzed by direct sequencing of the double-stranded
20 plasmid DNA by the thermocycling method using the Taq DyeDeoxy Terminator Cycle Sequencing Kit. The 3'-end sequence from nucleotide position 10290-10686 was also determined by direct sequencing of PCR-derived amplicon cDNA. Sequence analysis was performed using the automated
25 373A DNA sequencing machine. The color-coded sequence chromatograms were read by the investigator and the data was entered manually into a computer-based spreadsheet. The approximate locations of the cDNA errors identified in the PDK-53 cDNA clones are illustrated in Figure 12.

Our determination of the nucleotide sequence of DEN-2 PDK-53 virus differed significantly from the PDK-53 genomic sequence published by Blok *et al.* (1992). The latter investigators reported a total of 53 nucleotide differences that encoded 27 amino acid mutations between the nucleotide sequences of the genome of DEN-2 16681 virus and that of its vaccine derivative, PDK-53 virus. They reported the following nonsilent mutations: 1 in the capsid, 2 in prM, 1 in M, 3 in E, 3 in NS1, 3 in NS2A, 2 in NS2B, 3 in NS3, 3 in NS4A, 3 in NS4B, and 3 in NS5. We detected only 8 nucleotide mutations between the genomes of these two virus strains. One mutation occurred in the 5'-NC region of the genome, while 7 nucleotide mutations, 4 of which encoded amino acid substitutions, occurred in the coding region of the genome as shown in Figure 13 and the following table.

5 Table: Summary of nucleotide differences between the genomes of DEN-2 16681 virus and its vaccine derivative virus, strain PDK-53.

Genome	Gene	Genome				
		Nucleotide		Amino Acid		
Position	Gene	16681	PDK-53	16681	PDK-53	
10	57 ^a	5'-NC	C	T	-	-
	524 ^a	prM-29	A	T	Asp	Val
15	2055 ^a	E-373	C	T	Phe	Phe
	2579 ^a	NS1-53	G	A	Gly	Asp
	4018	NS2A-151	C	T	Leu	Phe
	5547	NS3-342	T	C	Arg	Arg
	6599 ^a	NS4A-75	G	C	Gly	Ala
20	8571 ^a	NS5-334	C	T	Val	Val

^a 16681 vs. PDK-53 difference agrees with Blok *et al.* (1992)

25 The few nucleotide positions where our data and those of Blok *et al.* (1992) agreed, in terms of sequence differences between the 16681 and PDK-53 viral genomes, were distributed throughout the genome. The entire genome of DEN-2 16681 virus was cloned and sequenced before we
30 received the PDK-53 vaccine virus at our laboratory.

Except for the 3'-terminal cDNA clones #17-#19, every PDK-53 virus-specific cDNA clone constructed in our laboratory contained at least one nucleotide position of 16681/PDK-53 sequence difference confirmed by both ourselves and Blok et al. (1992). Therefore, our PDK-53 virus-specific cDNA clones did not result from contamination of PDK-53-specific PCR reactions with 16681 virus-specific cDNA template. Our PDK-53 virus-specific cDNA clones, which also contained the many sequence discrepancies between our data and those of Blok et al. (1992), encoded the nucleotide sequence from the 5' terminus to nucleotide position 10337 of the genome of PDK-53 virus. The 3'-terminal 387 nucleotides (10337-10723) of DEN-2 PDK-53 virus were identical to those of the parental 16681 virus. Since none of the PDK-53 virus-specific cDNA clones covering this region of the genome contained a point of confirmed 16681/PDK-53 sequence difference, we repeated the PCR amplification of the 3' terminus of the PDK-53 virus genome. This was done to ensure that the 3'-terminal cDNA clones #17-#19 did not result from PCR reactions contaminated by 16681 virus-specific DNA template. The PCR reaction components were pipetted in a room in which DEN cloning had not been performed previously, using new micropipetors, newly opened pipet tips with aerosol barrier, and freshly made stock reagents. Direct sequencing of the resulting double-stranded PCR cDNA amplicon confirmed that the 3'-387 nucleotides of DEN-2 PDK-53 virus was indeed identical to the 3' terminus of the 16681 parent.

The finalized nucleotide sequence of DEN-2 virus, strain 16681, including the nucleotide and amino acid mutations identified for DEN-2 PDK-53 virus, is shown in Figure 14.

5

Construction of DEN-16681 Full-Length Clone in pUC19:

For the construction of the full genome-length cDNA clone of DEN-2 16681 virus, 5 of the sequence-
10 characterized PCR-amplified cDNA subclones were selected for splicing. However, clone #5 contained a cDNA "error" that was not readily spliced out with the existing clones. This error, which was a C-to-T mutation at nucleotide position 1730 and encoded a nonsilent Thr-to-Ile amino
15 acid substitution at E-265, was incorporated into the F2 construct. The intermediate F2 construct was the result of splicing the F2-Sal clone (#5) SphI/HpaI fragment to the Sal-F2 clone (#7) HpaI/KpnI fragment in the MCS of plasmid pUC18 (Figure 15). To correct the error, a new
20 PCR amplicon was made using primers D2-1261 and cD2-2955. Resulting clones in the TA-vector were sequenced, and the correct SphI/HpaI fragment of a new clone was substituted for the faulty SphI/HpaI fragment of the original F2 construct (Figure 16). The corrected F2 clone was
25 designated F2-C.

The relevant cDNA clones of DEN-2 16681 virus were spliced together via a series of intermediate ligation products in the MCS of pUC18 to yield F1/3/4/5, which contained all of the genome except for the SphI-KpnI 1380-
30 4493 region present in clone F2-C. Multiple attempts to

ligate the F2-C SphI/KpnI cDNA fragment into F1/3/4/5 in pUC18 failed. The cDNA insert of F1/3/4/5-pUC18 was then transferred to the MCS of pUC19, resulting in F1/3/4/5-pUC19. This operation simply reversed the orientation of the cDNA insert within the context of the pUC plasmid. Ligation of SphI/KpnI-cut F1/3/4/5-pUC19 and F2-C SphI/KpnI insert readily yielded transformants in *E. coli* Xl1-Blue that contained the full-length cDNA clone F1/2/3/4/5-pUC19, which was designated pD2/IC-20. The detailed splicing procedures for pD2/IC-20 are illustrated in Figure 17. The orientation-specific cloning of the full genome-length cDNA in pUC19 rather than pUC18 is diagrammed in Figure 18.

The full genome-length cDNA of DEN-2 16681 virus was cloned into the MCS of pUC19. Apparent full genome-length viral mRNA was transcribed from linearized pD2/IC-20. This transcribed product failed to yield infectious virus following electroporation of BHK-21 cells. Most of the cDNA in the pD2/IC-20 clone was resequenced, and several cloning artifacts, including a fatal single-nucleotide deletion, were identified. Original subunit intermediate cDNA constructs in pUC18 were resequenced to confirm that they possessed the correct sequence and corrected where necessary. The corrected primary cDNA clones F1, F2-C, and F3/4/5 were then ligated into the low-copy plasmid pBR322, rather than the high copy-number pUC18 plasmid. It was envisioned that the cDNA would be more stable in a slower-replicating plasmid in *E. coli*.

To enable more straightforward cloning into pBR322, the MCS of pUC19 was spliced into the pBR322 plasmid

(Figure 19). This resulted in plasmids pBRUC-138 and pBRUC-139 containing the pUC MCS in both orientations within the pBR322 plasmid backbone. The SphI site was removed from both pBRUC plasmids by cutting with SphI, blunt ending of the cut ends using T4 DNA polymerase, and then ligating the ends back together. This was necessary for the construction of the full-length cDNA clone because SphI is one of the cDNA restriction/splicing sites for the clone.

10 The F3/4/5-F cDNA clone of DEN-2 16681 virus, which had been verified by sequence analysis, was cloned into pBRUC-139 (SphI⁻) (Figure 20). Following this ligation, the F1-E and F2-C cDNA clone fragments were also moved into the pBR322 backbone to construct the full genome-length cDNA clone, pD2/IC-30P (Figure 20). This recombinant plasmid was replicated successfully in both TB-1 and MC-1061 strains of *E. coli*.

Construction of DEN-2 PDK-53 Infectious cDNA Clone:

20 The full-length infectious clone of DEN-2 16681 virus was used in the construction of the infectious clone for PDK-53 virus. Since the 3'-noncoding regions of the genomes of both viruses are identical, and the amino acid sequences of the translated precursor polyproteins encoded by genome nucleotide positions 6646-10269 are identical in both viruses, the infectious clone of PDK-53 virus was constructed using the 16681 3'-end cDNA from the NheI site at nucleotide position 6646 to the 3' terminus of the genome (Figure 21). After correcting a cDNA error in the

30

PDK-53 F3-3C subunit clone, this fragment and the F2-16B cDNA fragment were ligated into the infectious clone backbone to construct the DEN-2 PDK-53 virus-specific full-length cDNA clone, pD2/IC-130V (Figure 21).

5

Transcription of Viral mRNA from DEN-2 Infectious cDNA

Clones:

Viral genomic RNA extracted from gradient-purified virions was analyzed by nondenaturing RNA agarose gel electrophoresis to observe the level of RNA degradation and the limits of detectability by ethidium bromide staining. Figure 22 shows an agarose gel electropherogram for 22-383 ng of viral genomic RNA obtained from purified preparations of wild-type DEN-2 16681 virus and wild-type Venezuelan equine encephalitis (VEE) virus, strain Trinidad donkey. Although degradation of the RNA is visible as a spectrum of smaller molecular weight nucleic acid (smear in Figure 22), definite full-genome length RNA bands are clearly visible. This smear of nucleic acid is probably also due, in part, to multiple conformations of the single-stranded RNA molecules which migrate through the gel at different rates. The relative gel migration of the single-stranded RNA does not correlate directly with the sizes of the double-stranded molecular weight marker DNA bands (MW, Figure 22); the VEE and DEN-2 viral genomes are 11,447 and 10,723 nucleotides in length, respectively. BHK-21 and C6/36 cells were transfected successfully by electroporation with 2000, 500, 100, 10, 1, and 0.1 ng of viral genomic RNA extracted from purified VEE or DEN-2

16681 virus, as indicated by development of CPE, expression of viral proteins detected by indirect immunofluorescence tests using virus-specific antibody, and/or by plaque titration of infectious virus from the transfected-cell culture medium. RNA quantities of 1 ng or less were essentially undetectable in the ethidium bromide-stained agarose gel system we used. Therefore, authentic RNA transcripts derived from full genome-length cDNA and visualized in agarose gel electropherograms of transcription reactions should be infectious for BHK-21 cells by electroporation.

Investigators previously constructed an infectious cDNA clone for VEE virus as reported by Kinney *et al.* (1989). RNA transcription reaction conditions that yielded high quantity and quality of infectious mRNA transcripts from the pVE/IC-92 infectious clone of VEE virus failed in multiple attempts to transcribe RNA from the pD2/IC-20 clone of DEN-2 16681 virus. Figure 23 shows an agarose gel electropherogram that demonstrates successful transcription of RNA from the VEE clone, but not pD2/IC-20.

In an attempt to improve RNA transcription from the DEN-2 clone, commercial transcription kits were purchased. The Megascript transcription kit supplied by Ambion also failed to transcribe RNA from the DEN clone. However, the Ampliscribe kit obtained from Epicentre Technologies enabled efficient transcription of RNA from the DEN-2 clone (Figure 24).

The success of the Ampliscribe kit apparently was due to the high concentration of ribonucleotides and a very

high, but proprietary, concentration of T7 RNA polymerase.
The RNA transcribed from pD2/IC-20 was not infectious.
However, viral mRNA transcribed from DEN-2 16681 clone
pD2/2-IC30P and PDK-53 clone pD2/IC-130V was infectious
5 (Figure 25).

Viral mRNA transcripts from both replicates of
pD2/IC-30P (A and D) and pD2/IC-130V (F and J) were
infectious, producing viable infectious virus in
electroporated BHK-21 cells. Figure 26 shows RNA
10 transcripts from pD2/IC-20, pD2/IC-30P, and pD2/IC-130V.

Construction of DEN-2 16681/PDK-53 Chimeric cDNA Clones:

Several chimeric full-length cDNA clones were derived
15 from the pD2/IC-30P and pD2/IC-130V clones. All clones
were constructed in the pBRUC-139 derivative of the pBR322
plasmid vector. *E. coli* strains XL1-Blue, MC-1061, and
TB-1 were successfully transformed with ligated
recombinant plasmids containing full genome-length cDNA.
20 Viable virus was derived from all of the indicated clones.
The evolutionary tree for the chimeric viruses is
diagrammed in Figure 27.

Details concerning the splicing strategies for the
chimeric clones are shown in Figure 28. Appropriate cDNA
25 fragments were cut and ligated together at the internal
SalI, SphI, KpnI, and NheI sites as well as at the 5'-SstI
and 3'-XbaI sites.

Viable prototype and chimeric viruses were derived
from each of the clones indicated in Figure 28 by
30 electroporation of BHK-21 cells with viral genome-length

mRNA transcribed from linearized plasmids. Seed stocks of these viruses were prepared by centrifuge-clarification of the cell culture medium, adjustment of the FBS concentration to 10%, and freezing of seed aliquots at 5 -70°C. Virus concentrations were determined by plaque titration of the virus seeds in monolayer cultures of Vero cells. The results of these virus titrations are shown in the following table.

Table. Plaque titration of DEN-2 16681 and PDK-53 stock seed viruses and chimeric viruses recovered from BHK-21 cells transfected with infectious clone-derived viral mRNA transcripts.

	<u>Virus</u>	<u>(PFU/ml)</u>	<u>Genotype^a</u>
10	DEN-2 16681	8.0 X 10 ⁷	c D F G L R G V
	DEN-2 PDK-53	5.1 x 10 ³	t V . D F . A .
15	D2/IC-30P-A	3.6 X 10 ⁵
	D2/IC-30P-A2	1.7 X 10 ⁵
	D2/IC-130V-F	4.0 X 10 ⁵	t V . D F . A .
	D2/IC-130V-J	2.2 X 10 ⁵	t V . D F . A .
20	D2/IC-130V2-1	2.8 X 10 ⁵	t V A .
	D2/IC-130V2-7	8.8 X 10 ⁴	t V A .
	D2/IC-31-12	2.1 X 10 ⁵	t V
	D2/IC-31-15	3.2 X 10 ⁵	t V
25	D2/IC-32-A	1.4 X 10 ⁶	. . . D F
	D2/IC-32-G	1.2 X 10 ⁶	. . . D F
30	D2/IC-33-C	9.6 X 10 ⁴ A .
	D2/IC-33-P	1.9 X 10 ⁵ A .
	D2/IC-321-L	1.1 X 10 ⁶	t V . D F
	D2/IC-321-N	7.6 X 10 ⁵	t V . D F
35	D2/IC-323-B	7.2 X 10 ⁵	. . . D F . A .
	D2/IC-323-I	8.8 X 10 ⁵	. . . D F . A .
	D2/IC-31-57-5	2.4 X 10 ⁵	t
40	D2/IC31-524-D	3.2 X 10 ⁴	c V

^a Genotype is designated in small case for the virus-specific 5'-noncoding nucleotide and in upper case single-letter amino acid abbreviation for amino acids encoded by virus -specific nucleotide mutations. Dots represent nucleotide or amino acid sequence identity with DEN-2 16681 virus.

To establish the validity of the clone-derived chimeric viruses, relevant genomic cDNA fragments were amplified directly from seed viruses by PCR and spot-sequenced. The results are shown in Figure 29. This validation process is ongoing. Except for D2/IC-31-524 virus, appropriate cDNA insert regions in chimeric viruses have been confirmed by sequence analysis. Except for D2/IC-30P, D2/IC-130V, and D2/IC-31-57, which have been fully confirmed, clone-derived chimeric viruses have yet to be spot-sequenced in a recipient clone-derived cDNA region to definitely establish the chimeric nature of the virus. The recipient clone is the recombinant plasmid backbone into which a cDNA fragment, the insert fragment, from a heterologous donor clone is spliced. Where duplicate clone-derived viruses were obtained, both viruses of a given genotype were spot-sequenced, and both gave the same result, which is shown in Figure 29.

Submission of pD2/IC-30P and pD2/IC-130V to ATCC:

20

Patent deposits of the full genome-length cDNA clones of DEN-2 16681 and PDK-53 viruses were submitted to the American Type Culture Collection (ATCC), Rockville, Maryland, U.S.A. Both pD2/IC-30P-A and pD2/IC-130V-F were grown overnight in *E. coli* TB-1 cells. Six cryogenic vials containing 1 ml each of frozen cell culture in 10% glycerol were submitted by dry ice shipment. Prior to shipment, plasmid was extracted from a 1 ml aliquot of each virus-specific culture. The recombinant full-length

plasmid was recovered from the cells as shown in Figure 30.

The pD2/IC-30P-A deposit with the ATCC was assigned accession number ATCC 69826, and the pD2/IC-130V-F deposit with the ATCC was assigned accession number ATCC 69825. Date of deposit was May 25, 1995.

Construction of Chimeric DEN-2/1, -2/3, and -2/4

Infectious Clones:

10 We contemplate deriving chimeric DEN-2/1, DEN-2/3, and DEN-2/4 viruses from recombinant full genome-length cDNA clones containing the genetic background of DEN-2 PDK-53 virus and the prM and E genes of the DEN-1, DEN-3, and DEN-4 candidate vaccine viruses, respectively. To
15 accomplish this, the prM and E genes of the vaccine viruses were amplified by PCR. Because our laboratory has been establishing a sequence database to analyze the molecular epidemiology of several flaviviruses, including all of the serotypes of dengue virus, the primers used for
20 cDNA amplification in the PCR were readily available at our laboratory. The amplified cDNA molecules were sequenced directly, thus providing the sequence of the population of virions in the virus seed. The amplified cDNA amplicons for the DEN-1, DEN-3, and DEN-4 vaccine
25 viruses have all been cloned into the pGEM-5Zf TA-vector. The cloned cDNA has not been analyzed by sequencing, since it will be necessary to rederive the cDNA amplicons by PCR to incorporate appropriate RENZ cleavage sites within the amplicon for splicing into the full-length cDNA backbone
30 of DEN-2 PDK-53 virus. The partial nucleotide sequences

of the genomes of the DEN-1, DEN-3, and DEN-4 vaccine viruses were aligned with the DEN-2 PDK-53 sequence. All four sequences are aligned with the nucleotide sequence of DEN-2 16681 virus and its deduced amino acid sequence in Figure 31. The deduced amino acid sequences of the DEN viruses are aligned in Figure 32.

It is readily evident from the aligned nucleotide sequence data that useful restriction enzyme sites in the DEN-2 virus-specific cDNA are not conserved in the DEN-1, DEN-3, and DEN-4 viruses. Therefore, splicing sites must be engineered into the cDNA to enable the splicing of heterotypic DEN-1, DEN-3, and DEN-4 prM and E genes into the DEN-2 backbone. It is not yet clear precisely how the nonstructural proteins of flaviviruses interact with the structural proteins during intracellular maturation of the virus. Furthermore, the interaction of the capsid protein with the genomic mRNA molecule in the nucleocapsid of the virion has not been defined. However, coexpression of the E and prM proteins has been more successful than expression of E alone in expression systems *in vitro*. The DEN-2 nonstructural proteins are involved in all virus-specific intracellular polyprotein processing and replication of viral mRNA, and the predominant portion of the mRNA genome interacting with the capsid protein is presumably, but not necessarily, DEN-2 virus-specific. For these reasons, our strategy is to splice in the prM and E genes of DEN-1, DEN-3, and DEN-4 viruses very precisely, while maintaining the DEN-2 context of the bracketing capsid and NS1 protein regions.

The strategies for creating XhoI and XbaI splice sites at the 5' end of the prM gene and near the 3' end of the E gene are illustrated in detail in Figures 33 and 34, respectively. Briefly, mutagenic primers containing the appropriate RENZ site are utilized in PCR reactions to synthesize new cDNA for the prM and E genes of all four viruses. A DEN-2 PDK-53 virus-specific cDNA cassette plasmid, designated pD2V-CAS12, containing the genome region from the 5' terminus through nucleotide position 4696 is constructed via intermediate plasmid constructs pF1-Xho and pF2-Xba as illustrated in Figures 35 and 36. The XhoI/XbaI cDNA fragments cut directly from DEN-1, DEN-3, and DEN-4 virus-specific amplicons synthesized by PCR using the mutagenic primers are ligated into the pD2V-CAS12 cassette plasmid to create subclone chimeras. The SstI/KpnI fragment of the resulting pD1V-CAS12, pD3V-CAS12, and pD4V-CAS12 cassettes are moved into pD2/IC-130V restricted with SstI/KpnI to create the chimeric full genome-length cDNA clones (Figure 36).

20

Discussion:

Infectious cDNA clones permit the directed engineering of viral genomes. Depending on their viability in terms of ability to replicate in cell culture, infectious clone-derived viruses can be modified by incorporating point mutations, multiple mutations, deletions, gene regions of related or heterologous viruses, or nonviral genes. Infectious cDNA clones have been developed for many RNA viruses, including flaviviruses DEN-4 (Lai et al., 1991), yellow fever (Rice

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et al., 1989), Kunjin (Khromykh and Westaway, 1994), Japanese encephalitis (Sumiyoshi et al., 1992), and TBE (unpublished data). We describe herein the development of infectious cDNA clones for DEN-2 16681 virus and its candidate vaccine derivative, strain PDK-53. We also describe the construction of chimeric viruses, incorporating the prM and E genes of candidate DEN-1, DEN-3, and DEN-4 vaccine viruses within the genetic background of the DEN-2 PDK-53 vaccine virus.

10 Although the candidate vaccine viruses developed at Mahidol University are currently the best live DEN virus vaccine candidates in terms of immunogenicity and safety in adult humans, the DEN-1, DEN-3, and DEN-4 vaccine viruses replicate poorly in cell culture and possess low infectivity in humans, requiring up to 2000-fold more PFU of virus to infect and immunize humans than is needed for the DEN-2 PDK-53 vaccine virus. The low infectivities of these viruses have significant implications for vaccine production in cell culture, potentially decreased immunogenic efficacy, and more rapid inactivation under conditions of a poorly maintained cold chain in tropical countries where dengue viruses are endemic.

 The purpose of engineering chimeric DEN vaccine viruses is to enhance the replicative ability and immunogenicity of the DEN-1, DEN-3, and DEN-4 vaccine viruses. A primary assumption has been that the attenuated DEN-2 PDK-53 vaccine virus replicates to appropriate levels in cell culture. In fact, it does appear that the genome of DEN-2 PDK-53 virus is eminently suited to serve as the genetic backbone for chimeric

viruses containing the prM and E genes of DEN-1, DEN-3, and DEN-4 vaccine viruses. We have recently completed growth curves for DEN-2 16681 virus, DEN-2 PDK-53 virus, and their infectious clone derivative viruses in LLC-MK₂ cells.

The viruses were titrated in Vero cell monolayers. These data are shown in the following table:

	Virus	Maximum Titer (PFU/ml)	Maximum Titer at Day
10	DEN-2 16681	2.6×10^8	10
	D2/IC-30P-A	1.7×10^7	8
	D2/IC-30P-A2	6.6×10^7	7
15	DEN-2 PDK-53	3.8×10^7	9
	D2/IC-130V-F	2.9×10^7	7
	D2/IC-130V-J	1.7×10^7	7

The DEN-2 PDK-53 virus and its infectious clone derivative viruses grow to approximately 10^7 PFU/ml in LLC-MK₂ cells, about as well as the DEN-2 16681 virus.

A second assumption is that the chimeric DEN viruses will be viable and the DEN-2 PDK-53 virus-specific replication machinery will significantly increase replication of the chimeric viruses in cell culture and increase their infectivity and immunogenicity in humans relative to the wild-type vaccine viruses. The high degree of conservation of amino acid sequences among the polyproteins of the four DEN viruses should ensure that the chimeric viruses will be viable. The level of

replication attained by the chimeric DEN viruses is determined empirically, as was determined for the DEN-2 PDK-53 infectious clone derivative virus.

Bray et al. (1991) constructed chimeric DEN-4/1 and DEN-4/2 viruses that appeared to appropriately express DEN-1 and DEN-2 structural protein antigens in the genetic background of DEN-4 virus. These investigators spliced much of the 5'-noncoding region, and the capsid, prM and E genes of DEN-1 or DEN-2 virus into the full-length cDNA clone of DEN-4 virus. The near 3'-terminal splice site they chose in the E gene is very close to that proposed by us in our project. These chimeric viruses replicated very slowly relative to the wild-type viruses. The authors attributed this slow replication to possible suboptimal gene expression, assembly, and/or maturation due to incompatibility of heterotypic genes or RNA packaging in the nucleocapsid. Another possibility is that cDNA errors may have been incorporated into their constructs. In contrast, Pletnev et al. (1993) engineered chimeric viruses between DEN-4 virus and tick-borne encephalitis (TBE) virus, which is a very distant flavivirus relative of DEN viruses. Thus, DEN virus chimeras may be derived that are viable.

A third assumption is that our chimeric DEN viruses will express the appropriate structural protein antigens of DEN-1, DEN-3, and DEN-4 viruses, and that vaccinees will respond with development of appropriate serum titers of DEN-1, DEN-3, and DEN-4 neutralizing antibodies following immunization with the chimeric viruses. We describe the insertion of the prM and E genes of DEN-1,

DEN-3, and DEN-4 viruses into the DEN-2 clone. Thr-to-Ser amino acid substitutions near the amino terminus of the prM protein in DEN-2, DEN-2/1, DEN-2/3, and DEN-2/4 viruses resulting from mutagenesis to create the XhoI site of the cassettes should be conservative in nature and affect the phenotype of derived viruses minimally, if at all. Alternatively, a unique MluI site (ACGCGT) could be created via a single, silent A-to-G point mutation at nucleotide position 453 in the DEN-2 clone. The MluI site immediately preceding the T7 promoter could easily be eliminated by cutting the clone with MluI, blunt-ending, and religation. The clone-derived DEN-2 and chimeric viruses would then have the prM amino-terminal sequence "FHLTTR."

The carboxyl-terminal 24 amino acids of the E glycoprotein of all of the infectious clone-derived viruses will be those of the DEN-2 PDK-53 virus. Therefore, the E protein of all of the chimeric viruses will have amino acid mutations in this region. Yet, the carboxyl-terminal 39 amino acids of the DEN virus E protein comprise membrane-spanning, transmembrane domains. In all enveloped viruses, the transmembrane domains of the integral viral proteins of related viruses are quite variable in amino acid sequence. It has often been noted that the important conserved feature of amino acids in this domain lies in their hydrophobic, "lipid-loving" nature rather than in the absolute sequence. Creation of a MroI site (TCCGGA) or a unique AgeI site (ACCGGT) at nucleotide positions 2281-2286 in the DEN-2 clone would

result in amino acids "SG" or "TG", respectively, at positions E-449 and E-450 in the clone-derived viruses.

The E protein of all flaviviruses share a similar gross tertiary structure that is indicated by the absolute conservation of the 6 Cys residues in the prM protein and in the 12 Cys residues in the ectodomain (the region located on environment side of the viral lipid envelope) of the E protein of DEN, Japanese encephalitis, West Nile, Murray Valley encephalitis, St. Louis encephalitis, Kunjin, yellow fever, TBE, Langat, and Powasson flaviviruses (data not shown). Cys residues are involved in intrachain Cys-Cys disulfide bonds that determine the overall structure of the protein. We fully expect the DEN-2/1, DEN-2/3, and DEN-2/4 chimeric viruses to be viable and to replicate more efficiently than the wild-type DEN-1, DEN-3, and DEN-4 vaccine viruses, respectively. Furthermore, chimeric recombinants involving the genetic backbone of one flavivirus and the structural genes of a variety of different flaviviruses may also be viable, as has been demonstrated for DEN-4/TBE virus recombinants (Pictnev et al., 1993). Such recombinant viruses offer the potential opportunity to engineer chimeric vaccine viruses for a number of flavivirus-associated diseases within the genetic background of a single flavivirus. The X-ray crystallographic structure of the E glycoprotein of TBE flavivirus has recently been published (Rey et al., 1995). This development has significant implications for the future design of flavivirus molecular vaccines.

A fourth assumption is that the chimeric DEN viruses will retain the attenuated phenotype of the wild-type DEN-1, DEN-3, and DEN-4 vaccine viruses, despite enhanced replicative efficacy provided by the more efficient nonstructural genes and 5' and 3' noncoding regions of the DEN-2 PDK-53 virus. This presupposes that DEN-2 PDK-53 virus has attenuating mutations in the noncoding regions or in the nonstructural genes and/or that attenuating mutations occur in the prM/E region of the genomes of DEN-1, DEN-3, and DEN-4 viruses. Mutations in essentially any region of the viral genome may be capable of attenuating a virulent virus. This has been demonstrated for a number of viruses including polio virus, VEE virus, and Theiler's virus. Noncoding as well as protein coding regions may be involved in attenuation. Attenuating mutations in the envelope proteins of enveloped viruses are common (Barrett et al., 1990).

The nucleotide mutations in DEN-2 PDK-53 virus at genome nucleotide positions 57 (5'-noncoding region), 524 (prM), 2579 (NS1), 4018 (NS2A), and 6599 (NS4A) may be involved in attenuation of the virus. Unless the prM amino acid mutation is the only mutation affecting virulence of the virus, the DEN-2 PDK-53 genetic background, within which the structural genes from heterologous viruses will be expressed, does itself possess genotypic markers of attenuation. We can determine the genetic loci involved in the attenuation of the DEN-2 PDK-53 virus by analyzing DEN-2 16681/PDK-53 recombinant viruses derived from chimeric 16681/PDK-53

full-length clones. The E gene of DEN-2 PDK-53 virus contains no attenuating mutations.

Although investigators have sequenced the structural genes of numerous DEN-3 virus strains (e.g., Lanciotti et al., 1994), none have sequenced the DEN-3 16562 virus, parent to the DEN-3 PCMK-30/FRhL-3 vaccine virus. After determining the sequences of the prM and E genes of this virus, we can establish if any amino acid mutations have occurred within these genes in the DEN-3 vaccine virus.

10 By comparison, nucleotide sequence information for the parental DEN-1 and DEN-4 viruses have been determined (unpublished data (parental DEN-1 virus); Lanciotti et al., submitted for publication (parental DEN-4 virus)). The nucleotide sequences of the E gene of DEN-4 1036 virus

15 and both prM and E genes of DEN-1 16007 virus have been determined. The following amino acid mutations were identified:

	Virus type	E Protein Amino Acid Position	Amino Acid	
			Parent Strain	Vaccine Strain
5	DEN-1	E-130	Val	Ala
		E-203	Glu	Lys
		E-204	Arg	Lys
10		E-225	Ser	Leu
		E-384	Ala	Glu
		E-477	Met	Val
15	DEN-4	E-345	Glu	Lys
		E-364	Val	Ala

There were six amino acid mutations in the E protein of DEN-1 16007 PDK-13 virus and 2 mutations in that of DEN-4 1036 PDK-48 virus. There were no amino acid substitutions in the prM protein of the DEN-1 vaccine virus. Glu-to-Lys and Lys-to-Glu amino acid substitutions, as occur at DEN-1 E-203 and DEN-4 E-345, are common motifs in sequence comparisons between parent viruses and their vaccine derivatives. It is likely that the heterologous prM/E cDNA inserts in recombinant full-length cDNA clones will transport genetic loci of attenuation into the chimeric DEN-2/1, DEN-2/3, and DEN-2/4 virus derivatives. The optimum scenario for the chimeric viruses involves increased replication ability in the presence of genetic loci of attenuation in the heterologous DEN-1, DEN-3, and

DEN-4 structural gene inserts within the genetic background of the DEN-2 PDK-53 virus.

Nucleotide sequence analysis of expressed genes is essential. The error rate in the original RT/PCR derived cDNA clones of DEN-2 16681 virus was 8.2×10^{-4} , that is 1 cDNA error for every 1227 nucleotides of cloned, sequenced cDNA. In a previous sequencing project involving VEE virus and employing classical, non-PCR cDNA synthesis methodology, the error rate was calculated to be 3.9×10^{-4} or 1 error for every 2543 nucleotides of cloned, sequenced cDNA. These errors are due to nucleotide incorporation errors by reverse transcriptase during first strand cDNA synthesis and perhaps to the cloning of individual variants within the original population of virions.

Unlike many DNA polymerases, RNA polymerases and reverse transcriptase have no editing function. Incorrect nucleotides incorporated during strand elongation are not detected or removed before continuing. The Taq DNA polymerase is also known to incorporate errors into PCR amplicons. Thus, at least 4-8 cDNA "errors" can be expected to occur in 10 kb of cloned cDNA. We have observed the incorporation of spurious in-frame termination codons (TAA, TAG, TGA) in cDNA clones derived from both VEE and DEN viruses. Premature termination of amino acid translation would result in a truncated protein and would undoubtedly be a lethal mutation for a candidate infectious clone. Much of the utility of genes expressed *in vitro* is compromised when those genes are not characterized by sequence analysis. If cDNA errors occur in candidate infectious cDNA clones, it may be difficult

to determine if phenotypic effects of directed mutations are due to the engineered mutation, to cDNA errors, or to synergistic action or compensation between errors and engineered mutations.

5 Wiktor et al. (1984) reported that two cDNA errors caused spurious amino acid substitutions in rabies virus glycoprotein expressed in recombinant vaccinia virus and resulted in expression of non-authentic rabies glycoprotein. After sequence analysis and correction of
10 the cDNA, expression of authentic rabies glycoprotein was obtained. A faulty cDNA clone may behave as expected in one circumstantial context, yet behave very inappropriately and be highly misleading in a different context. A faulty structural gene cDNA clone of the
15 virulent VEE Trinidad donkey (TRD) virus that was expressed in recombinant vaccinia virus was essentially authentic by monoclonal antibody analysis of expressed VEE virus-specific proteins and by protection of immunized mice from challenge with virulent VEE virus (Kinney et
20 al., 1988a; Kinney et al., 1988b). However, incorporation of this cDNA clone into an infectious cDNA clone of VEE virus completely abrogated the virulence of the clone-derived virus, whereas the corrected cDNA fragment resulted in derivation of virulent virus (Kinney et al.,
25 1993).

Although Lai et al. (1991) originally derived their infectious clone of DEN-4 virus from sequence characterized subunit cDNA clones (Zhao et al., 1986; Mackow et al., 1987), the original full-length clone was
30 not infectious (Lai et al., 1991). While these

investigators indicated that they sequenced both strands of much of the cloned genomic cDNA, they did not indicate that they sequenced more than a single clone for a given cDNA region. Nucleotides encoding cDNA errors will be confirmed on both cDNA strands, but will not be identified as errors unless the sequences of two or more independent cDNA clones covering the same region of the genome are sequenced. The functional full-length clone of DEN-4 virus was obtained by repeated splicing of large new cDNA fragments into the full-length clone until a functional clone was obtained. The authors did not indicate that the newly cloned regions were characterized by nucleotide sequence analysis (Lai et al., 1991). It is probable that the slowed replication of the DEN-4/1 and DEN-4/2 chimeric viruses relative to wild-type viruses reported by Bray et al. (1991) is due to the presence of cDNA artifacts within the full-length cDNA clone. The critical importance of accurate nucleotide sequence characterization of genes expressed *in vitro*, particularly when those genes are expressed in the form of infectious cDNA clones, is still not widely appreciated by many in the molecular biology field.

Although putative nucleotide sequences for the genomes of DEN-2 16681 and DEN-2 PDK-53 viruses have been reported in the literature (Blok et al., 1992), our sequence results indicate that the published data is highly flawed. Blok et al. (1992) reported 53 nucleotide mutations between the two viruses; we determined only 8 mutations. We analyzed at least two independent cDNA clones for regions covering the entire genomes of both

viruses. The DEN-16681 sequencing project was completed prior to receiving the DEN-2 PDK-53 virus in our laboratory, and the nucleotide sequence of the PDK-53 virus was determined from cDNA amplified directly from virus present in vaccine vials.

There are now only two classes of infectious clones developed for vaccine flaviviruses that have themselves been administered to humans: the infectious clone of yellow fever virus, vaccine strain 17D (Rice et al., 1989; Hahn et al., 1987; Rice et al., 1985), and the DEN-1, DEN-2, DEN-3, and DEN-4 vaccine derivative infectious clones described herein. Both classes of infectious clones have the important advantage of being derived from vaccine viruses that have been tested for efficacy and safety in humans. The yellow fever 17D virus vaccine has long been one of the most effective human vaccines developed; immunization with this virus provides lifelong immunity. In the case of DEN virus, it is essential that vaccines provide immunity against infection by all four serotypes of the virus. DEN-1, DEN-2, DEN-3, and DEN-4 vaccine viruses have been developed at Mahidol University, Bangkok, Thailand. All four vaccine viruses have been tested in humans and have been demonstrated to be immunogenic and safe for human adults.

Replicating vaccines in the form of live, attenuated viruses offer distinct advantages in terms of immunogenic efficacy due to replicative amplification of viral antigens (antigenic mass) in the vaccinees and replication in appropriate target tissues. Inactivated or subunit antigens usually suffer from a lack of sufficient

antigenic mass and subsequent failure to stimulate an effective immune response. Expression of proteins in recombinant vaccinia virus, which replicates primarily at the site of inoculation, may provide protection against parenteral challenge with virulent virus, but may not protect against an aerosol challenge. This was demonstrated for VEE virus when it was shown that recombinant vaccinia virus expressing the structural proteins of VEE virus protected mice from intraperitoneal challenge, but not intranasal challenge, with virulent VEE virus (Kinney et al., 1988b). Immunization with the live, attenuated VEE TC-83 vaccine virus, on the other hand, provided immunity against both parenteral challenge (immunity provided by circulating serum IgG antibody) and intranasal challenge (mucosal, IgA-base immunity) with virulent VEE virus. Furthermore, the level of immunity, as measured by titers of VEE virus-specific neutralizing antibody, were considerably higher in TC-83 virus-immunized mice and horses (the natural epidemic host for VEE virus) than in animals immunized with recombinant vaccinia/VEE virus (Kinney et al., 1988b; Bowen et al., 1992). Similar results have been reported for vaccinia/influenza A virus recombinants in rodents (Smith et al., 1986). Furthermore, a replicating vaccine virus provides the appropriate T-cell epitopes to stimulate cell-mediated immunity as well as humoral immunity. T-cell epitopes may be lacking in subunit vaccines. In short, vaccination with a safe live, attenuated vaccine virus provides the optimal immunization of a natural infection in terms of the type and level of immunity

elicited and the repertoire of viral antigens involved in generating the immune response.

To use the DEN viruses described herein as vaccine candidates, it is necessary to rederive the viruses by
5 transfection of a cell line, such as primary dog kidney, certified for human use under conditions of good laboratory practice and management to ensure the avoidance of potential adventitious agents that might be present in uncertified cell lines. Although the cDNA-derived viruses
10 originate from candidate vaccine viruses that have undergone testing in humans, they require recertification by analysis for possible *in vitro* phenotypic markers of attenuation and by safety testing in small animals and probably nonhuman primates. All investigative studies
15 involving the pathogenesis of DEN virus are hampered by the unavailability of a suitable animal model. Certain *in vitro* characteristics are apparently associated with attenuation of DEN viruses, but the only definitive test is vaccine trial in human volunteers. Vaccine trials
20 would presumably follow those of the original wild-type vaccine viruses developed at Mahidol University. The protocol includes titration of the individual vaccine virus candidates in adult human volunteers to determine the minimal infectious/immunogenic dose for each virus.
25 This is followed by immunization trials with different bivalent and trivalent combinations of vaccine virus. The final test is the quadravalent vaccine composed of appropriate doses of all four vaccine viruses. If the preliminary trials are successful, larger trials are
30 scheduled, and the vaccine viruses are tested in children,

who are the primary target for vaccine delivery.

We describe herein a preferred method to develop an infectious cDNA clone for a flavivirus. Optimally, a wild-type vaccine virus serves as the template for the clone construction. Large cDNA fragments are amplified from the genomic mRNA by PCR using virus-specific primers and directly cloned into a TA-vector or into the MCS of a low-copy number plasmid following restriction of the amplicon cDNA. The low-copy pBRUC-139 vector contains the MCS of pUC19 to permit convenient cloning of cDNA using a variety of RENZ sites. Other low-copy plasmids are available. The bacteriophage T7 or SP6 promoter is usually engineered into the 5'-terminal mRNA-sense amplicon, and a unique RENZ site for linearization of the recombinant plasmid containing the full-length cDNA must be engineered into the 3-terminal complementary (negative)-sense amplicon. Exhaustive nucleotide analysis of the cDNA clones is desirable.

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APPENDIX APRIMERS DESIGNED FOR DEN-2 CLONING/SEQUENCING PROJECT:

SEQ. ID	PRIMER	MER/SENSE	SEQUENCE
3	pUC/M13-P5	25/+	5'-CCCAGTCACGACGTTGTAAAACGAC-3'
4	pUC/M13-P5B	27/+	5'-GGATGTGCTGCAAGGCGATTAAGTTGG-3'
5	pUC/M13-P3	25/+	5'-TGAGCGGATAACAATTCACACAGG-3'
6	pUC/M13-P3B	27/-	5'-GGCTTTACACTTTATGCTTCCGGCTCG-3'
7	D2-1-ECO.T7 75/+		5'-GCGGATATTG/GAATTC/TCTAGA/ AATTTAATACGACTCACTATA/ AGTTGTTAGTCTACGTGGACCGACAAAGACAG-3' (5'-Fill /EcoRI /XbaI/T7 Promoter/ 5'-end of DEN-2)
8	D2-SMT71	77/+	5'-CCAGT/GAATTC/GAGCTC/ACGCGT/ AAATTTAATACGACTCACTATA/ AGTTGTTAGTCTACGTGGACCGACAAAGACAG-3' (5'-Fill/EcoRI/SstI/MluI/T7 Promoter/ 5'-end of DEN-2)
9	D2-1	24/+	5'-AGTTGTTAGTCTACGTGGACCGAC-3'
10	D2-28	34/+	5'-GACAGATTCTTTGAGGGAGCTGAGCTCAACGTAG-3'
11	D2-134	28/+	5'-TCAATATGCTGAAACGCGAGAGAAACCG-3'
12	cD2-250	26/-	5'-GGGATTGTTAGGAAACGAAGGAACGC-3'
13	D2-274	32/+	5'-CCACCAACAGCAGGGATACTGAAAAGATGGGG-3'
14	cD2-378	25/-	5'-TGCAGATCTGCGTCTCCTATTCAAG-3'
15	D2-528	25/+	5'-CGTGAACATGTGTACCCTCATGGCC-3'
16	cD2-616	26/-	5'-TTGCACCAACAGTCAATGTCTTCAGG-3'
17	D2-616	25/+	5'-ACCAGAAGACATAGATTGTTGGTGC-3'
18	cD2-618	25/-	5'-GCACCAACAGTCTATGTCTTCTGGC-3'
19	cD2-771	25/-	5'-ATGTTTCCAGGCCCTTCTGATGAC-3'
20	D2-847	25/+	5'-GCAGCAATCCTGGCATAACCCATAG-3'
21	D2-996	27/+	5'-GGTTGACATAGTCTTAGAACATGGAAG-3'
22	cD2-996	27/-	5'-CTTCCATGTTCTAAGACTATGTCAACC-3'

SEQ. ID NO:	PRIMER	MER/SENSE	101 SEQUENCE
23	D2-1005	35/+	5'-GTCTTAGAACATGGAAGTTGTGTGACGACGATGGC-3'
24	D2-1141	25/+	5'-ACAACAGAATCTCGCTGCCCAACAC-3'
25	D2-1211	25/+	5'-GCAAACACTCCATGGTAGACAGAGG-3'
26	cD2-1211	25/-	5'-CCTCTGTCTACCATGGAGTGTTC-3'
27	cD2-1227	27/-	5'-CCACATCCATTTCCCCTCTGTCT-3'
28	D2-1261	30/+	5'-GGAAAGGGAGGCATTGTGACCTGTGCTATG-3'
29	D2-1416	28/+	5'-GGAAATCAAATAACACCACAGAGTCC-3'
30	cD2-1503	34/-	5'-CTGCAGCAACACCATCTCATTGAAGTCGAGGCC-3'
31	D2-1510	25/+	5'-GACTTCAATGAGATGGTGTGCTGC-3'
32	cD2-1510	25/+	5'-GCAGCAGCACCATCTCATTGAAGTC-3'
33	D2-1546	28/+	5'-AAGCTTGGCTGGTGCACAGGCAATGGTT-3'
34	cD2-1567	27/-	5'-TGTAACGGCAGGTCTAGGAACCATTG-3'
35	D2-1777	23/+	5'-GGACATCTCAAGTCAGGCTGAG-3'
36	cD2-1777	23/+	5'-CTCAGCCTGCACTTGAGATGTCC-3'
37	D2-1863	27/+	5'-GAAGGAAATAGCAGAAACACAACATGG-3'
38	cD2-1888	33/-	5'-CCCTTCATATTGTA CTGATAACTATTGTTCC-3'
39	D2-2047	32/+	5'-CCTCCATTCGGAGACAGCTACATCATATAGG-3'
40	cD2-2047	32/-	5'-CCTATGATGATGTAGCTGTCTCCGAATGGAGG-3'
41	D2-2170	29/+	5'-ATGGCCATTTTAGGTGACACAGCCTGGGA-3'
42	cD2-2200	27/-	5'-TGTAACACTCCTCCCAGGGATCCAAA-3'
43	D2-2308	29/+	5'-CTCATAGGAGTCATTATCACATGGATAGG-3'
44	cD2-2504	35/-	5'-GGGGATTCTGGTTGGAAC TTATATGTTCTGTCC-3'
45	cD2-2622	30/-	5'-TGATTCAATTCTGGTGTATTTGTTCCAC-3'
46	D2-2702	25/+	5'-AAGGAATCATGCAGGCAGGAAAACG-3'
47	cD2-2864	22/-	5'-ACTTCCAGCGAGTTCCAAGCTC-3' A A
48	D2-2992	25/+	5'-AACAGAGCCGTCCATGCCGATATGG-3'
49	cD2-3105	22/-	5'-TCCATTGCTCCAAAGGGTGTGT-3' G
50	D2-3236	25/+	5'-AGCTTGAGATGGACTTTGATTTCTG-3'

SEQ. ID NO:	PRIMER	MER/SENSE	SEQUENCE
51	cD2-3410	22/-	5'-GGTCTGATTTCCATCCCCTACC-3'
52	D2-3621	23/+	5'-GTCCTTTAGAGACCTGGGAAGAG-3'
53	cD2-3739	25/-	5'-GTTTTCTCAAGAGTAGTCCAGCTGC-3' C
54	D2-3905	25/+	5'-ATCAATTGGCAGTGACTATCATGGC-3'
55	cD2-4002	25/-	5'-TGTTAAGAGCAGTGGAGAAACGGAC-3' A G
56	cD2-4060	25/-	5'-GATTGAGACCTTTGATCGTCAACGC-3'
57	D2-4214	25/+	5'-TGACAGGACCATTAGTGGCTGGAGG-3'
58	D2-4257	34/+	5'-CGTGCTCACTGGACGATCGGCCGATTTGGAAGTGC-3'
59	cD2-4323	24/-	5'-GGGCTGCTTCTGATATTTCTGCC-3' C
60	D2-4497	25/+	5'-CCTGTGGGAAGTGAAGAAACAACGG-3'
61	cD2-4557	30/-	5'-GCTCCATCTTCCAGTTCAGCCTTTCCCATG-3'
62	cD2-4615	25/-	5'-CTCCGGCTCCAATCTGAGAGTATCC-3' G G A
63	D2-4746	25/+	5'-CCTAATATCATATGGAGGAGGCTGG-3'
64	D2-4792	25/+	5'-GAAGGAGAAGAAGTCCAGGTATTGG-3'
65	cD2-4922	25/-	5'-CTGTGCAACAATTGGAGATCCTGACG-3' T T
66	D2-4994	25/+	5'-GTGGAGCATATGTGAGTGCTATAGC-3'
67	D2-5124	25/+	5'-TCTGACTATGGCCGGAAGGTATCTC-3'
68	D2-5173	25/+	5'-ACATTAATCTTGGCCCCACTAGAG-3'
69	cD2-5272	19/-	5'-CGATCTCCCGCCCGGTGTG-3' A
70	cD2-5318	25/-	5'-CTAACTGGTGATAGCAGCCTCATGG-3'
71	cD2-5656	27/-	5'-CCTACTGAGTTGTATCACTTTCTTTCC-3'
72	cD2-5891	26/-	5'-TGGATTTCTTCCATTCTCCCTCTTC-3'
73	D2-5770	25/+	5'-TTCAAGGCTGAGAGGGTTATAGACC-3'
74	D2-6152	25/+	5'-TCTGGTTGGCCTACAGAGTGGCAGC-3'
75	cD2-6252	27/-	5'-CCTTCTTTGTCCAGATTTCCACTTCC-3' A

SEQ. ID NO:	PRIMER	MER/SENSE	103 SEQUENCE
76	D2-6493	35/+	5'-GCGTACAACCATGCTCTCAGTGAAGTCCGGAGAC-3'
77	cD2-6605	24/-	5'-TTCCAGGGTCATCTTCCCTATAC-3' G
78	cD2-6624	31/-	5'-GATGCTAGCCGTGATTATGCAGCACATTCCC-3'
79	D2-6748	25/+	5'-AAACAGAGAACACCCCAAGACAACC-3'
80	cD2-6932	21/-	5'-CGGCATACAGCGTCCATGCTG-3'
81	D2-7055	25/+	5'-GTCTCGGAAAGGATGGCCATTGTC-3'
82	cD2-7195	25/-	5'-CTCTGGTTGCTTTTGCTTGAAGTCC-3' A G G
83	cD2-7217	27/-	5'-CCGCCGCTGCTCTTTTCTGAGCTTCTC-3'
84	D2-7378	25/+	5'-AGGACTACATGGGCTCTGTGTGAGG-3'
85	cD2-7515	19/-	5'-GAGAAGTCCAGCTCCGGCC-3'
86	D2-7769	25/+	5'-AGAGAAACATGGTCACACCAGAAGG-3'
87	cD2-7885	22/-	5'-GTTCTTCGTGTCCTGGTCCTCC-3'
88	D2-8165	25/+	5'-GGAAATATGGAGGAGCCTAGTGAGG-3'
89	cD2-8210	22/-	5'-ACCCAGTACATCTCATGTGTGG-3'
90	D2-8428	28/+	5'-GAGCATGAAACATCATGGCACTATGACC-3'
91	D2-8440	25/+	5'-TCATGGCACTATGACCAAGACCACC-3'
92	cD2-8529	22/-	5'-CAGTCTGACCACTCCGTTCAACC-3' C A G
93	D2-8773	25/+	5'-AAGGTGAGAAGCAATGCAGCCTTGG-3'
94	D2-8798	29/+	5'-GGCCATATTCCTGATGAGAACAAGTGG-3'
95	cD2-8865	22/-	5'-TCTTTCCCTGTCAACCAGCTCC-3' C T
96	D2-9046	25/+	5'-AATGAAGATCACTGGTTCTCCAGAG-3'
97	D2-9131	25/+	5'-ACGTGAGCAAGAAAGAGGGAGGAGC-3'
98	cD2-9166	22/-	5'-TGTCCCATCCTGCTGTGTCATC-3' A G
99	cD2-9234	30/-	5'-GCTAGTTTCTTGTGTTCTCCTTCCATGTGG-3'
100	D2-9344	25/+	5'-TCATATCGAGAAGAGACCAAAGAGG-3'
101	cD2-9429	24/-	5'-ACTCCTTCTCCCTCCATCTGTCTG-3'

SEQ. ID NO:	PRIMER	MER/SENSE	104 SEQUENCE
102	cD2-9438	27/-	5'-ATGCTTTTGAAGATTCCTTCTCCCTCC-3' A C
103	cD2-9468	32/-	5'-GCACAGCGATTTCTTCTGTGATTGTTAGGTGC-3'
104	D2-9645	25/+	5'-ACAATGGGAACCTTCAAGAGGATGG-3'
105	D2-9656.BAM	45/+	5'-TTATCACATT/GGATCC/TTCAAGAGGATGGA ATGATTGGACACAAG-3' (5'-Fill/BamHI/DEN-2 Sequence)
106	cD2-9668	28/-	5'-CAGAAGGGCACTTGTGTCCAATCATTCC-3'
107	cD2-9779	21/-	5'-CTCCCTGGGAAATTCGGGCTC-3' T G
108	cD2-9796	28/-	5'-CCGTCTCCCGCAAAGACCACCCTGCTCC-3'
109	cD2-9796.XBA	44/-	5'-TTATCACCTA/TCTAGA/CCGTCTCCC GCAAAGACCACCCTGCTCC-3'
110	cD2-9913	26/-	5'-GTTGGAACCCAATGTGATGGTACTGC-3'
111	D2-9937	25/+	5'-ACAAGTCGAACAACCTGGTCCATAC-3'
112	cD2-9977	21/-	5'-GCATGTCTTCCGTGGTCATCC-3' T
113	cD2-10003	25/-	5'-CTTGAATCCACACCCTGTTCCAGAC-3'
114	D2-10203	25/+	5'-ATACACAGATTACATGCCATCCATG-3'
115	cD2-10261	21/-	5'-TTTTGCCTTCTACCACAGGAC-3' T A
116	D2-10289	25/-	5'-GAAACAAGGCTAGAAGTCAGGTCCG-3'
117	cD2-10337	23/-	5'-GACGGGGCTCACAGGTAGCATAG-3'
118	D2-10418	25/+	5'-GCCTGTAGCTCCACCTGAGAAGGTG-3'
119	D2-10470	25/+	5'-GGAAGCTGTACGCATGGCGTAGTGG-3'
120	cD2-10530	19/-	5'-GGGCCCCCTTGTGCTGC-3' A
121	cD2-10687	59/-	5'-AGAACCTGTTGATTCAACAGCACCATTCCATTTTCTG-3'
122	cD2-10687.XBA	59/-	5'-TTATCACCTA/GCATGC/TCTAGA/ AGAACCTGTTGATTCAACAGCACCATTCCATTTTCTG-3' (5'-Fill/SphI/XbaI/ 3'-End DEN-2 Sequence)
123	cD2-10687.X2	52/-	5'-TTATCACCTA/TCTAGA/ GAACCTGTTGATTCAACAGCACCATTCCATTTTCTG-3' (5'-Fill/XbaI/ 3'-End DEN-2 Sequence)

While particular embodiments of the invention have been described in detail, it will be apparent to those skilled in the art that these embodiments are exemplary rather than limiting, and the true scope of the invention is that defined within the attached claims.

