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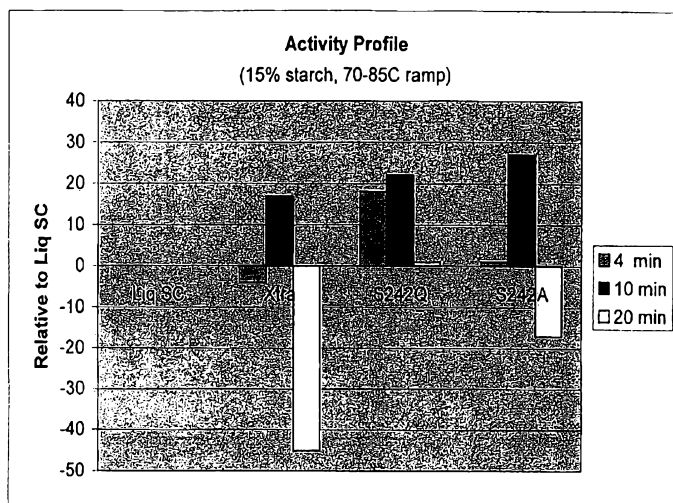


Figure 7

(57) Abstract: Disclosed are compositions comprising variants of alpha-amylase that have alpha-amylase activity and that exhibit altered properties relative to a parent AmyS-like alpha-amylase from which they are derived. The compositions generally comprise at least one of an additional enzyme, a detergent, a surfactant, a chelator, an oxidizing agent, an acidulant, an alkalinizing agent, a source of peroxide, a source of hardness, a salt, a detergent complexing agent, a polymer, a stabilizing agent, or a fabric conditioner. Also disclosed are detergent formulations comprising the variants. Methods of using the compositions for desizing woven material and washing or cleaning items, such as dishes or laundry, are disclosed. Kits related thereto are also provided.

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## **Alpha-Amylase Variants With Altered Properties**

### **SEQUENCE LISTING**

Attached hereto is a sequence listing comprising SEQ ID NOS 1-30, each of  
5 which is herein incorporated by reference in its entirety.

### **CROSS-REFERENCE TO RELATED APPLICATIONS**

This claims benefit of U.S. Provisional Applications 60/985,619, filed November  
5, 2007, 61/026,579, filed February 6, 2008, 61/041,075, filed March 31, 2008, and  
10 61/059,411, filed June 6, 2008, the disclosures of each of which are incorporated herein  
by reference in their entireties, for all purposes.

### **FIELD OF THE DISCLOSURE**

This disclosure relates to novel alpha-amylases. In particular, it relates to  
15 methods of using certain variant alpha-amylase activities, and blends thereof for stain  
removal and as a component of detergent compositions for washing.

### **BACKGROUND**

Alpha-amylases (alpha-1,4-glucan-4-glucohydrolases, E.C. 3.2.1.1) constitute a  
20 group of enzymes that catalyze hydrolysis of starch and related linear or branched 1,4-  
glucosidic oligo- and polysaccharides.

Amylases can be used for a variety of purposes. For examples, amylases are used  
commercially in the initial stages of starch processing (e.g., liquefaction); in wet milling  
processes; and in alcohol production from carbohydrate sources. They are also used as  
25 cleaning agents or adjuncts in detergent matrices; in the textile industry for starch  
desizing; in baking applications; in the beverage industry; in oilfields in drilling  
processes; in recycling processes, e.g. for de-inking paper, and in animal feed.

Attempts have been made to construct alpha-amylase variants with improved  
properties for specific uses, such as starch liquefaction and textile desizing.

There is a need for the creation and improvement of amylases that provide, e.g., manufacturing and/or performance advantages over the industry standard enzymes (e.g., from *Bacillus licheniformis*), for various uses, including commercial desizing as well as cleaning/washing and stain or starch removal processes. There is also a need for  
5 detergents and cleaning aids or formulations comprising improved amylases and additional components, such as surfactant, chelators, and the like.