SEQUENCE LISTING

(1) GENERAL INFORMATION

- (i) APPLICANT: MAHIDOL UNIVERSITY
Bangkok, Thailand

The United States of
America, as represented by the Secretary,
Department of Health and Human Services
c/o Centers for Disease Control and
Prevention
Technology Transfer Office
Mail Stop E-67
1600 Clifton Road
Atlanta, Georgia 30333

- (ii) TITLE OF THE INVENTION: INFECTIOUS CDNA CLONES FOR DENGUE 2
VIRUS ...

- (iii) NUMBER OF SEQUENCES: 137

(iv) CORRESPONDENCE ADDRESS:

- (A) ADDRESSEE: NEEDLE & ROSENBERG, P.C.
(B) STREET: Suite 1200, 127 Peachtree Street, NE
(C) CITY: Atlanta
(D) STATE: GA
(E) COUNTRY: USA
(F) ZIP: 30303

(v) COMPUTER READABLE FORM:

- (A) MEDIUM TYPE: Diskette
(B) COMPUTER: IBM Compatible
(C) OPERATING SYSTEM: DOS
(D) SOFTWARE: FastSEQ Version 1.5

(vi) CURRENT APPLICATION DATA:

- (A) APPLICATION NUMBER: U.S. Serial No. 08/483,292
(B) FILING DATE: 7 Jun 1995
(C) CLASSIFICATION:

(vii) PRIOR APPLICATION DATA:

- (A) APPLICATION NUMBER:
(B) FILING DATE:

(viii) ATTORNEY/AGENT INFORMATION:

- (A) NAME: Spratt, Gwendolyn D.
(B) REGISTRATION NUMBER: 36,016
(C) REFERENCE/DOCKET NUMBER: 14114.0179/P

(ix) TELECOMMUNICATION INFORMATION:

- (A) TELEPHONE: 404-688-0770
(B) TELEFAX: 404-688-9880
(C) TELEX:

(2) INFORMATION FOR SEQ ID NO:1:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 10723 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:
- (ix) FEATURE:

- (A) NAME/KEY: Coding Sequence
- (B) LOCATION: 97...10269
- (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

AGTTGTTAGT CTACGTGGAC CGACAAAGAC AGATTCTTTG AGGGAGCTAA GCTCAACGTA	60
GTTCTAACAG TTTTSTAATT AGAGAGCAGA TCTCTG ATG AAT AAC CAA CGG AAA	114
Met Asn Asn Gln Arg Lys	
1 5	
AAG GCG AAA AAC ACG CCT TTC AAT ATG CTG AAA CGC GAG AGA AAC CGC	162
Lys Ala Lys Asn Thr Pro Phe Asn Met Leu Lys Arg Glu Arg Asn Arg	
10 15 20	
GTG TCG ACT GTG CAA CAG CTG ACA AAG AGA TTC TCA CTT GGA ATG CTG	210
Val Ser Thr Val Gln Gln Leu Thr Lys Arg Phe Ser Leu Gly Met Leu	
25 30 35	
CAG GGA CGA GGA CCA TTA AAA CTG TTC ATG GCC CTG GTG GCG TTC CTT	258
Gln Gly Arg Gly Pro Leu Lys Leu Phe Met Ala Leu Val Ala Phe Leu	
40 45 50	
CGT TTC CTA ACA ATC CCA CCA ACA GCA GGG ATA TTG AAG AGA TGG GGA	306
Arg Phe Leu Thr Ile Pro Pro Thr Ala Gly Ile Leu Lys Arg Trp Gly	
55 60 65 70	
ACA ATT AAA AAA TCA AAA GCT ATT AAT GTT TTG AGA GGG TTC AGG AAA	354
Thr Ile Lys Lys Ser Lys Ala Ile Asn Val Leu Arg Gly Phe Arg Lys	
75 80 85	
GAG ATT GGA AGG ATG CTG AAC ATC TTG AAT AGG AGA CGC AGA TCT GCA	402
Glu Ile Gly Arg Met Leu Asn Ile Leu Asn Arg Arg Arg Arg Ser Ala	
90 95 100	
GGC ATG ATC ATT ATG CTG ATT CCA ACA GTG ATG GCG TTC CAT TTA ACC	450
Gly Met Ile Ile Met Leu Ile Pro Thr Val Met Ala Phe His Leu Thr	
105 110 115	
ACA CGT AAC GGA GAA CCA CAC ATG ATC GTC AGC AGA CAA GAG AAA GGG	498
Thr Arg Asn Gly Glu Pro His Met Ile Val Ser Arg Gln Glu Lys Gly	
120 125 130	

108

AAA Lys 135	AGT Ser	CTT Leu	CTG Leu	TTT Phe	AAA Lys 140	ACA Thr	GAG Glu	GAT Asp	GGC Gly	GTG Val 145	AAC Asn	ATG Met	TGT Cys	ACC Thr	CTC Leu 150	546
ATG Met	GCC Ala	ATG Met	GAC Asp	CTT Leu 155	GGT Gly	GAA Glu	TTG Leu	TGT Cys	GAA Glu 160	GAC Asp	ACA Thr	ATC Ile	ACG Thr	TAC Tyr 165	AAG Lys	594
TGT Cys	CCC Pro	CTT Leu	CTC Leu 170	AGG Arg	CAG Gln	AAT Asn	GAG Glu	CCA Pro 175	GAA Glu	GAC Asp	ATA Ile	GAC Asp	TGT Cys 180	TGG Trp	TGC Cys	642
AAC Asn	TCT Ser	ACG Thr 185	TCC Ser	ACG Thr	TGG Trp	GTA Val	ACT Thr 190	TAT Tyr	GGG Gly	ACG Thr	TGT Cys	ACC Thr 195	ACC Thr	ATG Met	GGA Gly	690
GAA Glu 200	CAT His	AGA Arg	AGA Arg	GAA Glu	AAA Lys 205	AGA Arg	TCA Ser	GTG Val	GCA Ala	CTC Leu	GTT Val 210	CCA Pro	CAT His	GTG Val	GGA Gly	738
ATG Met 215	GGA Gly	CTG Leu	GAG Glu	ACA Thr	CGA Arg 220	ACT Thr	GAA Glu	ACA Thr	TGG Trp	ATG Met 225	TCA Ser	TCA Ser	GAA Glu	GGG Gly	GCC Ala 230	786
TGG Trp	AAA Lys	CAT His	GTC Val	CAG Gln 235	AGA Arg	ATT Ile	GAA Glu	ACT Thr	TGG Trp 240	ATC Ile	TTG Leu	AGA Arg	CAT His	CCA Pro 245	GGC Gly	834
TTC Phe	ACC Thr	ATG Met	ATG Met 250	GCA Ala	GCA Ala	ATC Ile	CTG Leu	GCA Ala 255	TAC Tyr	ACC Thr	ATA Ile	GGA Gly	ACG Thr 260	ACA Thr	CAT His	882
TTC Phe	CAA Gln 265	AGA Arg	GCC Ala	CTG Leu	ATT Ile	TTC Phe	ATC Ile 270	TTA Leu	CTG Leu	ACA Thr	GCT Ala	GTC Val 275	ACT Thr	CCT Pro	TCA Ser	930
ATG Met 280	ACA Thr	ATG Met	CGT Arg	TGC Cys	ATA Ile 285	GGA Gly	ATG Met	TCA Ser	AAT Asn	AGA Arg	GAC Asp 290	TTT Phe	GTG Val	GAA Glu	GGG Gly	978
GTT Val 295	TCA Ser	GGA Gly	GGA Gly	AGC Ser	TGG Trp 300	GTT Val	GAC Asp	ATA Ile	GTC Val	TTA Leu 305	GAA Glu	CAT His	GGA Gly	AGC Ser 310	TGT Cys	1026
GTG Val	ACG Thr	ACG Thr	ATG Met	GCA Ala 315	AAA Lys	AAC Asn	AAA Lys	CCA Pro	ACA Thr 320	TTG Leu	GAT Asp	TTT Phe	GAA Glu 325	CTG Leu	ATA Ile	1074
AAA Lys	ACA Thr	GAA Glu	GCC Ala 330	AAA Lys	CAG Gln	CCT Pro	GCC Ala	ACC Thr 335	CTA Leu	AGG Arg	AAG Lys	TAC Tyr	TGT Cys 340	ATA Ile	GAG Glu	1122
GCA Ala	AAG Lys	CTA Leu 345	ACC Thr	AAC Asn	ACA Thr	ACA Thr	ACA Thr 350	GAA Glu	TCT Ser	CGC Arg	TGC Cys	CCA Pro 355	ACA Thr	CAA Gln	GGG Gly	1170

GAA	CCC	AGC	CTA	AAT	GAA	GAG	CAG	GAC	AAA	AGG	TTC	GTC	TGC	AAA	CAC	1218
Glu	Pro	Ser	Leu	Asn	Glu	Glu	Gln	Asp	Lys	Arg	Phe	Val	Cys	Lys	His	
	360					365					370					
TCC	ATG	GTA	GAC	AGA	GGA	TGG	GGA	AAT	GGA	TGT	GGA	CTA	TTT	GGA	AAG	1266
Ser	Met	Val	Asp	Arg	Gly	Trp	Gly	Asn	Gly	Cys	Gly	Leu	Phe	Gly	Lys	
	375				380					385					390	
GGA	GGC	ATT	GTG	ACC	TGT	GCT	ATG	TTC	AGA	TGC	AAA	AAG	AAC	ATG	GAA	1314
Gly	Gly	Ile	Val	Thr	Cys	Ala	Met	Phe	Arg	Cys	Lys	Lys	Asn	Met	Glu	
				395					400					405		
GGA	AAA	GTT	GTG	CAA	CCA	GAA	AAC	TTG	GAA	TAC	ACC	ATT	GTG	ATA	ACA	1362
Gly	Lys	Val	Val	Gln	Pro	Glu	Asn	Leu	Glu	Tyr	Thr	Ile	Val	Ile	Thr	
			410					415					420			
CCT	CAC	TCA	GGG	GAA	GAG	CAT	GCA	GTC	GGA	AAT	GAC	ACA	GGA	AAA	CAT	1410
Pro	His	Ser	Gly	Glu	Glu	His	Ala	Val	Gly	Asn	Asp	Thr	Gly	Lys	His	
		425					430					435				
GGC	AAG	GAA	ATC	AAA	ATA	ACA	CCA	CAG	AGT	TCC	ATC	ACA	GAA	GCA	GAA	1458
Gly	Lys	Glu	Ile	Lys	Ile	Thr	Pro	Gln	Ser	Ser	Ile	Thr	Glu	Ala	Glu	
	440					445					450					
TTG	ACA	GGT	TAT	GGC	ACT	GTC	ACA	ATG	GAG	TGC	TCT	CCA	AGA	ACG	GGC	1506
Leu	Thr	Gly	Tyr	Gly	Thr	Val	Thr	Met	Glu	Cys	Ser	Pro	Arg	Thr	Gly	
	455			460						465					470	
CTC	GAC	TTC	AAT	GAG	ATG	GTG	TTG	CTG	CAG	ATG	GAA	AAT	AAA	GCT	TGG	1554
Leu	Asp	Phe	Asn	Glu	Met	Val	Leu	Leu	Gln	Met	Glu	Asn	Lys	Ala	Trp	
				475					480					485		
CTG	GTG	CAC	AGG	CAA	TGG	TTC	CTA	GAC	CTG	CCG	TTA	CCA	TGG	TTG	CCC	1602
Leu	Val	His	Arg	Gln	Trp	Phe	Leu	Asp	Leu	Pro	Leu	Pro	Trp	Leu	Pro	
			490					495					500			
GGA	GCG	GAC	ACA	CAA	GGG	TCA	AAT	TGG	ATA	CAG	AAA	GAG	ACA	TTG	GTC	1650
Gly	Ala	Asp	Thr	Gln	Gly	Ser	Asn	Trp	Ile	Gln	Lys	Glu	Thr	Leu	Val	
		505				510						515				
ACT	TTC	AAA	AAT	CCC	CAT	GCG	AAG	AAA	CAG	GAT	GTT	GTT	GTT	TTA	GGA	1698
Thr	Phe	Lys	Asn	Pro	His	Ala	Lys	Lys	Gln	Asp	Val	Val	Val	Leu	Gly	
	520					525					530					
TCC	CAA	GAA	GGG	GCC	ATG	CAC	ACA	GCA	CTT	ACA	GGG	GCC	ACA	GAA	ATC	1746
Ser	Gln	Glu	Gly	Ala	Met	His	Thr	Ala	Leu	Thr	Gly	Ala	Thr	Glu	Ile	
	535				540					545					550	
CAA	ATG	TCA	TCA	GGA	AAC	TTA	CTC	TTC	ACA	GGA	CAT	CTC	AAG	TGC	AGG	1794
Gln	Met	Ser	Ser	Gly	Asn	Leu	Leu	Phe	Thr	Gly	His	Leu	Lys	Cys	Arg	
				555					560					565		
CTG	AGA	ATG	GAC	AAG	CTA	CAG	CTC	AAA	GGA	ATG	TCA	TAC	TCT	ATG	TGC	1842
Leu	Arg	Met	Asp	Lys	Leu	Gln	Leu	Lys	Gly	Met	Ser	Tyr	Ser	Met	Cys	
			570					575					580			
ACA	GGA	AAG	TTT	AAA	GTT	GTG	AAG	GAA	ATA	GCA	GAA	ACA	CAA	CAT	GGA	1890
Thr	Gly	Lys	Phe	Lys	Val	Val	Lys	Glu	Ile	Ala	Glu	Thr	Gln	His	Gly	
		585					590					595				

110

ACA Thr 600	ATA Ile	GTT Val	ATC Ile	AGA Arg	GTG Val	CAA Gln 605	TAT Tyr	GAA Glu	GGG Gly	GAC Asp	GGC Gly 610	TCT Ser	CCA Pro	TGC Cys	AAG Lys	1938
ATC Ile 615	CCT Pro	TTT Phe	GAG Glu	ATA Ile	ATG Met 620	GAT Asp	TTG Leu	GAA Glu	AAA Lys	AGA Arg 625	CAT His	GTC Val	TTA Leu	GGT Gly 630	CGC Arg	1986
CTG Leu	ATT Ile	ACA Thr	GTC Val	AAC Asn 635	CCA Pro	ATT Ile	GTG Val	ACA Thr	GAA Glu 640	AAA Lys	GAT Asp	AGC Ser	CCA Pro	GTC Val 645	AAC Asn	2034
ATA Ile	GAA Glu	GCA Ala	GAA Glu 650	CCT Pro	CCA Pro	TTC Phe	GGA Gly	GAC Asp 655	AGC Ser	TAC Tyr	ATC Ile	ATC Ile	ATA Ile 660	GGA Gly	GTA Val	2082
GAG Glu	CCG Pro	GGA Gly 665	CAA Gln	CTG Leu	AAG Lys	CTC Leu 670	AAC Asn	TGG Trp	TTT Phe	AAG Lys	AAA Lys	GGA Gly 675	AGT Ser	TCT Ser	ATC Ile	2130
GGC Gly 680	CAA Gln	ATG Met	TTT Phe	GAG Glu	ACA Thr 685	ACA Thr	ATG Met	AGG Arg	GGG Gly	GCG Ala	AAG Lys 690	AGA Arg	ATG Met	GCC Ala	ATT Ile	2178
TTA Leu 695	GGT Gly	GAC Asp	ACA Thr	GCC Ala	TGG Trp 700	GAT Asp	TTT Phe	GGA Gly	TCC Ser	TTG Leu 705	GGA Gly	GGA Gly	GTG Val	TTT Phe 710	ACA Thr	2226
TCT Ser	ATA Ile	GGA Gly	AAG Lys	GCT Ala 715	CTC Leu	CAC His	CAA Gln	GTC Val	TTT Phe 720	GGA Gly	GCA Ala	ATC Ile	TAT Tyr	GGA Gly 725	GCT Ala	2274
GCC Ala 730	TTC Phe	AGT Ser	GGG Gly 730	GTT Val	TCA Ser	TGG Trp	ACT Thr	ATG Met 735	AAA Lys	ATC Ile	CTC Leu	ATA Ile	GGA Gly 740	GTC Val	ATT Ile	2322
ATC Ile	ACA Thr	TGG Trp 745	ATA Ile	GGA Gly	ATG Met	AAT Asn 750	TCA Ser	CGC Arg	AGC Ser	ACC Thr	TCA Ser	CTG Leu 755	TCT Ser	GTG Val	ACA Thr	2370
CTA Leu 760	GTA Val	TTG Leu	GTG Val	GGA Gly	ATT Ile	GTG Val 765	ACA Thr	CTG Leu	TAT Tyr	TTG Leu 770	GGA Gly	GTC Val	ATG Met	GTG Val	CAG Gln	2418
GCC Ala 775	GAT Asp	AGT Ser	GGT Gly	TGC Cys 780	GTT Val	GTG Val	AGC Ser	TGG Trp	AAA Lys	AAC Asn 785	AAA Lys	GAA Glu	CTG Leu	AAA Lys	TGT Cys 790	2466
GGC Gly	AGT Ser	GGG Gly	ATT Ile	TTC Phe 795	ATC Ile	ACA Thr	GAC Asp	AAC Asn	GTG Val 800	CAC His	ACA Thr	TGG Trp	ACA Thr	GAA Glu 805	CAA Gln	2514
TAC Tyr	AAG Lys	TTC Phe	CAA Gln 810	CCA Pro	GAA Glu	TCC Ser	CCT Pro	TCA Ser	AAA Lys 815	CTA Leu	GCT Ala	TCA Ser	GCT Ala 820	ATC Ile	CAG Gln	2562
AAA Lys	GCC Ala	CAT His 825	GAA Glu	GAG Glu	GGC Gly	ATT Ile	TGT Cys 830	GGA Gly	ATC Ile	CGC Arg	TCA Ser	GTA Val 835	ACA Thr	AGA Arg	CTG Leu	2610

GAG Glu	AAT Asn	CTG Leu	ATG Met	TGG Trp	AAA Lys	CAA Gln	ATA Ile	ACA Thr	CCA Pro	GAA Glu	TTG Leu	AAT Asn	CAC His	ATT Ile	CTA Leu	2658
	840					845					850					
TCA Ser	GAA Glu	AAT Asn	GAG Glu	GTG Val	AAG Lys	TTA Leu	ACT Thr	ATT Ile	ATG Met	ACA Thr	GGA Gly	GAC Asp	ATC Ile	AAA Lys	GGA Gly	2706
	855				860					865					870	
ATC Ile	ATG Met	CAG Gln	GCA Ala	GGA Gly	AAA Lys	CGA Arg	TCT Ser	CTG Leu	CGG Arg	CCT Pro	CAG Gln	CCC Pro	ACT Thr	GAG Glu	CTG Leu	2754
				875					880					885		
AAG Lys	TAT Tyr	TCA Ser	TGG Trp	AAA Lys	ACA Thr	TGG Trp	GGC Gly	AAA Lys	GCA Ala	AAA Lys	ATG Met	CTC Leu	TCT Ser	ACA Thr	GAG Glu	2802
			890				895						900			
TCT Ser	CAT His	AAC Asn	CAG Gln	ACC Thr	TTT Phe	CTC Leu	ATT Ile	GAT Asp	GGC Gly	CCC Pro	GAA Glu	ACA Thr	GCA Ala	GAA Glu	TGC Cys	2850
		905					910						915			
CCC Pro	AAC Asn	ACA Thr	AAT Asn	AGA Arg	GCT Ala	TGG Trp	AAT Asn	TCG Ser	TTG Leu	GAA Glu	GTT Val	GAA Glu	GAC Asp	TAT Tyr	GGC Gly	2898
		920				925					930					
TTT Phe	GGA Gly	GTA Val	TTC Phe	ACC Thr	ACC Thr	AAT Asn	ATA Ile	TGG Trp	CTA Leu	AAA Lys	TTG Leu	AAA Lys	GAA Glu	AAA Lys	CAG Gln	2946
	935				940					945					950	
GAT Asp	GTA Val	TTC Phe	TGC Cys	GAC Asp	TCA Ser	AAA Lys	CTC Leu	ATG Met	TCA Ser	GCG Ala	GCC Ala	ATA Ile	AAA Lys	GAC Asp	AAC Asn	2994
				955					960					965		
AGA Arg	GCC Ala	GTC Val	CAT His	GCC Ala	GAT Asp	ATG Met	GGT Gly	TAT Tyr	TGG Trp	ATA Ile	GAA Glu	AGT Ser	GCA Ala	CTC Leu	AAT Asn	3042
			970				975						980			
GAC Asp	ACA Thr	TGG Trp	AAG Lys	ATA Ile	GAG Glu	AAA Lys	GCC Ala	TCT Ser	TTC Phe	ATT Ile	GAA Glu	GTT Val	AAA Lys	AAC Asn	TGC Cys	3090
		985				990						995				
CAC His	TGG Trp	CCA Pro	AAA Lys	TCA Ser	CAC His	ACC Thr	CTC Leu	TGG Trp	AGC Ser	AAT Asn	GGA Gly	GTG Val	CTA Leu	GAA Glu	AGT Ser	3138
	1000				1005						1010					
GAG Glu	ATG Met	ATA Ile	ATT Ile	CCA Pro	AAG Lys	AAT Asn	CTC Leu	GCT Ala	GGA Gly	CCA Pro	GTG Val	TCT Ser	CAA Gln	CAC His	AAC Asn	3186
	1015				1020					1025					1030	
TAT Tyr	AGA Arg	CCA Pro	GGC Gly	TAC Tyr	CAT His	ACA Thr	CAA Gln	ATA Ile	ACA Thr	GGA Gly	CCA Pro	TGG Trp	CAT His	CTA Leu	GGT Gly	3234
			1035						1040					1045		
AAG Lys	CTT Leu	GAG Glu	ATG Met	GAC Asp	TTT Phe	GAT Asp	TTC Phe	TGT Cys	GAT Asp	GGA Gly	ACA Thr	ACA Thr	GTG Val	GTA Val	GTG Val	3282
			1050				1055						1060			
ACT Thr	GAG Glu	GAC Asp	TGC Cys	GGA Gly	AAT Asn	AGA Arg	GGA Gly	CCC Pro	TCT Ser	TTG Leu	AGA Arg	ACA Thr	ACC Thr	ACT Thr	GCC Ala	3330
	1065					1070						1075				

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TCT Ser 1080	GGA Gly	AAA Lys	CTC Leu	ATA Ile	ACA Thr	GAA Glu	TGG Trp	TGC Cys	TGC Cys	CGA Arg	TCT Ser	TGC Cys	ACA Thr	TTA Leu	CCA Pro	3378
CCG Pro 1095	CTA Leu	AGA Arg	TAC Tyr	AGA Arg	GGT Gly	GAG Glu	GAT Asp	GGG Gly	TGC Cys	TGG Trp	TAC Tyr	GGG Gly	ATG Met	GAA Glu	ATC Ile	3426
AGA Arg	CCA Pro	TTG Leu	AAG Lys	GAG Glu	AAA Lys	GAA Glu	GAG Glu	AAT Asn	TTG Leu	GTC Val	AAC Asn	TCC Ser	TTG Leu	GTC Val	ACA Thr	3474
GCT Ala	GGA Gly	CAT His	GGG Gly	CAG Gln	GTC Val	GAC Asp	AAC Asn	TTT Phe	TCA Ser	CTA Leu	GGA Gly	GTC Val	TTG Leu	GGA Gly	ATG Met	3522
GCA Ala	TTG Leu	TTC Phe	CTG Leu	GAG Glu	GAA Glu	ATG Met	CTT Leu	AGG Arg	ACC Thr	CGA Arg	GTA Val	GGA Gly	ACG Thr	AAA Lys	CAT His	3570
GCA Ala	ATA Ile	CTA Leu	CTA Leu	GTT Val	GCA Ala	GTT Val	TCT Ser	TTT Phe	GTG Val	ACA Thr	TTG Leu	ATC Ile	ACA Thr	GGG Gly	AAC Asn	3618
ATG Met 1175	TCC Ser	TTT Phe	AGA Arg	GAC Asp	CTG Leu	GGA Gly	AGA Arg	GTG Val	ATG Met	GTT Val	ATG Met	GTA Val	GGC Gly	GCC Ala	ACT Thr	3666
ATG Met	ACG Thr	GAT Asp	GAC Asp	ATA Ile	GGT Gly	ATG Met	GGC Gly	GTG Val	ACT Thr	TAT Tyr	CTT Leu	GCC Ala	CTA Leu	CTA Leu	GCA Ala	3714
GCC Ala	TTC Phe	AAA Lys	GTC Val	AGA Arg	CCA Pro	ACT Thr	TTT Phe	GCA Ala	GCT Ala	GGA Gly	CTA Leu	CTC Leu	TTG Leu	AGA Arg	AAG Lys	3762
CTG Leu	ACC Thr	TCC Ser	AAG Lys	GAA Glu	TTG Leu	ATG Met	ATG Met	ACT Thr	ACT Thr	ATA Ile	GGA Gly	ATT Ile	GTA Val	CTC Leu	CTC Leu	3810
TCC Ser	CAG Gln	AGC Ser	ACC Thr	ATA Ile	CCA Pro	GAG Glu	ACC Thr	ATT Ile	CTT Leu	GAG Glu	TTG Leu	ACT Thr	GAT Asp	GCG Ala	TTA Leu	3858
GCC Ala 1255	TTA Leu	GGC Gly	ATG Met	ATG Met	GTC Val	CTC Leu	AAA Lys	ATG Met	GTG Val	AGA Arg	AAT Asn	ATG Met	GAA Glu	AAG Lys	TAT Tyr	3906
CAA Gln	TTG Leu	GCA Ala	GTG Val	ACT Thr	ATC Ile	ATG Met	GCT Ala	ATC Ile	TTG Leu	TGC Cys	GTC Val	CCA Pro	AAC Asn	GCA Ala	GTG Val	3954
ATA Ile	TTA Leu	CAA Gln	AAC Asn	GCA Ala	TGG Trp	AAA Lys	GTG Val	AGT Ser	TGC Cys	ACA Thr	ATA Ile	TTG Leu	GCA Ala	GTG Val	GTG Val	4002
TCC Ser	GTT Val	TCC Ser	CCA Pro	CTG Leu	CTC Leu	TTA Leu	ACA Thr	TCC Ser	TCA Ser	CAG Gln	CAA Gln	AAA Lys	ACA Thr	GAT Asp	TGG Trp	4050

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ATA CCA TTA GCA TTG ACG ATC AAA GGT CTC AAT CCA ACA GCT ATT TTT	4098
Ile Pro Leu Ala Leu Thr Ile Lys Gly Leu Asn Pro Thr Ala Ile Phe	
1320 1325 1330	
CTA ACA ACC CTC TCA AGA ACC AGC AAG AAA AGG AGC TGG CCA TTA AAT	4146
Leu Thr Thr Leu Ser Arg Thr Ser Lys Lys Arg Ser Trp Pro Leu Asn	
1335 1340 1345 1350	
GAG GCT ATC ATG GCA GTC GGG ATG GTG AGC ATT TTA GCC AGT TCT CTC	4194
Glu Ala Ile Met Ala Val Gly Met Val Ser Ile Leu Ala Ser Ser Leu	
1355 1360 1365	
CTA AAA AAT GAT ATT CCC ATG ACA GGA CCA TTA GTG GCT GGA GGG CTC	4242
Leu Lys Asn Asp Ile Pro Met Thr Gly Pro Leu Val Ala Gly Gly Leu	
1370 1375 1380	
CTC ACT GTG TGC TAC GTG CTC ACT GGA CGA TCG GCC GAT TTG GAA CTG	4290
Leu Thr Val Cys Tyr Val Leu Thr Gly Arg Ser Ala Asp Leu Glu Leu	
1385 1390 1395	
GAG AGA GCA GCC GAT GTC AAA TGG GAA GAC CAG GCA GAG ATA TCA GGA	4338
Glu Arg Ala Ala Asp Val Lys Trp Glu Asp Gln Ala Glu Ile Ser Gly	
1400 1405 1410	
AGC AGT CCA ATC CTG TCA ATA ACA ATA TCA GAA GAT GGT AGC ATG TCG	4386
Ser Ser Pro Ile Leu Ser Ile Thr Ile Ser Glu Asp Gly Ser Met Ser	
1415 1420 1425 1430	
ATA AAA AAT GAA GAG GAA GAA CAA ACA CTG ACC ATA CTC ATT AGA ACA	4434
Ile Lys Asn Glu Glu Glu Glu Gln Thr Leu Thr Ile Leu Ile Arg Thr	
1435 1440 1445	
GGA TTG CTG GTG ATC TCA GGA CTT TTT CCT GTA TCA ATA CCA ATC ACG	4482
Gly Leu Leu Val Ile Ser Gly Leu Phe Pro Val Ser Ile Pro Ile Thr	
1450 1455 1460	
GCA GCA GCA TGG TAC CTG TGG GAA GTG AAG AAA CAA CGG GCC GGA GTA	4530
Ala Ala Ala Trp Tyr Leu Trp Glu Val Lys Lys Gln Arg Ala Gly Val	
1465 1470 1475	
TTG TGG GAT GTT CCT TCA CCC CCA CCC ATG GGA AAG GCT GAA CTG GAA	4578
Leu Trp Asp Val Pro Ser Pro Pro Pro Met Gly Lys Ala Glu Leu Glu	
1480 1485 1490	
GAT GGA GCC TAT AGA ATT AAG CAA AAA GGG ATT CTT GGA TAT TCC CAG	4626
Asp Gly Ala Tyr Arg Ile Lys Gln Lys Gly Ile Leu Gly Tyr Ser Gln	
1495 1500 1505 1510	
ATC GGA GCC GGA GTT TAC AAA GAA GGA ACA TTC CAT ACA ATG TGG CAT	4674
Ile Gly Ala Gly Val Tyr Lys Glu Gly Thr Phe His Thr Met Trp His	
1515 1520 1525	
GTC ACA CGT GGC GCT GTT CTA ATG CAT AAA GGA AAG AGG ATT GAA CCA	4722
Val Thr Arg Gly Ala Val Leu Met His Lys Gly Lys Arg Ile Glu Pro	
1530 1535 1540	
TCA TGG GCG GAC GTC AAG AAA GAC CTA ATA TCA TAT GGA GGA GGC TGG	4770
Ser Trp Ala Asp Val Lys Lys Asp Leu Ile Ser Tyr Gly Gly Gly Trp	
1545 1550 1555	

AAG TTA GAA GGA GAA TGG AAG GAA GGA GAA GAA GTC CAG GTA TTG GCA Lys Leu Glu Gly Glu Trp Lys Glu Gly Glu Glu Val Gln Val Leu Ala 1560 1565 1570	4818
CTG GAG CCT GGA AAA AAT CCA AGA GCC GTC CAA ACG AAA CCT GGT CTT Leu Glu Pro Gly Lys Asn Pro Arg Ala Val Gln Thr Lys Pro Gly Leu 1575 1580 1585 1590	4866
TTC AAA ACC AAC GCC GGA ACA ATA GGT GCT GTA TCT CTG GAC TTT TCT Phe Lys Thr Asn Ala Gly Thr Ile Gly Ala Val Ser Leu Asp Phe Ser 1595 1600 1605	4914
CCT GGA ACG TCA GGA TCT CCA ATT ATC GAC AAA AAA GGA AAA GTT GTG Pro Gly Thr Ser Gly Ser Pro Ile Ile Asp Lys Lys Gly Lys Val Val 1610 1615 1620	4962
GGT CTT TAT GGT AAT GGT GTT GTT ACA AGG AGT GGA GCA TAT GTG AGT Gly Leu Tyr Gly Asn Gly Val Val Thr Arg Ser Gly Ala Tyr Val Ser 1625 1630 1635	5010
GCT ATA GCC CAG ACT GAA AAA AGC ATT GAA GAC AAC CCA GAG ATC GAA Ala Ile Ala Gln Thr Glu Lys Ser Ile Glu Asp Asn Pro Glu Ile Glu 1640 1645 1650	5058
GAT GAC ATT TTC CGA AAG AGA AGA CTG ACC ATC ATG GAC CTC CAC CCA Asp Asp Ile Phe Arg Lys Arg Arg Leu Thr Ile Met Asp Leu His Pro 1655 1660 1665 1670	5106
GGA GCG GGA AAG ACG AAG AGA TAC CTT CCG GCC ATA GTC AGA GAA GCT Gly Ala Gly Lys Thr Lys Arg Tyr Leu Pro Ala Ile Val Arg Glu Ala 1675 1680 1685	5154
ATA AAA CGG GGT TTG AGA ACA TTA ATC TTG GCC CCC ACT AGA GTT GTG Ile Lys Arg Gly Leu Arg Thr Leu Ile Leu Ala Pro Thr Arg Val Val 1690 1695 1700	5202
GCA GCT GAA ATG GAG GAA GCC CTT AGA GGA CTT CCA ATA AGA TAC CAG Ala Ala Glu Met Glu Glu Ala Leu Arg Gly Leu Pro Ile Arg Tyr Gln 1705 1710 1715	5250
ACC CCA GCC ATC AGA GCT GAG CAC ACC GGG CGG GAG ATT GTG GAC CTA Thr Pro Ala Ile Arg Ala Glu His Thr Gly Arg Glu Ile Val Asp Leu 1720 1725 1730	5298
ATG TGT CAT GCC ACA TTT ACC ATG AGG CTG CTA TCA CCA GTT AGA GTG Met Cys His Ala Thr Phe Thr Met Arg Leu Leu Ser Pro Val Arg Val 1735 1740 1745 1750	5346
CCA AAC TAC AAC CTG ATT ATC ATG GAC GAA GCC CAT TTC ACA GAC CCA Pro Asn Tyr Asn Leu Ile Ile Met Asp Glu Ala His Phe Thr Asp Pro 1755 1760 1765	5394
GCA AGT ATA GCA GCT AGA GGA TAC ATC TCA ACT CGA GTG GAG ATG GGT Ala Ser Ile Ala Ala Arg Gly Tyr Ile Ser Thr Arg Val Glu Met Gly 1770 1775 1780	5442
GAG GCA GCT GGG ATT TTT ATG ACA GCC ACT CCC CCG GGA AGC AGA GAC Glu Ala Ala Gly Ile Phe Met Thr Ala Thr Pro Pro Gly Ser Arg Asp 1785 1790 1795	5490

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CCA TTT CCT CAG AGC AAT GCA CCA ATC ATA GAT GAA GAA AGA GAA ATC	5538
Pro Phe Pro Gln Ser Asn Ala Pro Ile Ile Asp Glu Glu Arg Glu Ile	
1800 1805 1810	
CCT GAA CGC TCG TGG AAT TCC GGA CAT GAA TGG GTC ACG GAT TTT AAA	5586
Pro Glu Arg Ser Trp Asn Ser Gly His Glu Trp Val Thr Asp Phe Lys	
1815 1820 1825 1830	
GGG AAG ACT GTT TGG TTC GTT CCA AGT ATA AAA GCA GGA AAT GAT ATA	5634
Gly Lys Thr Val Trp Phe Val Pro Ser Ile Lys Ala Gly Asn Asp Ile	
1835 1840 1845	
GCA GCT TGC CTG AGG AAA AAT GGA AAG AAA GTG ATA CAA CTC AGT AGG	5682
Ala Ala Cys Leu Arg Lys Asn Gly Lys Lys Val Ile Gln Leu Ser Arg	
1850 1855 1860	
AAG ACC TTT GAT TCT GAG TAT GTC AAG ACT AGA ACC AAT GAT TGG GAC	5730
Lys Thr Phe Asp Ser Glu Tyr Val Lys Thr Arg Thr Asn Asp Trp Asp	
1865 1870 1875	
TTC GTG GTT ACA ACT GAC ATT TCA GAA ATG GGT GCC AAT TTC AAG GCT	5778
Phe Val Val Thr Thr Asp Ile Ser Glu Met Gly Ala Asn Phe Lys Ala	
1880 1885 1890	
GAG AGG GTT ATA GAC CCC AGA CGC TGC ATG AAA CCA GTC ATA CTA ACA	5826
Glu Arg Val Ile Asp Pro Arg Arg Cys Met Lys Pro Val Ile Leu Thr	
1895 1900 1905 1910	
GAT GGT GAA GAG CGG GTG ATT CTG GCA GGA CCT ATG CCA GTG ACC CAC	5874
Asp Gly Glu Glu Arg Val Ile Leu Ala Gly Pro Met Pro Val Thr His	
1915 1920 1925	
TCT AGT GCA GCA CAA AGA AGA GGG AGA ATA GGA AGA AAT CCA AAA AAT	5922
Ser Ser Ala Ala Gln Arg Arg Gly Arg Ile Gly Arg Asn Pro Lys Asn	
1930 1935 1940	
GAG AAT GAC CAG TAC ATA TAC ATG GGG GAA CCT CTG GAA AAT GAT GAA	5970
Glu Asn Asp Gln Tyr Ile Tyr Met Gly Glu Pro Leu Glu Asn Asp Glu	
1945 1950 1955	
GAC TGT GCA CAC TGG AAA GAA GCT AAA ATG CTC CTA GAT AAC ATC AAC	6018
Asp Cys Ala His Trp Lys Glu Ala Lys Met Leu Leu Asp Asn Ile Asn	
1960 1965 1970	
ACG CCA GAA GGA ATC ATT CCT AGC ATG TTC GAA CCA GAG CGT GAA AAG	6066
Thr Pro Glu Gly Ile Ile Pro Ser Met Phe Glu Pro Glu Arg Glu Lys	
1975 1980 1985 1990	
GTG GAT GCC ATT GAT GGC GAA TAC CGC TTG AGA GGA GAA GCA AGG AAA	6114
Val Asp Ala Ile Asp Gly Glu Tyr Arg Leu Arg Gly Glu Ala Arg Lys	
1995 2000 2005	
ACC TTT GTA GAC TTA ATG AGA AGA GGA GAC CTA CCA GTC TGG TTG GCC	6162
Thr Phe Val Asp Leu Met Arg Arg Gly Asp Leu Pro Val Trp Leu Ala	
2010 2015 2020	
TAC AGA GTG GCA GCT GAA GGC ATC AAC TAC GCA GAC AGA AGG TGG TGT	6210
Tyr Arg Val Ala Ala Glu Gly Ile Asn Tyr Ala Asp Arg Arg Trp Cys	
2025 2030 2035	

TTT GAT GGA GTC AAG AAC AAC CAA ATC CTA GAA GAA AAC GTG GAA GTT Phe Asp Gly Val Lys Asn Asn Gln Ile Leu Glu Glu Asn Val Glu Val 2040 2045 2050	6258
GAA ATC TGG ACA AAA GAA GGG GAA AGG AAG AAA TTG AAA CCC AGA TGG Glu Ile Trp Thr Lys Glu Gly Glu Arg Lys Lys Leu Lys Pro Arg Trp 2055 2060 2065 2070	6306
TTG GAT GCT AGG ATC TAT TCT GAC CCA CTG GCG CTA AAA GAA TTT AAG Leu Asp Ala Arg Ile Tyr Ser Asp Pro Leu Ala Leu Lys Glu Phe Lys 2075 2080 2085	6354
GAA TTT GCA GCC GGA AGA AAG TCT CTG ACC CTG AAC CTA ATC ACA GAA Glu Phe Ala Ala Gly Arg Lys Ser Leu Thr Leu Asn Leu Ile Thr Glu 2090 2095 2100	6402
ATG GGT AGG CTC CCA ACC TTC ATG ACT CAG AAG GCA AGA GAC GCA CTG Met Gly Arg Leu Pro Thr Phe Met Thr Gln Lys Ala Arg Asp Ala Leu 2105 2110 2115	6450
GAC AAC TTA GCA GTG CTG CAC ACG GCT GAG GCA GGT GGA AGG GCG TAC Asp Asn Leu Ala Val Leu His Thr Ala Glu Ala Gly Gly Arg Ala Tyr 2120 2125 2130	6498
AAC CAT GCT CTC AGT GAA CTG CCG GAG ACC CTG GAG ACA TTG CTT TTA Asn His Ala Leu Ser Glu Leu Pro Glu Thr Leu Glu Thr Leu Leu Leu 2135 2140 2145 2150	6546
CTG ACA CTT CTG GCT ACA GTC ACG GGA GGG ATC TTT TTA TTC TTG ATG Leu Thr Leu Leu Ala Thr Val Thr Gly Gly Ile Phe Leu Phe Leu Met 2155 2160 2165	6594
AGC GGA AGG GGC ATA GGG AAG ATG ACC CTG GGA ATG TGC TGC ATA ATC Ser Gly Arg Gly Ile Gly Lys Met Thr Leu Glu Met Cys Cys Ile Ile 2170 2175 2180	6642
ACG GCT AGC ATC CTC CTA TGG TAC GCA CAA ATA CAG CCA CAC TGG ATA Thr Ala Ser Ile Leu Leu Trp Tyr Ala Gln Ile Gln Pro His Trp Ile 2185 2190 2195	6690
GCA GCT TCA ATA ATA CTG GAG TTT TTT CTC ATA GTT TTG CTT ATT CCA Ala Ala Ser Ile Ile Leu Glu Phe Phe Leu Ile Val Leu Leu Ile Pro 2200 2205 2210	6738
GAA CCT GAA AAA CAG AGA ACA CCC CAA GAC AAC CAA CTG ACC TAC GTT Glu Pro Glu Lys Gln Arg Thr Pro Gln Asp Asn Gln Leu Thr Tyr Val 2215 2220 2225 2230	6786
GTC ATA GCC ATC CTC ACA GTG GTG GCC GCA ACC ATG GCA AAC GAG ATG Val Ile Ala Ile Leu Thr Val Val Ala Ala Thr Met Ala Asn Glu Met 2235 2240 2245	6834
GGT TTC CTA GAA AAA ACG AAG AAA GAT CTC GGA TTG GGA AGC ATT GCA Gly Phe Leu Glu Lys Thr Lys Lys Asp Leu Gly Leu Gly Ser Ile Ala 2250 2255 2260	6882
ACC CAG CAA CCC GAG AGC AAC ATC CTG GAC ATA GAT CTA CGT CCT GCA Thr Gln Gln Pro Glu Ser Asn Ile Leu Asp Ile Asp Leu Arg Pro Ala 2265 2270 2275	6930

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TCA GCA TGG ACG CTG TAT GCC GTG GCC ACA ACA TTT GTT ACA CCA ATG Ser Ala Trp Thr Leu Tyr Ala Val Ala Thr Thr Phe Val Thr Pro Met 2280 2285 2290	6978
TTG AGA CAT AGC ATT GAA AAT TCC TCA GTG AAT GTG TCC CTA ACA GCT Leu Arg His Ser Ile Glu Asn Ser Ser Val Asn Val Ser Leu Thr Ala 2295 2300 2305 2310	7026
ATA GCC AAC CAA GCC ACA GTG TTA ATG GGT CTC GGG AAA GGA TGG CCA Ile Ala Asn Gln Ala Thr Val Leu Met Gly Leu Gly Lys Gly Trp Pro 2315 2320 2325	7074
TTG TCA AAG ATG GAC ATC GGA GTT CCC CTT CTC GCC ATT GGA TGC TAC Leu Ser Lys Met Asp Ile Gly Val Pro Leu Leu Ala Ile Gly Cys Tyr 2330 2335 2340	7122
TCA CAA GTC AAC CCC ATA ACT CTC ACA GCA GCT CTT TTC TTA TTG GTA Ser Gln Val Asn Pro Ile Thr Leu Thr Ala Ala Leu Phe Leu Leu Val 2345 2350 2355	7170
GCA CAT TAT GCC ATC ATA GGG CCA GGA CTC CAA GCA AAA GCA ACC AGA Ala His Tyr Ala Ile Ile Gly Pro Gly Leu Gln Ala Lys Ala Thr Arg 2360 2365 2370	7218
GAA GCT CAG AAA AGA GCA GCG GCG GGC ATC ATG AAA AAC CCA ACT GTC Glu Ala Gln Lys Arg Ala Ala Ala Gly Ile Met Lys Asn Pro Thr Val 2375 2380 2385 2390	7266
GAT GGA ATA ACA GTG ATT GAC CTA GAT CCA ATA CCT TAT GAT CCA AAG Asp Gly Ile Thr Val Ile Asp Leu Asp Pro Ile Pro Tyr Asp Pro Lys 2395 2400 2405	7314
TTT GAA AAG CAG TTG GGA CAA GTA ATG CTC CTA GTC CTC TGC GTG ACT Phe Glu Lys Gln Leu Gly Gln Val Met Leu Leu Val Leu Cys Val Thr 2410 2415 2420	7362
CAA GTA TTG ATG ATG AGG ACT ACA TGG GCT CTG TGT GAG GCT TTA ACC Gln Val Leu Met Met Arg Thr Thr Trp Ala Leu Cys Glu Ala Leu Thr 2425 2430 2435	7410
TTA GCT ACC GGG CCC ATC TCC ACA TTG TGG GAA GGA AAT CCA GGG AGG Leu Ala Thr Gly Pro Ile Ser Thr Leu Trp Glu Gly Asn Pro Gly Arg 2440 2445 2450	7458
TTT TGG AAC ACT ACC ATT GCG GTG TCA ATG GCT AAC ATT TTT AGA GGG Phe Trp Asn Thr Thr Ile Ala Val Ser Met Ala Asn Ile Phe Arg Gly 2455 2460 2465 2470	7506
AGT TAC TTG GCC GGA GCT GGA CTT CTC TTT TCT ATT ATG AAG AAC ACA Ser Tyr Leu Ala Gly Ala Gly Leu Leu Phe Ser Ile Met Lys Asn Thr 2475 2480 2485	7554
ACC AAC ACA AGA AGG GGA ACT GGC AAC ATA GGA GAG ACG CTT GGA GAG Thr Asn Thr Arg Arg Gly Thr Gly Asn Ile Gly Glu Thr Leu Gly Glu 2490 2495 2500	7602
AAA TGG AAA AGC CGA TTG AAC GCA TTG GGA AAA AGT GAA TTC CAG ATC Lys Trp Lys Ser Arg Leu Asn Ala Leu Gly Lys Ser Glu Phe Gln Ile 2505 2510 2515	7650

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TAC Tyr 2520	AAG Lys 2520	AAA Lys 2520	AGT Ser 2520	GGA Gly 2520	ATC Ile 2525	CAG Gln 2525	GAA Glu 2530	GTG Val 2530	GAT Asp 2530	AGA Arg 2530	ACC Thr 2530	TTA Leu 2530	GCA Ala 2530	AAA Lys 2530	GAA Glu 2530	7698
GGC Gly 2535	ATT Ile 2535	AAA Lys 2540	AGA Arg 2540	GGA Gly 2540	GAA Glu 2540	ACG Thr 2545	GAC Asp 2545	CAT His 2545	CAC His 2545	GCT Ala 2545	GTG Val 2550	TCG Ser 2550	CGA Arg 2550	GGC Gly 2550	TCA Ser 2550	7746
GCA Ala 2555	AAA Lys 2555	CTG Leu 2555	AGA Arg 2555	TGG Trp 2555	TTC Phe 2560	GTT Val 2560	GAG Glu 2560	AGA Arg 2560	AAC Asn 2560	ATG Met 2565	GTC Val 2565	ACA Thr 2565	CCA Pro 2565	GAA Glu 2565	GGG Gly 2565	7794
AAA Lys 2570	GTA Val 2570	GTG Val 2570	GAC Asp 2570	CTC Leu 2575	GGT Gly 2575	TGT Cys 2575	GGC Gly 2575	AGA Arg 2575	GGA Gly 2580	GGC Gly 2580	TGG Trp 2580	TCA Ser 2580	TAC Tyr 2580	TAT Tyr 2580	TGT Cys 2580	7842
GGA Gly 2585	GGA Gly 2585	CTA Leu 2585	AAG Lys 2590	AAT Asn 2590	GTA Val 2590	AGA Arg 2590	GAA Glu 2590	GTC Val 2595	AAA Lys 2595	GGC Gly 2595	CTA Leu 2595	ACA Thr 2595	AAA Lys 2595	GGA Gly 2595	GGA Gly 2595	7890
CCA Pro 2600	GGA Gly 2600	CAC His 2600	GAA Glu 2605	GAA Glu 2605	CCC Pro 2605	ATC Ile 2605	CCC Pro 2610	ATG Met 2610	TCA Ser 2610	ACA Thr 2610	TAT Tyr 2610	GGG Gly 2610	TGG Trp 2610	AAT Asn 2610	CTA Leu 2610	7938
GTG Val 2615	CGT Arg 2615	CTT Leu 2620	CAA Gln 2620	AGT Ser 2620	GGA Gly 2620	GTT Val 2625	GAC Asp 2625	GTT Val 2625	TTC Phe 2625	TTC Phe 2625	ATC Ile 2630	CCG Pro 2630	CCA Pro 2630	GAA Glu 2630	AAG Lys 2630	7986
TGT Cys 2635	GAC Asp 2635	ACA Thr 2635	TTA Leu 2635	TTG Leu 2640	TGT Cys 2640	GAC Asp 2640	ATA Ile 2640	GGG Gly 2645	GAG Glu 2645	TCA Ser 2645	TCA Ser 2645	CCA Pro 2645	AAT Asn 2645	CCC Pro 2645	ACA Thr 2645	8034
GTG Val 2650	GAA Glu 2650	GCA Ala 2650	GGA Gly 2650	CGA Arg 2650	ACA Thr 2655	CTC Leu 2655	AGA Arg 2655	GTC Val 2655	CTT Leu 2660	AAC Asn 2660	TTA Leu 2660	GTA Val 2660	GAA Glu 2660	AAT Asn 2660	TGG Trp 2660	8082
TTG Leu 2665	AAC Asn 2665	AAC Asn 2665	ACT Thr 2670	CAA Gln 2670	TTT Phe 2670	TGC Cys 2670	ATA Ile 2675	AAG Lys 2675	GTT Val 2675	CTC Leu 2675	AAC Asn 2675	CCA Pro 2675	TAT Tyr 2675	ATG Met 2675		8130
CCC Pro 2680	TCA Ser 2680	GTC Val 2685	ATA Ile 2685	GAA Glu 2685	AAA Lys 2685	ATG Met 2690	GAA Glu 2690	GCA Ala 2690	CTA Leu 2690	CAA Gln 2690	AGG Arg 2690	AAA Lys 2690	TAT Tyr 2690	GGA Gly 2690	GGA Gly 2690	8178
GCC Ala 2695	TTA Leu 2695	GTG Val 2700	AGG Arg 2700	AAT Asn 2700	CCA Pro 2705	CTC Leu 2705	TCA Ser 2705	CGA Arg 2705	AAC Asn 2705	TCC Ser 2705	ACA Thr 2710	CAT His 2710	GAG Glu 2710	ATG Met 2710	TAC Tyr 2710	8226
TGG Trp 2715	GTA Val 2715	TCC Ser 2715	AAT Asn 2715	GCT Ala 2715	TCC Ser 2720	GGG Gly 2720	AAC Asn 2720	ATA Ile 2725	GTG Val 2725	TCA Ser 2725	TCA Ser 2725	GTG Val 2725	AAC Asn 2725	ATG Met 2725	ATT Ile 2725	8274
TCA Ser 2730	AGG Arg 2730	ATG Met 2730	TTG Leu 2730	ATC Ile 2735	AAC Asn 2735	AGA Arg 2735	TTT Phe 2735	ACA Thr 2735	ATG Met 2735	AGA Arg 2735	TAC Tyr 2740	AAG Lys 2740	AAA Lys 2740	GCC Ala 2740	ACT Thr 2740	8322
TAC Tyr 2745	GAG Glu 2745	CCG Pro 2745	GAT Asp 2750	GTT Val 2750	GAC Asp 2750	CTC Leu 2750	GGA Gly 2750	AGC Ser 2750	GGA Gly 2755	ACC Thr 2755	CGT Arg 2755	AAC Asn 2755	ATC Ile 2755	GGG Gly 2755	ATT Ile 2755	8370

GAA AGT GAG ATA CCA AAC CTA GAT ATA ATT GGG AAA AGA ATA GAA AAA Glu Ser Glu Ile Pro Asn Leu Asp Ile Ile Gly Lys Arg Ile Glu Lys 2760 2765 2770	8418
ATA AAG CAA GAG CAT GAA ACA TCA TGG CAC TAT GAC CAA GAC CAC CCA Ile Lys Gln Glu His Glu Thr Ser Trp His Tyr Asp Gln Asp His Pro 2775 2780 2785 2790	8466
TAC AAA ACG TGG GCA TAC CAT GGT AGC TAT GAA ACA AAA CAG ACT GGA Tyr Lys Thr Trp Ala Tyr His Gly Ser Tyr Glu Thr Lys Gln Thr Gly 2795 2800 2805	8514
TCA GCA TCA TCC ATG GTC AAC GGA GTG GTC AGG CTG CTG ACA AAA CCT Ser Ala Ser Ser Met Val Asn Gly Val Val Arg Leu Leu Thr Lys Pro 2810 2815 2820	8562
TGG GAC GTC GTC CCC ATG GTG ACA CAG ATG GCA ATG ACA GAC ACG ACT Trp Asp Val Val Pro Met Val Thr Gln Met Ala Met Thr Asp Thr Thr 2825 2830 2835	8610
CCA TTT GGA CAA CAG CGC GTT TTT AAA GAG AAA GTG GAC ACG AGA ACC Pro Phe Gly Gln Gln Arg Val Phe Lys Glu Lys Val Asp Thr Arg Thr 2840 2845 2850	8658
CAA GAA CCG AAA GAA GGC ACG AAG AAA CTA ATG AAA ATA ACA GCA GAG Gln Glu Pro Lys Glu Gly Thr Lys Lys Leu Met Lys Ile Thr Ala Glu 2855 2860 2865 2870	8706
TGG CTT TGG AAA GAA TTA GGG AAG AAA AAG ACA CCC AGG ATG TGC ACC Trp Leu Trp Lys Glu Leu Gly Lys Lys Lys Thr Pro Arg Met Cys Thr 2875 2880 2885	8754
AGA GAA GAA TTC ACA AGA AAG GTG AGA AGC AAT GCA GCC TTG GGG GCC Arg Glu Glu Phe Thr Arg Lys Val Arg Ser Asn Ala Ala Leu Gly Ala 2890 2895 2900	8802
ATA TTC ACT GAT GAG AAC AAG TGG AAG TCG GCA CGT GAG GCT GTT GAA Ile Phe Thr Asp Glu Asn Lys Trp Lys Ser Ala Arg Glu Ala Val Glu 2905 2910 2915	8850
GAT AGT AGG TTT TGG GAG CTG GTT GAC AAG GAA AGG AAT CTC CAT CTT Asp Ser Arg Phe Trp Glu Leu Val Asp Lys Glu Arg Asn Leu His Leu 2920 2925 2930	8898
GAA GGA AAG TGT GAA ACA TGT GTG TAC AAC ATG ATG GGA AAA AGA GAG Glu Gly Lys Cys Glu Thr Cys Val Tyr Asn Met Met Gly Lys Arg Glu 2935 2940 2945 2950	8946
AAG AAG CTA GGG GAA TTC GGC AAG GCA AAA GGC AGC AGA GCC ATA TGG Lys Lys Leu Gly Glu Phe Gly Lys Ala Lys Gly Ser Arg Ala Ile Trp 2955 2960 2965	8994
TAC ATG TGG CTT GGA GCA CGC TTC TTA GAG TTT GAA GCC CTA GGA TTC Tyr Met Trp Leu Gly Ala Arg Phe Leu Glu Phe Glu Ala Leu Gly Phe 2970 2975 2980	9042
TTA AAT GAA GAT CAC TGG TTC TCC AGA GAG AAC TCC CTG AGT GGA GTG Leu Asn Glu Asp His Trp Phe Ser Arg Glu Asn Ser Leu Ser Gly Val 2985 2990 2995	9090

120

GAA GGA GAA GGG CTG CAC AAG CTA GGT TAC ATT CTA AGA GAC GTG AGC Glu Gly Glu Gly Leu His Lys Leu Gly Tyr Ile Leu Arg Asp Val Ser 3000 3005 3010	9138
AAG AAA GAG GGA GGA GCA ATG TAT GCC GAT GAC ACC GCA GGA TGG GAT Lys Lys Glu Gly Gly Ala Met Tyr Ala Asp Asp Thr Ala Gly Trp Asp 3015 3020 3025 3030	9186
ACA AGA ATC ACA CTA GAA GAC KKA AAA AAT GAA GAA ATG GTA ACA AAC Thr Arg Ile Thr Leu Glu Asp Xaa Lys Asn Glu Glu Met Val Thr Asn 3035 3040 3045	9234
CAC ATG GAA GGA GAA CAC AAG AAA CTA GCC GAG GCC ATT TTC AAA CTA His Met Glu Gly Glu His Lys Lys Leu Ala Glu Ala Ile Phe Lys Leu 3050 3055 3060	9282
ACG TAC CAA AAC AAG GTG GTG CGT GTG CAA AGA CCA ACA CCA AGA GGC Thr Tyr Gln Asn Lys Val Val CGT Val Val Gln Arg Pro Thr Pro Arg Gly 3065 3070 3075	9330
ACA GTA ATG GAC ATC ATA TCG AGA AGA GAC CAA AGA GGT AGT GGA CAA Thr Val Met Asp Ile Ile Ser Arg Arg Asp Gln Arg Gly Ser Gly Gln 3080 3085 3090	9378
GTT GGC ACC TAT GGA CTC AAT ACT TTC ACC AAT ATG GAA GCC CAA CTA Val Gly Thr Tyr Gly Leu Asn Thr Phe Thr Asn Met Glu Ala Gln Leu 3095 3100 3105 3110	9426
ATC AGA CAG ATG GAG GGA GAA GGA GTC TTT AAA AGC ATT CAG CAC CTA Ile Arg Gln Met Glu Gly Glu Gly Val Phe Lys Ser Ile Gln His Leu 3115 3120 3125	9474
ACA ATC ACA GAA GAA ATC GCT GTG CAA AAC TGG TTA GCA AGA GTG GGG Thr Ile Thr Glu Glu Ile Ala Val Gln Asn Trp Leu Ala Arg Val Gly 3130 3135 3140	9522
CGC GAA AGG TTA TCA AGA ATG GCC ATC AGT GGA GAT GAT TGT GTT GTG Arg Glu Arg Leu Ser Arg Met Ala Ile Ser Gly Asp Asp Cys Val Val 3145 3150 3155	9570
AAA CCT TTA GAT GAC AGG TTC GCA AGC GCT TTA ACA GCT CTA AAT GAC Lys Pro Leu Asp Asp Arg Phe Ala Ser Ala Leu Thr Ala Leu Asn Asp 3160 3165 3170	9618
ATG GGA AAG ATT AGG AAA GAC ATA CAA CAA TGG GAA CCT TCA AGA GGA Met Gly Lys Ile Arg Lys Asp Ile Gln Gln Trp Glu Pro Ser Arg Gly 3175 3180 3185 3190	9666
TGG AAT GAT TGG ACA CAA GTG CCC TTC TGT TCA CAC CAT TTC CAT GAG Trp Asn Asp Trp Thr Gln Val Pro Phe Cys Ser His His Phe His Glu 3195 3200 3205	9714
TTA ATC ATG AAA GAC GGT CGC GTA CTC GTT GTT CCA TGT AGA AAC CAA Leu Ile Met Lys Asp Gly Arg Val Leu Val Val Pro Cys Arg Asn Gln 3210 3215 3220	9762
GAT GAA CTG ATT GGC AGA GCC CGA ATC TCC CAA GGA GCA GGG TGG TCT Asp Glu Leu Ile Gly Arg Ala Arg Ile Ser Gln Gly Ala Gly Trp Ser 3225 3230 3235	9810

TTG CGG GAG ACG GCC TGT TTG GGG AAG TCT TAC GCC CAA ATG TGG AGC 9858
 Leu Arg Glu Thr Ala Cys Leu Gly Lys Ser Tyr Ala Gln Met Trp Ser
 3240 3245 3250

TTG ATG TAC TTC CAC AGA CGC GAC CTC AGG CTG GCG GCA AAT GCT ATT 9906
 Leu Met Tyr Phe His Arg Arg Asp Leu Arg Leu Ala Ala Asn Ala Ile
 3255 3260 3265 3270

TGC TCG GCA GTA CCA TCA CAT TGG GTT CCA ACA AGT CGA ACA ACC TGG 9954
 Cys Ser Ala Val Pro Ser His Trp Val Pro Thr Ser Arg Thr Thr Trp
 3275 3280 3285

TCC ATA CAT GCT AAA CAT GAA TGG ATG ACA ACG GAA GAC ATG CTG ACA 10002
 Ser Ile His Ala Lys His Glu Trp Met Thr Thr Glu Asp Met Leu Thr
 3290 3295 3300

GTC TGG AAC AGG GTG TGG ATT CAA GAA AAC CCA TGG ATG GAA GAC AAA 10050
 Val Trp Asn Arg Val Trp Ile Gln Glu Asn Pro Trp Met Glu Asp Lys
 3305 3310 3315

ACT CCA GTG GAA TCA TGG GAG GAA ATC CCA TAC TTG GGG AAA AGA GAA 10098
 Thr Pro Val Glu Ser Trp Glu Glu Ile Pro Tyr Leu Gly Lys Arg Glu
 3320 3325 3330

GAC CAA TGG TGC GGC TCA TTG ATT GGG TTA ACA AGC AGG GCC ACC TGG 10146
 Asp Gln Trp Cys Gly Ser Leu Ile Gly Leu Thr Ser Arg Ala Thr Trp
 3335 3340 3345 3350

GCA AAG AAC ATC CAA GCA GCA ATA AAT CAA GTT AGA TCC CTT ATA GGC 10194
 Ala Lys Asn Ile Gln Ala Ala Ile Asn Gln Val Arg Ser Leu Ile Gly
 3355 3360 3365

AAT GAA GAA TAC ACA GAT TAC ATG CCA TCC ATG AAA AGA TTC AGA AGA 10242
 Asn Glu Glu Tyr Thr Asp Tyr Met Pro Ser Met Lys Arg Phe Arg Arg
 3370 3375 3380

GAA GAG GAA GAA GCA GGA GTT CTG TGG TAGAAAGCAA AACTAACATG AAACAAGG 10297
 Glu Glu Glu Glu Ala Gly Val Leu Trp
 3385 3390

CTAGAAGTCA GGTCCGATTA AGCCATAGTA CGGAAAAAAC TATGCTACCT GTGAGCCCCG 10357
 TCCAAGGACG TTAAGAAG TCAGGCCATC ATAAATGCCA TAGCTTGAGT AAACATATGCA 10417
 GCCTGTAGCT CCACCTGAGA AGGTGTAAAA AATCCGGGAG GCCACAAACC ATGGAAGCTG 10477
 TACGCATGGC GTAGTGGACT AGCGGTTAGA GAGGACCCCT CCCTTACAAA TCGCAGCAAC 10537
 AATGGGGGCC CAAGGCGAGA TGAAGCTGTA GTCTCGCTGG AAGGACTAGA GGTTAGAGGA 10597
 GACCCCCCG AAACAAAAA CAGCATATTG ACGCTGGGAA AGACCAGAGA TCCTGCTGTC 10657
 TCCTCAGCAT CATTCCAGGC ACAGAACGCC AGAAAATGGA ATGGTGCTGT TGAATCAACA 10717
 GGTCT 10723

(2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 10723 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

(ix) FEATURE:

- (A) NAME/KEY: Coding Sequence
- (B) LOCATION: 97...10269
- (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

AGTTGTTAGT	CTACGTGGAC	CGACAAAGAC	AGATTCTTTG	AGGGAGCTAA	GCTCAATGTA	60
GTTCTAACAG	TTTTTTAATT	AGAGAGCAGA	TCTCTG	ATG AAT AAC CAA CGG AAA		114
				Met Asn Asn Gln Arg Lys		
				1	5	
AAG GCG AAA AAC ACG CCT TTC AAT ATG CTG AAA CGC GAG AGA AAC CGC						162
Lys Ala Lys Asn Thr Pro Phe Asn Met Leu Lys Arg Glu Arg Asn Arg						
	10		15		20	
GTG TCG ACT GTG CAA CAG CTG ACA AAG AGA TTC TCA CTT GGA ATG CTG						210
Val Ser Thr Val Gln Gln Leu Thr Lys Arg Phe Ser Leu Gly Met Leu						
	25		30		35	
CAG GGA CGA GGA CCA TTA AAA CTG TTC ATG GCC CTG GTG GCG TTC CTT						258
Gln Gly Arg Gly Pro Leu Lys Leu Phe Met Ala Leu Val Ala Phe Leu						
	40		45		50	
CGT TTC CTA ACA ATC CCA CCA ACA GCA GGG ATA TTG AAG AGA TGG GGA						306
Arg Phe Leu Thr Ile Pro Pro Thr Ala Gly Ile Leu Lys Arg Trp Gly						
	55		60		65	70
ACA ATT AAA AAA TCA AAA GCT ATT AAT GTT TTG AGA GGG TTC AGG AAA						354
Thr Ile Lys Lys Ser Lys Ala Ile Asn Val Leu Arg Gly Phe Arg Lys						
	75		80		85	
GAG ATT GGA AGG ATG CTG AAC ATC TTG AAT AGG AGA CGC AGA TCT GCA						402
Glu Ile Gly Arg Met Leu Asn Ile Leu Asn Arg Arg Arg Arg Ser Ala						
	90		95		100	
GGC ATG ATC ATT ATG CTG ATT CCA ACA GTG ATG GCG TTC CAT TTA ACC						450
Gly Met Ile Ile Met Leu Ile Pro Thr Val Met Ala Phe His Leu Thr						
	105		110		115	
ACA CGT AAC GGA GAA CCA CAC ATG ATC GTC AGC AGA CAA GAG AAA GGG						498
Thr Arg Asn Gly Glu Pro His Met Ile Val Ser Arg Gln Glu Lys Gly						
	120		125		130	
AAA AGT CTT CTG TTT AAA ACA GAG GTT GGC GTG AAC ATG TGT ACC CTC						546
Lys Ser Leu Leu Phe Lys Thr Glu Val Gly Val Asn Met Cys Thr Leu						
	135		140		145	150
ATG GCC ATG GAC CTT GGT GAA TTG TGT GAA GAC ACA ATC ACG TAC AAG						594
Met Ala Met Asp Leu Gly Glu Leu Cys Glu Asp Thr Ile Thr Tyr Lys						
	155		160		165	
TGT CCC CTT CTC AGG CAG AAT GAG CCA GAA GAC ATA GAC TGT TGG TGC						642
Cys Pro Leu Leu Arg Gln Asn Glu Pro Glu Asp Ile Asp Cys Trp Cys						
	170		175		180	
NAC TCT ACG TCC ACG TGG GTA ACT TAT GGG ACG TGT ACC ACC ATG GGA						690
Xaa Ser Thr Ser Thr Trp Val Thr Tyr Gly Thr Cys Thr Thr Met Gly						
	185		190		195	

GAA Glu	CAT His	AGA Arg	AGA Arg	GAA Glu	AAA Lys	AGA Arg	TCA Ser	GTG Val	GCA Ala	CTC Leu	GTT Val	CCA Pro	CAT His	GTG Val	GGA Gly	738
200 205 210																
ATG Met	GGA Gly	CTG Leu	GAG Glu	ACA Thr	CGA Arg	ACT Thr	GAA Glu	ACA Thr	TGG Trp	ATG Met	TCA Ser	TCA Ser	GAA Glu	GGG Gly	GCC Ala	786
215 220 225 230																
TGG Trp	AAA Lys	CAT His	GTC Val	CAG Gln	AGA Arg	ATT Ile	GAA Glu	ACT Thr	TGG Trp	ATC Ile	TTG Leu	AGA Arg	CAT His	CCA Pro	GGC Gly	834
235 240 245																
TTC Phe	ACC Thr	ATG Met	ATG Met	GCA Ala	GCA Ala	ATC Ile	CTG Leu	GCA Ala	TAC Tyr	ACC Thr	ATA Ile	GGA Gly	ACG Thr	ACA Thr	CAT His	882
250 255 260																
TTC Phe	CAA Gln	AGA Arg	GCC Ala	CTG Leu	ATT Ile	TTC Phe	ATC Ile	TTA Leu	CTG Leu	ACA Thr	GCT Ala	GTC Val	ACT Thr	CCT Pro	TCA Ser	930
265 270 275																
ATG Met	ACA Thr	ATG Met	CGT Arg	TGC Cys	ATA Ile	GGA Gly	ATG Met	TCA Ser	AAT Asn	AGA Arg	GAC Asp	TTT Phe	GTG Val	GAA Glu	GGG Gly	978
280 285 290																
GTT Val	TCA Ser	GGA Gly	GGA Gly	AGC Ser	TGG Trp	GTT Val	GAC Asp	ATA Ile	GTC Val	TTA Leu	GAA Glu	CAT His	GGA Gly	AGC Ser	TGT Cys	1026
295 300 305 310																
GTG Val	ACG Thr	ACG Thr	ATG Met	GCA Ala	AAA Lys	AAC Asn	AAA Lys	CCA Pro	ACA Thr	TTG Leu	GAT Asp	TTT Phe	GAA Glu	CTG Leu	ATA Ile	1074
315 320 325																
AAA Lys	ACA Thr	GAA Glu	GCC Ala	AAA Lys	CAG Gln	CCT Pro	GCC Ala	ACC Thr	CTA Leu	AGG Arg	AAG Lys	TAC Tyr	TGT Cys	ATA Ile	GAG Glu	1122
330 335 340																
GCA Ala	AAG Lys	CTA Leu	ACC Thr	NAC Xaa	ACA Thr	ACA Thr	ACA Thr	GAA Glu	TCT Ser	CGC Arg	TGC Cys	CCA Pro	ACA Thr	CAA Gln	GGG Gly	1170
345 350 355																
GAA Glu	CCC Pro	AGC Ser	CTA Leu	AAT Asn	GAA Glu	GAG Glu	CAG Gln	GAC Asp	AAA Lys	AGG Arg	TTC Phe	GTC Val	TGC Cys	AAA Lys	CAC His	1218
360 365 370																
TCC Ser	ATG Met	GTA Val	GAC Asp	AGA Arg	GGA Gly	TGG Trp	GGA Gly	AAT Asn	GGA Gly	TGT Cys	GGA Gly	CTA Leu	TTT Phe	GGA Gly	AAG Lys	1266
375 380 385 390																
GGA Gly	GGC Gly	ATT Ile	GTG Val	ACC Thr	TGT Cys	GCT Ala	ATG Met	TTC Phe	AGA Arg	TGC Cys	AAA Lys	AAG Lys	AAC Asn	ATG Met	GAA Glu	1314
395 400 405																
GGA Gly	AAA Lys	GTT Val	GTG Val	CAA Gln	CCA Pro	GAA Glu	AAC Asn	TTG Leu	GAA Glu	TAC Tyr	ACC Thr	ATT Ile	GTG Val	ATA Ile	ACA Thr	1362
410 415 420																
CCT Pro	CAC His	TCA Ser	GGG Gly	GAA Glu	GAG Glu	CAT His	GCA Ala	GTC Val	GGA Gly	NAT Xaa	GAC Asp	ACA Thr	GGA Gly	AAA Lys	CAT His	1410
425 430 435																

124

GGC Gly	AAG Lys	GAA Glu	ATC Ile	AAA Lys	ATA Ile	ACA Thr	CCA Pro	CAG Gln	AGT Ser	TCC Ser	ATC Ile	ACA Thr	GAA Glu	GCA Ala	GAA Glu	1458
	440					445					450					
TTG Leu	ACA Thr	GGT Gly	TAT Tyr	GGC Gly	ACT Thr	GTC Val	ACA Thr	ATG Met	GAG Glu	TGC Cys	TCT Ser	CCA Pro	AGA Arg	ACG Thr	GGC Gly	1506
	455				460					465					470	
CTC Leu	GAC Asp	TTC Phe	AAT Asn	GAG Glu	ATG Met	GTG Val	TTG Leu	CTG Leu	CAG Gln	ATG Met	GAA Glu	AAT Asn	AAA Lys	GCT Ala	TGG Trp	1554
				475					480					485		
CTG Leu	GTG Val	CAC His	AGG Arg	CAA Gln	TGG Trp	TTC Phe	CTA Leu	GAC Asp	CTG Leu	CCG Pro	TTA Leu	CCA Pro	TGG Trp	TTG Leu	CCC Pro	1602
			490					495					500			
GGA Gly	GCG Ala	GAC Asp	ACA Thr	CAA Gln	GGG Gly	TCA Ser	AAT Asn	TGG Trp	ATA Ile	CAG Gln	AAA Lys	GAG Glu	ACA Thr	TTG Leu	GTC Val	1650
		505					510					515				
ACT Thr	TTC Phe	AAA Lys	AAT Asn	CCC Pro	CAT His	GCG Ala	AAG Lys	AAA Lys	CAG Gln	GAT Asp	GTT Val	GTT Val	GTT Val	TTA Leu	GGA Gly	1698
	520					525					530					
TCC Ser	CAA Gln	GAA Glu	GGG Gly	GCC Ala	ATG Met	CAC His	ACA Thr	GCA Ala	CTT Leu	ACA Thr	GGG Gly	GCC Ala	ACA Thr	GAA Glu	ATC Ile	1746
					540					545					550	
CAA Gln	ATG Met	TCA Ser	TCA Ser	GGA Gly	AAC Asn	TTA Leu	CTC Leu	TTC Phe	ACA Thr	GGA Gly	CAT His	CTC Leu	AAG Lys	TGC Cys	AGG Arg	1794
				555					560					565		
CTG Leu	AGA Arg	ATG Met	GAC Asp	AAG Lys	CTA Leu	CAG Gln	CTC Leu	AAA Lys	GGA Gly	ATG Met	TCA Ser	TAC Tyr	TCT Ser	ATG Met	TGC Cys	1842
			570					575					580			
ACA Thr	GGA Gly	AAG Lys	TTT Phe	AAA Lys	GTT Val	GTG Val	AAG Lys	GAA Glu	ATA Ile	GCA Ala	GAA Glu	ACA Thr	CAA Gln	CAT His	GGA Gly	1890
		585					590					595				
ACA Thr	ATA Ile	GTT Val	ATC Ile	AGA Arg	GTG Val	CAA Gln	TAT Tyr	GAA Glu	GGG Gly	GAC Asp	GGC Gly	TCT Ser	CCA Pro	TGC Cys	AAG Lys	1938
	600					605					610					
ATC Ile	CCT Pro	TTT Phe	GAG Glu	ATA Ile	ATG Met	GAT Asp	TTG Leu	GAA Glu	AAA Lys	AGA Arg	CAT His	GTC Val	TTA Leu	GGT Gly	CGC Arg	1986
	615				620					625					630	
CTG Leu	ATT Ile	ACA Thr	GTC Val	AAC Asn	CCA Pro	ATT Ile	GTG Val	ACA Thr	GAA Glu	AAA Lys	GAT Asp	AGC Ser	CCA Pro	GTC Val	AAC Asn	2034
				635					640					645		
ATA Ile	GAA Glu	GCA Ala	GAA Glu	CCT Pro	CCA Pro	TTT Phe	GGA Gly	GAC Asp	AGC Ser	TAC Tyr	ATC Ile	ATC Ile	ATA Ile	GGA Gly	GTA Val	2082
			650					655					660			
GAG Glu	CCG Pro	GGA Gly	CAA Gln	CTG Leu	AAG Lys	CTC Leu	AAC Asn	TGG Trp	TTT Phe	AAG Lys	AAA Lys	GGA Gly	AGT Ser	TCT Ser	ATC Ile	2130
		665					670					675				

125

GGC Gly	CAA Gln	ATG Met	TTT Phe	GAG Glu	ACA Thr	ACA Thr	ATG Met	AGG Arg	GGG Gly	GCG Ala	AAG Lys	AGA Arg	ATG Met	GCC Ala	ATT Ile	2178
	680					685					690					
TTA Leu	GGT Gly	GAC Asp	ACA Thr	GCC Ala	TGG Trp	GAT Asp	TTT Phe	GGA Gly	TCC Ser	TTG Leu	GGA Gly	GGA Gly	GTG Val	TTT Phe	ACA Thr	2226
695					700					705				710		
TCT Ser	ATA Ile	GGA Gly	AAG Lys	GCT Ala	CTC Leu	CAC His	CAA Gln	GTC Val	TTT Phe	GGA Gly	GCA Ala	ATC Ile	TAT Tyr	GGA Gly	GCT Ala	2274
				715					720					725		
GCC Ala	TTC Phe	AGT Ser	GGG Gly	GTT Val	TCA Ser	TGG Trp	ACT Thr	ATG Met	AAA Lys	ATC Ile	CTC Leu	ATA Ile	GGA Gly	GTC Val	ATT Ile	2322
			730					735					740			
ATC Ile	ACA Thr	TGG Trp	ATA Ile	GGA Gly	ATG Met	AAT Asn	TCA Ser	CGC Arg	AGC Ser	ACC Thr	TCA Ser	CTG Leu	TCT Ser	GTG Val	ACA Thr	2370
		745					750						755			
CTA Leu	GTA Val	TTG Leu	GTG Val	GGA Gly	ATT Ile	GTG Val	ACA Thr	CTG Leu	TAT Tyr	TTG Leu	GGA Gly	GTC Val	ATG Met	GTG Val	CAG Gln	2418
	760					765					770					
GCC Ala	GAT Asp	AGT Ser	GGT Gly	TGC Cys	GTT Val	GTG Val	AGC Ser	TGG Trp	AAA Lys	AAC Asn	AAA Lys	GAA Glu	CTG Leu	AAA Lys	TGT Cys	2466
775				780						785					790	
GGC Gly	AGT Ser	GGG Gly	ATT Ile	TTC Phe	ATC Ile	ACA Thr	GAC Asp	AAC Asn	GTG Val	CAC His	ACA Thr	TGG Trp	ACA Thr	GAA Glu	CAA Gln	2514
				795					800					805		
TAC Tyr	AAG Lys	TTC Phe	CAA Gln	CCA Pro	GAA Glu	TCC Ser	CCT Pro	TCA Ser	AAA Lys	CTA Leu	GCT Ala	TCA Ser	GCT Ala	ATC Ile	CAG Gln	2562
			810				815						820			
AAA Lys	GCC Ala	CAT His	GAA Glu	GAG Glu	GAC Asp	ATT Ile	TGT Cys	GGA Gly	ATC Ile	CGC Arg	TCA Ser	GTA Val	ACA Thr	AGA Arg	CTG Leu	2610
		825					830					835				
GAG Glu	AAT Asn	CTG Leu	ATG Met	TGG Trp	AAA Lys	CAA Gln	ATA Ile	ACA Thr	CCA Pro	GAA Glu	TTG Leu	AAT Asn	CAC His	ATT Ile	CTA Leu	2658
	840					845					850					
TCA Ser	GAA Glu	AAT Asn	GAG Glu	GTG Val	AAG Lys	TTA Leu	ACT Thr	ATT Ile	ATG Met	ACA Thr	GGA Gly	GAC Asp	ATC Ile	AAA Lys	GGA Gly	2706
855				860						865				870		
ATC Ile	ATG Met	CAG Gln	GCA Ala	GGA Gly	AAA Lys	CGA Arg	TCT Ser	CTG Leu	CGG Arg	CCT Pro	CAG Gln	CCC Pro	ACT Thr	GAG Glu	CTG Leu	2754
				875					880					885		
AAG Lys	TAT Tyr	TCA Ser	TGG Trp	AAA Lys	ACA Thr	TGG Trp	GGC Gly	AAA Lys	GCA Ala	AAA Lys	ATG Met	CTC Leu	TCT Ser	ACA Thr	GAG Glu	2802
			890					895					900			
TCT Ser	CAT His	NAC Xaa	CAG Gln	ACC Thr	TTT Phe	CTC Leu	ATT Ile	GAT Asp	GGC Gly	CCC Pro	GAA Glu	ACA Thr	GCA Ala	GAA Glu	TGC Cys	2850
			905					910						915		

CCC Pro	AAC Asn	ACA Thr	AAT Asn	AGA Arg	GCT Ala	TGG Trp	AAT Asn	TCG Ser	TTG Leu	GAA Glu	GTT Val	GAA Glu	GAC Asp	TAT Tyr	GGC Gly	2898
	920					925					930					
TTT Phe	GGA Gly	GTA Val	TTC Phe	ACC Thr	ACC Thr	AAT Asn	ATA Ile	TGG Trp	CTA Leu	AAA Lys	TTG Leu	AAA Lys	GAA Glu	AAA Lys	CAG Gln	2946
	935				940					945					950	
GAT Asp	GTA Val	TTC Phe	TGC Cys	GAC Asp	TCA Ser	AAA Lys	CTC Leu	ATG Met	TCA Ser	GCG Ala	GCC Ala	ATA Ile	AAA Lys	GAC Asp	AAC Asn	2994
				955					960					965		
AGA Arg	GCC Ala	GTC Val	CAT His	GCC Ala	GAT Asp	ATG Met	GGT Gly	TAT Tyr	TGG Trp	ATA Ile	GAA Glu	AGT Ser	GCA Ala	CTC Leu	NAT Xaa	3042
			970					975					980			
GAC Asp	ACA Thr	TGG Trp	AAG Lys	ATA Ile	GAG Glu	AAA Lys	GCC Ala	TCT Ser	TTC Phe	ATT Ile	GAA Glu	GTT Val	AAA Lys	AAC Asn	TGC Cys	3090
		985					990					995				
CAC His	TGG Trp	CCA Pro	AAA Lys	TCA Ser	CAC His	ACC Thr	CTC Leu	TGG Trp	AGC Ser	AAT Asn	GGA Gly	GTG Val	CTA Leu	GAA Glu	AGT Ser	3138
	1000					1005					1010					
GAG Glu	ATG Met	ATA Ile	ATT Ile	CCA Pro	AAG Lys	AAT Asn	CTC Leu	GCT Ala	GGA Gly	CCA Pro	GTG Val	TCT Ser	CAA Gln	CAC His	AAC Asn	3186
1015					1020					1025					1030	
TAT Tyr	AGA Arg	CCA Pro	GGC Gly	TAC Tyr	CAT His	ACA Thr	CAA Gln	ATA Ile	ACA Thr	GGA Gly	CCA Pro	TGG Trp	CAT His	CTA Leu	GGT Gly	3234
				1035					1040					1045		
AAG Lys	CTT Leu	GAG Glu	ATG Met	GAC Asp	TTT Phe	GAT Asp	TTC Phe	TGT Cys	GAT Asp	GGA Gly	ACA Thr	ACA Thr	GTG Val	GTA Val	GTG Val	3282
			1050					1055					1060			
ACT Thr	GAG Glu	GAC Asp	TGC Cys	GGA Gly	AAT Asn	AGA Arg	GGA Gly	CCC Pro	TCT Ser	TTG Leu	AGA Arg	ACA Thr	ACC Thr	ACT Thr	GCC Ala	3330
	1065						1070					1075				
TCT Ser	GGA Gly	AAA Lys	CTC Leu	ATA Ile	ACA Thr	GAA Glu	TGG Trp	TGC Cys	TGC Cys	CGA Arg	TCT Ser	TGC Cys	ACA Thr	TTA Leu	CCA Pro	3378
	1080					1085					1090					
CCG Pro	CTA Leu	AGA Arg	TAC Tyr	AGA Arg	GGT Gly	GAG Glu	GAT Asp	GGG Gly	TGC Cys	TGG Trp	TAC Tyr	GGG Gly	ATG Met	GAA Glu	ATC Ile	3426
1095					1100				1105						1110	
AGA Arg	CCA Pro	TTG Leu	AAG Lys	GAG Glu	AAA Lys	GAA Glu	GAG Glu	AAT Asn	TTG Leu	GTC Val	AAC Asn	TCC Ser	TTG Leu	GTC Val	ACA Thr	3474
				1115					1120					1125		
GCT Ala	GGA Gly	CAT His	GGG Gly	CAG Gln	GTC Val	GAC Asp	AAC Asn	TTT Phe	TCA Ser	CTA Leu	GGA Gly	GTC Val	TTG Leu	GGA Gly	ATG Met	3522
			1130					1135					1140			
GCA Ala	TTG Leu	TTC Phe	CTG Leu	GAG Glu	GAA Glu	ATG Met	CTT Leu	AGG Arg	ACC Thr	CGA Arg	GTA Val	GGA Gly	ACG Thr	AAA Lys	CAT His	3570
	1145						1150					1155				

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GCA ATA CTA CTA GTT GCA GTT TCT TTT GTG ACA TTG ATC ACA GGG AAC	3618
Ala Ile Leu Leu Val Ala Val Ser Phe Val Thr Leu Ile Thr Gly Asn	
1160 1165 1170	
ATG TCC TTT AGA GAC CTG GGA AGA GTG ATG GTT ATG GTA GGC GCC ACT	3666
Met Ser Phe Arg Asp Leu Gly Arg Val Met Val Met Val Gly Ala Thr	
1175 1180 1185 1190	
ATG ACG GAT GAC ATA GGT ATG GGC GTG ACT TAT CTT GCC CTA CTA GCA	3714
Met Thr Asp Asp Ile Gly Met Gly Val Thr Tyr Leu Ala Leu Leu Ala	
1195 1200 1205	
GCC TTC AAA GTC AGA CCA ACT TTT GCA GCT GGA CTA CTC TTG AGA AAG	3762
Ala Phe Lys Val Arg Pro Thr Phe Ala Ala Gly Leu Leu Leu Arg Lys	
1210 1215 1220	
CTG ACC TCC AAG GAA TTG ATG ATG ACT ACT ATA GGA ATT GTA CTC CTC	3810
Leu Thr Ser Lys Glu Leu Met Met Thr Thr Ile Gly Ile Val Leu Leu	
1225 1230 1235	
TCC CAG AGC ACC ATA CCA GAG ACC ATT CTT GAG TTG ACT GAT GCG TTA	3858
Ser Gln Ser Thr Ile Pro Glu Thr Ile Leu Glu Leu Thr Asp Ala Leu	
1240 1245 1250	
GCC TTA GGC ATG ATG GTC CTC AAA ATG GTG AGA AAT ATG GAA AAG TAT	3906
Ala Leu Gly Met Met Val Leu Lys Met Val Arg Asn Met Glu Lys Tyr	
1255 1260 1265 1270	
CAA TTG GCA GTG ACT ATC ATG GCT ATC TTG TGC GTC CCA AAC GCA GTG	3954
Gln Leu Ala Val Thr Ile Met Ala Ile Leu Cys Val Pro Asn Ala Val	
1275 1280 1285	
ATA TTA CAA AAC GCA TGG AAA GTG AGT TGC ACA ATA TTG GCA GTG GTG	4002
Ile Leu Gln Asn Ala Trp Lys Val Ser Cys Thr Ile Leu Ala Val Val	
1290 1295 1300	
TCC GTT TCC CCA CTG TTC TTA ACA TCC TCA CAG CAA AAA ACA GAT TGG	4050
Ser Val Ser Pro Leu Phe Leu Thr Ser Ser Gln Gln Lys Thr Asp Trp	
1305 1310 1315	
ATA CCA TTA GCA TTG ACG ATC AAA GGT CTC AAT CCA ACA GCT ATT TTT	4098
Ile Pro Leu Ala Leu Thr Ile Lys Gly Leu Asn Pro Thr Ala Ile Phe	
1320 1325 1330	
CTA ACA ACC CTC TCA AGA ACC AGC AAG AAA AGG AGC TGG CCA TTA AAT	4146
Leu Thr Thr Leu Ser Arg Thr Ser Lys Lys Arg Ser Trp Pro Leu Asn	
1335 1340 1345 1350	
GAG GCT ATC ATG GCA GTC GGG ATG GTG AGC ATT TTA GCC AGT TCT CTC	4194
Glu Ala Ile Met Ala Val Gly Met Val Ser Ile Leu Ala Ser Ser Leu	
1355 1360 1365	
CTA AAA AAT GAT ATT CCC ATG ACA GGA CCA TTA GTG GCT GGA GGG CTC	4242
Leu Lys Asn Asp Ile Pro Met Thr Gly Pro Leu Val Ala Gly Gly Leu	
1370 1375 1380	
CTC ACT GTG TGC TAC GTG CTC ACT GGA CGA TCG GCC GAT TTG GAA CTG	4290
Leu Thr Val Cys Tyr Val Leu Thr Gly Arg Ser Ala Asp Leu Glu Leu	
1385 1390 1395	

GAG AGA GCA GCC GAT GTC AAA TGG GAA GAC CAG GCA GAG ATA TCA GGA Glu Arg Ala Ala Asp Val Lys Trp Glu Asp Gln Ala Glu Ile Ser Gly 1400 1405 1410	4338
AGC AGT CCA ATC CTG TCA ATA ACA ATA TCA GAA GAT GGT AGC ATG TCG Ser Ser Pro Ile Leu Ser Ile Thr Ile Ser Glu Asp Gly Ser Met Ser 1415 1420 1425 1430	4386
ATA AAA AAT GAA GAG GAA GAA CAA ACA CTG ACC ATA CTC ATT AGA ACA Ile Lys Asn Glu Glu Glu Glu Gln Thr Leu Thr Ile Leu Ile Arg Thr 1435 1440 1445	4434
GGA TTG CTG GTG ATC TCA GGA CTT TTT CCT GTA TCA ATA CCA ATC ACG Gly Leu Leu Val Ile Ser Gly Leu Phe Pro Val Ser Ile Pro Ile Thr 1450 1455 1460	4482
GCA GCA GCA TGG TAC CTG TGG GAA GTG AAG AAA CAA CGG GCC GGA GTA Ala Ala Ala Trp Tyr Leu Trp Glu Val Lys Lys Gln Arg Ala Gly Val 1465 1470 1475	4530
TTG TGG GAT GTT CCT TCA CCC CCA CCC ATG GGA AAG GCT GAA CTG GAA Leu Trp Asp Val Pro Ser Pro Pro Pro Met Gly Lys Ala Glu Leu Glu 1480 1485 1490	4578
GAT GGA GCC TAT AGA ATT AAG CAA AAA GGG ATT CTT GGA TAT TCC CAG Asp Gly Ala Tyr Arg Ile Lys Gln Lys Gly Ile Leu Gly Tyr Ser Gln 1495 1500 1505 1510	4626
ATC GGA GCC GGA GTT TAC AAA GAA GGA ACA TTC CAT ACA ATG TGG CAT Ile Gly Ala Gly Val Tyr Lys Glu Gly Thr Phe His Thr Met Trp His 1515 1520 1525	4674
GTC ACA CGT GGC GCT GTT CTA ATG CAT AAA GGA AAG AGG ATT GAA CCA Val Thr Arg Gly Ala Val Leu Met His Lys Gly Lys Arg Ile Glu Pro 1530 1535 1540	4722
TCA TGG GCG GAC GTC AAG AAA GAC CTA ATA TCA TAT GGA GGA GGC TGG Ser Trp Ala Asp Val Lys Lys Asp Leu Ile Ser Tyr Gly Gly Gly Trp 1545 1550 1555	4770
AAG TTA GAA GGA GAA TGG AAG GAA GGA GAA GAA GTC CAG GTA TTG GCA Lys Leu Glu Gly Glu Trp Lys Glu Gly Glu Glu Val Gln Val Leu Ala 1560 1565 1570	4818
CTG GAG CCT GGA AAA AAT CCA AGA GCC GTC CAA ACG AAA CCT GGT CTT Leu Glu Pro Gly Lys Asn Pro Arg Ala Val Gln Thr Lys Pro Gly Leu 1575 1580 1585 1590	4866
TTC AAA ACC AAC GCC GGA ACA ATA GGT GCT GTA TCT CTG GAC TTT TCT Phe Lys Thr Asn Ala Gly Thr Ile Gly Ala Val Ser Leu Asp Phe Ser 1595 1600 1605	4914
CCT GGA ACG TCA GGA TCT CCA ATT ATC GAC AAA AAA GGA AAA GTT GTG Pro Gly Thr Ser Gly Ser Pro Ile Ile Asp Lys Lys Gly Lys Val Val 1610 1615 1620	4962
GGT CTT TAT GGT AAT GGT GTT GTT ACA AGG AGT GGA GCA TAT GTG AGT Gly Leu Tyr Gly Asn Gly Val Val Thr Arg Ser Gly Ala Tyr Val Ser 1625 1630 1635	5010

GCT ATA GCC CAG ACT GAA AAA AGC ATT GAA GAC AAC CCA GAG ATC GAA	5058
Ala Ile Ala Gln Thr Glu Lys Ser Ile Glu Asp Asn Pro Glu Ile Glu	
1640 1645 1650	
GAT GAC ATT TTC CGA AAG AGA AGA CTG ACC ATC ATG GAC CTC CAC CCA	5106
Asp Asp Ile Phe Arg Lys Arg Arg Leu Thr Ile Met Asp Leu His Pro	
1655 1660 1665 1670	
GGA GCG GGA AAG ACG AAG AGA TAC CTT CCG GCC ATA GTC AGA GAA GCT	5154
Gly Ala Gly Lys Thr Lys Arg Tyr Leu Pro Ala Ile Val Arg Glu Ala	
1675 1680 1685	
ATA AAA CGG GGT TTG AGA ACA TTA ATC TTG GCC CCC ACT AGA GTT GTG	5202
Ile Lys Arg Gly Leu Arg Thr Leu Ile Leu Ala Pro Thr Arg Val Val	
1690 1695 1700	
GCA GCT GAA ATG GAG GAA GCC CTT AGA GGA CTT CCA ATA AGA TAC CAG	5250
Ala Ala Glu Met Glu Glu Ala Leu Arg Gly Leu Pro Ile Arg Tyr Gln	
1705 1710 1715	
ACC CCA GCC ATC AGA GCT GAG CAC ACC GGG CGG GAG ATT GTG GAC CTA	5298
Thr Pro Ala Ile Arg Ala Glu His Thr Gly Arg Glu Ile Val Asp Leu	
1720 1725 1730	
ATG TGT CAT GCC ACA TTT ACC ATG AGG CTG CTA TCA CCA GTT AGA GTG	5346
Met Cys His Ala Thr Phe Thr Met Arg Leu Leu Ser Pro Val Arg Val	
1735 1740 1745 1750	
CCA AAC TAC AAC CTG ATT ATC ATG GAC GAA GCC CAT TTC ACA GAC CCA	5394
Pro Asn Tyr Asn Leu Ile Ile Met Asp Glu Ala His Phe Thr Asp Pro	
1755 1760 1765	
GCA AGT ATA GCA GCT AGA GGA TAC ATC TCA ACT CGA GTG GAG ATG GGT	5442
Ala Ser Ile Ala Ala Arg Gly Tyr Ile Ser Thr Arg Val Glu Met Gly	
1770 1775 1780	
GAG GCA GCT GGG ATT TTT ATG ACA GCC ACT CCC CCG GGA AGC AGA GAC	5490
Glu Ala Ala Gly Ile Phe Met Thr Ala Thr Pro Pro Gly Ser Arg Asp	
1785 1790 1795	
CCA TTT CCT CAG AGC AAT GCA CCA ATC ATA GAT GAA GAA AGA GAA ATC	5538
Pro Phe Pro Gln Ser Asn Ala Pro Ile Ile Asp Glu Glu Arg Glu Ile	
1800 1805 1810	
CCT GAA CGT TCG TGG AAT TCC GGA CAT GAA TGG GTC ACG GAT TTT AAA	5586
Pro Glu Arg Ser Trp Asn Ser Gly His Glu Trp Val Thr Asp Phe Lys	
1815 1820 1825 1830	
GGG AAG ACT GTT TGG TTC GTT CCA AGT ATA AAA GCA GGA AAT GAT ATA	5634
Gly Lys Thr Val Trp Phe Val Pro Ser Ile Lys Ala Gly Asn Asp Ile	
1835 1840 1845	
GCA GCT TGC CTG AGG AAA AAT GGA AAG AAA GTG ATA CAA CTC AGT AGG	5682
Ala Ala Cys Leu Arg Lys Asn Gly Lys Lys Val Ile Gln Leu Ser Arg	
1850 1855 1860	
AAG ACC TTT GAT TCT GAG TAT GTC AAG ACT AGA ACC AAT GAT TGG GAC	5730
Lys Thr Phe Asp Ser Glu Tyr Val Lys Thr Arg Thr Asn Asp Trp Asp	
1865 1870 1875	