### SUMMARY

In one aspect the present disclosure relates, inter alia, to novel  $\alpha$ -amylolytic  
10 enzymes variants of parent  $\alpha$ -amylase such as an AmyS-like  $\alpha$ -amylase, in particular variants exhibiting altered properties that are advantageous in connection with the cleaning or washing processes, or the removal of starch, for example in desizing woven material.

For example, the variant is altered, as compared to a parent AmyS-like alpha-  
15 amylase or a reference amylase, in one or more of net charge, substrate specificity, substrate cleavage, substrate binding, thermal stability, activity at one or more pH's, stability at one or more pH's, stability in oxidizing conditions,  $\text{Ca}^{2+}$  requirements, specific activity, catalytic rate, catalytic efficiency, activity in the presence of a chelator, thermal or pH stability in the presence of a chelator, utility for desizing, or utility for a  
20 cleaning process, or amount of expression in a protein expression system, and other properties of interest. For instance, one or more alterations may result in a variant that has reduced  $\text{Ca}^{2+}$  dependency and/or an altered pH/activity profile and/or altered thermostability, as compared to a parent  $\alpha$ -amylase, such as an AmyS-like amylase.

In one aspect, there is provided herein a variant of a parent *Geobacillus*  
25 *stearothermophilus* alpha-amylase, wherein the variant has an amino acid sequence which has at least about 95% homology to a parent *Geobacillus stearothermophilus* alpha-amylase and comprises a substitution of amino acid 242, wherein the amino acid positions in the peptide sequence are numbered relative to a reference amylase (e.g., SEQ ID NO: 1 or 2), and wherein the variant has alpha-amylase activity.

In another aspect, provided are compositions comprising: a) at least one variant alpha-amylase comprising an amino acid sequence at least about 95% identical to that of a parent AmyS-like alpha-amylase, and having a substitution at an amino acid position corresponding to position 242 of a reference alpha-amylase, said variant having detectable alpha-amylase activity, and b) at least one of an additional enzyme, a detergent, a surfactant, a chelator, an oxidizing agent, an acidulant, an alkalizing agent, a source of peroxide, a source of hardness, a salt, a detergent complexing agent, a polymer, a stabilizing agent, or a fabric conditioner. In preferred embodiments, the reference amylase is SEQ ID NO: 1 or 2, and the composition is a component of a product for use in laundry, dish, or hard-surface cleaning, desizing, or fabric or stain treatment.

In one embodiment, the composition comprises an additional enzyme is a protease, a lipase, an amylase, a cellulase, a peroxidase, an oxidase, a pectinase, a lyase, a cutinase, a laccase, or a combination thereof.

In various embodiments, the surfactant is nonionic, anionic, cationic, or zwitterionic. The variant alpha-amylase is preferably a S242Q variant. In some embodiments, the variant has altered stability to oxidation and the variant alpha-amylase further includes deletion or substitution of one or more methionine residues including residues located at amino positions 8, 9, 96, 200, 206, 284, 307, 311, 316, and 438 of a parent AmyS-like alpha-amylase, where the reference alpha-amylase in SEQ ID NO: 2.

In others, the variant alpha-amylase further comprises a sequence modification at one or more amino acid positions corresponding to amino acid positions 97, 179, 180, 193, 319, 349, 358, 416, 428, or 443 of the reference alpha-amylase. In yet other embodiments, the variant comprises one or more of substitution at positions as follows: a cysteine at 349, a cysteine at 428, a glutamic acid at 97, an arginine at 97, a glutamic acid at 319, an arginine at 319, a glutamic acid at 358, an arginine at 358, a glutamic acid at 443, or an arginine at 443.

Also useful herein are variant alpha-amylases comprising a substitution of an N193 or a V416 or both, e.g., a substitution of N193F or V416G, or both. In certain embodiments, the variants feature deletion of one or more amino acids, e.g., at positions F178, R179, G180, I181, G182 and K183.

Preferably, the variant alpha-amylase has altered metal ion dependence or altered stability or activity in an absence of added calcium or a presence of a chelator in certain embodiments.