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TTC GTG GTT ACA ACT GAC ATT TCA GAA ATG GGT GCC AAT TTC AAG GCT Phe Val Val Thr Thr Asp Ile Ser Glu Met Gly Ala Asn Phe Lys Ala 1880 1885 1890	5778
GAG AGG GTT ATA GAC CCC AGA CGC TGC ATG AAA CCA GTC ATA CTA ACA Glu Arg Val Ile Asp Pro Arg Arg Cys Met Lys Pro Val Ile Leu Thr 1895 1900 1905 1910	5826
GAT GGT GAA GAG CGG GTG ATT CTG GCA GGA CCT ATG CCA GTG ACC CAC Asp Gly Glu Glu Arg Val Ile Leu Ala Gly Pro Met Pro Val Thr His 1915 1920 1925	5874
TCT AGT GCA GCA CAA AGA AGA GGG AGA ATA GGA AGA AAT CCA AAA AAT Ser Ser Ala Ala Gln Arg Arg Gly Arg Ile Gly Arg Asn Pro Lys Asn 1930 1935 1940	5922
GAG AAT GAC CAG TAC ATA TAC ATG GGG GAA CCT CTG GAA AAT GAT GAA Glu Asn Asp Gln Tyr Ile Tyr Met Gly Glu Pro Leu Glu Asn Asp Glu 1945 1950 1955	5970
GAC TGT GCA CAC TGG AAA GAA GCT AAA ATG CTC CTA GAT AAC ATC AAC Asp Cys Ala His Trp Lys Glu Ala Lys Met Leu Leu Asp Asn Ile Asn 1960 1965 1970	6018
ACG CCA GAA GGA ATC ATT CCT AGC ATG TTC GAA CCA GAG CGT GAA AAG Thr Pro Glu Gly Ile Ile Pro Ser Met Phe Glu Pro Glu Arg Glu Lys 1975 1980 1985 1990	6066
GTG GAT GCC ATT GAT GGC GAA TAC CGC TTG AGA GGA GAA GCA AGG AAA Val Asp Ala Ile Asp Gly Glu Tyr Arg Leu Arg Gly Glu Ala Arg Lys 1995 2000 2005	6114
ACC TTT GTA GAC TTA ATG AGA AGA GGA GAC CTA CCA GTC TGG TTG GCC Thr Phe Val Asp Leu Met Arg Arg Gly Asp Leu Pro Val Trp Leu Ala 2010 2015 2020	6162
TAC AGA GTG GCA GCT GAA GGC ATC AAC TAC GCA GAC AGA AGG TGG TGT Tyr Arg Val Ala Ala Glu Gly Ile Asn Tyr Ala Asp Arg Arg Trp Cys 2025 2030 2035	6210
TTT GAT GGA GTC AAG AAC AAC CAA ATC CTA GAA GAA AAC GTG GAA GTT Phe Asp Gly Val Lys Asn Asn Gln Ile Leu Glu Glu Asn Val Glu Val 2040 2045 2050	6258
GAA ATC TGG ACA AAA GAA GGG GAA AGG AAG AAA TTG AAA CCC AGA TGG Glu Ile Trp Thr Lys Glu Gly Glu Arg Lys Lys Leu Lys Pro Arg Trp 2055 2060 2065 2070	6306
TTG GAT GCT AGG ATC TAT TCT GAC CCA CTG GCG CTA AAA GAA TTT AAG Leu Asp Ala Arg Ile Tyr Ser Asp Pro Leu Ala Leu Lys Glu Phe Lys 2075 2080 2085	6354
GAA TTT GCA GCC GGA AGA AAG TCT CTG ACC CTG AAC CTA ATC ACA GAA Glu Phe Ala Ala Gly Arg Lys Ser Leu Thr Leu Asn Leu Ile Thr Glu 2090 2095 2100	6402
ATG GGT AGG CTC CCA ACC TTC ATG ACT CAG AAG GCA AGA GAC GCA CTG Met Gly Arg Leu Pro Thr Phe Met Thr Gln Lys Ala Arg Asp Ala Leu 2105 2110 2115	6450

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GAC AAC TTA GCA GTG CTG CAC ACG GCT GAG GCA GGT GGA AGG GCG TAC Asp Asn Leu Ala Val Leu His Thr Ala Glu Ala Gly Gly Arg Ala Tyr 2120 2125 2130	6498
AAC CAT GCT CTC AGT GAA CTG CCG GAG ACC CTG GAG ACA TTG CTT TTA Asn His Ala Leu Ser Glu Leu Pro Glu Thr Leu Glu Thr Leu Leu Met 2135 2140 2145 2150	6546
CTG ACA CTT CTG GCT ACA GTC ACG GGA GGG ATC TTT TTA TTC TTG ATG Leu Thr Leu Leu Ala Thr Val Thr Gly Gly Ile Phe Leu Phe Leu Met 2155 2160 2165	6594
AGC GCA AGG GGC ATA GGG AAG ATG ACC CTG GGA ATG TGC TGC ATA ATC Ser Ala Arg Gly Ile Gly Lys Met Thr Leu Gly Met Cys Cys Ile Ile 2170 2175 2180	6642
ACG GCT AGC ATC CTC CTA TGG TAC GCA CAA ATA CAG CCA CAC TGG ATA Thr Ala Ser Ile Leu Leu Trp Tyr Ala Gln Ile Gln Pro His Trp Ile 2185 2190 2195	6690
GCA GCT TCA ATA ATA CTG GAG TTT TTT CTC ATA GTT TTG CTT ATT CCA Ala Ala Ser Ile Ile Leu Glu Phe Phe Leu Ile Val Leu Leu Ile Pro 2200 2205 2210	6738
GAA CCT GAA AAA CAG AGA ACA CCC CAA GAC AAC CAA CTG ACC TAC GTT Glu Pro Glu Lys Gln Arg Thr Pro Gln Asp Asn Gln Leu Thr Tyr Val 2215 2220 2225 2230	6786
GTC ATA GCC ATC CTC ACA GTG GTG GCC GCA ACC ATG GCA AAC GAG ATG Val Ile Ala Ile Leu Thr Val Val Ala Ala Thr Met Ala Asn Glu Met 2235 2240 2245	6834
GGT TTC CTA GAA AAA ACG AAG AAA GAT CTC GGA TTG GGA AGC ATT GCA Gly Phe Leu Glu Lys Thr Lys Lys Asp Leu Gly Leu Gly Ser Ile Ala 2250 2255 2260	6882
ACC CAG CAA CCC GAG AGC AAC ATC CTG GAC ATA GAT CTA CGT CCT GCA Thr Gln Gln Pro Glu Ser Asn Ile Leu Asp Ile Asp Leu Arg Pro Ala 2265 2270 2275	6930
TCA GCA TGG ACG CTG TAT GCC GTG GCC ACA ACA TTT GTT ACA CCA ATG Ser Ala Trp Thr Leu Tyr Ala Val Ala Thr Thr Phe Val Thr Pro Met 2280 2285 2290	6978
TTG AGA CAT AGC ATT GAA AAT TCC TCA GTG AAT GTG TCC CTA ACA GCT Leu Arg His Ser Ile Glu Asn Ser Ser Val Asn Val Ser Leu Thr Ala 2295 2300 2305 2310	7026
ATA GCC AAC CAA GCC ACA GTG TTA ATG GGT CTC GGG AAA GGA TGG CCA Ile Ala Asn Gln Ala Thr Val Leu Met Gly Leu Gly Lys Gly Trp Pro 2315 2320 2325	7074
TTG TCA AAG ATG GAC ATC GGA GTT CCC CTT CTC GCC ATT GGA TGC TAC Leu Ser Lys Met Asp Ile Gly Val Pro Leu Leu Ala Ile Gly Cys Tyr 2330 2335 2340	7122
TCA CAA GTC AAC CCC ATA ACT CTC ACA GCA GCT CTT TTC TTA TTG GTA Ser Gln Val Asn Pro Ile Thr Leu Thr Ala Ala Leu Phe Leu Leu Val 2345 2350 2355	7170

GCA CAT TAT GCC ATC ATA GGG CCA GGA CTC CAA GCA AAA GCA ACC AGA Ala His Tyr Ala Ile Ile Gly Pro Gly Leu Gln Ala Lys Ala Thr Arg 2360 2365 2370	7218
GAA GCT CAG AAA AGA GCA GCG GCG GGC ATC ATG AAA AAC CCA ACT GTC Glu Ala Gln Lys Arg Ala Ala Ala Gly Ile Met Lys Asn Pro Thr Val 2375 2380 2385 2390	7266
GAT GGA ATA ACA GTG ATT GAC CTA GAT CCA ATA CCT TAT GAT CCA AAG Asp Gly Ile Thr Val Ile Asp Leu Asp Pro Ile Pro Tyr Asp Pro Lys 2395 2400 2405	7314
TTT GAA AAG CAG TTG GGA CAA GTA ATG CTC CTA GTC CTC TGC GTG ACT Phe Glu Lys Gln Leu Gly Gln Val Met Leu Leu Val Leu Cys Val Thr 2410 2415 2420	7362
CAA GTA TTG ATG ATG AGG ACT ACA TGG GCT CTG TGT GAG GCT TTA ACC Gln Val Leu Met Met Arg Thr Thr Trp Ala Leu Cys Glu Ala Leu Thr 2425 2430 2435	7410
TTA GCT ACC GGG CCC ATC TCC ACA TTG TGG GAA GGA AAT CCA GGG AGG Leu Ala Thr Gly Pro Ile Ser Thr Leu Trp Glu Gly Asn Pro Gly Arg 2440 2445 2450	7458
TTT TGG AAC ACT ACC ATT GCG GTG TCA ATG GCT AAC ATT TTT AGA GGG Phe Trp Asn Thr Thr Ile Ala Val Ser Met Ala Asn Ile Phe Arg Gly 2455 2460 2465 2470	7506
AGT TAC TTG GCC GGA GCT GGA CTT CTC TTT TCT ATT ATG AAG AAC ACA Ser Tyr Leu Ala Gly Ala Gly Leu Leu Phe Ser Ile Met Lys Asn Thr 2475 2480 2485	7554
ACC AAC ACA AGA AGG GGA ACT GGC AAC ATA GGA GAG ACG CTT GGA GAG Thr Asn Thr Arg Arg Gly Thr Gly Asn Ile Gly Glu Thr Leu Gly Glu 2490 2495 2500	7602
AAA TGG AAA AGC CGA TTG AAC GCA TTG GGA AAA AGT GAA TTC CAG ATC Lys Trp Lys Ser Arg Leu Asn Ala Leu Gly Lys Ser Glu Phe Gln Ile 2505 2510 2515	7650
TAC AAG AAA AGT GGA ATC CAG GAA GTG GAT AGA ACC TTA GCA AAA GAA Tyr Lys Lys Ser Gly Ile Gln Glu Val Asp Arg Thr Leu Ala Lys Glu 2520 2525 2530	7698
GGC ATT AAA AGA GGA GAA ACG GAC CAT CAC GCT GTG TCG CGA GGC TCA Gly Ile Lys Arg Gly Glu Thr Asp His His Ala Val Ser Arg Gly Ser 2535 2540 2545 2550	7746
GCA AAA CTG AGA TGG TTC GTT GAG AGA AAC ATG GTC ACA CCA GAA GGG Ala Lys Leu Arg Trp Phe Val Glu Arg Asn Met Val Thr Pro Glu Gly 2555 2560 2565	7794
AAA GTA GTG GAC CTC GGT TGT GGC AGA GGA GGC TGG TCA TAC TAT TGT Lys Val Val Asp Leu Gly Cys Gly Arg Gly Gly Trp Ser Tyr Tyr Cys 2570 2575 2580	7842
GGA GGA CTA AAG AAT GTA AGA GAA GTC AAA GGC CTA ACA AAA GGA GGA Gly Gly Leu Lys Asn Val Arg Glu Val Lys Gly Leu Thr Lys Gly Gly 2585 2590 2595	7890

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CCA GGA CAC GAA GAA CCC ATC CCC ATG TCA ACA TAT GGG TGG AAT CTA	7938
Pro Gly His Glu Glu Pro Ile Pro Met Ser Thr Tyr Gly Trp Asn Leu	
2600 2605 2610	
GTG CGT CTT CAA AGT GGA GTT GAC GTT TTC TTC ATC CCG CCA GAA AAG	7986
Val Arg Leu Gln Ser Gly Val Asp Val Phe Phe Ile Pro Pro Glu Lys	
2615 2620 2625 2630	
TGT GAC ACA TTA TTG TGT GAC ATA GGG GAG TCA TCA CCA AAT CCC ACA	8034
Cys Asp Thr Leu Leu Cys Asp Ile Gly Glu Ser Ser Pro Asn Pro Thr	
2635 2640 2645	
GTG GAA GCA GGA CGA ACA CTC AGA GTC CTT AAC TTA GTA GAA AAT TGG	8082
Val Glu Ala Gly Arg Thr Leu Arg Val Leu Asn Leu Val Glu Asn Trp	
2650 2655 2660	
TTG AAC AAC AAC ACT CAA TTT TGC ATA AAG GTT CTC AAC CCA TAT ATG	8130
Leu Asn Asn Asn Thr Gln Phe Cys Ile Lys Val Leu Asn Pro Tyr Met	
2665 2670 2675	
CCC TCA GTC ATA GAA AAA ATG GAA GCA CTA CAA AGG AAA TAT GGA GGA	8178
Pro Ser Val Ile Glu Lys Met Glu Ala Leu Gln Arg Lys Tyr Gly Gly	
2680 2685 2690	
GCC TTA GTG AGG AAT CCA CTC TCA CGA AAC TCC ACA CAT GAG ATG TAC	8226
Ala Leu Val Arg Asn Pro Leu Ser Arg Asn Ser Thr His Glu Met Tyr	
2695 2700 2705 2710	
TGG GTA TCC AAT GCT TCC GGG AAC ATA GTG TCA TCA GTG AAC ATG ATT	8274
Trp Val Ser Asn Ala Ser Gly Asn Ile Val Ser Ser Val Asn Met Ile	
2715 2720 2725	
TCA AGG ATG TTG ATC AAC AGA TTT ACA ATG AGA TAC AAG AAA GCC ACT	8322
Ser Arg Met Leu Ile Asn Arg Phe Thr Met Arg Tyr Lys Lys Ala Thr	
2730 2735 2740	
TAC GAG CCG GAT GTT GAC CTC GGA AGC GGA ACC CGT AAC ATC GGG ATT	8370
Tyr Glu Pro Asp Val Asp Leu Gly Ser Gly Thr Arg Asn Ile Gly Ile	
2745 2750 2755	
GAA AGT GAG ATA CCA AAC CTA GAT ATA ATT GGG AAA AGA ATA GAA AAA	8418
Glu Ser Glu Ile Pro Asn Leu Asp Ile Ile Gly Lys Arg Ile Glu Lys	
2760 2765 2770	
ATA AAG CAA GAG CAT GAA ACA TCA TGG CAC TAT GAC CAA GAC CAC CCA	8466
Ile Lys Gln Glu His Glu Thr Ser Trp His Tyr Asp Gln Asp His Pro	
2775 2780 2785 2790	
TAC AAA ACG TGG GCA TAC CAT GGT AGC TAT GAA ACA AAA CAG ACT GGA	8514
Tyr Lys Thr Trp Ala Tyr His Gly Ser Tyr Glu Thr Lys Gln Thr Gly	
2795 2800 2805	
TCA GCA TCA TCC ATG GTC AAC GGA GTG GTC AGG CTG CTG ACA AAA CCT	8562
Ser Ala Ser Ser Met Val Asn Gly Val Val Arg Leu Leu Thr Lys Pro	
2810 2815 2820	
TGG GAC GTT GTC CCC ATG GTG ACA CAG ATG GCA ATG ACA GAC ACG ACT	8610
Trp Asp Val Val Pro Met Val Thr Gln Met Ala Met Thr Asp Thr Thr	
2825 2830 2835	

CCA TTT GGA CAA CAG CGC GTT TTT AAA GAG AAA GTG GAC ACG AGA ACC Pro Phe Gly Gln Gln Arg Val Phe Lys Glu Lys Val Asp Thr Arg Thr 2840 2845 2850	8658
CAA GAA CCG AAA GAA GGC ACG AAG AAA CTA ATG AAA ATA ACA GCA GAG Gln Glu Pro Lys Glu Gly Thr Lys Lys Leu Met Lys Ile Thr Ala Glu 2855 2860 2865 2870	8706
TGG CTT TGG AAA GAA TTA GGG AAG AAA AAG ACA CCC AGG ATG TGC ACC Trp Leu Trp Lys Glu Leu Gly Lys Lys Lys Thr Pro Arg Met Cys Thr 2875 2880 2885	8754
AGA GAA GAA TTC ACA AGA AAG GTG AGA AGC AAT GCA GCC TTG GGG GCC Arg Glu Glu Phe Thr Arg Lys Val Arg Ser Asn Ala Ala Leu Gly Ala 2890 2895 2900	8802
ATA TTC ACT GAT GAG AAC AAG TGG AAG TCG GCA CGT GAG GCT GTT GAA Ile Phe Thr Asp Glu Asn Lys Trp Lys Ser Ala Arg Glu Ala Val Glu 2905 2910 2915	8850
GAT AGT AGG TTT TGG GAG CTG GTT GAC AAG GAA AGG AAT CTC CAT CTT Asp Ser Arg Phe Trp Glu Leu Val Asp Lys Glu Arg Asn Leu His Leu 2920 2925 2930	8898
GAA GGA AAG TGT GAA ACA TGT GTG TAC AAC ATG ATG GGA AAA AGA GAG Glu Gly Lys Cys Glu Thr Cys Val Tyr Asn Met Met Gly Lys Arg Glu 2935 2940 2945 2950	8946
AAG AAG CTA GGG GAA TTC GGC AAG GCA AAA GGC AGC AGA GCC ATA TGG Lys Lys Leu Gly Glu Phe Gly Lys Ala Lys Gly Ser Arg Ala Ile Trp 2955 2960 2965	8994
TAC ATG TGG CTT GGA GCA CGC TTC TTA GAG TTT GAA GCC CTA GGA TTC Tyr Met Trp Leu Gly Ala Arg Phe Leu Glu Phe Glu Ala Leu Gly Phe 2970 2975 2980	9042
TTA AAT GAA GAT CAC TGG TTC TCC AGA GAG AAC TCC CTG AGT GGA GTG Leu Asn Glu Asp His Trp Phe Ser Arg Glu Asn Ser Leu Ser Gly Val 2985 2990 2995	9090
GAA GGA GAA GGG CTG CAC AAG CTA GGT TAC ATT CTA AGA GAC GTG AGC Glu Gly Glu Gly Leu His Lys Leu Gly Tyr Ile Leu Arg Asp Val Ser 3000 3005 3010	9138
AAG AAA GAG GGA GGA GCA ATG TAT GCC GAT GAC ACC GCA GGA TGG GAT Lys Lys Glu Gly Gly Ala Met Tyr Ala Asp Asp Thr Ala Gly Trp Asp 3015 3020 3025 3030	9186
ACA AGA ATC ACA CTA GAA GAC KKA AAA AAT GAA GAA ATG GTA ACA AAC Thr Arg Ile Thr Leu Glu Asp Xaa Lys Asn Glu Glu Met Val Thr Asn 3035 3040 3045	9234
CAC ATG GAA GGA GAA CAC AAG AAA CTA GCC GAG GCC ATT TTC AAA CTA His Met Glu Gly Glu His Lys Lys Leu Ala Glu Ala Ile Phe Lys Leu 3050 3055 3060	9282
ACG TAC CAA AAC AAG GTG GTG CGT GTG CAA AGA CCA ACA CCA AGA GGC Thr Tyr Gln Asn Lys Val Val Arg Val Gln Arg Pro Thr Pro Arg Gly 3065 3070 3075	9330

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ACA GTA ATG GAC ATC ATA TCG AGA AGA GAC CAA AGA GGT AGT GGA CAA Thr Val Met Asp Ile Ile Ser Arg Arg Asp Gln Arg Gly Ser Gly Gln 3080 3085 3090	9378
GTT GGC ACC TAT GGA CTC AAT ACT TTC ACC AAT ATG GAA GCC CAA CTA Val Gly Thr Tyr Gly Leu Asn Thr Phe Thr Asn Met Glu Ala Gln Leu 3095 3100 3105 3110	9426
ATC AGA CAG ATG GAG GGA GAA GGA GTC TTT AAA AGC ATT CAG CAC CTA Ile Arg Gln Met Glu Gly Glu Gly Val Phe Lys Ser Ile Gln His Leu 3115 3120 3125	9474
ACA ATC ACA GAA GAA ATC GCT GTG CAA AAC TGG TTA GCA AGA GTG GGG Thr Ile Thr Glu Glu Ile Ala Val Gln Asn Trp Leu Ala Arg Val Gly 3130 3135 3140	9522
CGC GAA AGG TTA TCA AGA ATG GCC ATC AGT GGA GAT GAT TGT GTT GTG Arg Glu Arg Leu Ser Arg Met Ala Ile Ser Gly Asp Asp Cys Val Val 3145 3150 3155	9570
AAA CCT TTA GAT GAC AGG TTC GCA AGC GCT TTA ACA GCT CTA AAT GAC Lys Pro Leu Asp Asp Arg Phe Ala Ser Ala Leu Thr Ala Leu Asn Asp 3160 3165 3170	9618
ATG GGA AAG ATT AGG AAA GAC ATA CAA CAA TGG GAA CCT TCA AGA GGA Met Gly Lys Ile Arg Lys Asp Ile Gln Gln Trp Glu Pro Ser Arg Gly 3175 3180 3185 3190	9666
TGG AAT GAT TGG ACA CAA GTG CCC TTC TGT TCA CAC CAT TTC CAT GAG Trp Asn Asp Trp Thr Gln Val Pro Phe Cys Ser His His Phe His Glu 3195 3200 3205	9714
TTA ATC ATG AAA GAC GGT CGC GTA CTC GTT GTT CCA TGT AGA AAC CAA Leu Ile Met Lys Asp Gly Arg Val Leu Val Val Pro Cys Arg Asn Gln 3210 3215 3220	9762
GAT GAA CTG ATT GGC AGA GCC CGA ATC TCC CAA GGA GCA GGG TGG TCT Asp Glu Leu Ile Gly Arg Ala Arg Ile Ser Gln Gly Ala Gly Trp Ser 3225 3230 3235	9810
TTG CGG GAG ACG GCC TGT TTG GGG AAG TCT TAC GCC CAA ATG TGG AGC Leu Arg Glu Thr Ala Cys Leu Gly Lys Ser Tyr Ala Gln Met Trp Ser 3240 3245 3250	9858
TTG ATG TAC TTC CAC AGA CGC GAC CTC AGG CTG GCG GCA AAT GCT ATT Leu Met Tyr Phe His Arg Arg Asp Leu Arg Ala Ala Asn Ala Ile 3255 3260 3265 3270	9906
TGC TCG GCA GTA CCA TCA CAT TGG GTT CCA ACA AGT CGA ACA ACC TGG Cys Ser Ala Val Pro Ser His Trp Val Pro Thr Ser Arg Thr Thr Trp 3275 3280 3285	9954
TCC ATA CAT GCT AAA CAT GAA TGG ATG ACA ACG GAA GAC ATG CTG ACA Ser Ile His Ala Lys His Glu Trp Met Thr Thr Glu Asp Met Leu Thr 3290 3295 3300	10002
GTC TGG AAC AGG GTG TGG ATT CAA GAA AAC CCA TGG ATG GAA GAC AAA Val Trp Asn Arg Val Trp Ile Gln Glu Asn Pro Trp Met Glu Asp Lys 3305 3310 3315	10050

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ACT CCA GTG GAA TCA TGG GAG GAA ATC CCA TAC TTG GGG AAA AGA GAA 10098
 Thr Pro Val Glu Ser Trp Glu Glu Ile Pro Tyr Leu Gly Lys Arg Glu
 3320 3325 3330

GAC CAA TGG TGC GGC TCA TTG ATT GGG TTA ACA AGC AGG GCC ACC TGG 10146
 Asp Gln Trp Cys Gly Ser Leu Ile Gly Leu Thr Ser Arg Ala Thr Trp
 3335 3340 3345 3350

GCA AAG AAC ATC CAA GCA GCA ATA AAT CAA GTT AGA TCC CTT ATA GGC 10194
 Ala Lys Asn Ile Gln Ala Ala Ile Asn Gln Val Arg Ser Leu Ile Gly
 3355 3360 3365

AAT GAA GAA TAC ACA GAT TAC ATG CCA TCC ATG AAA AGA TTC AGA AGA 10242
 Asn Glu Glu Tyr Thr Asp Tyr Met Pro Ser Met Lys Arg Phe Arg Arg
 3370 3375 3380

GAA GAG GAA GAA GCA GGA GTT CTG TGG TAGAAAGCAA AACTAACATG AAACAAGG 10297
 Glu Glu Glu Glu Ala Gly Val Leu Trp
 3385 3390

CTAGAAGTCA GGTCGGATTA AGCCATAGTA CGGAAAAAAC TATGCTACCT GTGAGCCCCG 10357
 TCCAAGGACG TTAAGAAG TCAGGCCATC ATAAATGCCA TAGCTTGAGT AACTATGCA 10417
 GCCTGTAGCT CCACCTGAGA AGGTGTAAAA AATCCGGGAG GCCACAAACC ATGGAAGCTG 10477
 TACGCATGGC GTAGTGGACT AGCGGTTAGA GAGGACCCCT CCCTTACAAA TCGCAGCAAC 10537
 AATGGGGGCC CAAGGCGAGA TGAAGCTGTA GTCTCGCTGG AAGGACTAGA GGTAGAGGA 10597
 GACCCCCCG AAACAAAAA CAGCATATTG ACGCTGGGAA AGACCAGAGA TCCTGCTGTC 10657
 TCCTCAGCAT CATTCCAGGC ACAGAACGCC AGAAAATGGA ATGGTGCTGT TGAATCAACA 10717
 GGTCT 10723

(2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

CCCAGTCACG ACGTTGTAAA ACGAC

25

(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 27 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

137

GGATGTGCTG CAAGGCGATT AAGTTGG

27

(2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

TGAGCGGATA ACAATTTAC ACAGG

25

(2) INFORMATION FOR SEQ ID NO:6:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 27 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

GGCTTTACAC TTTATGCTTC CGGCTCG

27

(2) INFORMATION FOR SEQ ID NO:7:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 75 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

GCGGATATTG GAATTCTCTA GAAATTTAAT ACGACTCACT ATAAGTTGTT AGTCTACGTG
GACCGACAAA GACAG

60

75

(2) INFORMATION FOR SEQ ID NO:8:

(i) SEQUENCE CHARACTERISTICS:

138

- (A) LENGTH: 77 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

CCAGTGAATT CGAGCTCAG CGTAAATTTA ATACGACTCA CTATAAGTTG TTAGTCTACG 60
TGGACCGACA AAGACAG 77

(2) INFORMATION FOR SEQ ID NO:9:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 24 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

AGTTGTTAGT CTACGTGGAC CGAC 24

(2) INFORMATION FOR SEQ ID NO:10:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 34 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

GACAGATTCT TTGAGGGAGC TGAGCTCAAC GTAG 34

(2) INFORMATION FOR SEQ ID NO:11:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 28 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA

139

(iii) HYPOTHETICAL: NO
(iv) ANTISENSE: NO
(v) FRAGMENT TYPE:
(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

TCAATATGCT GAAACGCGAG AGAAACCG

28

(2) INFORMATION FOR SEQ ID NO:12:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 26 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA
(iii) HYPOTHETICAL: NO
(iv) ANTISENSE: NO
(v) FRAGMENT TYPE:
(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

GGGATTGTTA GGAAACGAAG GAACGC

26

(2) INFORMATION FOR SEQ ID NO:13:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 32 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA
(iii) HYPOTHETICAL: NO
(iv) ANTISENSE: NO
(v) FRAGMENT TYPE:
(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

CCACCAACAG CAGGGATACT GAAAAGATGG GG

32

(2) INFORMATION FOR SEQ ID NO:14:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 25 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA
(iii) HYPOTHETICAL: NO
(iv) ANTISENSE: NO
(v) FRAGMENT TYPE:
(vi) ORIGINAL SOURCE:

140

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:
TGCAGATCTG CGTCTCCTAT TCAAG 25
- (2) INFORMATION FOR SEQ ID NO:15:
- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 25 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
(iii) HYPOTHETICAL: NO
(iv) ANTISENSE: NO
(v) FRAGMENT TYPE:
(vi) ORIGINAL SOURCE:
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:
CGTGAACATG TGTACCCTCA TGGCC 25
- (2) INFORMATION FOR SEQ ID NO:16:
- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 26 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
(iii) HYPOTHETICAL: NO
(iv) ANTISENSE: NO
(v) FRAGMENT TYPE:
(vi) ORIGINAL SOURCE:
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:
TTGCACCAAC AGTCAATGTC TTCAGG 26
- (2) INFORMATION FOR SEQ ID NO:17:
- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 25 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
(iii) HYPOTHETICAL: NO
(iv) ANTISENSE: NO
(v) FRAGMENT TYPE:
(vi) ORIGINAL SOURCE:
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:
ACCAGAAGAC ATAGATTGTT GGTGC 25
- (2) INFORMATION FOR SEQ ID NO:18:
- (i) SEQUENCE CHARACTERISTICS:

141

- (A) LENGTH: 25 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

GCACCAACAG TCTATGTCTT CTGGC

25

(2) INFORMATION FOR SEQ ID NO:19:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 25 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:

ATGTTTCCAG GCCCCTTCTG ATGAC

25

(2) INFORMATION FOR SEQ ID NO:20:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 25 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:

GCAGCAATCC TGGCATAAC CATAG

25

(2) INFORMATION FOR SEQ ID NO:21:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 27 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO

142

(iv) ANTISENSE: NO
(v) FRAGMENT TYPE:
(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:
GGTTGACATA GTCTTAGAAC ATGGAAG 27

(2) INFORMATION FOR SEQ ID NO:22:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 27 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA
(iii) HYPOTHETICAL: NO
(iv) ANTISENSE: NO
(v) FRAGMENT TYPE:
(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:
CTTCCATGTT CTAAGACTAT GTCAACC 27

(2) INFORMATION FOR SEQ ID NO:23:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 35 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA
(iii) HYPOTHETICAL: NO
(iv) ANTISENSE: NO
(v) FRAGMENT TYPE:
(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:
GTCTTAGAAC ATGGAAGTTG TGTGACGACG ATGCC 35

(2) INFORMATION FOR SEQ ID NO:24:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 25 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA
(iii) HYPOTHETICAL: NO
(iv) ANTISENSE: NO
(v) FRAGMENT TYPE:
(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:
ACAACAGAAT CTCGCTGCCC AACAC 25

143

(2) INFORMATION FOR SEQ ID NO:25:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:

GCAAACACTC CATGGTAGAC AGAGG

25

(2) INFORMATION FOR SEQ ID NO:26:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:

CCTCTGTCTA CCATGGAGTG TTTGC

25

(2) INFORMATION FOR SEQ ID NO:27:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 27 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:

CCACATCCAT TTCCCCATCC TCTGTCT

27

(2) INFORMATION FOR SEQ ID NO:28:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 30 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

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- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:

GGAAAGGGAG GCATTGTGAC CTGTGCTATG

30

(2) INFORMATION FOR SEQ ID NO:29:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 28 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:29:

GGAAATCAAA ATAACACCAC AGAGTTCC

28

(2) INFORMATION FOR SEQ ID NO:30:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 34 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:30:

CTGCAGCAAC ACCATCTCAT TGAAGTCGAG GCCC

34

(2) INFORMATION FOR SEQ ID NO:31:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 25 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

145

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:31:
GACTTCAATG AGATGGTGCT GCTGC 25
- (2) INFORMATION FOR SEQ ID NO:32:
- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 25 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
(iii) HYPOTHETICAL: NO
(iv) ANTISENSE: NO
(v) FRAGMENT TYPE:
(vi) ORIGINAL SOURCE:
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:32:
GCAGCAGCAC CATCTCATTG AAGTC 25
- (2) INFORMATION FOR SEQ ID NO:33:
- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 28 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
(iii) HYPOTHETICAL: NO
(iv) ANTISENSE: NO
(v) FRAGMENT TYPE:
(vi) ORIGINAL SOURCE:
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:33:
AAGCTTGGCT GGTGCACAGG CAATGGTT 28
- (2) INFORMATION FOR SEQ ID NO:34:
- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 27 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
(iii) HYPOTHETICAL: NO
(iv) ANTISENSE: NO
(v) FRAGMENT TYPE:
(vi) ORIGINAL SOURCE:
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:34:
TGGTAACGGC AGGTCTAGGA ACCATTG 27
- (2) INFORMATION FOR SEQ ID NO:35:

146

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 23 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:35:

GGACATCTCA AGTGCAGGCT GAG

23

(2) INFORMATION FOR SEQ ID NO:36:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 23 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:36:

CTCAGCCTGC ACTTGAGATG TCC

23

(2) INFORMATION FOR SEQ ID NO:37:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 27 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:37:

GAAGGAAATA GCAGAAACAC AACATGG

27

(2) INFORMATION FOR SEQ ID NO:38:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 33 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA

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- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:38:

CCCTTCATAT TGTACTCTGA TAACTATTGT TCC

33

(2) INFORMATION FOR SEQ ID NO:39:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 32 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:39:

CCTCCATTG GAGACAGCTA CATCATCATA GG

32

(2) INFORMATION FOR SEQ ID NO:40:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 32 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:40:

CCTATGATGA TGTAGCTGTC TCCGAATGGA GG

32

(2) INFORMATION FOR SEQ ID NO:41:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 29 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

148

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:41:

ATGGCCATTT TAGGTGACAC AGCCTGGGA

29

(2) INFORMATION FOR SEQ ID NO:42:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 27 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:42:

TGTAACACT CCTCCAGGG ATCCAAA

27

(2) INFORMATION FOR SEQ ID NO:43:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 29 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:43:

CTCATAGGAG TCATTATCAC ATGGATAGG

29

(2) INFORMATION FOR SEQ ID NO:44:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 34 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:44:

GGGGATTCTG GTTGGAACTT ATATTGTTCT GTCC

34

(2) INFORMATION FOR SEQ ID NO:45:

149

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 30 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:45:

TGATTCAATT CTGGTGTTAT TTGTTCCAC

30

(2) INFORMATION FOR SEQ ID NO:46:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:46:

AAGGAATCAT GCAGGCAGGA AAACG

25

(2) INFORMATION FOR SEQ ID NO:47:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 22 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:47:

ACTTCCAGCG AGTCCAAGC TC

22

(2) INFORMATION FOR SEQ ID NO:48:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

150

(iii) HYPOTHETICAL: NO
(iv) ANTISENSE: NO
(v) FRAGMENT TYPE:
(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:48:

AACAGAGCCG TCCATGCCGA TATGG

25

(2) INFORMATION FOR SEQ ID NO:49:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 22 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA
(iii) HYPOTHETICAL: NO
(iv) ANTISENSE: NO
(v) FRAGMENT TYPE:
(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:49:

TCCATTGCTC CAAAGGGTGT GT

22

(2) INFORMATION FOR SEQ ID NO:50:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 25 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA
(iii) HYPOTHETICAL: NO
(iv) ANTISENSE: NO
(v) FRAGMENT TYPE:
(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:50:

AGCTTGAGAT GGACTTTGAT TTCTG

25

(2) INFORMATION FOR SEQ ID NO:51:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 22 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA
(iii) HYPOTHETICAL: NO
(iv) ANTISENSE: NO
(v) FRAGMENT TYPE:
(vi) ORIGINAL SOURCE:

151

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:51:

GGTCTGATTT CCATCCCGTA CC

22

(2) INFORMATION FOR SEQ ID NO:52:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 23 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:52:

GTCCTTTAGA GACCTGGGAA GAG

23

(2) INFORMATION FOR SEQ ID NO:53:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:53:

GTTTTCTCAA GAGTAGTCCA GCTGC

25

(2) INFORMATION FOR SEQ ID NO:54:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:54:

ATCAATTGGC AGTGACTATC ATGGC

25

(2) INFORMATION FOR SEQ ID NO:55:

152

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 25 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:55:

TGTTAAGAGC AGTGGAGAAA CGGAC

25

(2) INFORMATION FOR SEQ ID NO:56:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 25 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:56:

GATTGAGACC TTTGATCGTC AACGC

25

(2) INFORMATION FOR SEQ ID NO:57:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 25 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:57:

TGACAGGACC ATTAGTGGCT GGAGG

25

(2) INFORMATION FOR SEQ ID NO:58:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 34 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA

153

- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:58:

CGTGCTCACT GGACGATCGG CCGATTTGGA ACTG

34

(2) INFORMATION FOR SEQ ID NO:59:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 24 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:59:

GGGCTGCTTC CTGATATTTTC TGCC

24

(2) INFORMATION FOR SEQ ID NO:60:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:60:

CCTGTGGGAA GTGAAGAAAC AACGG

25

(2) INFORMATION FOR SEQ ID NO:61:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 30 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:61:

GCTCCATCTT CCAGTTCAGC CTTTCCCATG

30

(2) INFORMATION FOR SEQ ID NO:62:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:62:

CTCCGGCTCC AATCTGAGAG TATCC

25

(2) INFORMATION FOR SEQ ID NO:63:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:63:

CCTAATATCA TATGGAGGAG GCTGG

25

(2) INFORMATION FOR SEQ ID NO:64:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:64:

GAAGGAGAAG AAGTCCAGGT ATTGG

25

(2) INFORMATION FOR SEQ ID NO:65:

155

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:65:

CTGTGACAA TTGGAGATCC TGACG

25

(2) INFORMATION FOR SEQ ID NO:66:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:66:

GTGGAGCATA TGTGAGTGCT ATAGC

25

(2) INFORMATION FOR SEQ ID NO:67:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:67:

TCTGACTATG GCCGGAAGGT ATCTC

25

(2) INFORMATION FOR SEQ ID NO:68:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

156

- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:68:

ACATTAATCT TGGCCCCAC TAGAG

25

(2) INFORMATION FOR SEQ ID NO:69:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 19 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:69:

CGATCTCCCG CCCGGTGTG

19

(2) INFORMATION FOR SEQ ID NO:70:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 25 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:70:

CTAACTGGTG ATAGCAGCCT CATGG

25

(2) INFORMATION FOR SEQ ID NO:71:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 27 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

157

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:71:

CCTACTGAGT TGTATCACTT TCTTCC

27

(2) INFORMATION FOR SEQ ID NO:72:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 26 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:72:

TGGATTCTT CCTATTCTCC CTCTTC

26

(2) INFORMATION FOR SEQ ID NO:73:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:73:

TTCAAGGCTG AGAGGGTTAT AGACC

25

(2) INFORMATION FOR SEQ ID NO:74:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:74:

TCTGGTTGGC CTACAGAGTG GCAGC

25

(2) INFORMATION FOR SEQ ID NO:75:

158

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 27 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:75:

CCTTCTTTTG TCCAGATTC CACTTCC

27

(2) INFORMATION FOR SEQ ID NO:76:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 35 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:76:

GCGTACAACC ATGCTCTCAG TGAAGTCCG GAGAC

35

(2) INFORMATION FOR SEQ ID NO:77:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 24 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:77:

TTCCCAGGGT CATCTCCCT ATAC

24

(2) INFORMATION FOR SEQ ID NO:78:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 31 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

159

- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:78:

GATGCTAGCC GTGATTATGC AGCACATTCC C

31

(2) INFORMATION FOR SEQ ID NO:79:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:79:

AAACAGAGAA CACCCAAGA CAACC

25

(2) INFORMATION FOR SEQ ID NO:80:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 21 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:80:

CGGCATACAG CGTCCATGCT G

21

(2) INFORMATION FOR SEQ ID NO:81:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

160

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:81:

GTCTCGGGAA AGGATGGCCA TTGTC

25

(2) INFORMATION FOR SEQ ID NO:82:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:82:

CTCTGGTTGC TTTTGCTTGA AGTCC

25

(2) INFORMATION FOR SEQ ID NO:83:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 27 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:83:

CCGCCGCTGC TCTTTTCTGA GCTTCTC

27

(2) INFORMATION FOR SEQ ID NO:84:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:84:

AGGACTACAT GGGCTCTGTG TGAGG

25

(2) INFORMATION FOR SEQ ID NO:85:

161

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 19 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:85:

GAGAAGTCCA GCTCCGGCC

19

(2) INFORMATION FOR SEQ ID NO:86:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:86:

AGAGAAACAT GGTACACCA GAAGG

25

(2) INFORMATION FOR SEQ ID NO:87:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 22 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:87:

GTTCTTCGTG TCCTGGTCCT CC

22

(2) INFORMATION FOR SEQ ID NO:88:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

162

- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:88:

GGAAATATGG AGGAGCCTAG TGAGG

25

(2) INFORMATION FOR SEQ ID NO:89:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 22 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:89:

ACCCAGTACA TCTCATGTGT GG

22

(2) INFORMATION FOR SEQ ID NO:90:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 28 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:90:

GAGCATGAAA CATCATGGCA CTATGACC

28

(2) INFORMATION FOR SEQ ID NO:91:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 25 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

163

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:91:

TCATGGCACT ATGACCAAGA CCACC

25

(2) INFORMATION FOR SEQ ID NO:92:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 22 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:92:

CAGTCTGACC ACTCCGTTCA CC

22

(2) INFORMATION FOR SEQ ID NO:93:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:93:

AAGGTGAGAA GCAATGCAGC CTTGG

25

(2) INFORMATION FOR SEQ ID NO:94:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 29 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:94:

GGGCCATATT CACTGATGAG AACAAAGTGG

29

(2) INFORMATION FOR SEQ ID NO:95:

(i) SEQUENCE CHARACTERISTICS:

164

- (A) LENGTH: 22 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:95:

TCTTTCCTG TCAACCAGCT CC

22

(2) INFORMATION FOR SEQ ID NO:96:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 25 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:96:

AATGAAGATC ACTGGTTCTC CAGAG

25

(2) INFORMATION FOR SEQ ID NO:97:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 25 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:97:

ACGTGAGCAA GAAAGAGGGA GGAGC

25

(2) INFORMATION FOR SEQ ID NO:98:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 22 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO

165

- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:98:

TGTCCCATCC TGCTGTGTCA TC

22

(2) INFORMATION FOR SEQ ID NO:99:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 30 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:99:

GCTAGTTTCT TGTGTTCTCC TTCCATGTGG

30

(2) INFORMATION FOR SEQ ID NO:100:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 25 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:100:

TCATATCGAG AAGAGACCAA AGAGG

25

(2) INFORMATION FOR SEQ ID NO:101:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 24 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:101:

ACTCCTTCTC CCTCCATCTG TCTG

24

166

(2) INFORMATION FOR SEQ ID NO:102:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 27 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:102:

ATGCTTTTGA AGATTCCTTC TCCCTCC

27

(2) INFORMATION FOR SEQ ID NO:103:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 32 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:103:

GCACAGCGAT TTCTTCTGTG ATTGTTAGGT GC

32

(2) INFORMATION FOR SEQ ID NO:104:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:104:

ACAATGGGAA CCTTCAAGAG GATGG

25

(2) INFORMATION FOR SEQ ID NO:105:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 45 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

167

(ii) MOLECULE TYPE: cDNA
(iii) HYPOTHETICAL: NO
(iv) ANTISENSE: NO
(v) FRAGMENT TYPE:
(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:105:

TTATCACATT GGATCCTTCA AGAGGATGGA ATGATTGGAC ACAAG

45

(2) INFORMATION FOR SEQ ID NO:106:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 28 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA
(iii) HYPOTHETICAL: NO
(iv) ANTISENSE: NO
(v) FRAGMENT TYPE:
(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:106:

CAGAAGGGCA CTTGTGTCCA ATCATTCC

28

(2) INFORMATION FOR SEQ ID NO:107:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 21 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA
(iii) HYPOTHETICAL: NO
(iv) ANTISENSE: NO
(v) FRAGMENT TYPE:
(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:107:

CTCCCTGGGA AATTCGGGCT C

21

(2) INFORMATION FOR SEQ ID NO:108:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 28 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA
(iii) HYPOTHETICAL: NO
(iv) ANTISENSE: NO
(v) FRAGMENT TYPE:
(vi) ORIGINAL SOURCE:

168

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:108:

CCGTCTCCCG CAAAGACCAC CCTGCTCC

28

(2) INFORMATION FOR SEQ ID NO:109:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 44 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:109:

TTATCACCTA TCTAGACCGT CTCCGCAA GACCACCCTG CTCC

44

(2) INFORMATION FOR SEQ ID NO:110:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 26 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:110:

GTTGGAACCC AATGTGATGG TACTGC

26

(2) INFORMATION FOR SEQ ID NO:111:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:111:

ACAAGTCGAA CAACCTGGTC CATA C

25

(2) INFORMATION FOR SEQ ID NO:112:

169

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 21 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:112:

GCATGTCTTC CGTCGTCATC C

21

- (2) INFORMATION FOR SEQ ID NO:113:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 25 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:113:

CTTGAATCCA CACCCTGTTC CAGAC

25

- (2) INFORMATION FOR SEQ ID NO:114:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 25 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:114:

ATACACAGAT TACATGCCAT CCATG

25

- (2) INFORMATION FOR SEQ ID NO:115:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 21 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA

170

- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:115:

TTTTGCCTTC TACCACAGGA C

21

(2) INFORMATION FOR SEQ ID NO:116:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:116:

GAAACAAGGC TAGAAGTCAG GTCGG

25

(2) INFORMATION FOR SEQ ID NO:117:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 23 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:117:

GACGGGGCTC ACAGGTAGCA TAG

23

(2) INFORMATION FOR SEQ ID NO:118:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

171

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:118:

GCCTGTAGCT CCACCTGAGA AGGTG

25

(2) INFORMATION FOR SEQ ID NO:119:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:119:

GGAAGCTGTA CGCATGGCGT AGTGG

25

(2) INFORMATION FOR SEQ ID NO:120:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 19 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:120:

GGGCCCCCGT TGTTGCTGC

19

(2) INFORMATION FOR SEQ ID NO:121:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 37 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:121:

AGAACCCTGTT GATTCAACAG CACCATTCCTCA TTTTCTG

37

(2) INFORMATION FOR SEQ ID NO:122:

172

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 59 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:122:

TTATCACCTA GCATGCTCTA GAAGAACCTG TTGATTCAAC AGCACCATTC CATTTCCTG 59

(2) INFORMATION FOR SEQ ID NO:123:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 52 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:123:

TTATCACCTA TCTAGAGAAC CTGTTGATTC AACAGCACCA TTCCATTTTC TG 52

(2) INFORMATION FOR SEQ ID NO:124:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 2394 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:
- (ix) FEATURE:

- (A) NAME/KEY: Coding Sequence
- (B) LOCATION: 1...2394
- (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:124:

AGA TTC TCA AAA GGA TTG CTC TCA GGC CAA GGA CCC ATG AAA TTG GTG 48
 Arg Phe Ser Lys Gly Leu Leu Ser Gly Gln Gly Pro Met Lys Leu Val
 1 5 10 15

173

ATG	GCT	TTC	ATA	GCA	TTC	TTA	AGA	TTT	CTA	GCC	ATA	CCC	CCA	ACA	GCA	96
Met	Ala	Phe	Ile	Ala	Phe	Leu	Arg	Phe	Leu	Ala	Ile	Pro	Pro	Thr	Ala	
			20					25					30			
GGA	ATT	TTG	GCT	AGA	TGG	GGC	TCA	TTC	AAG	AAG	AAT	GGA	GCG	ATT	AAA	144
Gly	Ile	Leu	Ala	Arg	Trp	Gly	Ser	Phe	Lys	Lys	Asn	Gly	Ala	Ile	Lys	
		35				40					45					
GTG	TTA	CGG	GGT	TTC	AAG	AGA	GAA	ATC	TCA	AAC	ATG	CTA	AAC	ATA	ATG	192
Val	Leu	Arg	Gly	Phe	Lys	Arg	Glu	Ile	Ser	Asn	Met	Leu	Asn	Ile	Met	
	50					55					60					
AAC	AGG	AGG	AAA	AGA	TCC	GTG	ACC	ATG	CTC	CTT	ATG	CTG	CTG	CCC	ACA	240
Asn	Arg	Arg	Lys	Arg	Ser	Val	Thr	Met	Leu	Leu	Met	Leu	Leu	Pro	Thr	
65					70					75					80	
GCC	CTG	GCG	TTC	CAT	CTG	ACG	ACA	CGA	GGG	GGA	GAG	CCG	CAT	ATG	ATA	288
Ala	Leu	Ala	Phe	His	Leu	Thr	Thr	Arg	Gly	Gly	Glu	Pro	His	Met	Ile	
				85					90					95		
GTT	AGC	AAG	CAG	GAA	AGA	GGA	AAG	TCA	CTT	TTG	TTC	AAG	ACC	TCT	GCA	336
Val	Ser	Lys	Gln	Glu	Arg	Gly	Lys	Ser	Leu	Leu	Phe	Lys	Thr	Ser	Ala	
			100					105					110			
GGT	GTC	AAC	ATG	TGC	ACC	CTC	ATT	GCG	ATG	GAT	TTG	GGA	GAG	TTG	TGT	384
Gly	Val	Asn	Met	Cys	Thr	Leu	Ile	Ala	Met	Asp	Leu	Gly	Glu	Leu	Cys	
		115				120						125				
GAG	GAC	ACG	ATG	ACC	TAC	AAA	TGC	CCC	CGG	ATC	ACT	GAG	GCG	GAA	CCA	432
Glu	Asp	Thr	Met	Thr	Tyr	Lys	Cys	Pro	Arg	Ile	Thr	Glu	Ala	Glu	Pro	
	130					135					140					
GAT	GAC	GTT	GAC	TGT	TGG	TGC	AAT	GCC	ACG	GAC	ACA	TGG	GTG	ACC	TAT	480
Asp	Asp	Val	Asp	Cys	Trp	Cys	Asn	Ala	Thr	Asp	Thr	Trp	Val	Thr	Tyr	
145				150						155					160	
GGA	ACG	TGC	TCT	CAA	ACT	GGC	GAA	CAC	CGA	CGA	GAC	AAA	CGT	TCC	GTC	528
Gly	Thr	Cys	Ser	Gln	Thr	Gly	Glu	His	Arg	Arg	Asp	Lys	Arg	Ser	Val	
				165					170					175		
GCA	TTG	GCC	CCA	CAC	GTG	GGG	CTT	GGC	CTA	GAA	ACA	AGA	GCC	GAA	ACG	576
Ala	Leu	Ala	Pro	His	Val	Gly	Leu	Gly	Leu	Glu	Thr	Arg	Ala	Glu	Thr	
			180					185					190			
TGG	ATG	TCC	TCT	GAA	GGT	GCT	TGG	AAA	CAG	ATA	CAA	AAA	GTA	GAG	ACT	624
Trp	Met	Ser	Ser	Glu	Gly	Ala	Trp	Lys	Gln	Ile	Gln	Lys	Val	Glu	Thr	
		195				200						205				
TGG	GCT	CTG	AGA	CAT	CCA	GGA	TTC	ACG	GTG	ATA	GCC	CTT	TTT	CTA	GCA	672
Trp	Ala	Leu	Arg	His	Pro	Gly	Phe	Thr	Val	Ile	Ala	Leu	Phe	Leu	Ala	
	210					215					220					
CAT	GCC	ATA	GGA	ACA	TCC	ATC	ACC	CAG	AAA	GGG	ATC	ATT	TTC	ATT	TTG	720
His	Ala	Ile	Gly	Thr	Ser	Ile	Thr	Gln	Lys	Gly	Ile	Ile	Phe	Ile	Leu	
225					230					235					240	
CTG	ATG	CTG	GTA	ACA	CCA	TCT	ATG	GCC	ATG	CGA	TGC	GTG	GGA	ATA	GGC	768
Leu	Met	Leu	Val	Thr	Pro	Ser	Met	Ala	Met	Arg	Cys	Val	Gly	Ile	Gly	
				245					250					255		

AAC Asn	AGA Arg	GAC Asp	TTC Phe	GTG Val	GAA Glu	GGA Gly	CTG Leu	TCA Ser	GGA Gly	GCA Ala	ACA Thr	TGG Trp	GTG Val	GAT Asp	GTG Val	816
			260					265					270			
GTA Val	CTG Leu	GAG Glu	CAT His	GGA Gly	AGT Ser	TGC Cys	GTC Val	ACC Thr	ACC Thr	ATG Met	GCA Ala	AAA Lys	AAC Asn	AAA Lys	CCA Pro	864
			275					280					285			
ACA Thr	CTG Leu	GAC Asp	ATT Ile	GAA Glu	CTC Leu	TTG Leu	AAG Lys	ACG Thr	GAG Glu	GTC Val	ACA Thr	AAC Asn	CCT Pro	GCA Ala	GTT Val	912
			290				295					300				
CTG Leu	CGT Arg	AAA Lys	TTG Leu	TGC Cys	ATT Ile	GAA Glu	GCT Ala	AAA Lys	ATA Ile	TCA Ser	AAC Asn	ACC Thr	ACC Thr	ACC Thr	GAT Asp	960
			305			310				315					320	
TCG Ser	AGA Arg	TGT Cys	CCA Pro	ACA Thr	CAA Gln	GGA Gly	GAA Glu	GCC Ala	ACA Thr	CTG Leu	GTG Val	GAA Glu	GAA Glu	CAA Gln	GAC Asp	1008
				325					330					335		
GCG Ala	AAC Asn	TTT Phe	GTG Val	TGC Cys	CGA Arg	CGA Arg	ACG Thr	TTC Phe	GTG Val	GAC Asp	AGA Arg	GGC Gly	TGG Trp	GGC Gly	AAT Asn	1056
			340					345					350			
GGC Gly	TGT Cys	GGG Gly	CTA Leu	TTC Phe	GGA Gly	AAA Lys	GGT Gly	AGT Ser	CTA Leu	ATA Ile	ACG Thr	TGT Cys	GCC Ala	AAG Lys	TTT Phe	1104
			355				360					365				
AAG Lys	TGT Cys	GTG Val	ACA Thr	AAA Lys	CTA Leu	GAA Glu	GGA Gly	AAG Lys	ATA Ile	GCT Ala	CAA Gln	TAT Tyr	GAA Glu	AAC Asn	CTA Leu	1152
						375					380					
AAA Lys	TAT Tyr	TCA Ser	GTG Val	ATA Ile	GTC Val	ACC Thr	GTC Val	CAC His	ACT Thr	GGA Gly	GAT Asp	CAG Gln	CAC His	CAG Gln	GTG Val	1200
			385			390					395				400	
GGA Gly	AAT Asn	GAG Glu	ACT Thr	ACA Thr	GAA Glu	CAT His	GGA Gly	ACA Thr	ACT Thr	GCA Ala	ACC Thr	ATA Ile	ACA Thr	CCT Pro	CAA Gln	1248
				405					410					415		
GCT Ala	CCT Pro	ACG Thr	TCG Ser	GAA Glu	ATA Ile	CAG Gln	CTG Leu	ACC Thr	GAC Asp	TAC Tyr	GGA Gly	ACC Thr	CTT Leu	ACA Thr	TTA Leu	1296
			420					425					430			
GAT Asp	TGT Cys	TCA Ser	CCT Pro	AGG Arg	ACA Thr	GGG Gly	CTA Leu	GAT Asp	TTT Phe	AAC Asn	GAG Glu	ATG Met	GTG Val	TTG Leu	CTG Leu	1344
			435				440					445				
ACA Thr	ATG Met	AAA Lys	AAG Lys	AAA Lys	TCA Ser	TGG Trp	CTT Leu	GTC Val	CAC His	AAA Lys	CAG Gln	TGG Trp	TTT Phe	CTA Leu	GAC Asp	1392
						455					460					
TTA Leu	CCA Pro	CTG Leu	CCT Pro	TGG Trp	ACC Thr	TCT Ser	GGG Gly	GCT Ala	TTA Leu	ACA Thr	TCC Ser	CAA Gln	GAG Glu	ACT Thr	TGG Trp	1440
					470					475				480		
AAC Asn	AGA Arg	CAA Gln	GAT Asp	TTA Leu	CTG Leu	GTC Val	ACA Thr	TTT Phe	AAG Lys	ACA Thr	GCT Ala	CAT His	GCA Ala	AAG Lys	AAG Lys	1488
				485					490					495		

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CAG Gln	GAA Glu	GTA Val	GTC Val	GTA Val	CTA Leu	GGA Gly	TCA Ser	CAA Gln	GAA Glu	GGA Gly	GCA Ala	ATG Met	CAC His	ACT Thr	GCG Ala	1536
			500					505					510			
CTG Leu	ACT Thr	GGA Gly	GCG Ala	ACA Thr	GAA Glu	ATC Ile	CAA Gln	ACG Thr	TCA Ser	GGA Gly	ACG Thr	ACA Thr	ACA Thr	ATT Ile	TTC Phe	1584
		515					520					525				
GCA Ala	GGA Gly	CAC His	CTA Leu	AAA Lys	TGC Cys	AGA Arg	CTA Leu	AAA Lys	ATG Met	GAC Asp	AAA Lys	CTA Leu	ACT Thr	TTA Leu	AAA Lys	1632
		530				535					540					
GGG Gly	ATG Met	TCA Ser	TAT Tyr	GTG Val	ATG Met	TGC Cys	ACA Thr	GGC Gly	TCA Ser	TTC Phe	AAG Lys	TTA Leu	GAG Glu	AAA Lys	GAA Glu	1680
					550					555					560	
GTG Val	GCT Ala	GAG Glu	ACC Thr	CAG Gln	CAT His	GGA Gly	ACT Thr	GTT Val	CTG Leu	GTG Val	CAG Gln	GTT Val	AAA Lys	TAT Tyr	GAA Glu	1728
				565					570					575		
GGA Gly	ACA Thr	GAC Asp	GCA Ala	CCA Pro	TGC Cys	AAG Lys	ATT Ile	CCC Pro	TTT Phe	TCG Ser	ACC Thr	CAA Gln	GAT Asp	GAG Glu	AAA Lys	1776
			580					585					590			
GGA Gly	GCA Ala	ACC Thr	CAG Gln	AAT Asn	GGG Gly	AGA Arg	TTA Leu	ATA Ile	ACA Thr	GCC Ala	AAC Asn	CCC Pro	ATA Ile	GTC Val	ACT Thr	1824
		595					600					605				
GAC Asp	AAA Lys	GAA Glu	AAA Lys	CCA Pro	GTC Val	AAT Asn	ATT Ile	GAG Glu	GCA Ala	GAA Glu	CCA Pro	CCC Pro	TTT Phe	GGT Gly	GAG Glu	1872
		610				615					620					
AGC Ser	TAC Tyr	ATC Ile	GTG Val	GTA Val	GGA Gly	GCA Ala	GGT Gly	GAA Glu	AAA Lys	GCT Ala	TTG Leu	AAA Lys	CTA Leu	AGC Ser	TGG Trp	1920
					630					635					640	
TTC Phe	AAG Lys	AAA Lys	GGA Gly	AGC Ser	AGC Ser	ATA Ile	GGG Gly	AAA Lys	ATG Met	TTT Phe	GAA Glu	GCA Ala	ACT Thr	GCC Ala	CGA Arg	1968
				645				650						655		
GGA Gly	GCA Ala	CGA Arg	AGG Arg	ATG Met	GCC Ala	ATT Ile	CTG Leu	GGA Gly	GAC Asp	ACC Thr	GCA Ala	TGG Trp	GAC Asp	TTC Phe	GGT Gly	2016
			660					665					670			
TCT Ser	ATA Ile	GGA Gly	GGA Gly	GTG Val	TTC Phe	ACG Thr	TCT Ser	ATG Met	GGA Gly	AAA Lys	CTG Leu	GTA Val	CAC His	CAG Gln	GTT Val	2064
		675					680					685				
TTT Phe	GGA Gly	ACT Thr	GCA Ala	TAT Tyr	GGA Gly	GTT Val	TTG Leu	TTT Phe	AGC Ser	GGA Gly	GTT Val	TCT Ser	TGG Trp	ACC Thr	ATG Met	2112
		690				695					700					
AAA Lys	ATA Ile	GGA Gly	ATA Ile	GGG Gly	ATT Ile	CTG Leu	CTG Leu	ACA Thr	TGG Trp	CTA Leu	GGA Gly	TTA Leu	AAT Asn	TCA Ser	AGG Arg	2160
		705			710					715					720	
AAC Asn	ACG Thr	TCC Ser	CTT Leu	TCG Ser	GTG Val	ATG Met	TGC Cys	ATC Ile	GCA Ala	GTT Val	GGC Gly	ATG Met	GTC Val	ACA Thr	CTG Leu	2208
				725					730					735		

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TAC	CTA	GGA	GTC	ATG	GTT	CAG	GCA	GAT	TCG	GGA	TGT	GTA	ATC	AAC	TGG	2256
Tyr	Leu	Gly	Val	Met	Val	Gln	Ala	Asp	Ser	Gly	Cys	Val	Ile	Asn	Trp	
			740					745					750			
AAA	GGC	AGA	GAA	CTT	AAA	TGT	GGA	AGC	GGC	ATT	TTT	GTC	ACT	AAT	GAA	2304
Lys	Gly	Arg	Glu	Leu	Lys	Cys	Gly	Ser	Gly	Ile	Phe	Val	Thr	Asn	Glu	
			755				760					765				
GTT	CAC	ACT	TGG	ACA	GAG	CAA	TAC	AAA	TTC	CAG	GCT	GAC	TCC	CCC	AAG	2352
Val	His	Thr	Trp	Thr	Glu	Gln	Tyr	Lys	Phe	Gln	Ala	Asp	Ser	Pro	Lys	
	770					775					780					
AGA	CTA	TCA	GCA	GCC	ATT	GGG	AAG	GCA	TGG	GAG	GAG	GGT	GTG			2394
Arg	Leu	Ser	Ala	Ala	Ile	Gly	Lys	Ala	Trp	Glu	Glu	Gly	Val			
785					790					795						

(2) INFORMATION FOR SEQ ID NO:125:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2145 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(ix) FEATURE:

(A) NAME/KEY: Coding Sequence

(B) LOCATION: 1...2145

(D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:125:

AAG	GTC	TTA	AAA	GGC	TTC	AAG	AAG	GAG	ATC	TCA	AAC	ATG	CTG	AGC	ATT	48
Lys	Val	Leu	Lys	Gly	Phe	Lys	Lys	Glu	Ile	Ser	Asn	Met	Leu	Ser	Ile	
1				5				10					15			
ATC	AAC	AAA	CGG	AAA	AAG	ACA	TCG	CTC	TGT	CTC	ATG	ATG	ATG	TTA	CCA	96
Ile	Asn	Lys	Arg	Lys	Lys	Thr	Ser	Leu	Cys	Leu	Met	Met	Met	Leu	Pro	
			20					25					30			
GCA	ACA	CTT	GCT	TTC	CAC	TTA	ACT	TCA	CGA	GAT	GGA	GAG	CCG	CGC	ATG	144
Ala	Thr	Leu	Ala	Phe	His	Leu	Thr	Ser	Arg	Asp	Gly	Glu	Pro	Arg	Met	
			35				40					45				
ATT	GTG	GGG	AAG	AAT	GAA	AGA	GGA	AAA	TCC	CTA	CTT	TTC	AAG	ACA	GCC	192
Ile	Val	Gly	Lys	Asn	Glu	Arg	Gly	Lys	Ser	Leu	Leu	Phe	Lys	Thr	Ala	
	50					55					60					
TCT	GGA	ATC	AAC	ATG	TGC	ACA	CTC	ATA	GCT	ATG	GAT	CTG	GGA	GAG	ATG	240
Ser	Gly	Ile	Asn	Met	Cys	Thr	Leu	Ile	Ala	Met	Asp	Leu	Gly	Glu	Met	
65					70					75					80	

TGT Cys	GAT Asp	GAC Asp	ACG Thr	GTC Val 85	ACT Thr	TAC Tyr	AAA Lys	TGC Cys	CCC Pro 90	CAC His	ATT Ile	ACC Thr	GAA Glu	GTG Val 95	GAG Glu	288
CCT Pro	GAA Glu	GAC Asp	ATT Ile 100	GAC Asp	TGC Cys	TGG Trp	TGC Cys	AAC Asn 105	CTT Leu	ACA Thr	TCG Ser	ACA Thr	TGG Trp 110	GTG Val	ACT Thr	336
TAT Tyr	GGA Gly	ACA Thr 115	TGC Cys	AAT Asn	CAA Gln	GCT Ala	GGA Gly 120	GAG Glu	CAT His	AGA Arg	CGC Arg	GAT Asp 125	AAG Lys	AGA Arg	TCA Ser	384
GTG Val	GCG Ala 130	TTA Leu	GCT Ala	CCC Pro	CAT His	GTT Val 135	GGC Gly	ATG Met	GGA Gly	CTG Leu	GAC Asp 140	ACA Thr	CGC Arg	ACT Thr	CAA Gln	432
ACC Thr 145	TGG Trp	ATG Met	TCG Ser	GCT Ala 150	GAA Glu	GGA Gly	GCT Ala	TGG Trp	AGA Arg	CAA Gln 155	GTC Val	GAG Glu	AAG Lys	GTA Val	GAG Glu 160	480
ACA Thr	TGG Trp	GCC Ala	CTT Leu 165	AGG Arg	CAC His	CCA Pro	GGG Gly	TTT Phe 170	ACC Thr	ATA Ile	CTA Leu	GCC Ala	CTA Leu	TTT Phe 175	CTT Leu	528
GCC Ala	CAT His	TAC Tyr	ATA Ile 180	GGC Gly	ACT Thr	TCC Ser	TTG Leu	ACC Thr 185	CAG Gln	AAA Lys	GTG Val	GTT Val	ATT Ile 190	TTT Phe	ATA Ile	576
CTA Leu	TTA Leu	ATG Met 195	CTG Leu	GTT Val	ACC Thr	CCA Pro	TCC Ser 200	ATG Met	ACA Thr	ATG Met	AGA Arg	TGT Cys 205	GTA Val	GGA Gly	GTA Val	624
GGA Gly	AAC Asn 210	AGA Arg	GAT Asp	TTT Phe	GTG Val	GAA Glu 215	GGC Gly	CTA Leu	TCG Ser	GGA Gly	GCT Ala 220	ACG Thr	TGG Trp	GTT Val	GAC Asp	672
GTG Val 225	GTG Val	CTC Leu	GAG Glu	CAC His	GGT Gly 230	GGG Gly	TGT Cys	GTG Val	ACT Thr 235	ACC Met	ATG Ala	GCT Ala	AAG Lys	AAC Asn	AAG Lys 240	720
CCC Pro	ACG Thr	CTG Leu	GAC Asp 245	ATA Ile	GAG Glu	CTT Leu	CAG Gln	AAG Lys	ACC Thr 250	GAG Glu	GCC Ala	ACC Thr	CAA Gln	CTG Leu 255	GCG Ala	768
ACC Thr	CTA Leu	AGG Arg	AAG Lys 260	CTA Leu	TGC Cys	ATT Ile	GAG Glu	GGA Gly 265	AAA Lys	ATT Ile	ACC Thr	AAC Asn	ATA Ile 270	ACA Thr	ACC Thr	816
GAC Asp	TCA Ser	AGA Arg 275	TGT Cys	CCC Pro	ACC Thr	CAA Gln	GGG Gly 280	GAA Glu	GCG Ala	ATT Ile	TTA Leu	CCT Pro 285	GAG Glu	GAG Glu	CAG Gln	864
GAC Asp	CAG Gln 290	AAC Asn	TAC Tyr	GTG Val	TGT Cys	AAG Lys 295	CAT His	ACA Thr	TAC Tyr	GTG Val	GAC Asp 300	AGA Arg	GGC Gly	TGG Trp	GGA Gly	912
AAC Asn 305	GGT Gly	TGT Cys	GGT Gly	TTG Leu	TTT Phe 310	GGC Gly	AAG Lys	GGA Gly	AGC Ser	TTG Leu 315	GTG Val	ACA Thr	TGC Cys	GCG Ala	AAA Lys 320	960

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TTT CAA TGT TTA GAA TCA ATA GAG GGA AAA GTG GTG CAA CAT GAG AAC	1008
Phe Gln Cys Leu Glu Ser Ile Glu Gly Lys Val Val Gln His Glu Asn	
325 330 335	
CTC AAA TAC ACC GTC ATC ATC ACA GTG CAC ACA GGA GAC CAA CAC CAG	1056
Leu Lys Tyr Thr Val Ile Ile Thr Val His Thr Gly Asp Gln His Gln	
340 345 350	
GTG GGA AAT GAA ACG CAG GGA GTC ACG GCT GAG ATA ACA CCC CAG GCA	1104
Val Gly Asn Glu Thr Gln Gly Val Thr Ala Glu Ile Thr Pro Gln Ala	
355 360 365	
TCA ACC GCT GAA GCC ATT TTA CCT GAA TAT GGA ACC CTC GGG CTA GAA	1152
Ser Thr Ala Glu Ala Ile Leu Pro Glu Tyr Gly Thr Leu Gly Leu Glu	
370 375 380	
TGC TCA CCA CGG ACA GGT TTG GAT TTC AAT GAA ATG ATC TCA TTG ACA	1200
Cys Ser Pro Arg Thr Gly Leu Asp Phe Asn Glu Met Ile Ser Leu Thr	
385 390 395 400	
ATG AAG AAC AAA GCA TGG ATG GTA CAT AGA CAA TGG TTC TTT GAC TTA	1248
Met Lys Asn Lys Ala Trp Met Val His Arg Gln Trp Phe Phe Asp Leu	
405 410 415	
CCC CTA CCA TGG ACA TCA GGA GCT ACA GCA GAA ACA CCA ACT TGG AAC	1296
Pro Leu Pro Trp Thr Ser Gly Ala Thr Ala Glu Thr Pro Thr Trp Asn	
420 425 430	
AGG AAA GAG CTT CTT GTG ACA TTT AAA AAT GCA CAT GCA AAA AAG CAA	1344
Arg Lys Glu Leu Leu Val Thr Phe Lys Asn Ala His Ala Lys Lys Gln	
435 440 445	
GAA GTA GTT GTT CTT GGA TCA CAA GAG GGA GCA ATG CAT ACA GCA CTG	1392
Glu Val Val Val Leu Gly Ser Gln Glu Gly Ala Met His Thr Ala Leu	
450 455 460	
ACA GGA GCT ACA GAG ATC CAA ACC TCA GGA GGC ACA AGT ATC TTT GCG	1440
Thr Gly Ala Thr Glu Ile Gln Thr Ser Gly Gly Thr Ser Ile Phe Ala	
465 470 475 480	
GGG CAC TTA AAA TGT AGA CTC AAG ATG GAC AAA TTG GAA CTC AAA GGG	1488
Gly His Leu Lys Cys Arg Leu Lys Met Asp Lys Leu Glu Leu Lys Gly	
485 490 495	
ATG AGC TAT GCA ATG TGC TTG GGT AGC TTT GTG TTG AAG AAA GAA GTC	1536
Met Ser Tyr Ala Met Cys Leu Gly Ser Phe Val Leu Lys Lys Glu Val	
500 505 510	
TCC GAA ACG CAG CAT GGG ACA ATA CTC ATT AAG GTT GAG TAC AAA GGG	1584
Ser Glu Thr Gln His Gly Thr Ile Leu Ile Lys Val Glu Tyr Lys Gly	
515 520 525	
AAA GAT GCA CCC TGC AAG ATT CCT TTC TCC ACG GAG GAT GGA CAA GGA	1632
Lys Asp Ala Pro Cys Lys Ile Pro Phe Ser Thr Glu Asp Gly Gln Gly	
530 535 540	
AAA GCT CAC AAT GGC AGA CTG ATC ACA GCC AAT CCA GTG GTG ACC AAG	1680
Lys Ala His Asn Gly Arg Leu Ile Thr Ala Asn Pro Val Val Thr Lys	
545 550 555 560	

179

AAG GAG GAG CCT GTC AAC ATT GAG GCT GAA CCT CCT TTT GGA GAA AGT	1728
Lys Glu Glu Pro Val Asn Ile Glu Ala Glu Pro Pro Phe Gly Glu Ser	
565	570
575	
AAC ATA GTA ATT GGA ATT GGA GAC AAA GCC CTG AAA ATC AAC TGG TAC	1776
Asn Ile Val Ile Gly Ile Gly Asp Lys Ala Leu Lys Ile Asn Trp Tyr	
580	585
590	
AAG AAG GGA AGC TCG ATT GGG AAG ATG TTC GAG GCT ACT GCC AGA GGT	1824
Lys Lys Gly Ser Ser Ile Gly Lys Met Phe Glu Ala Thr Ala Arg Gly	
595	600
605	
GCA AGG CGC ATG GCC ATC TTG GGA GAC ACA GCC TGG GAC TTT GGA TCA	1872
Ala Arg Arg Met Ala Ile Leu Gly Asp Thr Ala Trp Asp Phe Gly Ser	
610	615
620	
GTG GGT GGT GTT TTG AAT TCA TTA GGG AAA ATG GTC CAC CAA ATA TTT	1920
Val Gly Gly Val Leu Asn Ser Leu Gly Lys Met Val His Gln Ile Phe	
625	630
635	640
GGG AGT GCT TAC ACA GCC CTA TTT GGT GGA GTC TCC TGG ATG ATG AAA	1968
Gly Ser Ala Tyr Thr Ala Leu Phe Gly Gly Val Ser Trp Met Met Lys	
645	650
655	
ATT GGA ATA GGT GTC CTC TTA ACC TGG ATA GGG TTG AAC TCA AAA AAT	2016
Ile Gly Ile Gly Val Leu Leu Thr Trp Ile Gly Leu Asn Ser Lys Asn	
660	665
670	
ACT TCT ATG TCA TTT TCA TGC ATC GCG ATA GGA ATC ATT ACA CTC TAT	2064
Thr Ser Met Ser Phe Ser Cys Ile Ala Ile Gly Ile Ile Thr Leu Tyr	
675	680
685	
CTG GGA GCC GTG GTG CAA GCT GAC ATG GGG TGT GTC ATA AAC TGG AAA	2112
Leu Gly Ala Val Val Gln Ala Asp Met Gly Cys Val Ile Asn Trp Lys	
690	695
700	
GGC AAA GAA CTC AAA TGT GGA AGT GGA ATT TTC	2145
Gly Lys Glu Leu Lys Cys Gly Ser Gly Ile Phe	
705	710
715	

(2) INFORMATION FOR SEQ ID NO:126:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2175 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(ix) FEATURE:

- (A) NAME/KEY: Coding Sequence
- (B) LOCATION: 1...2175
- (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:126:

ATT Ile 1	CTG Leu	AAG Lys	AGA Arg	TGG Trp 5	GGA Gly	CAG Gln	TTG Leu	AAG Lys	AAA Lys 10	AAT Asn	AAG Lys	GCC Ala	ATC Ile	AGG Arg 15	ATA Ile	48
CTG Leu	ATT Ile	GGA Gly	TTC Phe 20	AGG Arg	AAG Lys	GAG Glu	ATA Ile	GGC Gly 25	CGC Arg	ATG Met	CTG Leu	AAC Asn 30	ATC Ile	TTG Leu	AAC Asn	96
GGG Gly	AGA Arg	AAA Lys 35	AGG Arg	TCA Ser	ACG Thr	ATA Ile	ACA Thr 40	TTG Leu	CTG Leu	TGC Cys	TTG Leu	ATT Ile 45	CCC Pro	ACC Thr	GTA Val	144
ATG Met 50	GCG Ala	TTT Phe	CAC His	TTG Leu	TCA Ser	ACA Thr 55	AGA Arg	GAT Asp	GGC Gly	GAA Glu	CCC Pro 60	CTC Leu	ATG Met	ATA Ile	GTG Val	192
GCA Ala 65	AAA Lys	CAT His	GAA Glu	AGG Arg	GGG Gly 70	AGA Arg	CCT Pro	CTC Leu	TTG Leu	TTT Phe 75	AAG Lys	ACA Thr	ACA Thr	GAG Glu	GGG Gly 80	240
ATC Ile	AAC Asn	AAA Lys	TGC Cys	ACT Thr 85	CTC Leu	ATT Ile	GCC Ala	ATG Met	GAC Asp 90	TTG Leu	GGT Gly	GAA Glu	ATG Met	TGT Cys 95	GAG Glu	288
GAC Asp	ACT Thr	GTC Val	ACG Thr 100	TAT Tyr	AAA Lys	TGC Cys	CCC Pro	TTA Leu 105	CTG Leu	GTC Val	AAT Asn	ACC Thr	GAA Glu 110	CCT Pro	GAA Glu	336
GAC Asp	ATT Ile	GAT Asp 115	TGC Cys	TGG Trp	TGC Cys	AAT Asn 120	CTC Leu	ACG Thr	TCT Ser	ACC Thr	TGG Trp 125	GTC Val	ACA Thr	TAT Tyr	GGG Gly	384
ACA Thr 130	TAC Tyr	ACC Thr	CAG Gln	AGC Ser	GGA Gly	GAA Glu 135	CGG Arg	AGA Arg	CGA Arg	GAG Glu	AAG Lys 140	CGC Arg	TCA Ser	GTA Val	GCT Ala	432
TTA Leu 145	ACA Thr	CCA Pro	CAT His	TCA Ser	GGA Gly 150	ATG Met	GGA Gly	TTG Leu	GAA Glu	ACA Thr 155	AGA Arg	GCT Ala	GAG Glu	ACA Thr	TGG Trp 160	480
ATG Met	TCA Ser	TCG Ser	GAA Glu	GGG Gly 165	GCT Ala	TGG Trp	AAG Lys	CAT His	GCT Ala 170	CAG Gln	AGA Arg	GTA Val	GAG Glu	AGC Ser 175	TGG Trp	528
ATA Ile	CTC Leu	AGA Arg	AAC Asn 180	CCA Pro	GGA Gly	TTC Phe	GCG Ala	CTC Leu 185	TTG Leu	GCA Ala	GGA Gly	TTT Phe 190	ATG Met	GCT Ala	TAT Tyr	576
ATG Met	ATT Ile	GGG Gly 195	CAA Gln	ACA Thr	GGA Gly	ATC Ile	CAG Gln	CGA Arg 200	ACT Thr	GTC Val	TTC Phe	TTT Phe 205	GTC Val	CTA Leu	ATG Met	624
ATG Met 210	CTG Leu	GTC Val	GCC Ala	CCA Pro	TCC Ser	TAC Tyr 215	GGA Gly	ATG Met	CGA Arg	TGC Cys	GTA Val 220	GGA Gly	GTA Val	GGA Gly	AAC Asn	672

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AGA Arg 225	GAC Asp	TTT Phe	GTG Val	GAA Glu	GGA Gly 230	GTC Val	TCA Ser	GGT Gly	GGA Gly	GCA Ala 235	TGG Trp	GTC Val	GAT Asp	CTG Leu	GTG Val 240	720
CTA Leu	GAA Glu	CAT His	GGA Gly 245	GGA Glu 245	TGC Cys	GTC Val	ACA Thr	ACC Thr	ATG Met 250	GCC Ala	CAG Gln	GGA Gly	AAA Lys	CCA Pro 255	ACC Thr	768
TTG Leu	GAT Asp	TTT Phe	GAA Glu 260	CTG Leu	ACT Thr	AAG Lys	ACA Thr	ACA Thr 265	GCC Ala	AAG Lys	GAA Glu	GTG Val	GCT Ala 270	CTG Leu	TTA Leu	816
AGA Arg	ACC Thr	TAT Tyr 275	TGC Cys	ATT Ile	GAA Glu	GCC Ala	TCA Ser 280	ATA Ile	TCA Ser	AAC Asn	ATA Ile	ACC Thr 285	ACG Thr	GCA Ala	ACA Thr	864
AGA Arg	TGT Cys 290	CCA Pro	ACG Thr	CAA Gln	GGA Gly 295	GAG Glu 295	CCT Pro	TAT Tyr	CTA Leu	AAA Lys	GAG Glu 300	GAA Glu	CAA Gln	GAC Asp	CAA Gln	912
CAG Gln 305	TAC Tyr	ATT Ile	TGC Cys	CGG Arg 310	AGA Arg 310	GAT Asp	GTG Val	GTA Val	GAC Asp 315	AGA Arg 315	GGG Gly	TGG Trp	GGC Gly	AAT Asn	GGC Gly 320	960
TGT Cys	GGC Gly	TTG Leu	TTT Phe	GGA Gly 325	AAA Lys	GGA Gly	GGA Gly	GTT Val	GTG Val 330	ACA Thr	TGT Cys	GCG Ala	AAG Lys	TTT Phe 335	TCA Ser	1008
TGT Cys	TCG Ser	GGG Gly	AAG Lys 340	ATA Ile	ACA Thr	GGC Gly	AAT Asn 345	TTG Leu	GTC Val	CAA Gln	ATT Ile	GAG Glu 350	AAC Asn	CTT Leu	GAA Glu	1056
TAC Tyr	ACA Thr	GTG Val 355	GTT Val	GTA Val	ACA Thr	GTC Val	CAC His 360	AAT Asn	GGA Gly	GAC Asp	ACC Thr 365	CAT His	GCA Ala	GTA Val	GGA Gly	1104
AAT Asn 370	GAC Asp	ACA Thr	TCC Ser	AAT Asn	CAT His	GGA Gly 375	GTT Val	ACA Thr	GCC Ala	ACG Thr	ATA Ile 380	ACT Thr	CCC Pro	AGG Arg	TCA Ser	1152
CCA Pro 385	TCG Ser	GTG Val	GAA Glu	GTC Val	AAA Lys 390	TTG Leu	CCG Pro	GAC Asp	TAT Tyr	GGA Gly 395	GAA Glu	CTA Leu	ACA Thr	CTC Leu	GAT Asp 400	1200
TGT Cys	GAA Glu	CCC Pro	AGG Arg	TCT Ser 405	GGA Gly	ATT Ile	GAC Asp	TTT Phe 410	AAT Asn 410	GAG Glu	ATG Met	ATT Ile	CTG Leu	ATG Met 415	AAA Lys	1248
ATG Met	AAA Lys	AAG Lys	AAA Lys 420	ACA Thr	TGG Trp	CTT Leu	GTG Val	CAT His 425	AAG Lys	CAA Gln	TGG Trp	TTT Phe 430	TTG Leu	GAT Asp	CTA Leu	1296
CCT Pro	CTA Leu	CCA Pro 435	TGG Trp	ACA Thr	GCA Ala	GGA Gly 440	GCA Ala	GAC Asp	ACA Thr	TCA Ser	GAG Glu	GTT Val 445	CAC His	TGG Trp	AAT Asn	1344
TAC Tyr	AAA Lys 450	GAG Glu	AGA Arg	ATG Met	GTG Val	ACA Thr 455	TTT Phe	AAG Lys	GTT Val	CCT Pro	CAT His 460	GCC Ala	AAG Lys	AGA Arg	CAG Gln	1392

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GAT Asp 465	GTG Val	ACA Thr	GTG Val	CTG Leu	GGA Gly 470	TCT Ser	CAG Gln	GAA Glu	GGA Gly 475	GCC Ala 475	ATG Met	CAT His	TCT Ser	GCC Ala 480	CTC Leu	1440
GCT Ala	GGA Gly	GCC Ala	ACA Thr	GAA Glu 485	GTG Val	GAC Asp	TCC Ser	GGT Gly	GAT Asp 490	GGA Gly	AAT Asn	CAC His	ATG Met	TTT Phe 495	GCA Ala	1488
GGA Gly	CAT His	CTC Leu	AAG Lys 500	TGC Cys	AAA Lys	GTC Val	CGT Arg	ATG Met 505	GAG Glu	AAA Lys	TTG Leu	AGA Arg	ATC Ile 510	AAG Lys	GGA Gly	1536
ATG Met	TCA Ser	TAC Tyr 515	ACG Thr	ATG Met	TGT Cys	TCA Ser	GGA Gly 520	AAG Lys	TTC Phe	TCA Ser	ATT Ile	GAC Asp 525	AAA Lys	GAG Glu	ATG Met	1584
GCA Ala	GAA Glu 530	ACA Thr	CAG Gln	CAT His	GGG Gly 535	ACA Thr	ACA Thr	GTG Val	GTG Val	AAA Lys 540	GTC Val	AAG Lys	TAT Tyr	GAA Glu	GGT Gly	1632
GCT Ala 545	GGA Gly	GCT Ala	CCG Pro	TGT Cys	AAA Lys 550	GTC Val	CCC Pro	ATA Ile	GAG Glu	ATA Ile 555	AGA Arg	GAT Asp	GTG Val	AAC Asn	AAG Lys 560	1680
AAA Lys	AAA Lys	GTG Val	GTT Val	GGG Gly 565	CGT Arg	ATC Ile	ATC Ile	TCA Ser	TCC Ser	ACC Thr 570	CCT Pro	TTG Leu	GCT Ala	GAG Glu 575	AAT Asn	1728
ACC Thr	AAC Asn	AGT Ser	GCA Ala 580	ACC Thr	AAC Asn	ATA Ile	GAG Glu	TTA Leu 585	GAA Glu	CCC Pro	CCC Pro	TTT Phe	GGG Gly 590	GAC Asp	AGC Ser	1776
TAC Tyr	ATA Ile	GTG Val 595	ATA Ile	GGT Gly	GTT Val	GGA Gly	AAC Asn 600	AGT Ser	GCA Ala	TTA Leu	ACA Thr	CTC Leu 605	CAT His	TGG Trp	TTC Phe	1824
AGG Arg 610	AAA Lys	GGG Gly	AGT Ser	TCC Ser	ATT Ile	GGC Gly 615	AAG Lys	ATG Met	TTT Phe	GAG Glu	TCC Ser 620	ACA Thr	TAC Tyr	AGA Arg	GGT Gly	1872
GCA Ala 625	AAA Lys	CGA Arg	ATG Met	GCC Ala	ATT Ile 630	CTA Leu	GGT Gly	GAA Glu	ACA Thr	GCT Ala 635	TGG Trp	GAT Asp	TTT Phe	GGT Gly 640	TCC Ser	1920
GTT Val	GGT Gly	GGA Gly	CTG Leu	TTC Phe 645	ACA Thr	TCA Ser	TTG Leu	GGA Gly	AAG Lys 650	GCT Ala	GTG Val	CAC His	CAG Gln	GTT Val 655	TTT Phe	1968
GGA Gly	AGT Ser	GTG Val	TAT Tyr 660	ACA Thr	ACC Thr	ATG Met	TTT Phe	GGA Gly 665	GGA Gly	GTC Val	TCA Ser	TGG Trp	ATG Met 670	ATT Ile	AGA Arg	2016
ATC Ile	CTA Leu	ATT Ile 675	GGG Gly	TTC Phe	CTA Leu	GTG Val	TTG Leu 680	TGG Trp	ATT Ile	GGC Gly	ACG Thr	AAC Asn	TCA Ser	AGG Arg	AAC Asn	2064
ACT Thr	TCA Ser	ATG Met	GCT Ala	ATG Met	ACG Thr	TGC Cys 695	ATA Ile	GCT Ala	GTT Val	GGA Gly	GGA Gly 700	ATC Ile	ACT Thr	CTG Leu	TTT Phe	2112

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CTG GGC TTC ACA GTT CAA GCA GAG ATG GGT TGT GTG GTG TCA TGG AGT 2160
 Leu Gly Phe Thr Val Gln Ala Glu Met Gly Cys Val Val Ser Trp Ser
 705 710 715 720

GGG AAA GAA TTG AGG 2175
 Gly Lys Glu Leu Arg
 725

(2) INFORMATION FOR SEQ ID NO:127:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 40 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:127:

CACTACCGCA AGGTAGAGAG CTCGGCATTG CCTCTTGGTG 40

(2) INFORMATION FOR SEQ ID NO:128:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 40 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:128:

GTGATGGCGT TCCATCTCTC GAGCCGTAAC GGAGAACCAC 40

(2) INFORMATION FOR SEQ ID NO:129:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 40 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

184

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:129:

GCCCTGGCGT TCCATCTCTC GAGCCGAGGG GGAGAGCCGC 40

(2) INFORMATION FOR SEQ ID NO:130:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 40 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:130:

ACACTTGCTT TCCACCTCTC GAGCCGAGAT GGAGAGCCGC 40

(2) INFORMATION FOR SEQ ID NO:131:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 40 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:131:

GTAATGGCGT TTCACCTCTC GAGCAGAGAT GGCGAACCCC 40

(2) INFORMATION FOR SEQ ID NO:132:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 36 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:132:

CCTATCCTTA CTTAAGATCT TCGTGGAGTG ACAGAC 36

185

(2) INFORMATION FOR SEQ ID NO:133:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 36 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:133:

GGATAGGAAT GAATTCTAGA AGCACCTCAC TGTCTG

36

(2) INFORMATION FOR SEQ ID NO:134:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 33 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:134:

CCGCAGAGAT CGTTTTCTG CCTGCATGAT TCC

33

(2) INFORMATION FOR SEQ ID NO:135:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 36 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:135:

CCGATCCTAA TTTAAGATCT TTGTGCAGGG AAAGCC

36

(2) INFORMATION FOR SEQ ID NO:136:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 36 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

186

- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:136:

CCTATCCCAA CTTGAGATCT TTATGAAGAT ACAGTA

36

(2) INFORMATION FOR SEQ ID NO:137:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 36 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:137:

CCTAACCGTG CTTGAGATCT TTGTGAAGTT ACCGAC

36

WHAT IS CLAIMED IS:

1. A quadravalent vaccine providing immunity against all four serotypes of dengue virus comprising a DEN-2 PDK-53 infectious clone-derived virus.
- 5 2. A quadravalent vaccine providing immunity against all four serotypes of dengue virus comprising a chimeric DEN-2/1 virus.
3. A quadravalent vaccine providing immunity against all four serotypes of dengue virus comprising a
10 chimeric DEN-2/3 virus.
4. A quadravalent vaccine providing immunity against all four serotypes of dengue virus comprising a chimeric DEN-2/4 virus.
5. A quadravalent vaccine providing immunity
15 against all four serotypes of dengue virus comprising DEN-2 PDK-53 infectious clone-derived and chimeric DEN-2/1, DEN-2/3, and DEN-2/4 viruses.
6. A method of immunization in which a desired immune response is produced against all four serotypes of
20 dengue virus comprising the step of administering to a subject a quadravalent vaccine comprising DEN-2 PDK-53 infectious clone-derived and chimeric DEN-2/1, DEN-2/3, and DEN-2/4 viruses.
7. A composition of matter comprising a full
25 genome-length infectious cDNA clone for a DEN-2 virus, strain 16681.
8. A composition of matter comprising a full genome-length infectious cDNA clone for a DEN-2 virus of a

strain characterized as replicating to high titer in cell culture.

9. A composition of matter comprising a full genome-length infectious cDNA clone for a DEN-2 virus, strain 16681, having the identifying characteristics of ATCC 69826.

10. A composition of matter comprising a full genome-length infectious cDNA clone for a DEN-2 virus, strain 16681, attenuated derivative, PDK-53.

11. A composition of matter comprising a full genome-length infectious cDNA clone for a DEN-2 virus attenuated derivative, characterized as replicating to high titer in cell culture.

12. A composition of matter comprising a full genome-length infectious cDNA clone for a DEN-2 virus, strain 16681, attenuated derivative, PDK-53, having the identifying characteristics of ATCC 69825.

13. A composition of matter comprising a full genome-length infectious cDNA clone of a chimeric DEN-2/1 virus, wherein said virus is characterized as the expressing prM and E genes of a DEN-1 attenuated virus in the context of the nonstructural genes of the DEN-2 PDK-53 virus.

14. The composition of matter of Claim 13, wherein said DEN-1 attenuated virus is DEN-1 PDK-13.

15. A composition of matter comprising a full genome-length infectious cDNA clone of a chimeric DEN-2 virus, wherein said virus is characterized as expressing the antigenicity of a DEN-1 attenuated virus.

16. A composition of matter comprising a full genome-length infectious cDNA clone of a chimeric DEN-2/3 virus, wherein said virus is characterized as expressing the prM and E genes of a DEN-3 attenuated virus in the context of the nonstructural genes of the DEN-2 PDK-53 virus.

17. The composition of matter of Claim 16, wherein said DEN-3 attenuated virus is DEN-3 PGMK30/FRhL-3.

18. A composition of matter comprising a full genome-length infectious cDNA clone of a chimeric DEN-2 virus, wherein said virus is characterized as expressing the antigenicity of a DEN-3 attenuated virus.

19. A composition of matter comprising a full genome-length infectious cDNA clone of a chimeric DEN-2/4 virus, wherein said virus is characterized as expressing the prM and E genes of a DEN-4 attenuated virus in the context of the nonstructural genes of the DEN-2 PDK-53 virus.

20. The composition of matter of Claim 19, wherein said DEN-4 attenuated virus is DEN-4 PDK-48.

21. A composition of matter comprising a full genome-length infectious cDNA clone of a chimeric DEN-2 virus, wherein said virus is characterized as expressing the antigenicity of a DEN-4 attenuated virus.

22. A genetic construct comprising a DNA sequence operably encoding the polyprotein of DEN-2 virus, strain 16681.

23. The genetic construct of Claim 22, wherein said polyprotein is the polyprotein encoded by the nucleotide sequence of SEQ ID NO:1.

24. A genetic construct comprising a DNA sequence operably encoding at least one protein of DEN-2 virus, strain 16681.

25. The genetic construct of Claim 24, wherein said
5 protein is a protein encoded by the nucleotide sequence of SEQ ID NO: 1.

26. A genetic construct comprising a DNA sequence operably encoding the polyprotein of DEN-2 virus, strain 16681, attenuated derivative, PDK-53.

10 27. The genetic construct of Claim 26, wherein said polyprotein is the polyprotein encoded by the nucleotide sequence of SEQ ID NO:2.

28. A genetic construct comprising a DNA sequence operably encoding at least one protein of DEN-2 virus,
15 strain 16681, attenuated derivative, PDK-53.

29. The genetic construct of Claim 28, wherein said protein is a protein encoded by the nucleotide sequence of SEQ ID NO: 2.

30. A genetic construct comprising a DNA sequence
20 operably encoding at least one structural protein of DEN-1 PDK-13.

31. The genetic construct of Claim 30, wherein said structural protein is a structural protein encoded by the nucleotide sequence of SEQ ID NO: 124.

25 32. A genetic construct comprising a DNA sequence operably encoding at least one structural protein of DEN-3 PGMK30/FRhL-3.

33. The genetic construct of Claim 32, wherein said structural protein is a structural protein encoded by the
30 nucleotide sequence of SEQ ID NO: 125.

34. A genetic construct comprising a DNA sequence operably encoding at least one structural protein of DEN-4 PDK-48.

35. The genetic construct of Claim 34, wherein said
5 structural protein is a structural protein encoded by the nucleotide sequence of SEQ ID NO: 126.