The variant alpha-amylase preferably has at least 95%, 98%, or even 99% or more homology to SEQ ID NO: 2, and comprises a substitution of amino acid 242 relative to numbering in a reference alpha-amylase comprising SEQ ID NO: 1, and wherein the variant alpha-amylase has alpha-amylase activity.

The parent AmyS-like alpha-amylase is SEQ ID NO: 1, 2, 6, 7, 8, 9, 10, 11, 12, 15, or 16, and the reference alpha-amylase is SEQ ID NO: 1 or 2 in one embodiment.

Preferably, the variant alpha-amylase has improved performance in a wash process at a  $\text{pH} \geq$  about 8, relative to the parent AmyS-like alpha-amylase.

The variant alpha-amylase can feature, in various embodiments, a set of substitutions of a) Q97E, Q319E, Q358E, Q443E; b) Q97E, Q319R, Q358E, Q443R; c) Q97E, Q319R, Q358E; d) Q97E, Q319R, Q443E; e) Q97E, Q319R, Q443R; f) Q97E, Q358R; g) Q97E, Q443E; h) Q319R, Q358E, Q443E; or i) Q319R, Q358R, Q443E.

In another of its several aspects, the disclosure provides compositions that are detergent or cleaning formulations comprising at least one variant amylase comprising an amino acid sequence at least about 95% identical to that of a parent AmyS-like alpha-amylase, and having a substitution at an amino acid position corresponding to position 242 of a reference alpha-amylase, wherein the variant has detectable alpha-amylase activity; wherein the reference amylase is SEQ ID NO: 1 or 2. In some embodiments, the variant is an S242 variant comprising at least a S242Q substitution.

In another of its several aspects, this disclosure provided methods of desizing a woven material subsequent to a weaving process comprising contacting the woven material with a variant alpha-amylase comprising an amino acid sequence at least about 95% identical to that of a parent AmyS-like alpha-amylase, and having a substitution at an amino acid position corresponding to position 242 of a reference alpha-amylase. The variant preferably has detectable alpha-amylase activity. The contacting is performed under conditions and for a time that are effective for at least partially removing sizing from the woven material.

In various embodiments, the variant alpha-amylase is altered, as compared to a parent AmyS-like alpha-amylase or a reference alpha-amylase, in one or more of: (a) net charge, (b) substrate specificity, (c) substrate cleavage, (d) substrate binding, (e) thermal stability, (f) activity at one or more pH's, (g) stability at one or more pH's, (h) stability in oxidizing conditions, (i)  $\text{Ca}^{2+}$  requirements, (j) specific activity, (k) catalytic rate, (l) catalytic efficiency,

(m) activity in a presence of a chelator, (n) thermal or pH stability in the presence of a chelator, (o) effectiveness for desizing, or (p) amount of expression in a protein expression system.

The parent AmyS-like alpha-amylase is SEQ ID NO: 1, 2, 6, 7, 8, 9, 10, 11, 12, 15, or 16, and the reference alpha-amylase is SEQ ID NO: 1 or 2 in various embodiments. Preferably, the variant alpha-amylase is a S242Q variant.

In certain embodiments, the variant alpha-amylase further comprises one or more of substitution at positions as follows: a cysteine at 349, a cysteine at 428, a glutamic acid at 97, an arginine at 97, a glutamic acid at 319, an arginine at 319, a glutamic acid at 358, an arginine at 358, a glutamic acid at 443, or an arginine at 443, wherein the reference alpha-amylase is SEQ ID NO: 1 or 2.

Methods of washing or cleaning are also provided. The methods comprise contacting one or more items to be washed or cleaned with a composition comprising a variant alpha-amylase comprising an amino acid sequence at least about 95% identical to that of a parent AmyS-like alpha-amylase, and having a substitution at an amino acid position corresponding to position 242 of a reference alpha-amylase. The contacting is performed under conditions and for a time effective for at least partially washing or cleaning the one or more items. The variant has detectable alpha-amylase activity. In exemplary methods, at least one item is soiled with at least one starch-containing material, the removal of which is aided by the variant amylase. In various embodiments of these methods, the composition further comprises one or more of an additional enzyme, a detergent, a surfactant, a chelator, an oxidizing agent, an acidulant, an alkalizing agent, a source of peroxide, a source of hardness, a salt, a detergent complexing agent, a polymer, a stabilizing agent, or a fabric conditioner.