36. A host cell comprising the genetic construct of any of Claims 22-35.

Construction of DEN-2 Infectious cDNA Clone

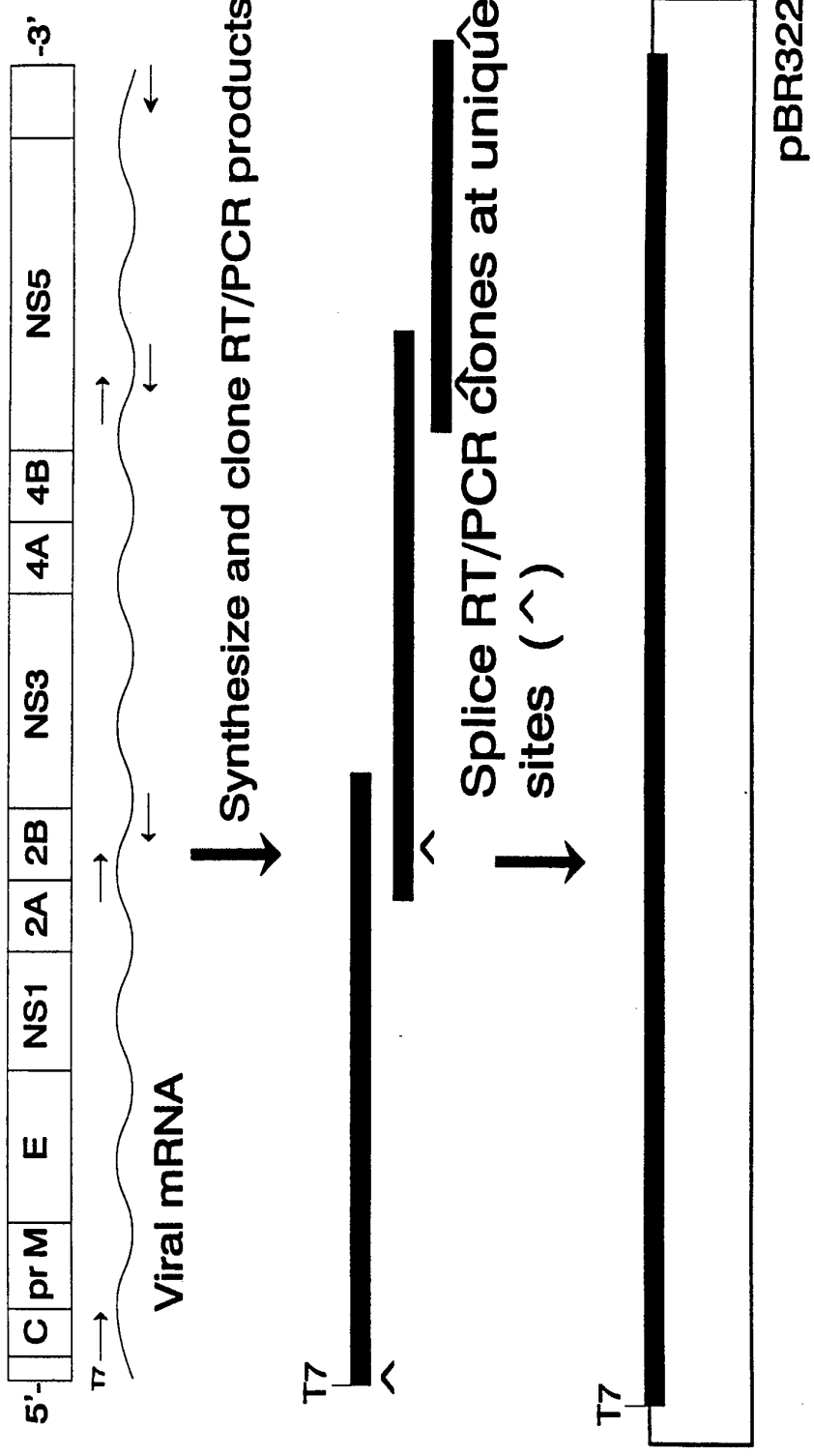
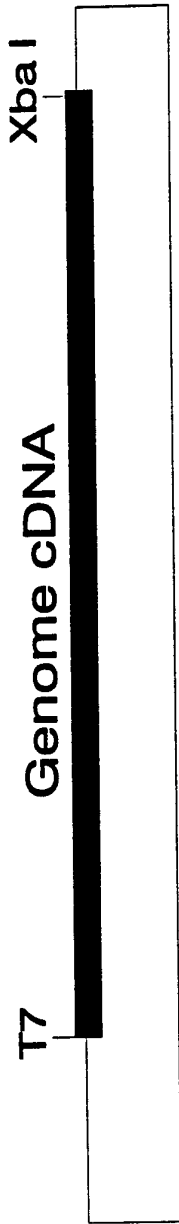


FIGURE 1

Transcription of DEN-2 RNA from Infectious Clone



pBR322

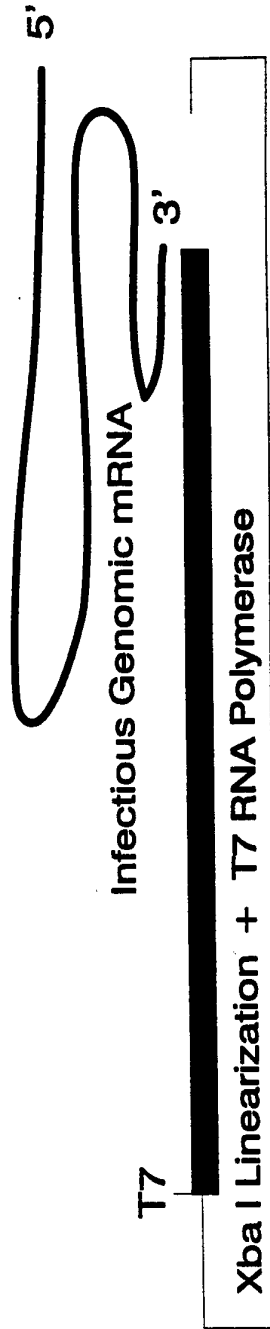
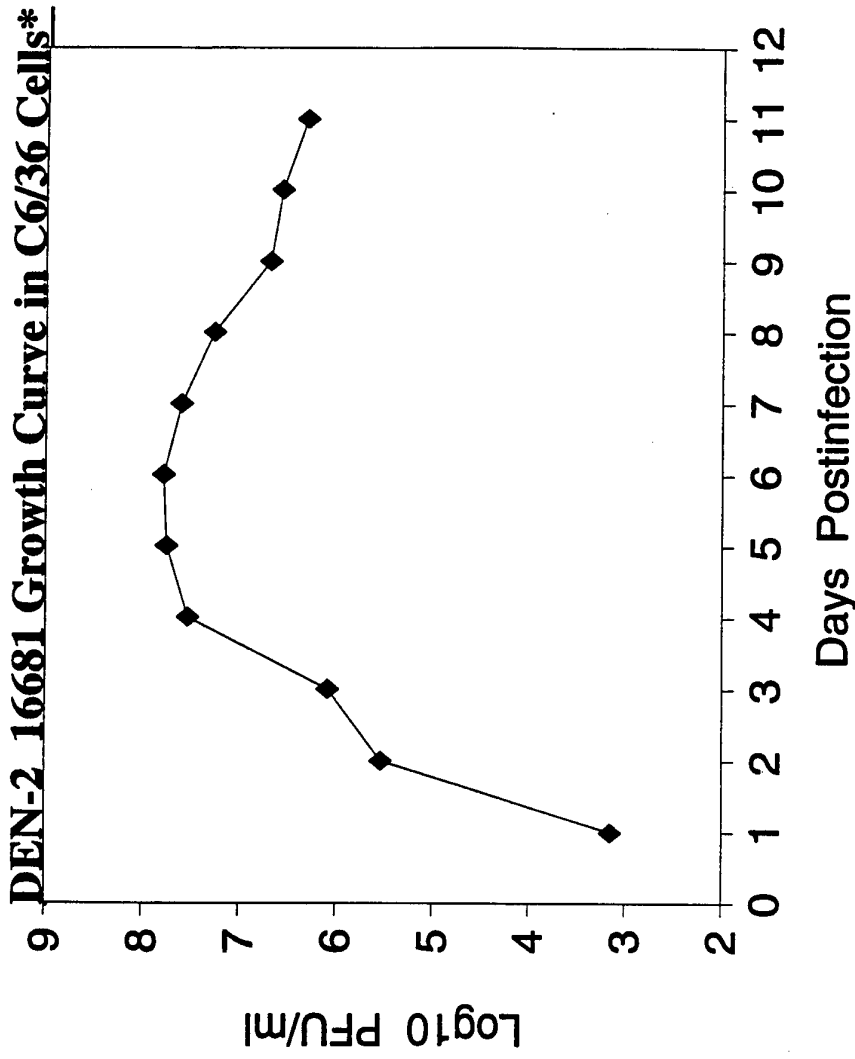


FIGURE 2

C	M	E	NS1	2A	2B	NS3	4A	4B	NS5	3'		
			1	2	3	4	5	6	7	8	9	0
PmlI	208	398	1531									
PstI		2244										
XcmI						5426						
XhoI												
MunI		2003	3907									
AseI	328			5175								
AvaI		3551		5426	6892							
BsaHI			3658	4732				8566				
BanI			3658	4493	5761				9382			
BglII	88	394								6858	6915	7646

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FIGURE 3B



* M.O.I. = 0.004, 150cm², 40 ml

FIGURE 4

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FIGURE 5A



HMC ↓

FIGURE 5B

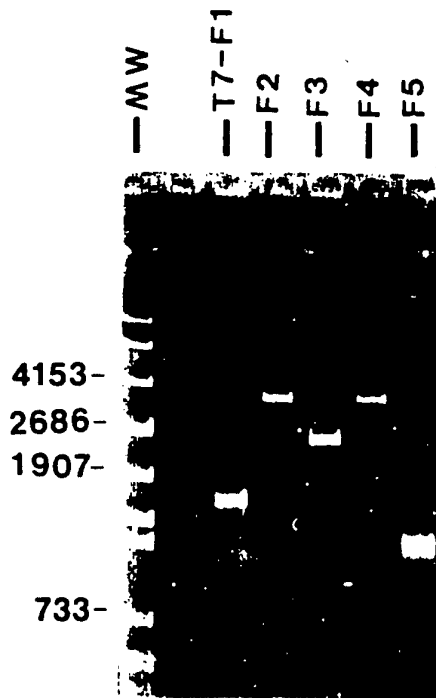
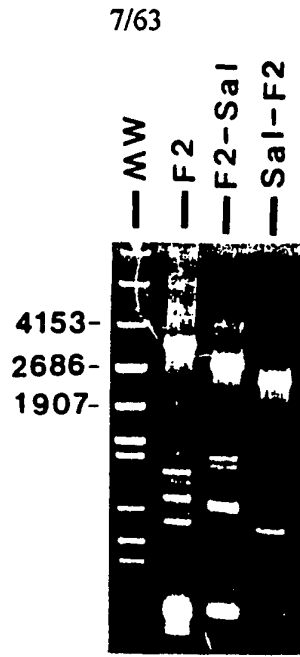
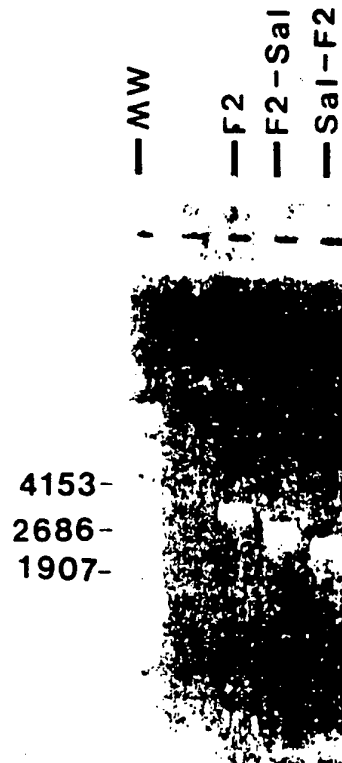


FIGURE 5C



HMC

FIGURE 5D



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B	<u>RT/PCR Amplicon</u>	<u>Expected Amplicon Length</u>	<u>Up-Amplimer</u>	<u>Down-Amplimer</u>
	T7-F1	1552-bp	D2-SMT71	cD2-1503
	F2	3327-bp	D2-1261	cD2-4557
	F2-Sal	2742-bp	D2-1261	cD2-4002
	Sal-F2	2388-bp	D2-2170	cD2-4557
	F3	2368-bp	D2-4257	cD2-6624
	F4	3304-bp	D2-6493	cD2-9796
	F5	1032-bp	D2-9656	cD2-10687.Xba

FIGURE 5E

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Plasmid Eco RI Digests

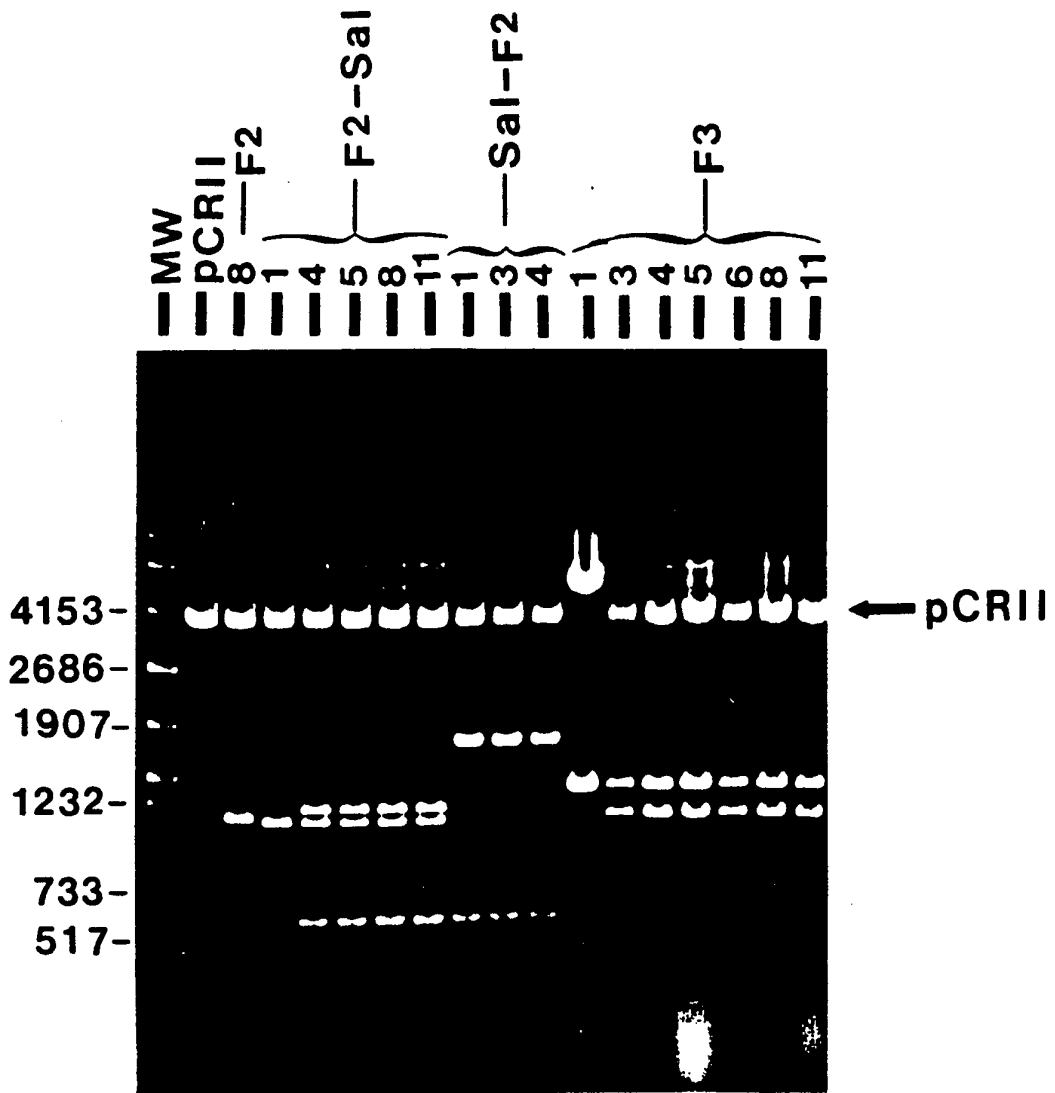


FIGURE 6

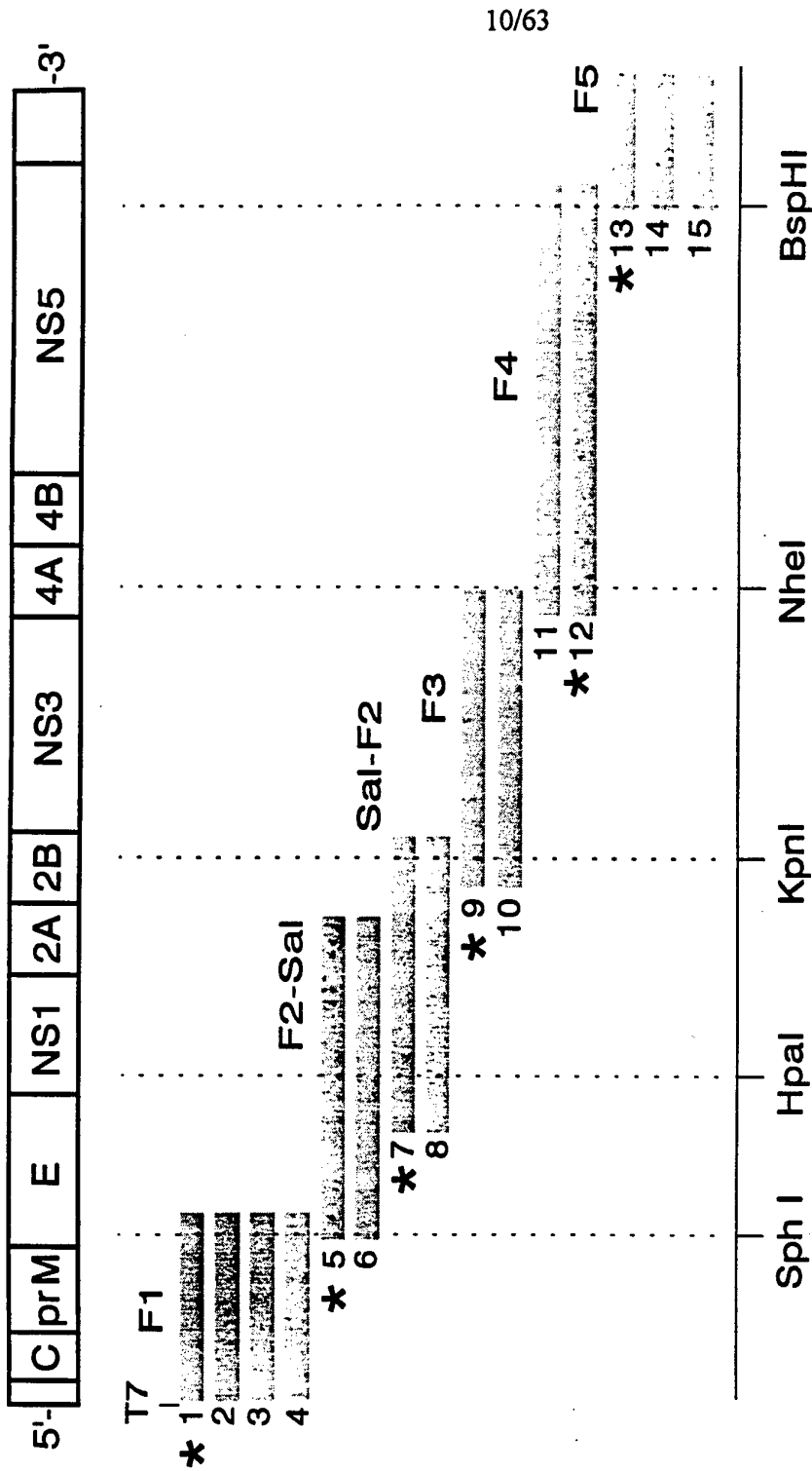
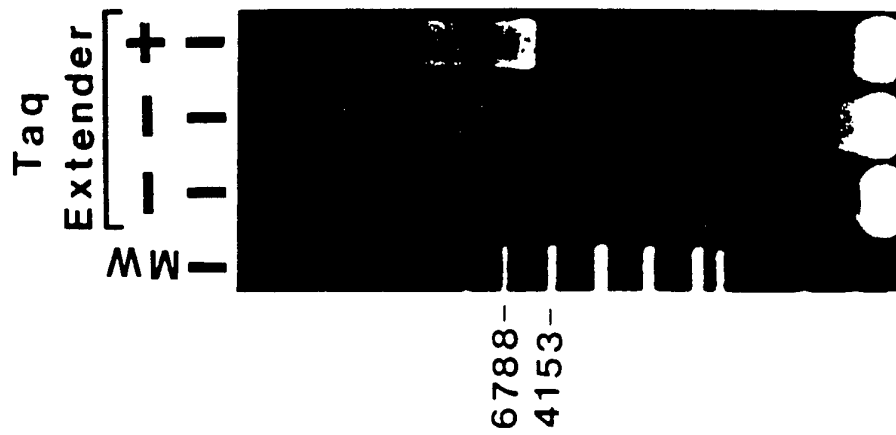
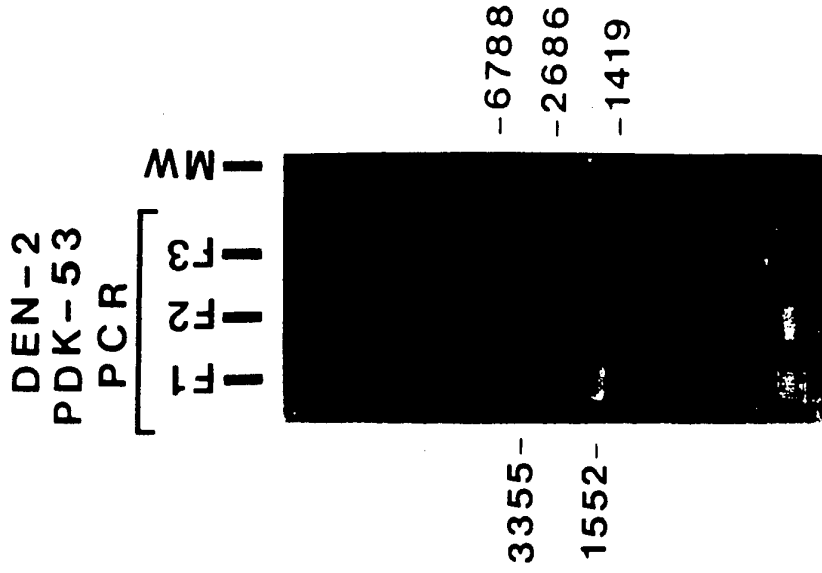


FIGURE 7

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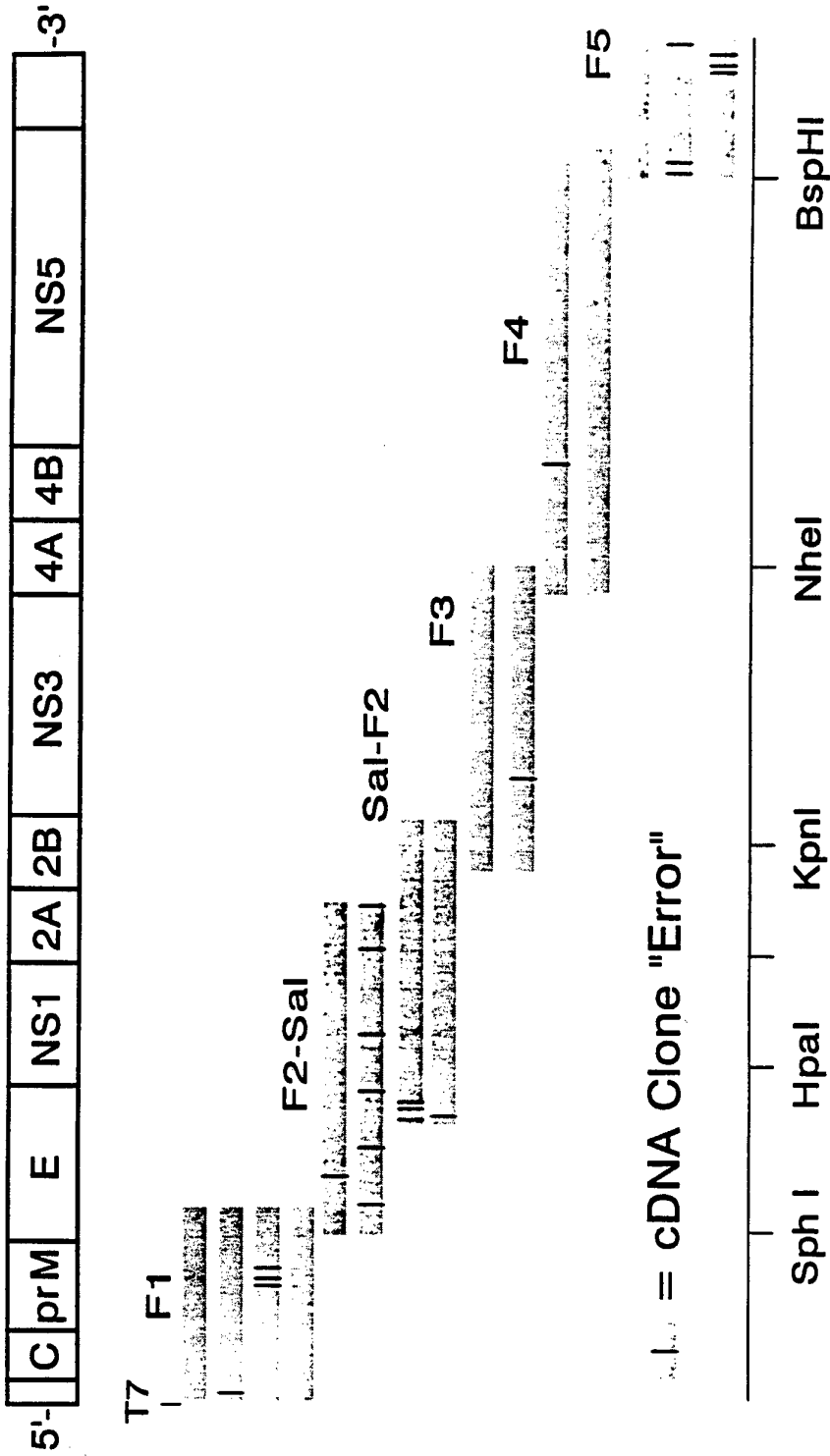


FIGURE 9

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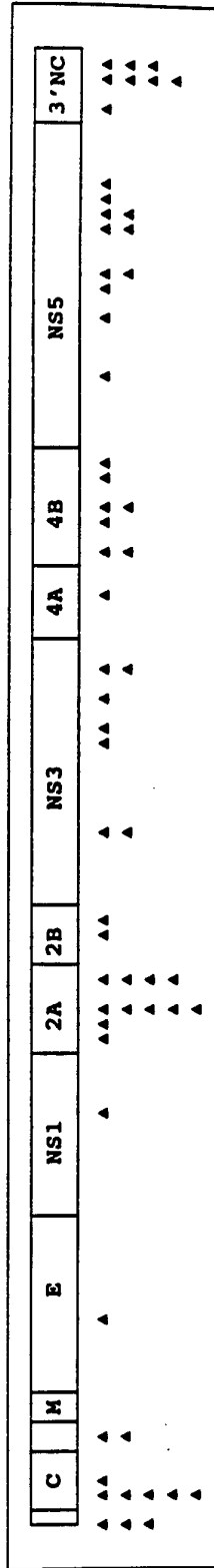


FIGURE 10

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	<u>5'-NC</u>	<u>Capsid</u>	<u>prM</u>	<u>E</u>	<u>NS1</u>	<u>NS2A</u>
		1 1 2 2 2 3	6 7	4	1 3	3 3 3 3 3 3 3
	4 6 7	0 3 1 2 3 2	5 0	4	0 8	6 6 7 8 8 8 8 9
	9 9 7	3 2 0 6 6 1	1 3	5	1	1 4 7 0 0 2 2 8
DEN-2-16681.RK	A A A	A T G T T A	G G	T	T	C G A C T C T A
DEN-2-16681.BLOK	G T G	G A A C A C	C C	C	C	T A C T C T C G
		■ ■ ■ ■ ■	■ ■ ■ ■ ■	■ ■ ■ ■ ■	■ ■ ■ ■ ■	■ ■ ■ ■ ■
	<u>NS2A</u>	<u>NS2B</u>	<u>NS3</u>	<u>NS4A</u>	<u>NS4B</u>	
	4 4 4	4 4	5 5 5 5 6 6	6	6 6 7 7 7 7 7	
	0 0 0	2 3	0 0 6 7 9 1 1	5	8 8 1 1 2 4 5	
	1 4 6	4 4	3 7 4 0 5 1 1	8	6 7 3 9 1 0 6	
	8 4 2	1 4	4 6 9 5 5 3 4	0	4 0 9 6 3 3 1	
DEN-2-16681.RK	C A A	T T	C G G T T A A	T	C G T G A C A	
DEN-2-16681.BLOK	T G G	C C	T A C C C C C	C	A A C C T T G	
	■ ■ ■	■ ■ ■	■ ■ ■ ■ ■ ■ ■	■ ■ ■	■ ■ ■ ■ ■ ■ ■	
	<u>NS5</u>			<u>3'-NC</u>		
	8 8 9 9 9 9 9 9 9 9			1 1 1 1 1 1 1 1		
	5 9 2 2 3 5 5 6 6 7 8			0 0 0 0 0 0 0 0		
	4 3 0 2 0 9 9 2 2 3 4			3 6 6 6 6 6 6 6		
	2 1 7 1 3 1 2 0 1 2 5			3 3 4 5 5 5 5 7		
				0 7 6 1 2 5 9 3		
DEN-2-16681.RK	G G C A G C G T G T C			G A G T G G C C		
DEN-2-16681.BLOK	T C T C C G C C A C A			C C A A A A A T		
	■ ■ ■ ■ ■ ■ ■ ■ ■ ■			■ ■ ■ ■ ■ ■ ■ ■ ■ ■		

FIGURE 11

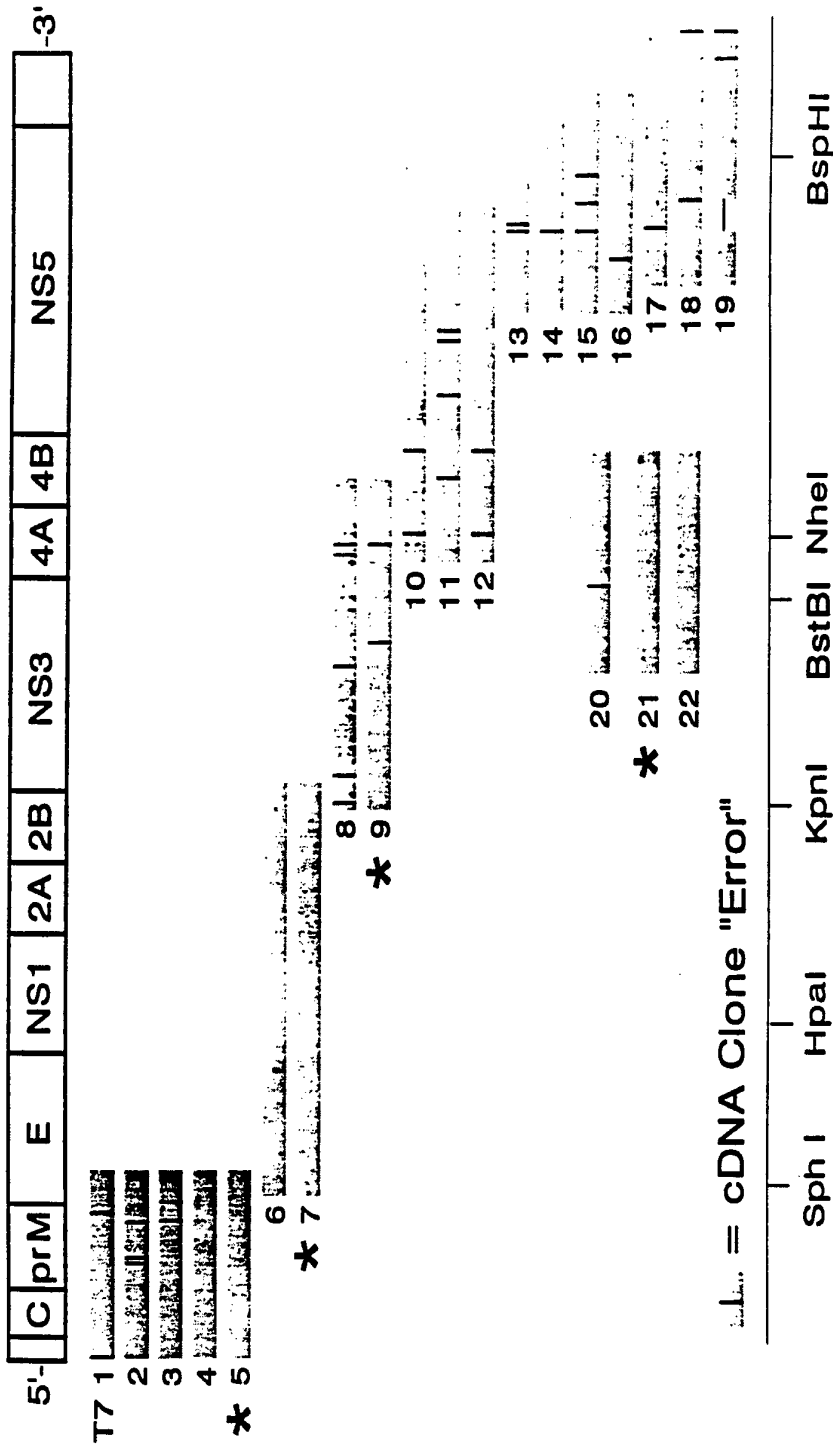


FIGURE 12

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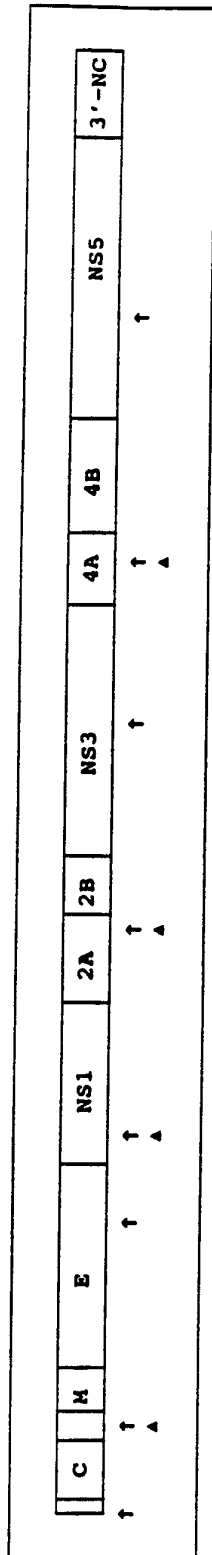


FIGURE 13

EcoRI MluI T7 Promoter
 Amplimer D2-SMT71 = CCAGTGAATTCGAGCTCAGCGTAAATTTAATACGACTCACTATA
 SstI

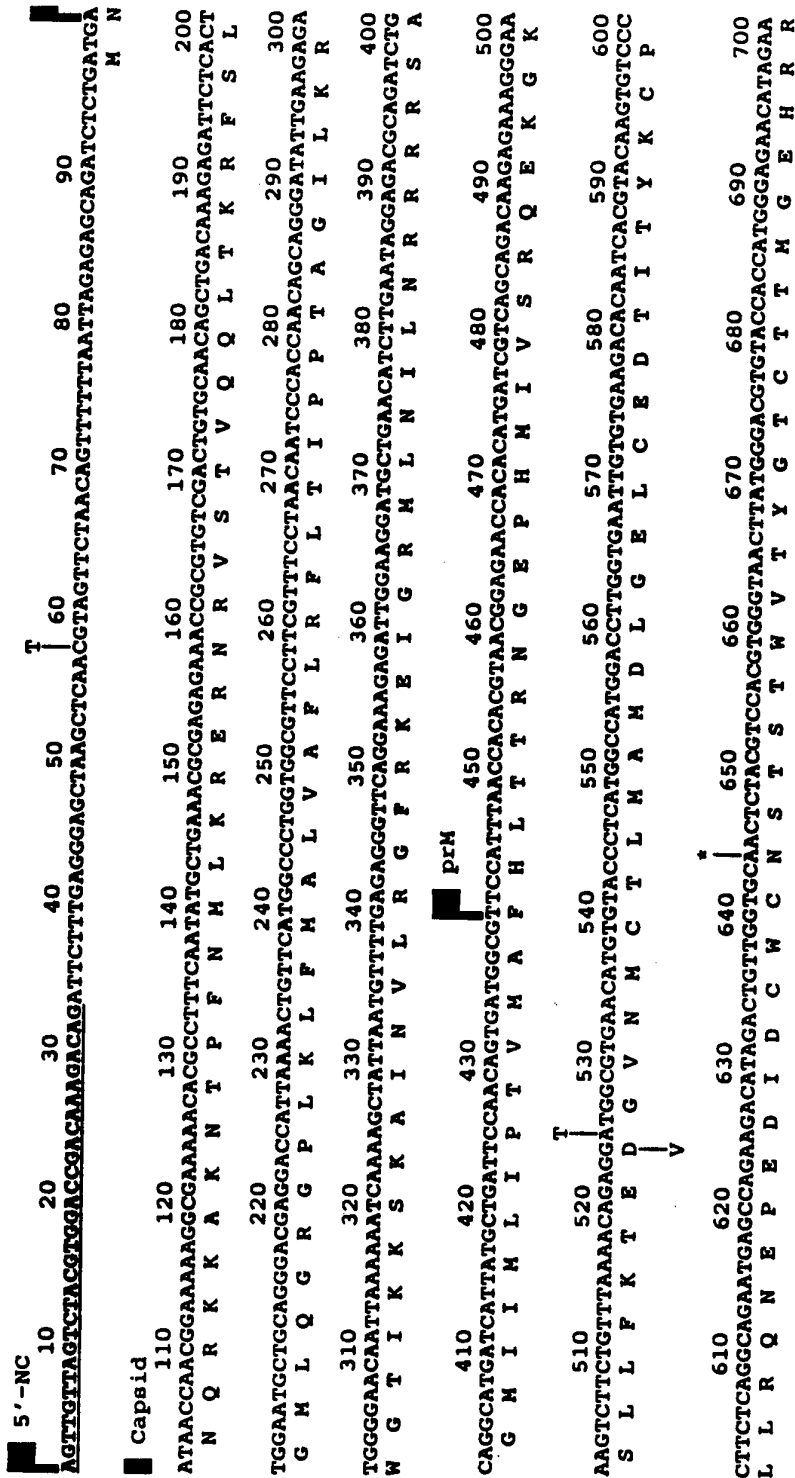


FIGURE 14A

M
 710 [REDACTED] 720 730 740 750 760 770 780 790 800
 GAGAAAAGATCAGTGGCACTCGTCCACATGTGGGAATGGGACTGGAGACACCAACTGAAACATGGATGTCATCAGAAAGGGCCCTGGAACATGTCCA
 E K R S V A L V P H V G H G L E T R T E T W M S S E G A W K H V Q
 810 820 830 840 850 860 870 880 890 900
 GAGAAATGGAACTTGGATCTGAGACATCCAGGCTTCCACCATGATGGCAGCAATCCCTGGCATACCACTAGGAACCAACACATTTCCAAAGAGCCCTGATT
 R I E T W I L R H P G F T M H A A I L A Y T I G T T H F Q R A L I
 E
 910 920 930 950 960 970 980 990 1000
 TTCATCTTACTGACAGCTGTCACCTTCAATGACAAATGCCCTGCTGATGGAATGCAATAGACTTTGTGAGGGGTTTTCAGGAGGAAGCTGGGTTG
 F I L L T A V T P S M T M R C I G M S N R D F V E G V S G G S W V D
 1010 1020 1030 1040 1050 1060 1070 1080 1090 1100
 ACATAGTCTAGAACATGGAACTGTGTGACGACGATGGCAAAAACAAACCAACATTTGAACTGATATAAACAGAAAGCCAAACAGCCTGCCCAC
 I V L E H G S C V T T M A K N K P T L D F E L I K T E A K Q P A T
 1110 1120 1130 1140 1150 1160 1170 1180 1190 1200
 CCTAAGGAAGTACTGTATAGAGGCAAGCTTCAATGACAAACACACAGAACTCTGGCTGCCAACACAGGGGAAACCCAGCCTAAATGAAGAGCAGGACAAA
 L R K Y C I E A K L T N T T T E S R C P T Q G E P S L N E E Q D K
 1210 1220 1230 1240 1250 1260 1270 1280 1290 1300
 AGGTTCTGCAACACTCCATGGTAGACAGAGGATGGGAAATGGATGGACTATTTGAAAGGGAGGCATTTGACCTGTGCTATGTTCCAGATGCA
 R F V C K H S M V D R G W G N G C G L F G K G I V T C A M F R C K
 Sphi
 1310 1320 1330 1340 1350 1360 1370 1380 1390 1400
 AAAAGAACATGGAAAGTTGTGCCAACAGAACTTGAATACACCTTGTGATACACTCAGGGGAAAGAGCATGCGAGTCGGAATGACAC
 K N M E G K V V Q P E N L E Y T I V I T P H S G E E H A V G N D T
 1410 1420 1430 1440 1450 1460 1470 1490 1500
 AGGAAACATGGCAAGGAATCAAAATAACACACAGAGTCCATCCAGAGCAATTTGACAGTTATGGCACTGTCCAAATGGAGTCTCTCCAAGA
 G K H G K E I K I T P Q S S I T E A E L T G Y G T V T M E C S P R
 1520 1530 1540 1550 1560 1570 1580 1590 1600
 ACGGCTCGACTTCAATGAGATGGTGTGCTGCAGATGGAATAAAGCTGGTGGTGCACAGGCAATGGTCTCCAGCCTGCGCTTCCATGGTTGC
 T G L D F N E M V L L Q M E N K A W L V H R Q W F L D L P L P W L P

FIGURE 14B

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1610 1620 1630 1640 1650 1660 1670 1680 1690 1700
 CCGAGCGGACACACAAAGGTCAAATGGATACAGAAAGAGACATTTGGTCACTTCAAAAATCCCATCCCGAAGAACAGGATGTTGTTTTAGGATC
 G A D T Q G S N W I Q K E T L V T F K N P H A K K Q D V V V L G S

1710 1720 1730 1740 1750 1760 1770 1780 1790 1800
 CCRAAGGGGCCATGCACACAGCACTACAGGGCCACAGAAATCCAAATGTCCATCAGGAAACTTACTCTTCACAGGACATCTCAAGTCCAGGCTGAGA
 Q E G A M H T A L T G A T E I Q M S S G N L L F T G H L K C R L R

1810 1820 1830 1840 1850 1860 1870 1880 1890 1900
 ATGACACAGCTACAGCTCAAGGATGTCTACTCTATGTGCACAGGAAGTTAAAGTTGTAAGGAAATAGCAGAAACACCAACATGGAACAATAGTTA
 M D K L Q L K G M S Y S M C T G K F K V V K E I A E T Q H G T I V I

1910 1920 1930 1940 1950 1960 1970 1980 1990 2000
 TCAGAGTGCNATATGAAGGGGACGGCTCCATGCCAAGATCCCTTTTGAATATGATGGAAATGGAAGAAACATGTCTTAGTGGCCTGATTACAGTCAA
 R V Q Y E G D G S P C K I P F E I M D L E K R H V L G R L I T V N

2010 2020 2030 2040 2050 2060 2070 2080 2090 2100
 CCCAATGTGACAGAAAAGATAGCCAGTCAACATAGAAAGCAGAACCTCCATTCGGAGACAGCTACATCATAGGATAGGATAGCCGGACRACTGAAG
 P I V T E K D S P V N I E A E P P F G D S Y I I I G V E P G Q L K

2110 2120 2130 2140 2150 2160 2170 2180 2190 2200
 CTCAACTGGTTTAAAGAAAGTCTATCGGCCAAATGTTGAGACAACTAGGGGGCGAAGAAATGGCCATTTTAGGTGACACAGCCCTGGGATT
 L N W F F K K G S S I G Q M F E T T M R G A K R H A I L G D T A W D F

2210 2220 2230 2240 2250 2260 2270 2280 2290 2300
 TTGGATCCTTGGAGGAGTGTTFACATCTATAGGAAGGCTCTCCACCAGTCTTTGGAGCAATCTATGGAGTGCCTTCAGTGGGTTTCATGGACTAT
 G S L G G V F F T S I G K A L H Q V F G A I Y G A A F S G V S W T M

2310 2320 2330 2340 2350 2360 2370 2380 2390 2400
 GAAATCCTCATAGGAGTCAATATACATGGAT
 K I L I G V I I T W I G M N S R S T S L S V T L V L V G I V T L Y

2410 2420 2430 2440 2450 2460 2470 2480 2490 2500
 TTGGGAGTCTGCTGCGCCGATAGTGGTTGGCTTGTAGCTGGAATAAACAAGAACTGAATGTCAGTGGGATTTTCATCATCAGACRACTGCACA
 L G V M V Q A D S G C V V S W K N K E L K C G S G I F I T D N V H T

NS1

FIGURE 14C

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2510 2520 2530 2540 2550 2560 2570 2590 2600
 CATGGACAGAACAAATACAGTCCCAACCCAGTCCCAAACTAGCTTCCAGAAAGCCCATGAGGGGCAATTTGTGGAAATCCGGCTCAGT
 W T E Q Y K F Q P E S P S K L A S A I Q K A H E E G I C G I R S V
 A
 |
 D

2610 2620 2630 2640 2650 2660 2670 2680 2690 2700
 AACRAGACTGGAGAACTGTGGAAACAATAACACCCAGAAATGAAATCACATTTATCAGAAATGAGGTGAAGTTAACTTATGACAGGAGACATC
 T R L E N L M W K Q I T P E L N H I L S E N E V K L T T I M T G D I
 HpaI
 |

2710 2720 2730 2740 2750 2760 2770 2780 2790 2800
 AAAGGAATCATGCAGCAGGAAACGATCTCTGGGCCTCAGCCCACTGAGCTGAAGTATTCATGGAARACATGGGCAAGCCAAAATGCTCTCTACAG
 K G I M Q A G K R S L R P Q P T E L K Y S W K T W G K A K M L S T E

* |
 2820 2830 2840 2850 2860 2870 2880 2890 2900
 AGTCTCATAACCCAGACCTTCTCATTTGATGSCCCCGAACACAGCAGAAATGCCCAACACAAATAGAGCTTGGAAATTCGTTGGAAGTTGAAGACTATGGCTT
 S H N Q T F L I D G P E T A E C P N T N R A W N S L E V E D Y G F

2910 2920 2930 2940 2950 2960 2970 2980 2990 3000
 TGGAGTATTCACCAACCAATATGGCTAAATGAAAGAAACAGGATGATTTCTGGACTCAAAACTCATGTCAGCGGCCATAAAGCAACACAGAGCC
 G V F T T N I W L K L K E K Q D V F C D S K L M S A A I K D N R A

* |
 3010 3020 3030 3040 3050 3060 3070 3080 3090 3100
 GTCCATGCCGATATGGGTTATTGGATAGAAAGTGCACCTCAATGACACATGGAAAGATAGAGAAAGCCCTCTTTCATTTGAAGTTAAAAACTGCCACTGGCCAA
 V H A D M G Y W I E S A L N D T W K I E K A S F I E V K N C H W P K

3110 3120 3130 3140 3150 3160 3170 3180 3190 3200
 AATCACACCCCTCTGGAGCAATGGAGTGTAGAAAGTGAATGATATTCAAAGAAATCTCGCTGGACCAGTGTCTCAACACAACTATAGACCAGGCTA
 S H T L W S N G V L E S E M I I P K N L A G P V S Q H N Y R P G Y

3210 3220 3230 3240 3250 3260 3270 3280 3290 3300
 CCATACAAATAACAGGACCATGGCATCTAGGTAAGCTTGGATGGACTTTGATTTCTGTGATGGAACAACAGTGGTAGTACTGAGGACTGGCGGAAT
 H T Q I T G P W H L G K L E M D F D F C D G T T V V V T E D C G N

FIGURE 14D

3310 3320 3330 3340 3350 3360 3370 3380 3390 3400
 AGAGACCCTCTTTGAGAACAAACCACCTGCTCTGGAAACTCATAACAGAAATGGTCTGCCGATCTTGACAAATFACCAACCGCTAAGATACAGAGGTGAGG
 R G P S L R T T A S G K L I T E W C C R S C T L P P L R Y R G E D

3410 3420 3430 3440 3450 3460 3470 3480 3490 3500
 ATGGGTGCTGGTAGGGATGGAAATCAGACCATTGGAGGAGAAAGAGAAATTTGGTCACTCCCTTGGTCAAGCTGGACATGGCGAGTGGCAACAATT
 G C W Y G H E I R P L K E K E E N L V N S L V T A G H G Q V D N F

3510 3520 3530 3540 3550 3560 3570 3580 3590 3600
 TTCACTAGGAGTCTGGGAATGGCAATGCTCCTGGAGGAATGCTTAGACCCCGAGTAGGAACGAAACATGCAACTACTACTAGTTGCAGTTCTTTTGTG
 S L G V L G M A L F L E E M L R T R V G T K H A I L L V A V S F V

3610 3620 3630 3640 3650 3660 3670 3680 3690 3700
 ACATTGATCACAGGGAAACATGCTCCTTAGAGACCCTGGAGAGTGTGTTATGGTAGGCCCCACTATGACGGATGACATAGGTATGGGCGTGACTTATC
 T L I T G N M S F R D L G R V M V G A T M T D I G M G V T Y L

3710 3720 3730 3740 3750 3760 3770 3780 3790 3800
 TTGCCCTACTAGCAGCCTTCAAGTCAAGCCACTTTGCGAGCTGACTCTTTGAGAAAGCTGACCTCCAAAGGAAATGATGACTACTATAGGAAT
 A L L A A F K V R P T F A A G L L L R K L T S K E L M T T I G I

3810 3820 3830 3840 3850 3860 3870 3880 3890 3900
 TGTACTCTCCAGACCAATACAGAGACCATCTTGGAGTTGACTGATCGGTAGCCTTAGCCCTAGGATGGTCCCAAAATGGTGAGAAATATGGAA
 V L L S Q S T I P E T I L E L T D A L A L G M H V L K M V R N M E

3910 3920 3930 3940 3950 3960 3970 3980 3990 4000
 AAGTATCAATGGCAGTACTATCATGGCTATCTGTGCTCCCAACGCGAGTGATATACAAAACCCATGGAAAGTGGACATGGCAAAATTTGGCAGTGG
 K Y Q L A V T I M A I L C V P N A V I L Q N A W K V S C T I L A V V

4010 4030 4040 4050 4060 4070 4080 4090 4100
 TGTCCGTTTCCCCACTGCTTAAACATCCCTCACAGCAAAAACAGATGGATACCATGGATCAGGATCAAAAGTCTCAATCCAAACAGCTATTTTCT
 S V S P L L L T S S Q Q K T D W I P L A L T I K G L N P T A I F L

4110 4120 4130 4140 4150 4160 4170 4180 4190 4200
 AACAACTCTCAGAACCCAGCAAGAAAGGAGCTGGCCATTAATGAGGCTATCATGGCAGTCCGGATGGTGAGCAATTTAGCCAGTTCTCTCCTAAA
 T T L S R T S K K R S W P L N E A I M A V G M V S I L A S S L L K

FIGURE 14E

5110 5120 5130 5140 5150 5160 5170 5180 5190 5200
 CACCAGGCGGGAAGACGAAGATACCTCCGGCCATAGTCAGAGAGCTATAAACCGGGTTGGAGACATTAATCTGGCCCCCCTAGAGTTG
 H P G A G K T K R Y L P A I V R E A I K R G L R T L I L A P T R V V

5210 5220 5230 5240 5250 5260 5270 5280 5290 5300
 TGGCAGCTGAAATGAGGAAGCCCTTAGAGGACTTCCAAATAAGATACCAGACCAGCCATCAGAGCTGAGCAGACCGGGCGGAGATTGGACCTAAT
 A A E H E E A L R G L P I R Y Q T P A I R A E H T G R E I V D L M

5310 5320 5330 5340 5350 5360 5370 5380 5390 5400
 GTGTCATGCCACATTTACCATTAGGCTGCTATCACCCAGTTAGAGTGCJAACTACAACCTGATTATCATGGACGAAGCCCATTTCCACAGACCCAGCAAGT
 C H A T F T M R L L S P V R V P N Y N L I I M D E A H F T D P A S

5410 5420 5430 5440 5450 5460 5470 5480 5490 5500
 ATAGCAGCTAGAGGATACATCTCAACTCGAGTGGAGATGGGTGAGGCAGCTGGGATTTTATGACAGCCACTCCCGGGAAGCAGAGACCCATTTCCCTC
 I A A R G Y I S T R V E H G E A A G I F M T A T P P G S R D P F P Q

5510 5520 5530 5540 5550 5560 5570 5580 5590 5600
 AGAGCAATGCACCAATCATAGTAGAAGAGAAATCCCTGAACCGCTCCGTGGAATCCCGACATGAATGGTCAACGGATTTTAAAGGGGAAGACTGTTTG
 S N A P I I D E E R E I P E R S W N S G H E W V T D F K G K T V W

5610 5620 5630 5640 5650 5660 5670 5680 5690 5700
 GTTCGTTCCAGTATAAAGCAGGAATGATATAGCAGCTTCCCTGAGGAAATGGAAAGAAAGTATACAACACTCAGTAGGAAGACCTTTGATCTGAG
 F V P S I K A G N D I A A C L R K N G K K V I Q L S R K T F D S E

5710 5720 5730 5740 5750 5760 5770 5780 5790 5800
 TATGTCAGACTAGAACCAATGATGGACTTCGTGGTTACAACACTGACATTTCCAGAAATGGGTGCCAATTTCAAGGCTGAGAGGGTTATAGACCCAGAC
 Y V K T R T N D W D F V V T T D I S E M G A N F K A E R V I D P R R

5810 5820 5830 5840 5850 5860 5870 5880 5890 5900
 GCTGCATGAACCACTACTAACAGATGGTGAAGCGGGTGTATTTGGCAGGACCTATGCCAGTCCACTCTGTGTCAGCACNAAGAAAGAGGGAG
 C H K P V I L T D G E E R V I L A G P H P V T H S S A A Q R R G R

5910 5920 5930 5940 5950 5960 5970 5980 5990 6000
 AATAGGAAGAAATCCAAAATGAGAAATGACCCAGTACATATACATGAGGGAACCTCTGGAATGATGAAGACTGTGCACACTGGAAGAAAGCTAAATG
 I G R N P K N E N D Q Y I Y M G E P L E N D E D C A H W K E A K M

6010 6020 6030 6040 6050 6060 6070 6080 6090 6100
 CTCCTAGATAACATCAACCGCCAGGAAGAAATCATCTCCTAGCATGTTCCGACCCAGGCTGAAAGGTGGATGCCATTTGATGGCGAATACCCGCTTGAGAG
 L L D N I N T P E G I I P S M F E P E R E K V D A I D G E Y R L R G

FIGURE 14G

6110 6120 6130 6140 6150 6160 6170 6180 6190 6200
GAGAAGCAAGGAAACCTTTGTAGACTTAATGAGAAAGGAGGACCTACAGTCTGGTGGCCCTACAGACTGGCAGCTGAAGGCATCAACTAGCCAGACAG
E A R K T F V D L H R R G D L P V W L A Y R V A A E G I N Y A D R

6210 6220 6230 6240 6250 6260 6270 6280 6290 6300
AAGTGGTGTGGTGGAGTCAAGAACCAACCAATCCTAGAAAGAAACGTGGAAGTTGAAATCTGGACAAAAGAAAGGAAAGAAATGAAACCC
R W C F D G V K N N Q I L E E N V E I W T K E G E R K K L K P

6310 6320 6330 6340 6350 6360 6370 6390 6400
AGATGGTTGGATGCTAGGATCTATTCTGACCCACTGGCGCTAAAAGAAATTAAGGAATTTGACGCCGGAAAGAAAGTCTCTGACCCCTGAACCTAATCACAG
R W L D A R I Y S D P L A L K E F K E F A A G R K S L T L N L I T E

6410 6420 6430 6440 6450 6460 6470 6480 6490 6500
AAATGGTAGGCTCCCAACCTTCATGACTCAGAAAGCAAGGACGCACTGGACAACTTAGCAGTGTGCACACGGCTGAGCCAGGTGGRAAGCGGTACAA
M G R L P T F M T Q K A R D A L D N L A V L H T A E A G G R A Y N

6510 6520 6530 6540 6550 6560 6570 6580 6590
CCATGCTCTCAGTGAACCTCCGAGACCCCTGGAGACATTTGCTTTTACTGACACTTCTGGCTACAGTCCAGGGAGGATCTTTTATCTTGTGAGCGGA
H A L S E L P E T L E T L L L L T L L A T V T G G I F L F L M S G
C

6610 6620 6630 6640 6650 6660 6670 6680 6690 6700
AGGGCANTAGGGAAGATGACCCCTGGAAATGTGCTGATATACCGCTAGCATCTCCTATGGTACGCACAAATACGCCACACTGGATAGCAGCTTCAA
R G I G K H T L G M C C I I T A S I L L W Y A Q I Q P H W I A A S I

6710 6720 6730 6740 6750 6760 6770 6780 6790 6800
TAATACTGGAGTTTTTCTCATAGTTTTGCTTATCCAGAACCTGAAACAGAGACACCCCAAGACAACTGACCTACGTTGTGCATAGCCATCCT
I L E F F L I V L L I P E P E K Q R T P Q D N Q L T Y V V I A I L

6810 6820 6830 6840 6850 6860 6870 6880 6890 6900
CACAGTGGTGGCCCAACCATGGCAACCGAGATGGGTTTCTAGAAAACGAAAGAAAGATCTCGGATTTGGAAAGCATTGCAACCCCAACCCCGAGAGC
T V V A A T M A N E M G F L E K T K K D L G L G S I A T Q Q P E S

6910 6920 6930 6940 6950 6960 6970 6980 6990 7000
AACATCCGGACATAGATCTACCTCCTGCATCAGCATGGACGGCTGTATGCCGTGGCCACAACTTTGTTACCAATTTGTTAGACATAGCATTGAAATTT
N I L D I D L R P A S A W T L Y A V A T T F V T P H L R H S I E N S

NS4A

NS4B

NheI

FIGURE 14H

8010 8020 8030 8040 8050 8060 8070 8080 8090 8100
 GTGTGACATAGGGGAGTCAATCCCAATCCACAGTGGAAAGCAGGACGAAACACTCAGAGTCCTTAAGTCTAGTAGAAAATGGTTGAACCAACACTCA
 CDIGESSSSPNPTEAAGRTLRVLLVENEWLNNTQ

8110 8120 8130 8140 8150 8160 8170 8180 8190 8200
 TTTTGCATAAAGTTCTCAACCATATATGCCCTCAGTCATAGAAAATGGAAGCCTACAAGGMAATATGGAGGCTTAGTGAGGAATCCACTCT
 FCIKVLPYHPSVIEKHAEALQRYGALVLRNPLS

8210 8220 8230 8240 8250 8260 8270 8280 8290 8300
 CAGGAACTCCACACATGAGATGACTGGTATCCATGTTCCGGGAACATAGTGTCAATCAGTGAACATGTTTCAAGGATGTTGATCAACAGATTTAC
 RNSTHEMYWVSNASGNIVSSVNMHISRLINRFT

8310 8320 8330 8340 8350 8360 8370 8380 8390 8400
 AATGATACAAGAAGCCACTTACGAGCCGATGTTGACCTCGGAAGCGGAACCGTAACATCGGATTAAGTGAAGTACCAACCTAGATATAAT
 MRYKKAATTAAGCAAGCATGAACATATGACCTGACAAACCTTGGACCTAACAACCTGAGTACCAACCTAGATATAAT

8410 8420 8430 8440 8450 8460 8470 8480 8490 8500
 GGAAGAATAAGAAATAAGCAAGCATGAACATATGACCTGACAAACCTTGGACCTAACAACCTGAGTACCAACCTAGATATAAT
 GKRIEKIKQEHETSWHYDQDHPYKTAACCTGAGTACCAACCTAGATATAAT

8510 8520 8530 8540 8550 8560 8570 8580 8590 8600
 CAACACAGACTGGTCCATCCATGTCACCGAGTGTGTCAGGCTGTGACAAACCTTGGACCTGACAAACCTTGGACCTGACAAACCTTGGACCTGACAAACCT
 KQTSASMSMVGVRLLTKPVDVPTQMAMT

8610 8620 8630 8640 8650 8660 8670 8680 8690 8700
 AGACACACTCCATTTGACACACAGCGGTTTTTAAGAGAAGTGGACAGAGAACCCAGAACCGAAGAGCAGCAGGAACTAATAAATAACA
 DTFPGQQRVFKVKVDTRTQEPKEGTKLMIIT

8710 8720 8730 8740 8750 8760 8770 8780 8790 8800
 GCAGAGTGGCTTTGGAAAGATTAGGGAAGAAAAGACACCCAGGATGTGACCCAGAGAAATCAAGAAAGTGAAGCAATGCAGCCTTGGGG
 AEWLWKELGKKKTKTPRMCTREEFTRKVRSNALGA

8810 8820 8830 8840 8850 8860 8870 8880 8890 8900
 CCATATTCATGATGAGAACAAGTGAAGTGGCCCTGAGGCTGTTAAGATAGTGGTGGGCTGTGACAAAGGAAAGAAATCTCCATCTTGA
 IFIDEWKWKSAREAVEDSRFVELVDKERNLHLE

8910 8920 8930 8940 8950 8960 8970 8980 8990 9000
 AGGAAAGTGAACATGTGTACAACATGATGGGAAAGAGAGAGAGCTAGGGGAATTCGGCAAGGCAAAAGGCGAGGCAATATGGTACATG
 GKCEITCVYNMHMGGKREKRLGEEFGKAKGSRARIWYM

FIGURE 14J

9010 9020 9030 9040 9050 9060 9070 9080 9090 9100
 TGGCTGAGCAGCGCTTCTTAGAGTTTGAAGCCCTAGGATTTAAATGAAGTACTGTTCTCCAGAGAGAACTCCCTGAGTGGAGTGGAGGAGAG
 W L G A R F L E F A L G F L N E D H W F S R E N S L S G V E G E G

9110 9120 9130 9140 9150 9160 9170 9180 9190 9200
 GGCTGCACAGCTAGGTACATTTCTAGAGCGTGGCCAGAGAGGGAGGACCAATGTATGCCGATGACACCCGAGGATGGGATACAAAGATCACA
 L H K L G Y I L R D V S K K E G G A M Y A D D T A G W D T R I T L

9210 9220 9230 9240 9250 9260 9270 9280 9290 9300
 AGAAGACKKAAAATGAAGAAATGGTACAAAACCATGGAGGAGACACACAGAACTAGCCGAGGCCATTTCAAACTAACGTACCAAACAAGGTG
 E D ? K N E E M V T N H M E G E H K K L A E A I F K L T Y Q N K V

9310 9320 9330 9340 9350 9360 9370 9380 9390 9400
 GTCCGTGCAAGACCAACCAAGAGGACAGTAAATGGACATCATTCGRAGAGACCAACAGAGGTAGTGGACAAGTTGGCCCTATGGACTCAATA
 V R V Q R P T P R G T V M D I I S R R D Q R G S G Q V G T Y G L N T

9410 9420 9430 9440 9450 9460 9470 9480 9490 9500
 CTTTCAACCAATATGGAGCCCAACTAATCAGACAGATGGAGGAGGAGTCTTAAAGCATTTCAGCACCTTCAACATCACAGAAATCGCTGTGCA
 F T N H E A Q L I R Q M E G E G V F K S I Q H L T I T E E I A V Q

9510 9520 9530 9540 9550 9560 9570 9580 9590 9600
 ARACTGTTAGCAAGAGTGGCGGAAAGTTATCAAGAAATGGCCATCAGTGGAGATGTTGTGTGTAACCTTAGATGACAGGTTCGCAAGCCCT
 N W L A R V G R E R L S R H A I S G D D C V V K P L D D R F A S A

9610 9620 9630 9640 9650 9660 9670 9680 9690 9700
 TTAACGCTTAATGACATGGGAAAGATAGGAAAGACATACAACTTCAAGAGGATGGAAATGGACACACACAGTCCCTTCTGTTCAC
 L T A L N D M G K I R K D I Q Q W E P S R G W N D W T Q V P F C S H

9710 9720 9730 9740 9750 9760 9770 9780 9790 9800
 ACCATTCCATGATTAATCATGAAGACGGTCCGCTACTCGTTGTTCCATGTAGAAACCAAGATGAATGATGGCAGGCCCCGAATCTCCCAAGGAGC
 H F H E L I M K D G R V L V P C R N Q D E L I G R A R I S Q G A

9810 9820 9830 9840 9850 9860 9870 9880 9890 9900
 AGGGTGTCTTTGGGAGACGGCCCTGTTGGGGAAGTCTTACGCCCAATGTGGAGCTTGTATGTTCTCCACAGACCGGACCTCAGGCTGGCGGCAAT
 G W S L R E T A C L G K S Y A Q M W S L M Y F H R R D L R L A A N

9910 9920 9930 9940 9950 9960 9970 9980 9990 10000
 GCTATTGCTCGGACATACCATCACATTTGGTTCCCAACAGTCCGAACTGGTCCATACATGCTAAACATGAATGGATGACAAACGGAGACATGCTGA
 A I C S A V P S H W V P T S R T T W S I H A K H E W M T T E D M L T

FIGURE 14K

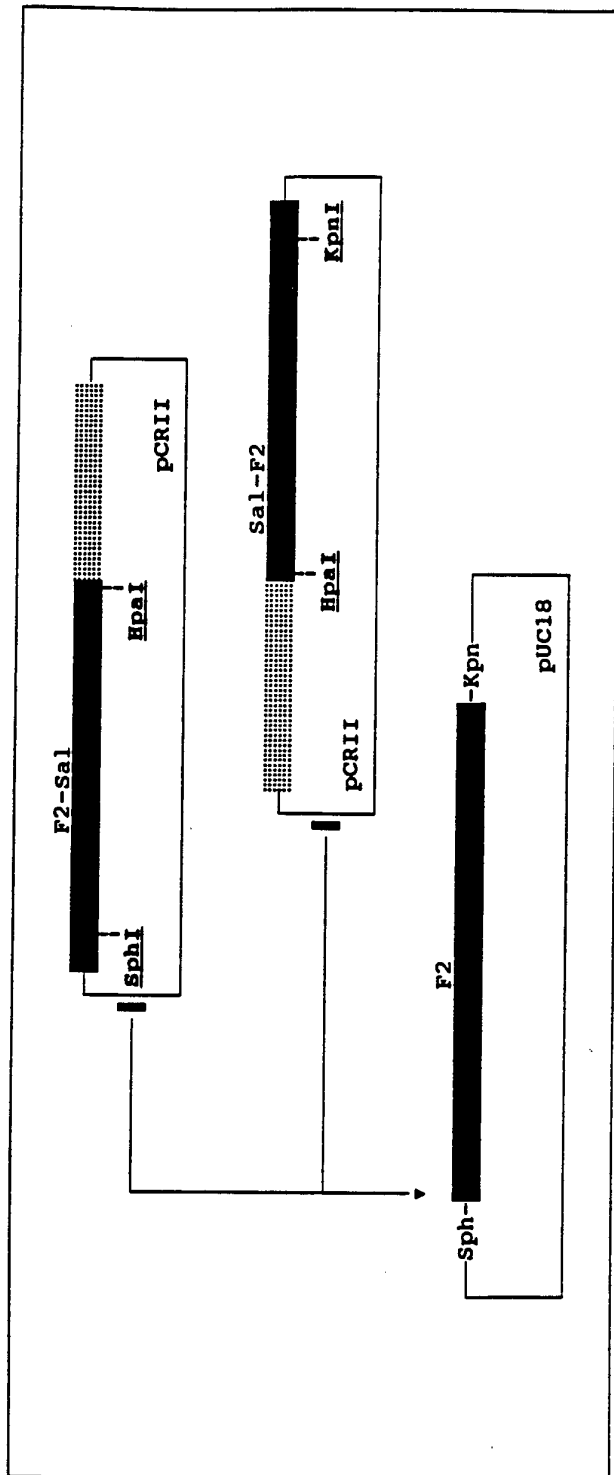


FIGURE 15

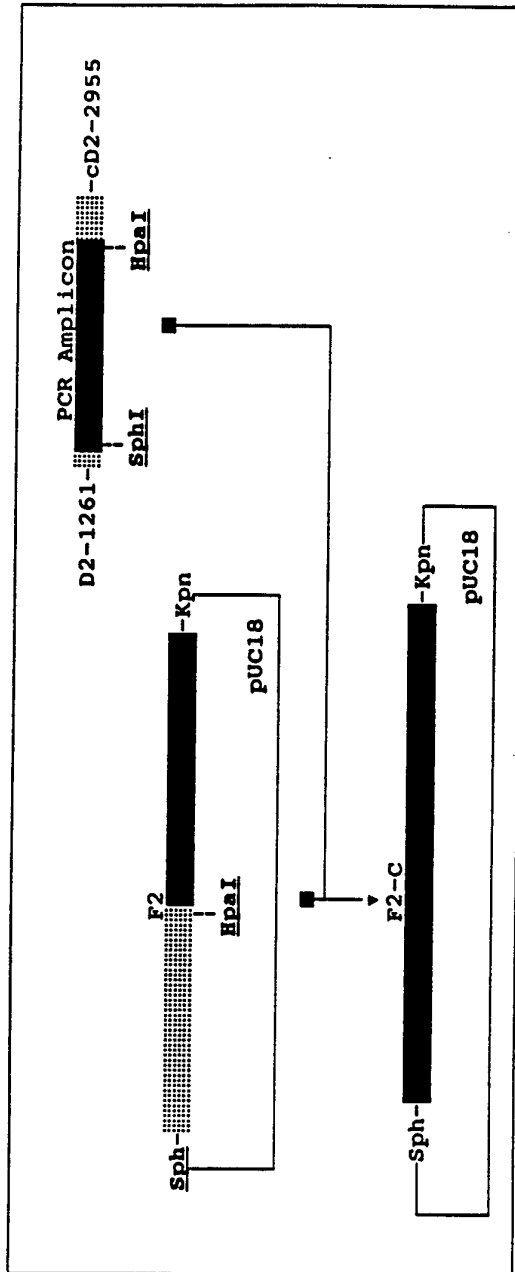


FIGURE 16

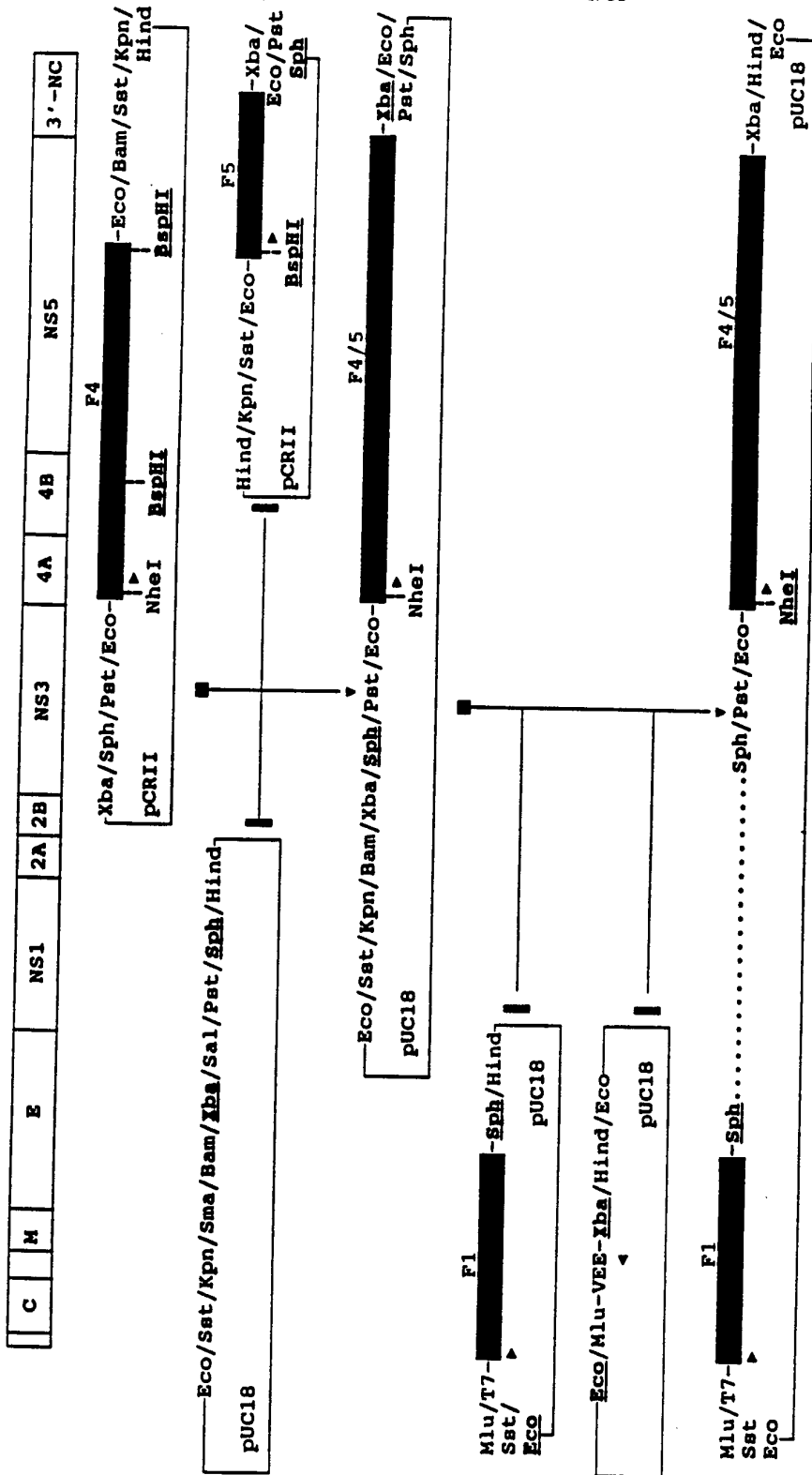


FIGURE 17A

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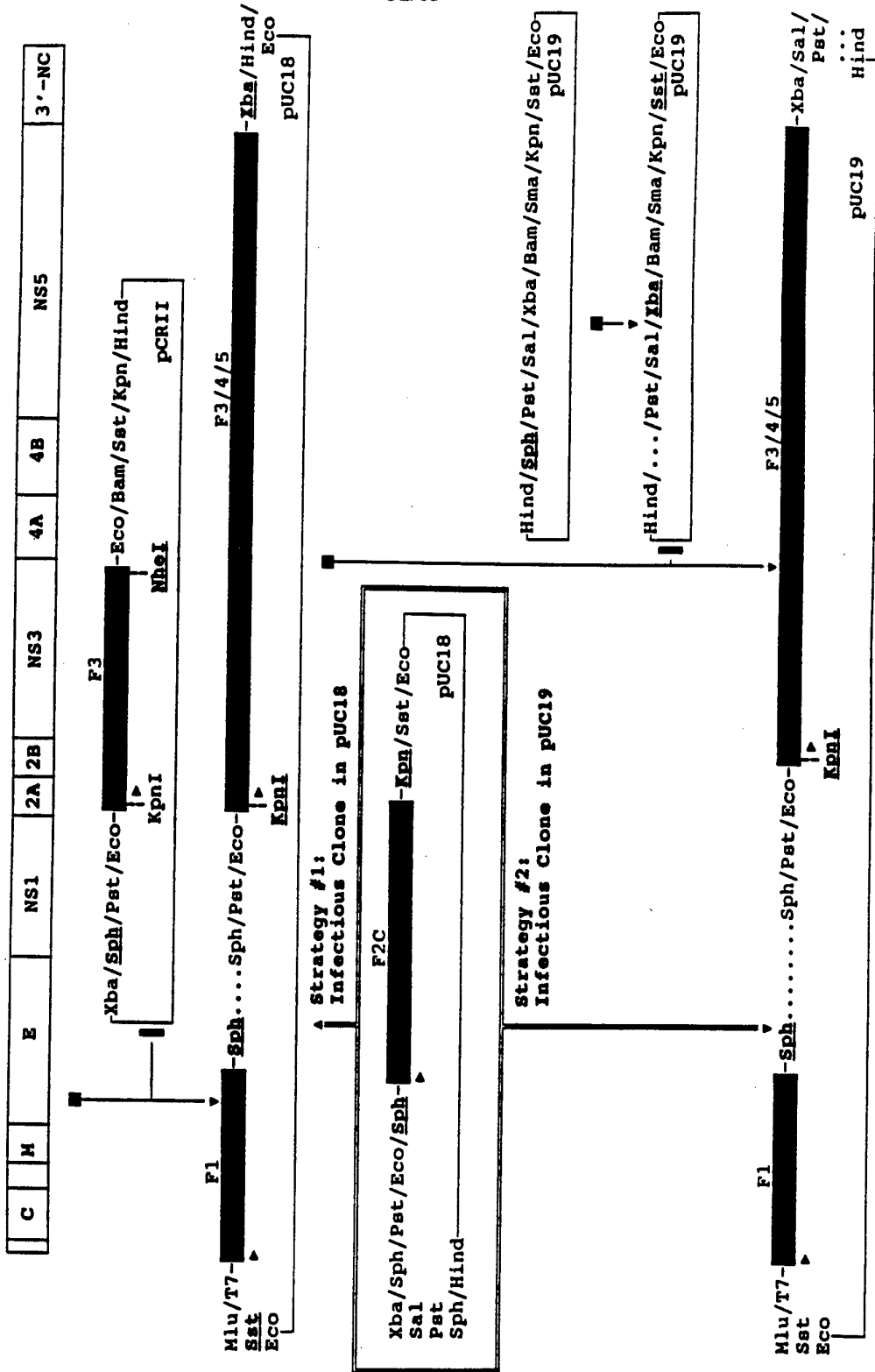


FIGURE 17B

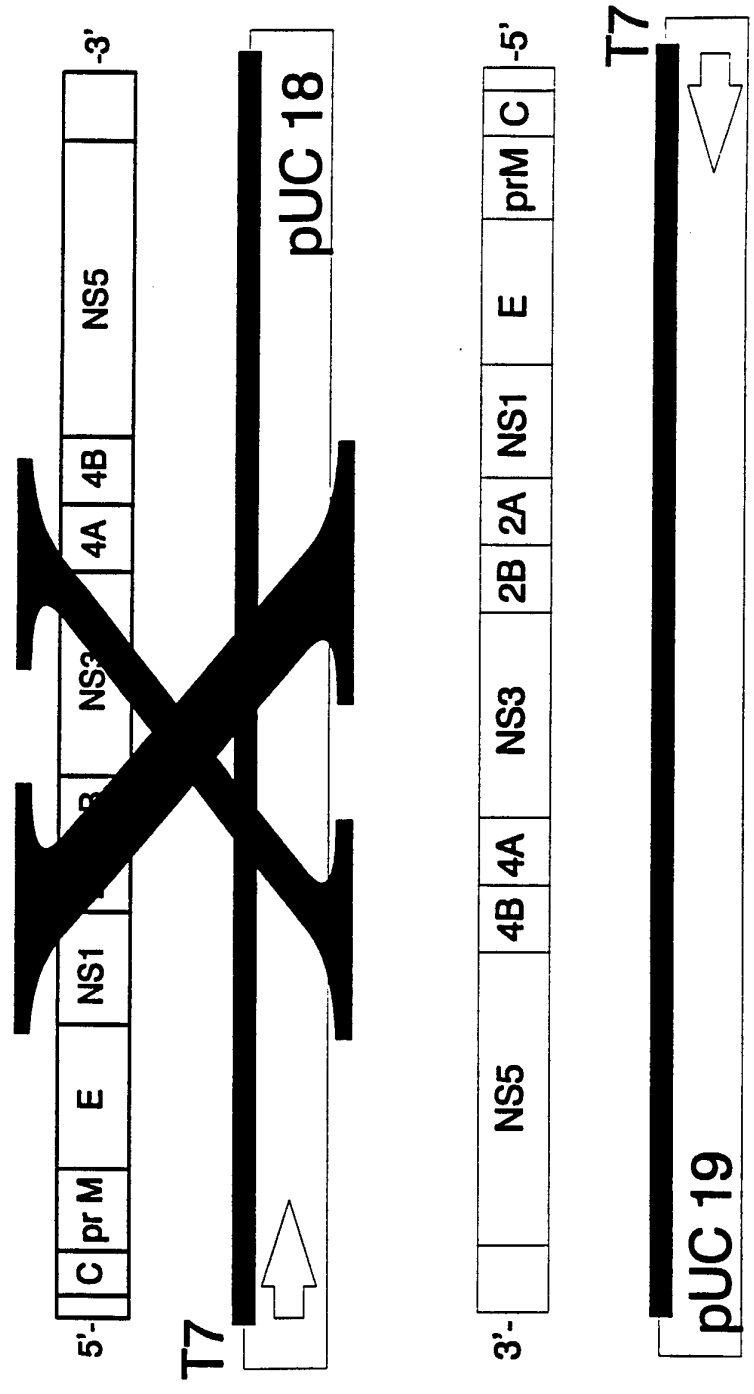


FIGURE 18

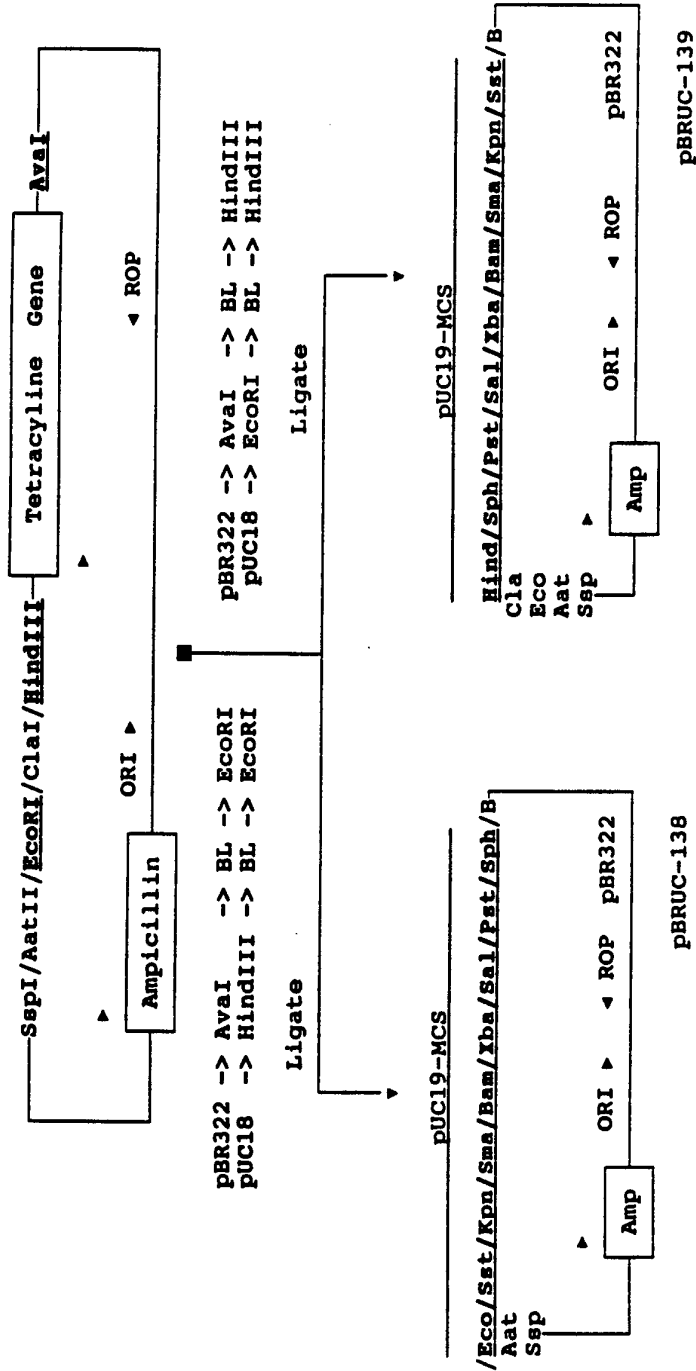


FIGURE 19

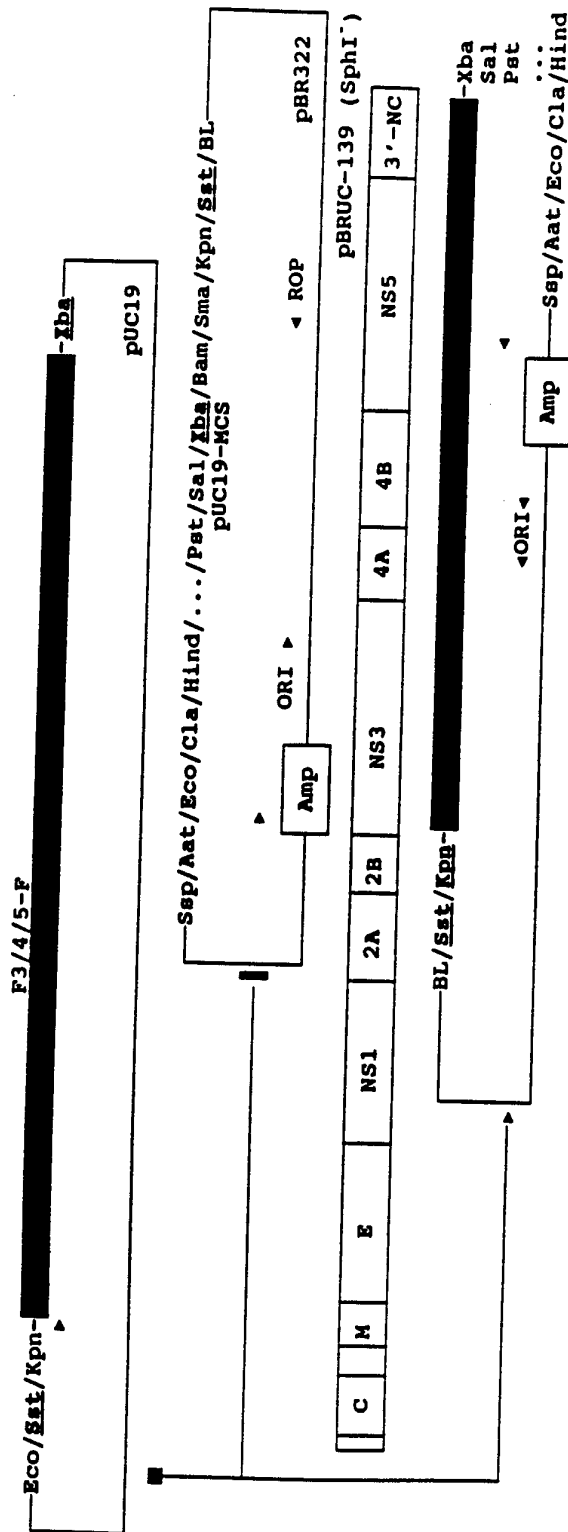


FIGURE 20A

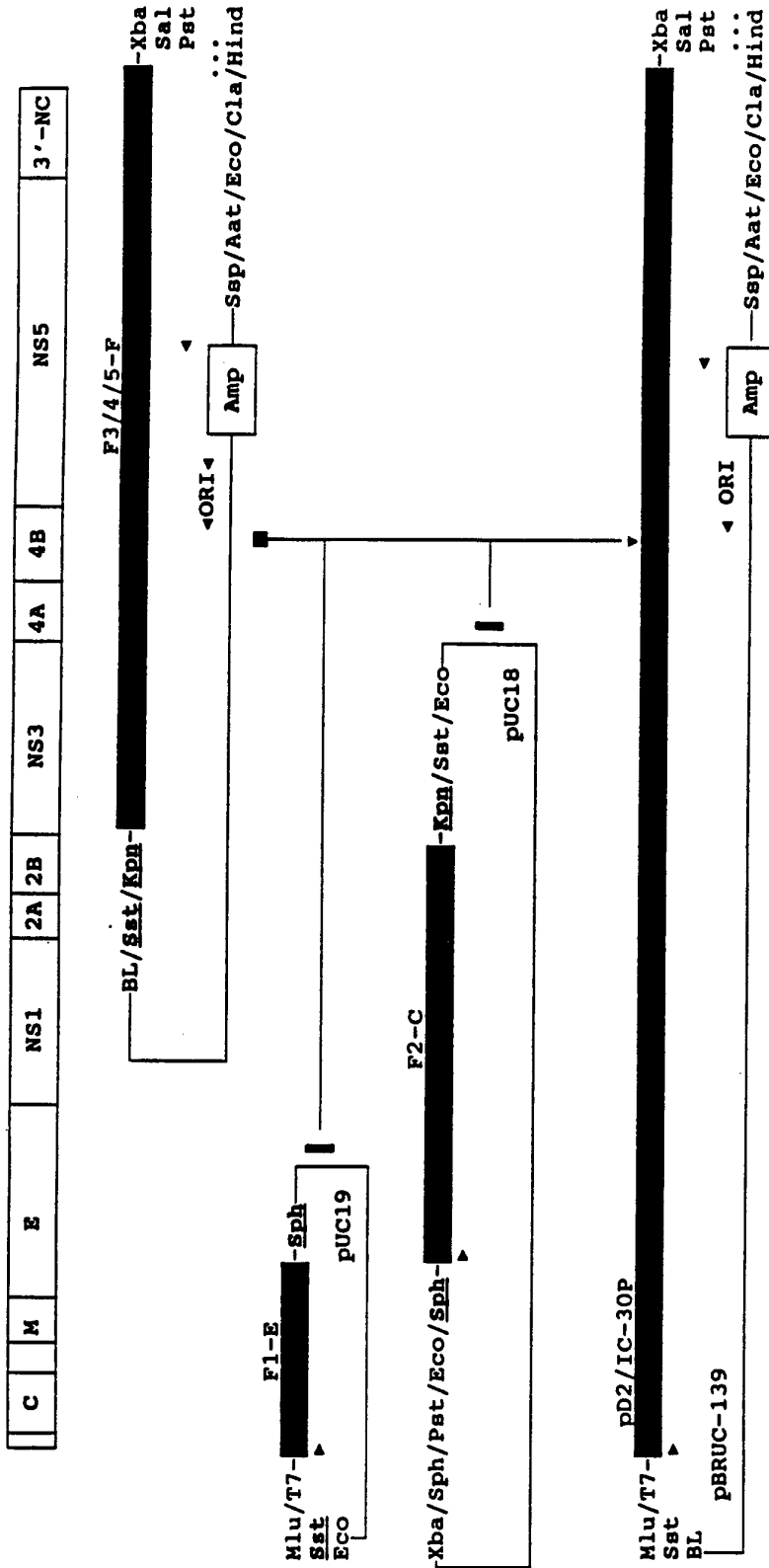


FIGURE 20B

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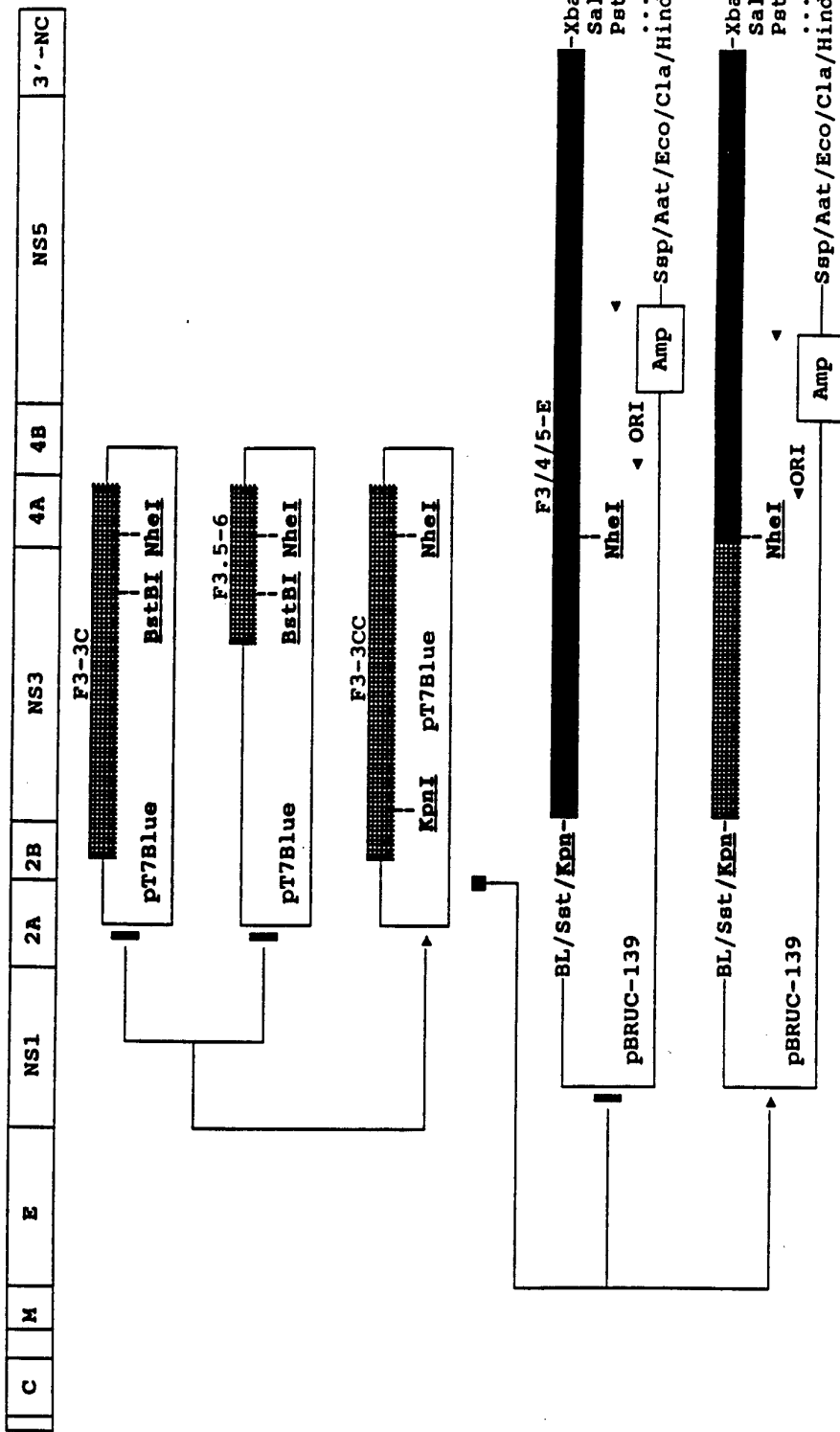
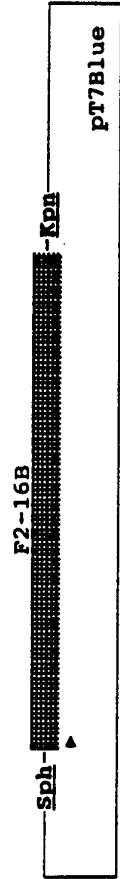
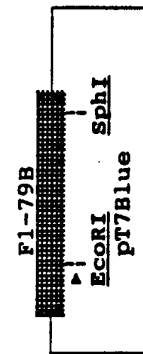
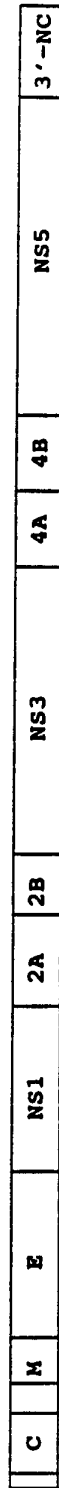


FIGURE 21A



pD2/IC-130-V



FIGURE 21B

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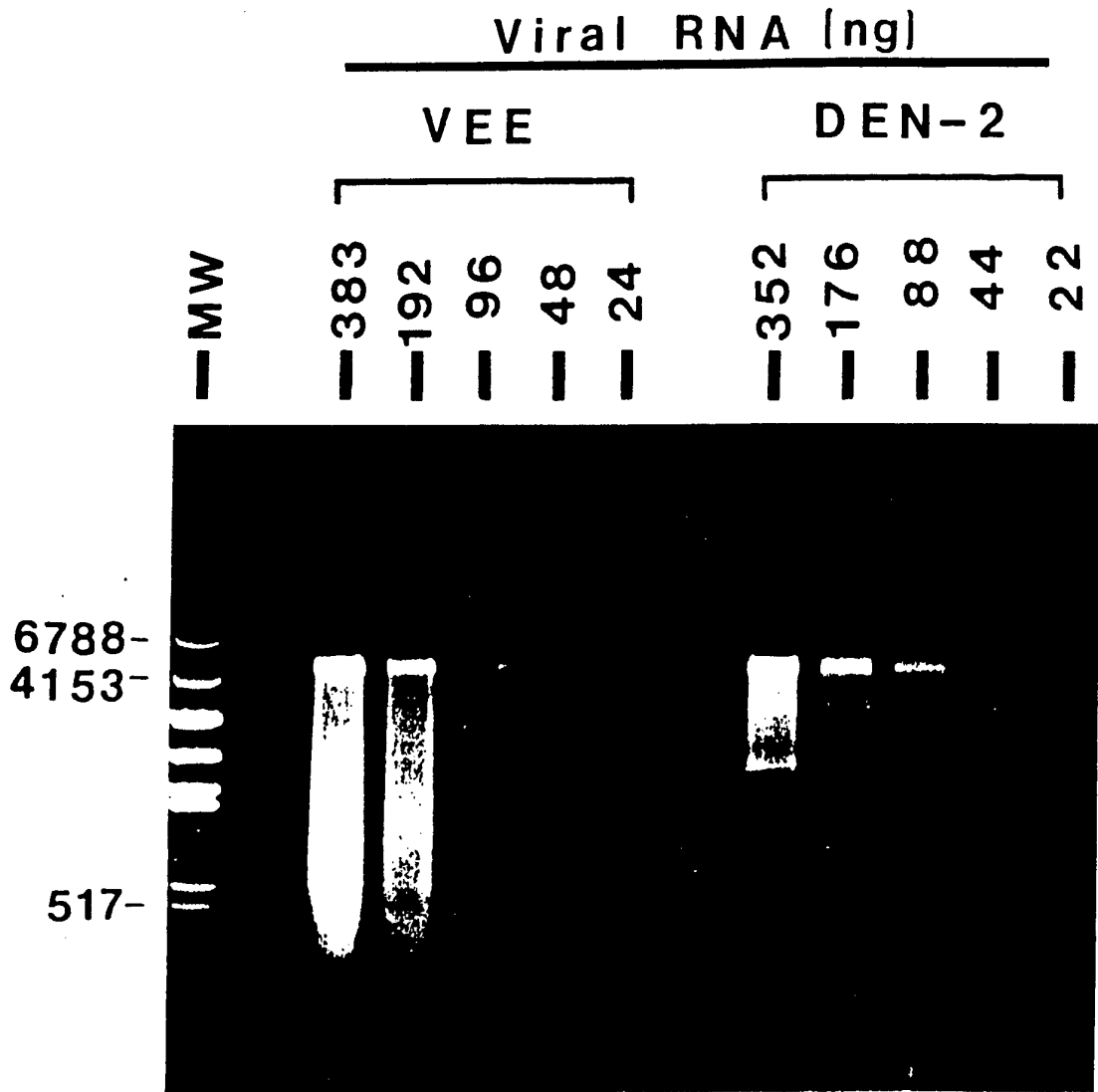


FIGURE 22

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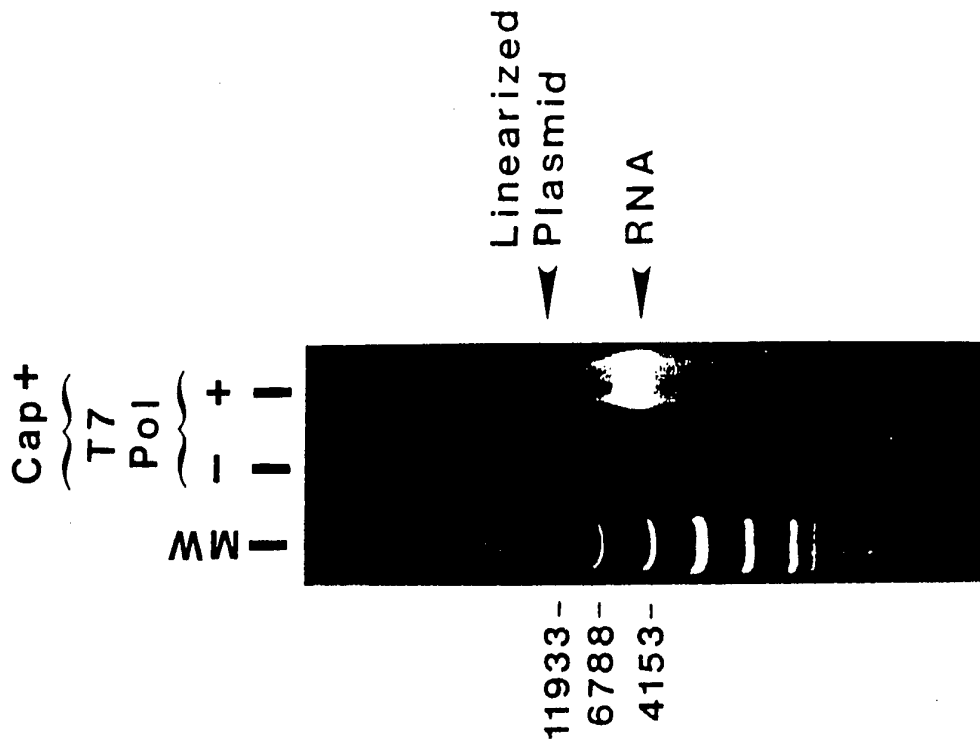


FIGURE 24B

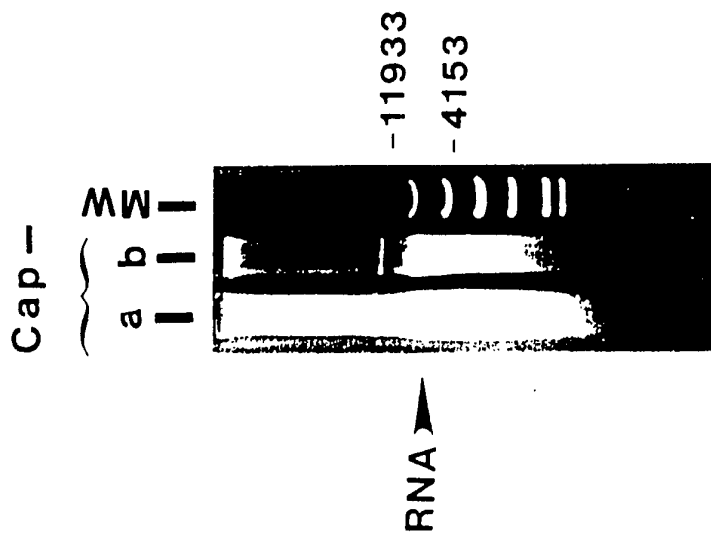


FIGURE 24A

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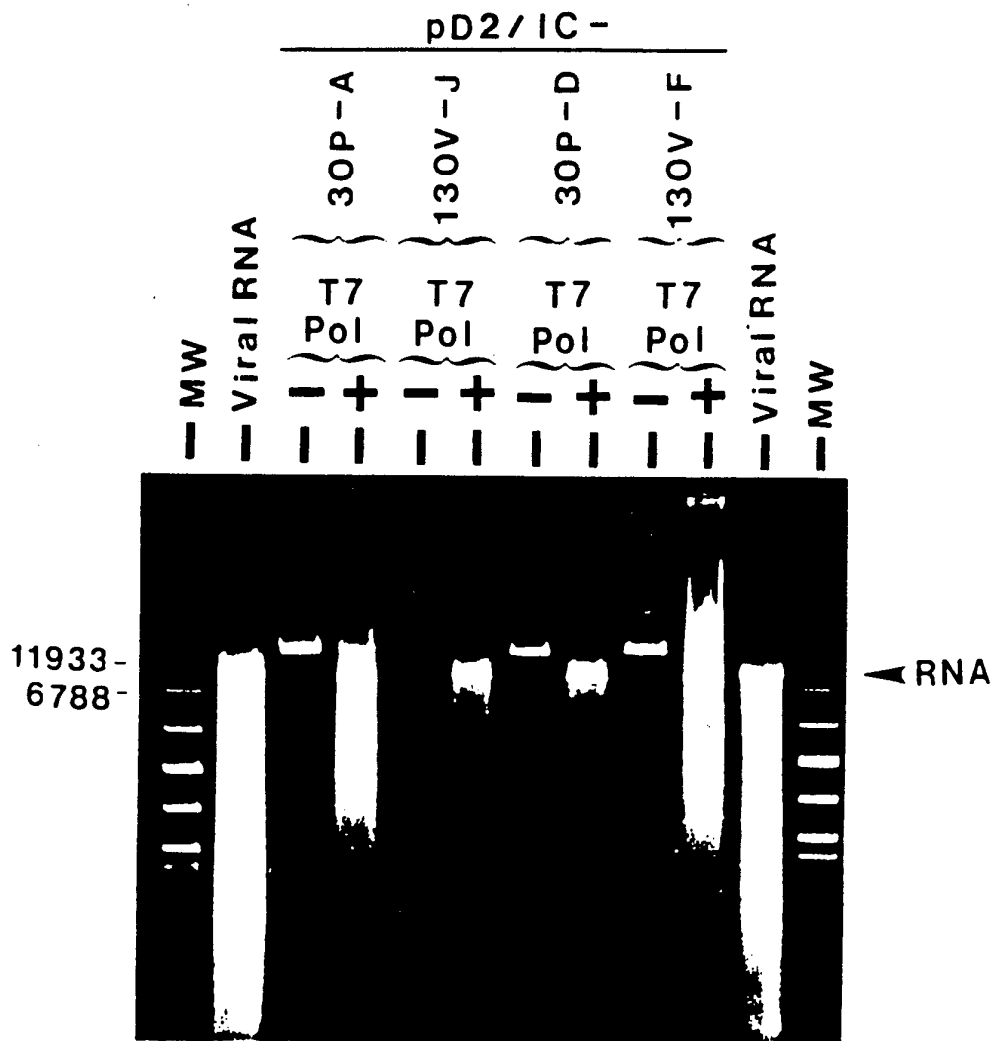