In one embodiment of the methods, the parent AmyS-like alpha-amylase is SEQ ID NO: 1, 2, 6, 7, 8, 9, 10, 11, 12, 15, or 16, and the reference alpha-amylase is SEQ ID NO: 1 or 2. Preferably, the variant alpha-amylase is a S242Q variant.

In various embodiments, the variant alpha-amylase has improved performance in a wash process at a pH  $\geq$  about 8, relative to the parent AmyS-like alpha-amylase.

In one embodiment, the variant alpha-amylase comprises one or more of substitution at positions as follows: a cysteine at 349, a cysteine at 428, a glutamic acid at 97, an arginine at 97, a glutamic acid at 319, an arginine at 319, a glutamic acid at 358, an arginine at 358, a glutamic acid at 443, or an arginine at 443. In others, the variant alpha-amylase comprises a set of



substitutions of a) Q97E, Q319E, Q358E, Q443E; b) Q97E, Q319R, Q358E, Q443R; c) Q97E, Q319R, Q358E; d) Q97E, Q319R, Q443E; e) Q97E, Q319R, Q443R; f) Q97E, Q358R; g) Q97E, Q443E; h) Q319R, Q358E, Q443E; or i) Q319R, Q358R, Q443E.

The method can also comprise use of variant alpha-amylases comprising deletion of one or more amino acids at positions F178, R179, G180, I181, G182, or K183.

In certain embodiments, the variant alpha-amylase has altered metal ion dependence or altered stability, or activity in an absence of added calcium or the presence of a chelator.

Also provided herein are kits comprising a) one or more variant alpha-amylases comprising an amino acid sequence at least about 95% identical to that of a parent AmyS-like alpha-amylase, and having a substitution at an amino acid position corresponding to position 242 of a reference alpha-amylase, said variant having detectable alpha-amylase activity, and b) at least one of an additional enzyme, a detergent, a surfactant, a chelator, an oxidizing agent, an acidulant, an alkalizing agent, a source of peroxide, a source of hardness, a salt, a detergent complexing agent, a polymer, a stabilizing agent, or a fabric conditioner.

In one embodiment, the kit further comprises instructions for use, e.g., for using the kit components in a process for desizing a woven material, or for washing or cleaning one or more items soiled with a starch-containing substance.

These and other features of the disclosure will be described in more detail below.

## 5 **BRIEF DESCRIPTION OF THE DRAWINGS**

Figure 1 shows alignment of amino acid sequences among several candidate parent alpha-amylases (AmyS-like amylases) for use herein. Positions corresponding to any amino acid position (e.g., 1 through 520) of the amylase from *Geobacillus stearothermophilus* (SEQ ID NO: 1) can be readily determined. SEQ ID NO: 1, alpha-amylase from *G. stearothermophilus* "BSG"; SEQ ID NO: 2, truncated amylase from *G. stearothermophilus* (AmyS, SPEZYME XTRA); SEQ ID NO: 3, *G. stearothermophilus* (S242A variant amylase); SEQ ID NO: 4, *G. stearothermophilus* (S242Q variant amylase); SEQ ID NO: 5, *G. stearothermophilus* (S242E variant amylase); SEQ ID NO: 6, Yamane 707 amylase; SEQ ID NO: 7, mature LAT amylase; SEQ ID NO: 8, *Bacillus licheniformis* wild-type amylase [TERMAMYL (NOVOZYMES) = SEQ ID NO: 8 in WO 02/10355A2]; SEQ ID NO: 9, *B. amyloliquefaciens* amylase, BAN; SEQ ID NO: 10, STAINZYME = AA560 which is SEQ ID NO: 2 in WO 0060060 or SEQ ID NO: 24 in US 6,528,298; SEQ ID NO: 11, *B. halmapalus* amylase (NATALASE); SEQ ID NO: 12, KSM-1378 (KAO CORP., SEQ ID NO: 3 in EP1199356); SEQ ID NO: 13, *Bacillus* spp. KSM-K38 (KAO CORP., SEQ ID NO: 4 in US 6,403,355 B1); SEQ ID NO: 14, *Bacillus* spp. KSM-K36 (KAO CORP., SEQ ID NO: 2 in US 6,403,355 B1); SEQ ID NO: 15, LIQUOZYME SC (NOVOZYMES); and SEQ ID NO: 16, Consensus Parent Alpha-Amylase Sequence #1.