FIGURE 25

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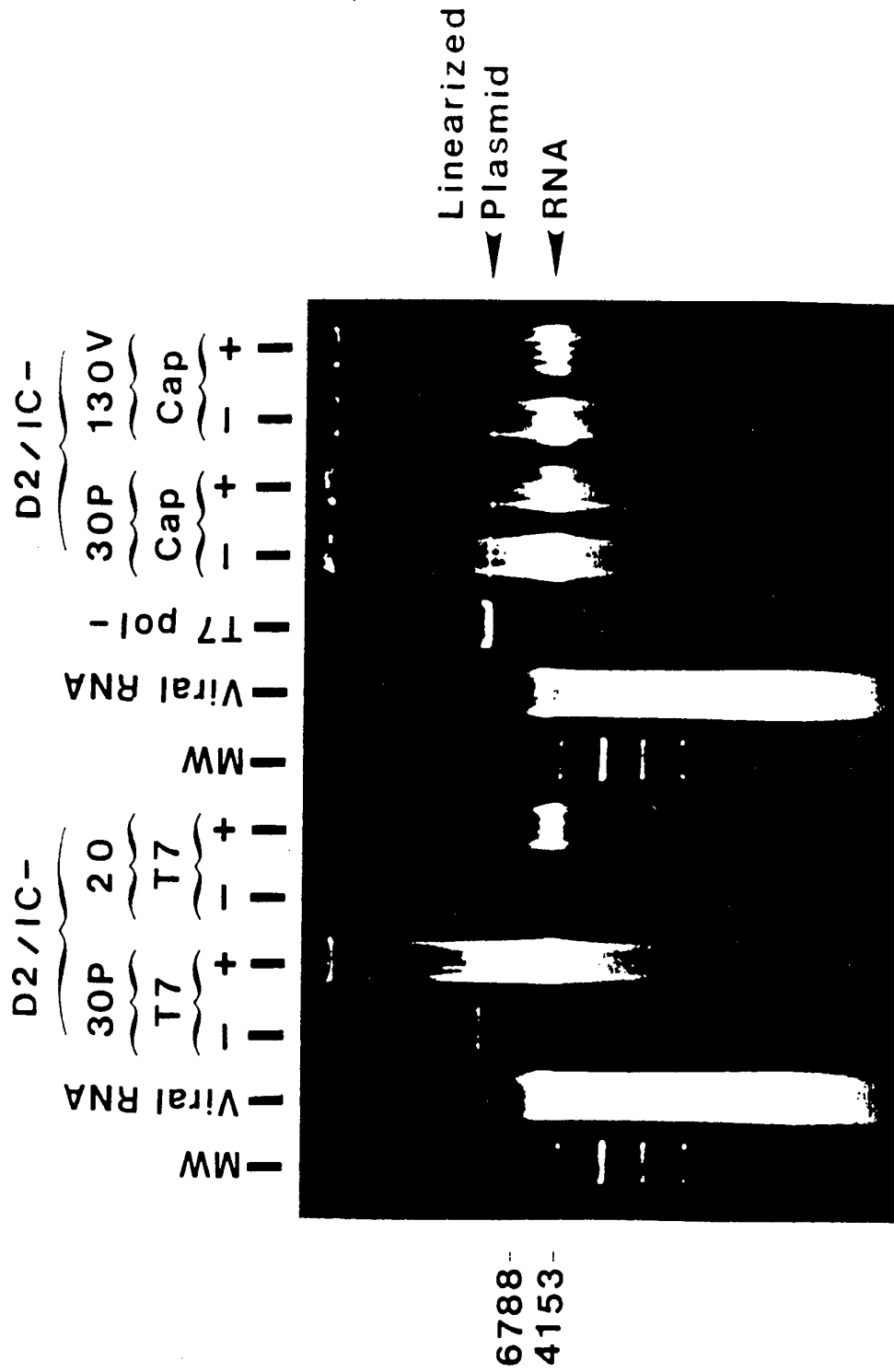


FIGURE 26

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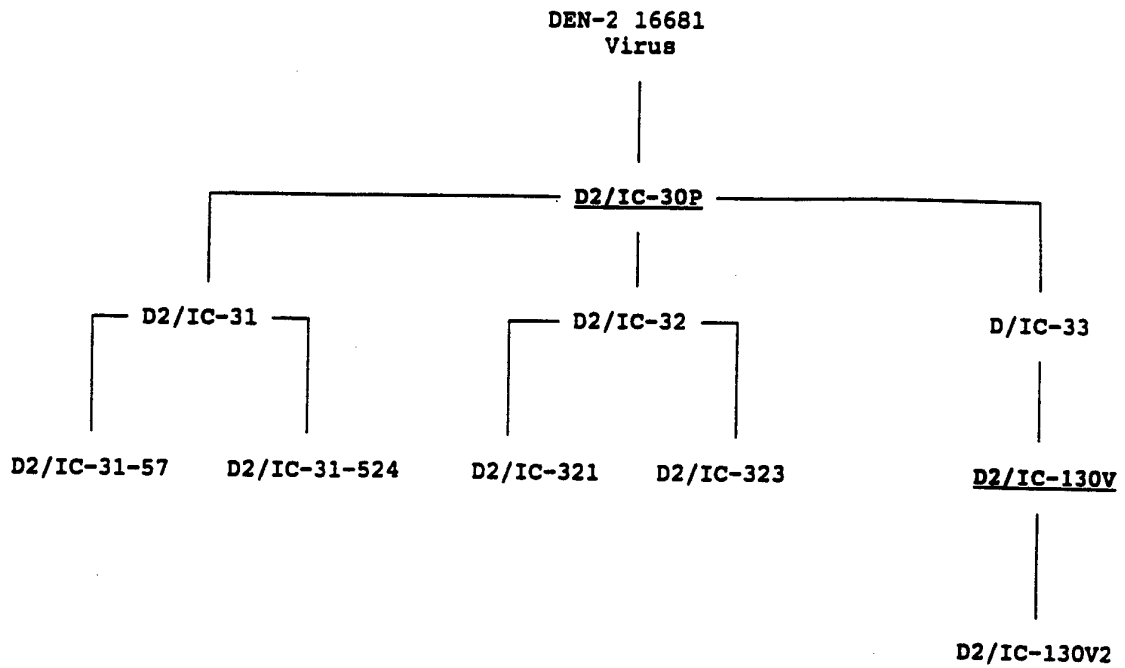
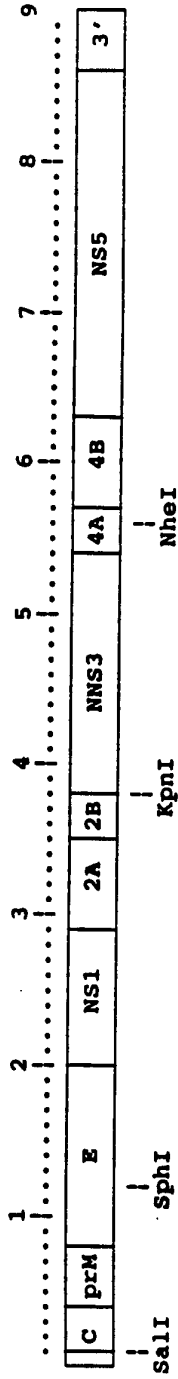


FIGURE 27



pd2/IC-

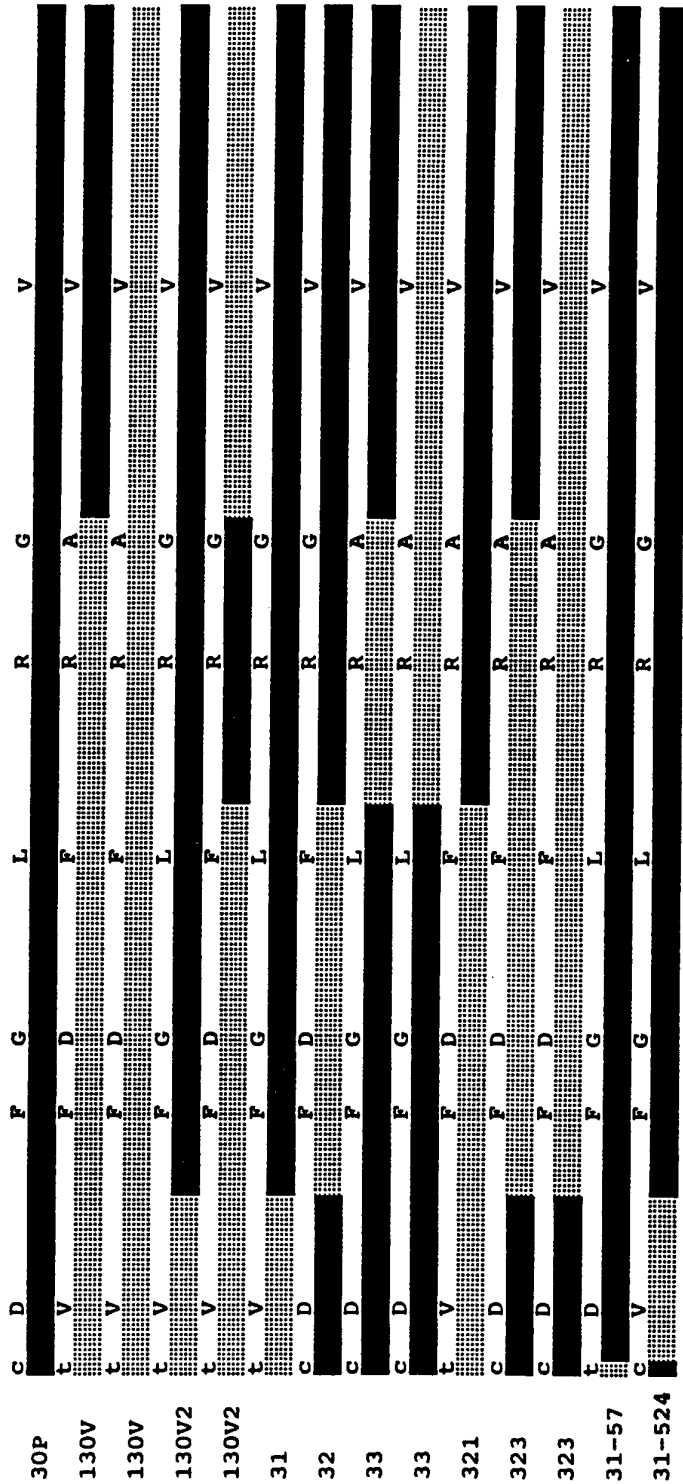


FIGURE 28

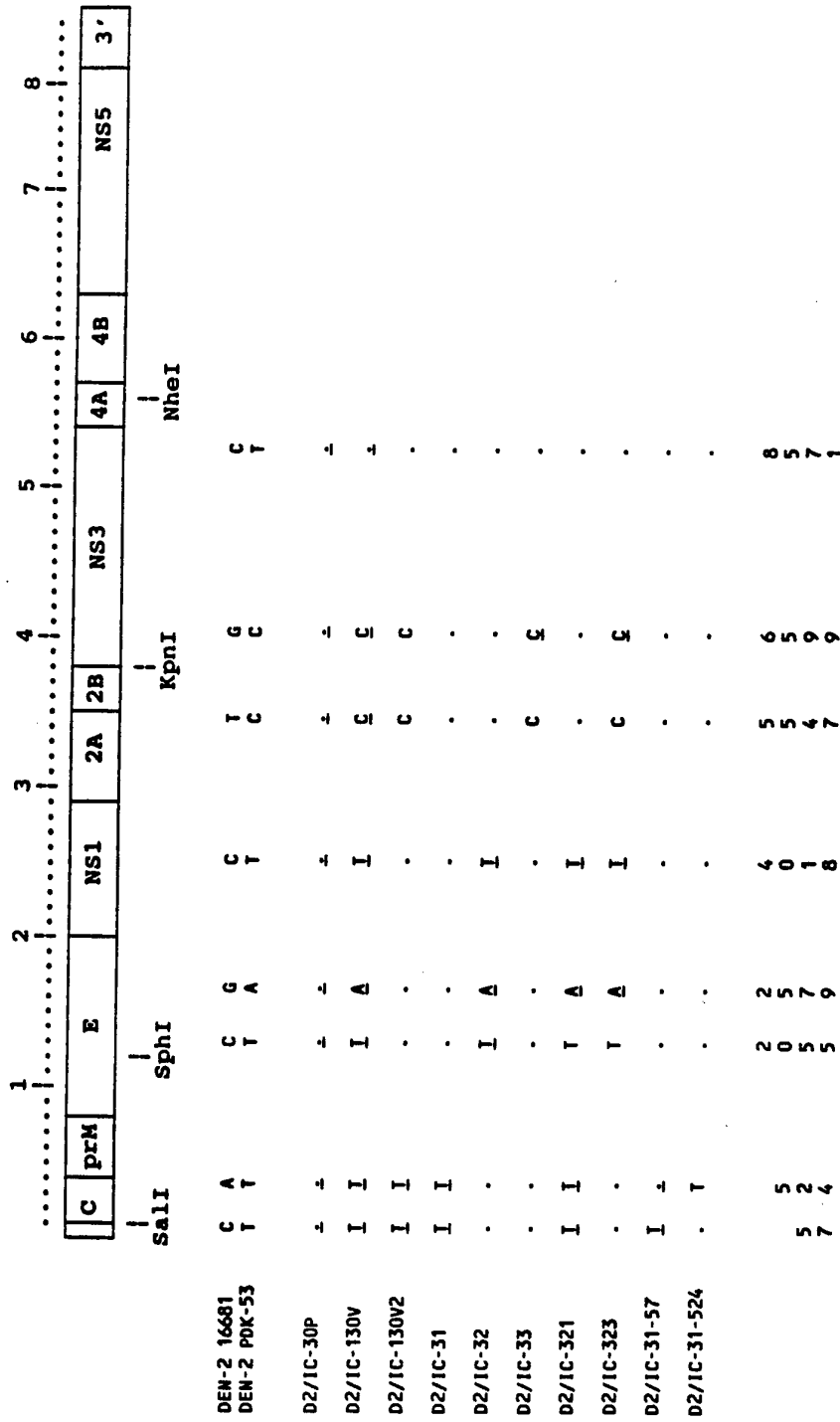


FIGURE 29

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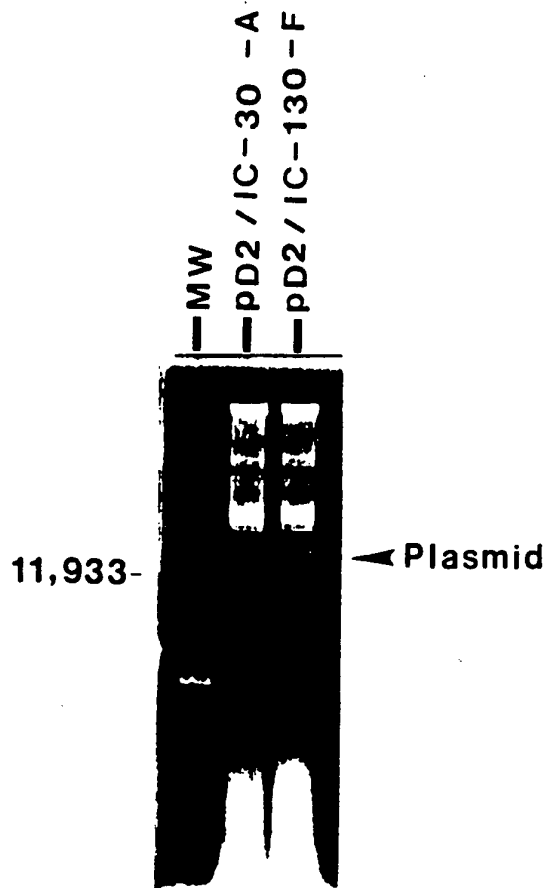


FIGURE 30

EcoRI MluI T7 Promoter
 Amplimer D2-SMT71 = GCAGTGAATTCGAGCTCACGCCTAATTTAATACGACTCACTATA
 Sati

5'-NC
 10 20 30 40 50 60 70 80 90
 AGTGTGATGCTACGTCGACCCGACAAAGACAGATTCCTTTGAGGGAGCTAAGCTCAACGCTAGTCTCTAACAGTTTTTTAATTAGACAGCAGATCTCTGATGA
 D2
 D1
 D3
 D4

Capsid
 110 120 130 140 150 160 170 180 190 200
 ATAAACCAACGGAAAGGCGAATAACACGCTTTCAATATGCTGAACCGGAGAAACCGGCTGTCAGCTGTGCAACAGCTGACAAAGAGATTTCTCACT
 N Q R K K A K N T P F N M L K R E R N R V S T V Q Q L T K R F S L
 D2
 D1
 D3
 D4

210 220 230 240 250 260 270 280 290 300
 TGGATGCTGCAGGGACGAGGACCATTAARAACCTGTTCAATGGCCCTGGTGGCGTTCCTTCGTTTCCCTAACAATCCCACCAACAGCAGGGATATTGAAGAGA
 G M L Q G R G P L K L F M A L V A F L R F L T I P P T A G I L K R
 D2
 D1
 D3
 D4

...TC.....

FIGURE 31A

1410 1420 1430 1440 1450 1460 1470 1480 1490 1500
D2 AGGAACAATGGCAAGGAAATCAAAATACACCACAGAGTTCCATCACAGAAGCAGAATTGACAGGTTATGGCAGCTGTCAACAATGGAGTGTCTCCAGA
G K H G K E I K I T P Q S S I T E A E L T G Y G T V T M E C S P R

1510 1520 1530 1540 1550 1560 1570 1580 1590 1600
D2 ACGGCTCGACTTCAATGAGATGGTGTGTCAGATGGAAATAAAGCTGGTGGTGCACAGGCAATGGTTCCCTAGACCTGCCGTTACCATGGTTGC
T G L D F N E M V L L Q M E N K A W L V H R Q W F L D L P L P W L P

1610 1620 1630 1640 1650 1660 1670 1680 1690 1700
D2 CCGGACGGACACACAGGGTCAAAATGGATACAGAAGAGACATGGTCACTTTCAAAATCCCCTATCGAAGAACAAGGATGTTGTTGTTTAGGATC
G A D T Q G S N W I Q K E T L V T F K N P H A K K Q D V V V L G S

1710 1720 1730 1740 1750 1760 1770 1780 1790 1800
D2 CCAAGAAGGGCCATGCACACAGCCTTACAGGGCCACAGAAATCCAAATGTATCAGGAAACTTACTTTCACAGGACATCTCAAGTGCAGGCTGAGA
Q E G A M H T A L T G A T E I Q M S S G N L L F T G H L K C R L R

1410 1420 1430 1440 1450 1460 1470 1480 1490 1500
D2 AGGAACAATGGCAAGGAAATCAAAATACACCACAGAGTTCCATCACAGAAGCAGAATTGACAGGTTATGGCAGCTGTCAACAATGGAGTGTCTCCAGA
G K H G K E I K I T P Q S S I T E A E L T G Y G T V T M E C S P R

1510 1520 1530 1540 1550 1560 1570 1580 1590 1600
D2 ACGGCTCGACTTCAATGAGATGGTGTGTCAGATGGAAATAAAGCTGGTGGTGCACAGGCAATGGTTCCCTAGACCTGCCGTTACCATGGTTGC
T G L D F N E M V L L Q M E N K A W L V H R Q W F L D L P L P W L P

1610 1620 1630 1640 1650 1660 1670 1680 1690 1700
D2 CCGGACGGACACACAGGGTCAAAATGGATACAGAAGAGACATGGTCACTTTCAAAATCCCCTATCGAAGAACAAGGATGTTGTTGTTTAGGATC
G A D T Q G S N W I Q K E T L V T F K N P H A K K Q D V V V L G S

1710 1720 1730 1740 1750 1760 1770 1780 1790 1800
D2 CCAAGAAGGGCCATGCACACAGCCTTACAGGGCCACAGAAATCCAAATGTATCAGGAAACTTACTTTCACAGGACATCTCAAGTGCAGGCTGAGA
Q E G A M H T A L T G A T E I Q M S S G N L L F T G H L K C R L R

1410 1420 1430 1440 1450 1460 1470 1480 1490 1500
D2 AGGAACAATGGCAAGGAAATCAAAATACACCACAGAGTTCCATCACAGAAGCAGAATTGACAGGTTATGGCAGCTGTCAACAATGGAGTGTCTCCAGA
G K H G K E I K I T P Q S S I T E A E L T G Y G T V T M E C S P R

1510 1520 1530 1540 1550 1560 1570 1580 1590 1600
D2 ACGGCTCGACTTCAATGAGATGGTGTGTCAGATGGAAATAAAGCTGGTGGTGCACAGGCAATGGTTCCCTAGACCTGCCGTTACCATGGTTGC
T G L D F N E M V L L Q M E N K A W L V H R Q W F L D L P L P W L P

1610 1620 1630 1640 1650 1660 1670 1680 1690 1700
D2 CCGGACGGACACACAGGGTCAAAATGGATACAGAAGAGACATGGTCACTTTCAAAATCCCCTATCGAAGAACAAGGATGTTGTTGTTTAGGATC
G A D T Q G S N W I Q K E T L V T F K N P H A K K Q D V V V L G S

1710 1720 1730 1740 1750 1760 1770 1780 1790 1800
D2 CCAAGAAGGGCCATGCACACAGCCTTACAGGGCCACAGAAATCCAAATGTATCAGGAAACTTACTTTCACAGGACATCTCAAGTGCAGGCTGAGA
Q E G A M H T A L T G A T E I Q M S S G N L L F T G H L K C R L R

FIGURE 31E

1810 1820 1830 1840 1850 1860 1870 1880 1890 1900
D2 ATGGACAAAGCTACAGCTCAAGGAATGTCATCTATGTGCACAGAAAGTTTAAAGTTGTAAGGAAATAGCAGAAACACAAACATGGAAACAATAGTTA
M D K L Q L K G H S Y S M C T G K F K V V K E I A E T Q H G T I V I

1910 1920 1930 1940 1950 1960 1970 1980 1990 2000
D2 TCAGAGTGCATATGAAGGGACGGCTCTCCATGCAAGATCCCTTTGAGATAATGGATTTGGAAAAGAACATGTCTTAGGTCGCCCTGATACAGTCAA
R V Q Y E G D G S P C K I P F E I M D L E K R H V L G R L I T V N

2010 2020 2030 2040 2050 2060 2070 2080 2090 2100
D2 CCCAATTGTGACAGAAAAGATAGCCAGTCAACATAGAGCAGACCCCTCCATTCGGAGACAGCTACATCATCATAGAGTAGAGCCGGGACAACTGAAG
P I V T E K D S P V N I E A E P P F G D S Y I I I G V E P G Q L K

2110 2120 2130 2140 2150 2160 2170 2180 2190 2200
D2 CTCRACTGGTTAAGAAGGAAAGTCTATCGGCCAAATGTTGAGACAAATGAGGGGGGCGGAGAGAAATGGCCATTTAGGTGACACAGCCCTGGGATT
L N W F K K G S S I G Q H F E T T H R G A K R M A I L G D T A W D F

1810 1820 1830 1840 1850 1860 1870 1880 1890 1900
D2 ATGGACAAAGCTACAGCTCAAGGAATGTCATCTATGTGCACAGAAAGTTTAAAGTTGTAAGGAAATAGCAGAAACACAAACATGGAAACAATAGTTA
M D K L Q L K G H S Y S M C T G K F K V V K E I A E T Q H G T I V I

1910 1920 1930 1940 1950 1960 1970 1980 1990 2000
D2 TCAGAGTGCATATGAAGGGACGGCTCTCCATGCAAGATCCCTTTGAGATAATGGATTTGGAAAAGAACATGTCTTAGGTCGCCCTGATACAGTCAA
R V Q Y E G D G S P C K I P F E I M D L E K R H V L G R L I T V N

2010 2020 2030 2040 2050 2060 2070 2080 2090 2100
D2 CCCAATTGTGACAGAAAAGATAGCCAGTCAACATAGAGCAGACCCCTCCATTCGGAGACAGCTACATCATCATAGAGTAGAGCCGGGACAACTGAAG
P I V T E K D S P V N I E A E P P F G D S Y I I I G V E P G Q L K

2110 2120 2130 2140 2150 2160 2170 2180 2190 2200
D2 CTCRACTGGTTAAGAAGGAAAGTCTATCGGCCAAATGTTGAGACAAATGAGGGGGGCGGAGAGAAATGGCCATTTAGGTGACACAGCCCTGGGATT
L N W F K K G S S I G Q H F E T T H R G A K R M A I L G D T A W D F

1810 1820 1830 1840 1850 1860 1870 1880 1890 1900
D2 ATGGACAAAGCTACAGCTCAAGGAATGTCATCTATGTGCACAGAAAGTTTAAAGTTGTAAGGAAATAGCAGAAACACAAACATGGAAACAATAGTTA
M D K L Q L K G H S Y S M C T G K F K V V K E I A E T Q H G T I V I

1910 1920 1930 1940 1950 1960 1970 1980 1990 2000
D2 TCAGAGTGCATATGAAGGGACGGCTCTCCATGCAAGATCCCTTTGAGATAATGGATTTGGAAAAGAACATGTCTTAGGTCGCCCTGATACAGTCAA
R V Q Y E G D G S P C K I P F E I M D L E K R H V L G R L I T V N

2010 2020 2030 2040 2050 2060 2070 2080 2090 2100
D2 CCCAATTGTGACAGAAAAGATAGCCAGTCAACATAGAGCAGACCCCTCCATTCGGAGACAGCTACATCATCATAGAGTAGAGCCGGGACAACTGAAG
P I V T E K D S P V N I E A E P P F G D S Y I I I G V E P G Q L K

2110 2120 2130 2140 2150 2160 2170 2180 2190 2200
D2 CTCRACTGGTTAAGAAGGAAAGTCTATCGGCCAAATGTTGAGACAAATGAGGGGGGCGGAGAGAAATGGCCATTTAGGTGACACAGCCCTGGGATT
L N W F K K G S S I G Q H F E T T H R G A K R M A I L G D T A W D F

FIGURE 31F

2510 2520 2530 2540 2550 2560 2570 2590 2600
 D2 CATGGACAGAACAAATCAAGTCCAAAGTCCCGCTTCAAACTAGCTTCAGCTATCCAGAAAGCCCATGAAAGGGGCATTTGTGGAATCCGCTCAGT
 W T E Q Y K F Q P E S P S K L A S A I Q K A H E E G I C G I R S V
 A
 D

D2 CATGGACAGAACAAATCAAGTCCAAAGTCCCGCTTCAAACTAGCTTCAGCTATCCAGAAAGCCCATGAAAGGGGCATTTGTGGAATCCGCTCAGT
 D1 .T.....G.....A.....GG.T..C.....CAAG.G....T.AG....C..TGG...G..ATGG..G....GTG.G
 D3
 D4

HpaI
 2610 2620 2630 2640 2650 2660 2670 2680 2690 2700
 AACAACTGGAGAACTGATGTGGAACCAATCAACCAACCAATGGAATGCAATTCATCAGAAATGAGGTGAACTTACTATTATGACAGGAGACATC
 T R L E N L M W K Q I T P E L N H I L S E N E V K L T I M T G D I

FIGURE 31H

D2	510	520	530	540	550	560	570	580	590	600			
D1	LPGADTQGSNWIQK	ETLVTFKPNPHAKKQDVVVLG	SOEGAMHTALTCATEIQ	SSGNLLFTGHLKCRRLRMDK	LQKGMYSMCTGKFKVVK	EIAETQHG	TI						
D3	TS..L.SQET.NRQDL	..TA.....E.....T.GITTI.A.....	K.....T.....	V.....S..LE..V.....								
D4	TA....SEVH.NY..RM	..V.....R....T.....	S..A.....VDSGD..HM.A.....	KV..E..RI.....T..S.....	SID..M.....T								
D2	610	620	630	640	650	660	670	680	690	700			
D1	VIRVOYEGDGPCKIP	PEIMDLKRRHVLGR	LITVNP	IVTEKDS	SPVNI	EAPPF	GDSYIIIG	VEP	QKLN	WFKKSS	IGOMFET	THRGAKR	MAILGDTAW
D3	L.K.E.K.KDASTE.GQK	AHN.....A.V..K.EE.....	E.N.V..IGDKA..I.Y.....	K...A.A...R.....								
D4	.VK.K...A.A...V.I..R.VN.KK.V..I.SST.LAENTN.AT	..L.....V.....GNSA.T.H.R.....	K...S.Y.....E....										
D2	710	720	730	740	750	760	770	790	800				
D1	DFGSLGGVFTSIGKALHQV	FGAIFYGAARFSGVSW	TKILIGVITWIGMNSR	STLSVTLV	LVGIVTLV	LGVMVQADSGCVVSW	KNKELKCGS	GIFITD	NV				
D3	...I.....M..LV	...TA..VL.....G..ILL..L.L..N.....	MCIA..M.....IN..GR.....	V..NE.									
D4	...V..LN.L..MV..I..SA.T.L.G..M..G..LL..L..KN..M.FSCIAI..I.....	AV...M..IN..G.....											
D2	810	820	830	840	850	860	870	880	900				
D1	HTWTEQYKFP	ESPKLASAIOKAHEED	ICGIRSVTRLENLHWKQITPEL	NHILSENEV	KLTIMTGD	IKIMQAGK	RSRLR	PQPT	ELKYSWKTW	GKAKHLS			
D3AD..KR.SA..G..W..GV												
D4													

NS1

FIGURE 32B

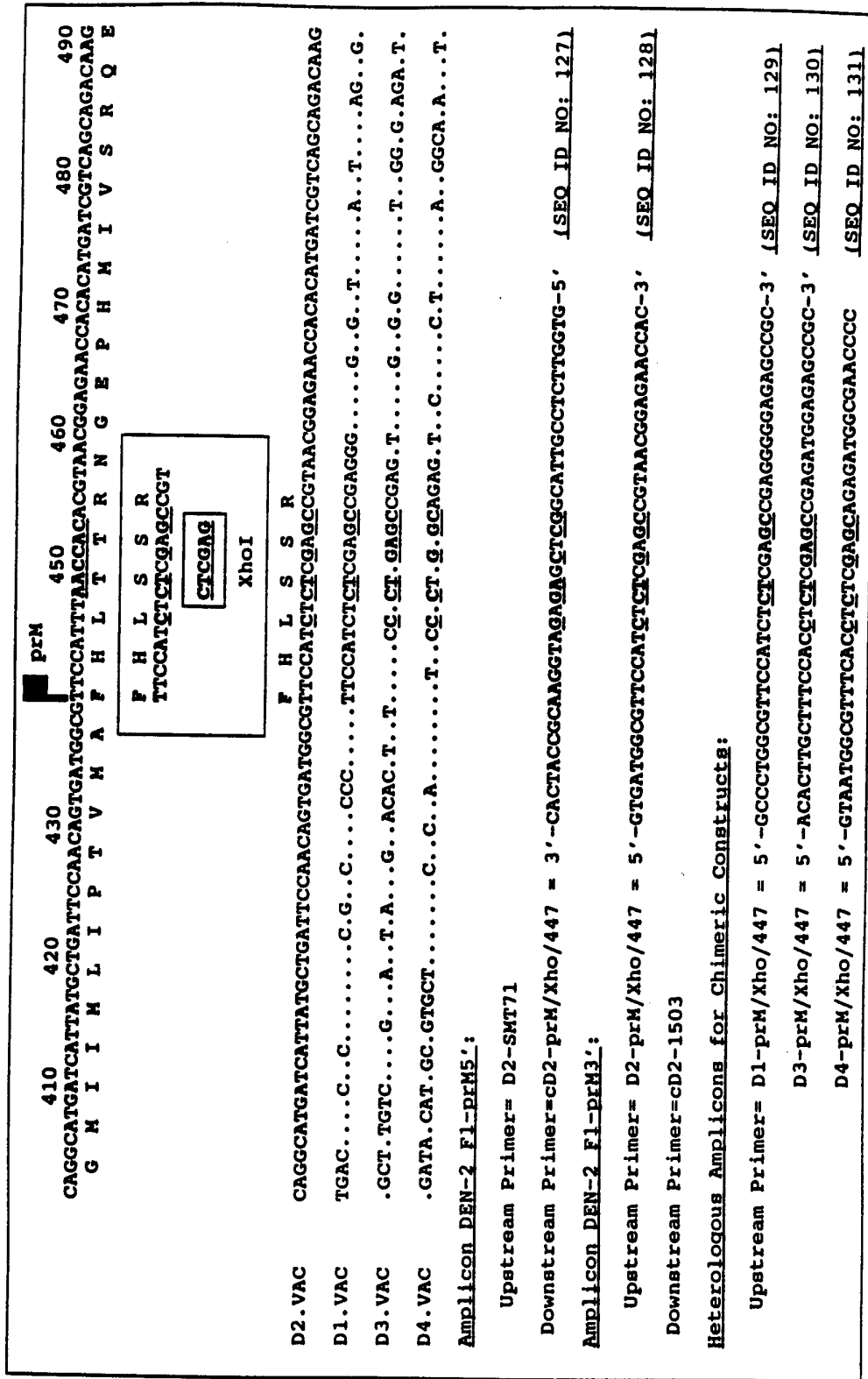


FIGURE 33

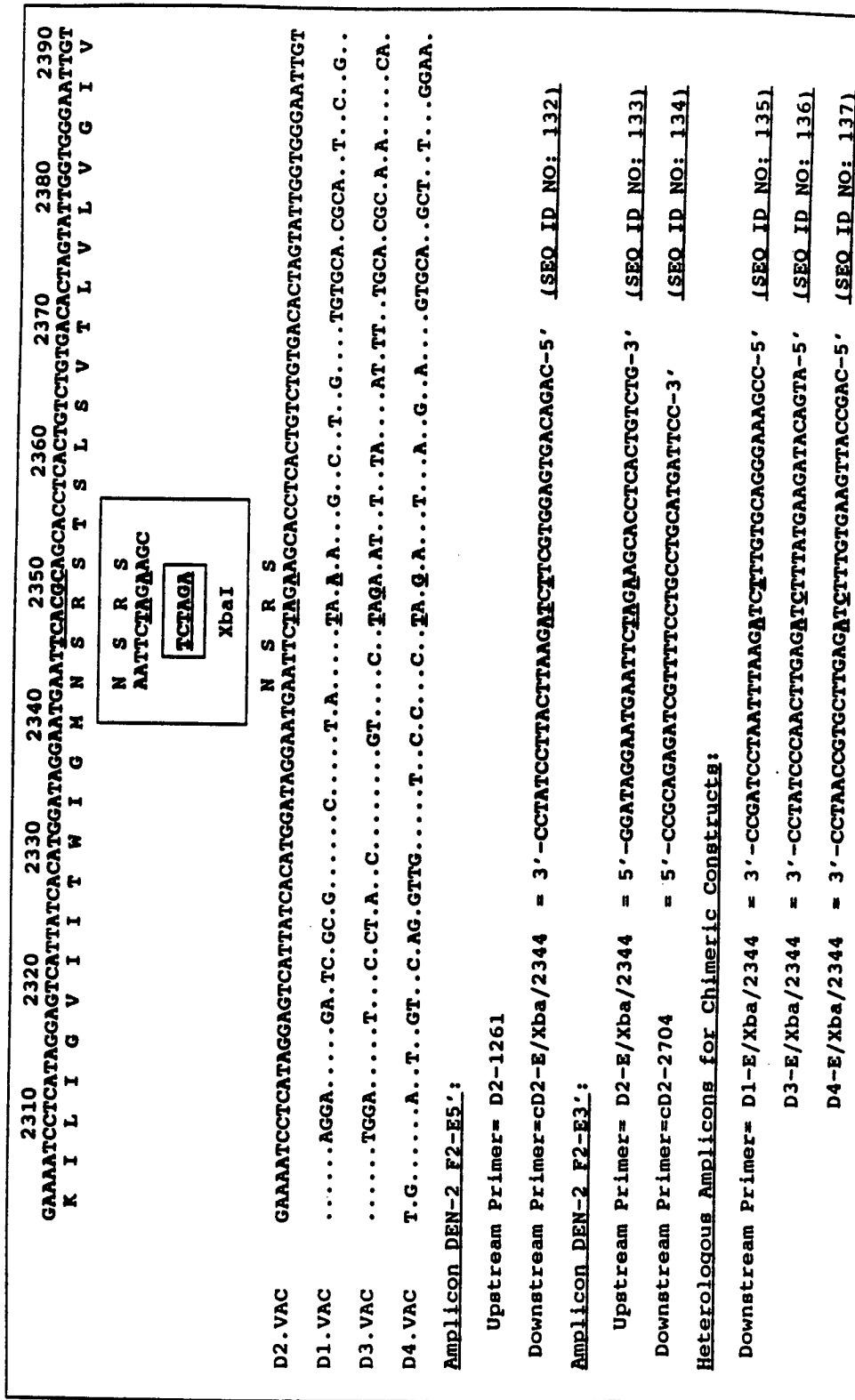


FIGURE 34A

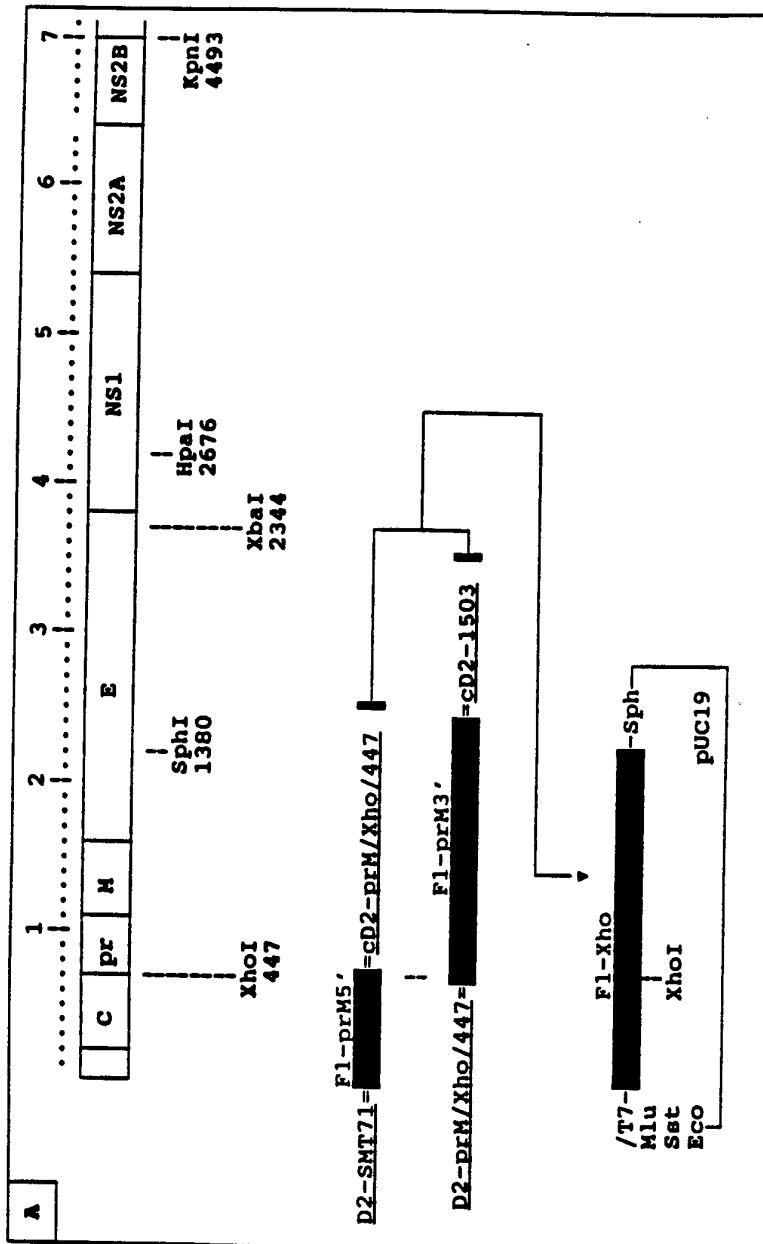


FIGURE 35A

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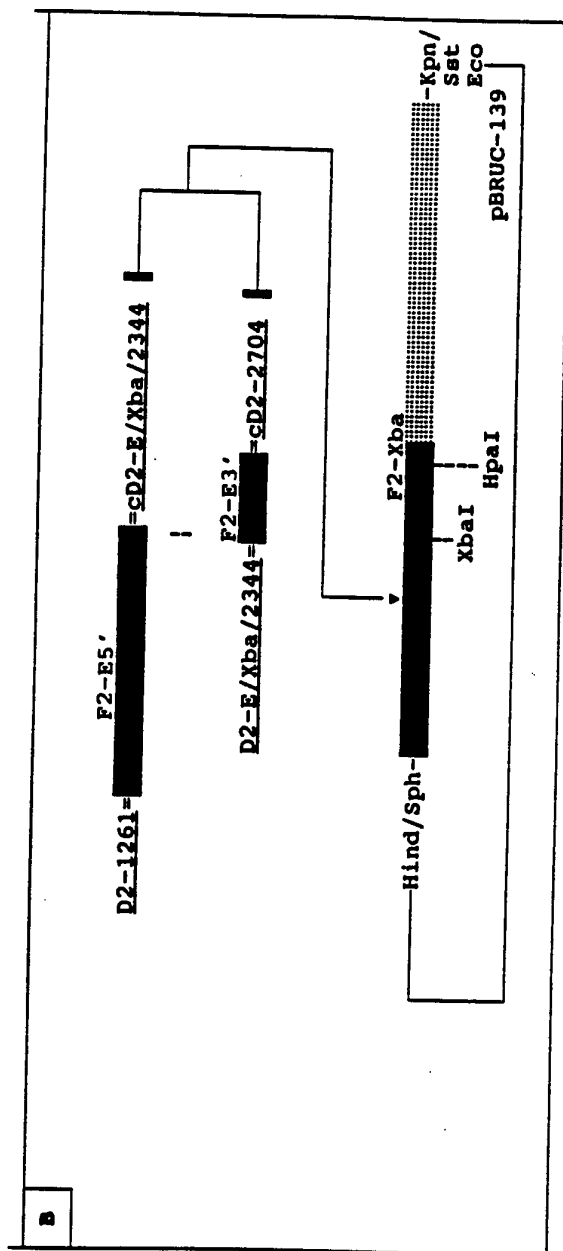


FIGURE 35B

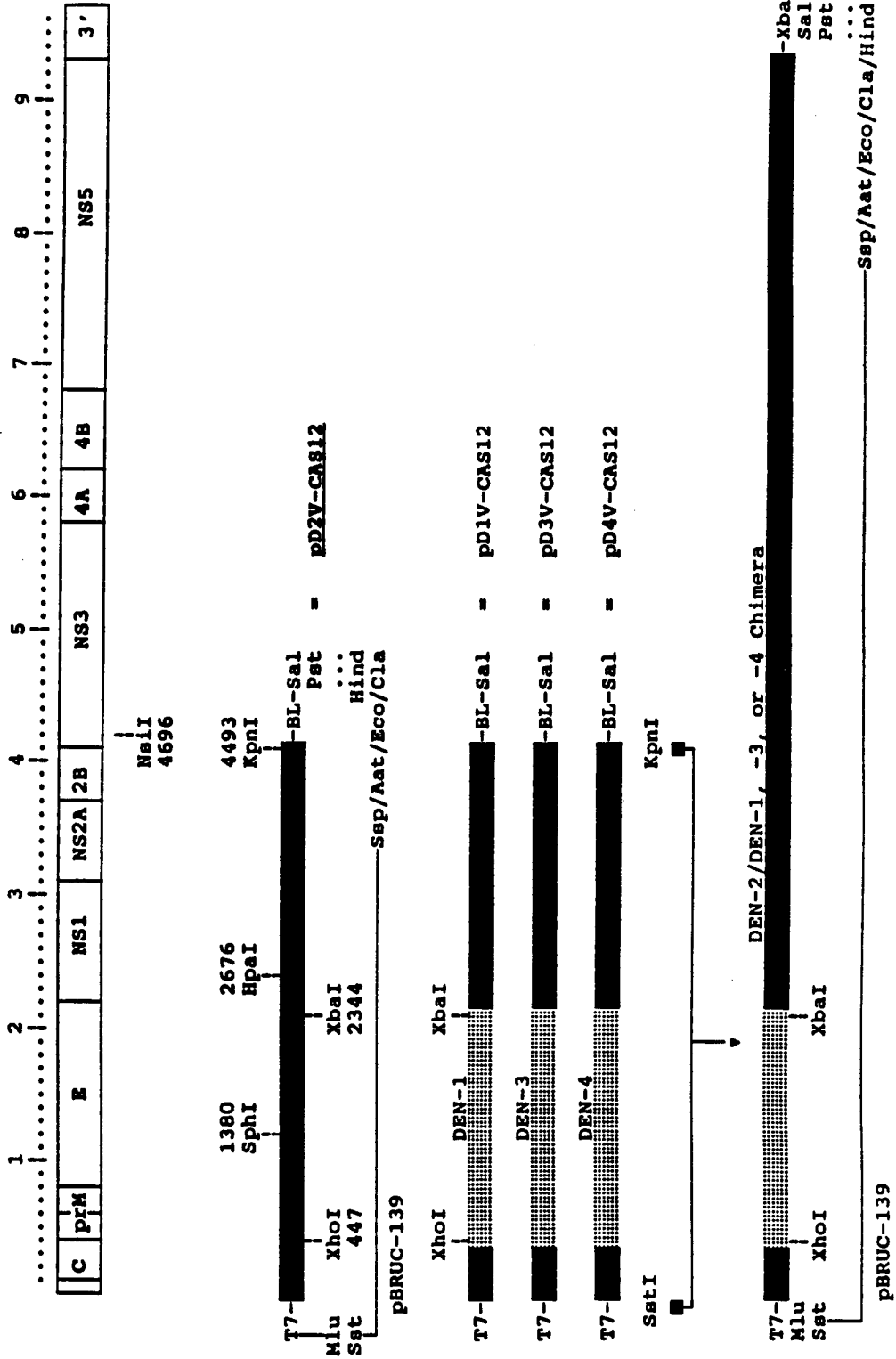


FIGURE 36

INTERNATIONAL SEARCH REPORT

International Application No
PLI/US 96/09209

<p>A. CLASSIFICATION OF SUBJECT MATTER IPC 6 C12N15/40 C12N15/86 C07K14/18 A61K39/12 C12N7/01 C12N7/00 C12N5/10</p>		
<p>According to International Patent Classification (IPC) or to both national classification and IPC</p>		
<p>B. FIELDS SEARCHED</p>		
<p>Minimum documentation searched (classification system followed by classification symbols) IPC 6 C07K C12N A61K</p>		
<p>Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched</p>		
<p>Electronic data base consulted during the international search (name of data base and, where practical, search terms used)</p>		
<p>C. DOCUMENTS CONSIDERED TO BE RELEVANT</p>		
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
P,X	VACCINE, vol. 14, no. 4, March 1996, GUILDFORD GB, pages 329-336, XP000579824 VAUGHN, D.W. ET AL.: "Testing of a Dengue 2 live-attenuated vaccine (strain 16681 PDK 53) in ten american volunteers" see the whole document ---	1
X	VIROLOGY, vol. 187, no. 4, April 1992, ORLANDO US, pages 573-590, XP000601641 BLOK, J. ET AL.: "Comparaison of Dengue -2 virus and its candidate vaccine derivative: sequence relationships with the Flaviviruses and other viruses" see the whole document ---	7-12, 22-29,36
Y	---	1
	-/--	
<p><input checked="" type="checkbox"/> Further documents are listed in the continuation of box C. <input checked="" type="checkbox"/> Patent family members are listed in annex.</p>		
<p>* Special categories of cited documents :</p> <p>"A" document defining the general state of the art which is not considered to be of particular relevance</p> <p>"E" earlier document but published on or after the international filing date</p> <p>"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)</p> <p>"O" document referring to an oral disclosure, use, exhibition or other means</p> <p>"P" document published prior to the international filing date but later than the priority date claimed</p> <p>"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention</p> <p>"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone</p> <p>"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.</p> <p>"&" document member of the same patent family</p>		
<p>Date of the actual completion of the international search</p> <p style="text-align: center;">6 September 1996</p>		<p>Date of mailing of the international search report</p> <p style="text-align: center;">23. 10. 96</p>
<p>Name and mailing address of the ISA</p> <p>European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+ 31-70) 340-2040, Tx. 31 651 epo nl, Fax: (+ 31-70) 340-3016</p>		<p>Authorized officer</p> <p style="text-align: center;">Chambonnet, F</p>

INTERNATIONAL SEARCH REPORT

International Application No
PCT/US 96/09209

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	WO,A,93 06214 (US ARMY) 1 April 1993 see claims 40,63-68 ---	1
A	AMERICAN JOURNAL OF TROPICAL MEDICINE AND HYGIENE, vol. 47, no. 4 sup, 1992, pages 99-100, XP000600344 VAUGHN, D.W. ET AL.: "Phase I testing of a dengue-2 live-attenuated vaccine strain 16681 PDK 53 in american volunteers" see the whole document & 41st Annual Meeting of the American Society of Tropical Medicine and Hygiene Washington, USA November 15-19 1992 ---	1
A	WO,A,92 03161 (US GOVERNMENT) 5 March 1992 see the whole document ---	1
A	WO,A,93 22440 (UNIV SINGAPORE ;TAN YIN HWEE (SG); FU JIANLIN (SG); TAN BOON HUAN) 11 November 1993 see the whole document ---	1,2,5,6, 13
A	WO,A,92 03545 (VIROGENETICS CORP) 5 March 1992 see claims 1,9,10,16-23,26; example 13 ---	1
A	VIROLOGY, vol. 174, no. 2, February 1990, ORLANDO US, pages 479-493, XP002012813 RICO-HESSE, R.: "Molecular evolution and distribution of Dengue Viruses type 1 and 2 in nature" see the whole document ---	1
A	JOURNAL OF GENERAL VIROLOGY, vol. 69, no. 6, June 1988, pages 1391-1398, XP000600928 GRUENBERG, A. ET AL.: "Partial nucleotide sequence and deduced amino acid sequence of the structural proteins of Dengue virus type 2, New Guinea C and PU0-218 strains" see the whole document ---	1
	-/--	

INTERNATIONAL SEARCH REPORT

International Application No

PCT/US 96/09209

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	<p>VIROLOGY, vol. 162, no. 1, January 1988, ORLANDO US, pages 167-180, XP000600931 HAHN, Y.S. ET AL.: "Nucleotide sequence of Dengue 2 RNA and comparison of the encoded proteins with those of other flaviviruses" see the whole document -----</p>	1

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US 96/09209

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This international search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. Claims Nos.: 6
because they relate to subject matter not required to be searched by this Authority, namely:
Remark: Although this claim is directed to a method of treatment of the human/animal body, the search has been carried out and based on the alleged effects of the compound/composition.
2. Claims Nos.:
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
3. Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
4. No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

- The additional search fees were accompanied by the applicant's protest.
- No protest accompanied the payment of additional search fees.

INTERNATIONAL SEARCH REPORT

Information on patent family members

Int ional Application No PCT/US 96/09209

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO-A-9306214	01-04-93	AU-B- 667836	18-04-96
		AU-B- 2691492	27-04-93
		CA-A- 2116980	01-04-93
		EP-A- 0604566	06-07-94
		JP-T- 6511150	15-12-94
WO-A-9203161	05-03-92	AU-B- 8762591	17-03-92
		US-A- 5494671	27-02-96
WO-A-9322440	11-11-93	AU-B- 4257593	29-11-93
		CA-A- 2134666	11-11-93
		EP-A- 0638122	15-02-95
WO-A-9203545	05-03-92	US-A- 5514375	07-05-96
		AU-B- 657711	23-03-95
		AU-B- 8728791	17-03-92
		GB-A,B 2269820	23-02-94
		JP-T- 6503227	14-04-94