Figure 2 shows the pHPLT-AmyS plasmid.

25 Figure 3 shows percent residual activity of S242 variants after heat stress at 95°C for 30 minutes. Variant positions P, S, W, and Y were missing and replaced by wild-type AmyS (Spezyme® Xtra (labeled "Z")). A positive control, *G. stearothermophilus* with  $\Delta$ 179-180 with the C-terminus truncated by 29 amino acids (i.e., SEQ ID NO: 2) is also shown. Lines indicate 2 $\times$  and 3 $\times$  above the standard deviation of the percent residual

activity of the wild-type enzyme. S242A and S242Q clearly show higher residual activities than the wild-type.

Figure 4: Panels A, B, C, D, E, F, G, H, and I show pair-wise alignments and consensus sequences for several sequences from Figure 1, and feature, respectively,  
5 Consensus Sequences 2, 3, 4, 5, 6, 7, 8, 9, and 10, or SEQ ID NOs: 22, 23, 24, 25, 26, 27, 28, 29, and 30, respectively.

Figure 5 shows the thermal melting curves and the melting points for the wild-type and amylase variants without added calcium.

Figure 6 shows the thermal melting curves and the melting points in the presence  
10 of 2 mM added calcium for both the wild-type and the amylase variants.

Figure 7 shows the activity profile at 4, 10, and 20 minutes for Spezyme Xtra and two variants, relative to Liquozyme SC.

Figure 8 shows the activity profile of four variants relative to the S242Q variant for three time points.

15 Figure 9 is a graph depicting the performance of S242Q (filled circles) and its variants (open circles), as a function of charge, in the rice starch microswatch assay under North American laundry conditions using S242Q combinatorial charge library, rice starch microswatch cleaning in Tide 2x, at 20°C. Reference is made to Example 10.

Figure 10 is a graph depicting the performance of a truncated *Bacillus sp.* TS-23  
20 amylase (closed circles) with the following mutations: Q98R, M201L, S243Q R309A, Q320R, Q359E, and K444E and its charge variants (open circles) (see co-pending U.S. Patent Application No. PCT/US2008/007103, filed 6 June 2008) in the rice starch microswatch assay as a function of charge under Western European laundry conditions with TS23t combinatorial charge library, rice starch microswatch cleaning in Persil at  
25 40°C. Reference is made to Example 10.

Figure 11 is a graph depicting the performance of S242Q (closed circles) and its variants (open circles) in the BODIPY-starch assay as a function of charge. S242Q combinatorial charge library (CCL), specific activity on BODIPY-starch, standard assay conditions Reference is made to Example 10.

Figure 12: Panel A is a graph depicting the relative BODIPY-starch hydrolysis as a function of relative shake tube expression (i.e., relative BODIPY-starch hydrolysis vs. relative shake tube expression); Panel B is a graph depicting the relative microswatch-starch hydrolysis as a function of relative shake tube expression (i.e., relative microswatch-starch hydrolysis vs. relative shake tube expression). Reference is made to Example 13.

Figure 13: Panel A is a graph depicting the relative shake tube expression as a function of charge; Panel B is a graph depicting the relative BODIPY-starch hydrolysis as a function of charge. Reference is made to Example 13.

Figure 14: Panel A is a graph depicting the relative shake tube expression as a function of charge; Panel B is a graph depicting the relative microswatch cleaning activity as a function of charge. Reference is made to Example 13.

Figure 15 shows the effects of added  $\text{Ca}^{2+}$  on desizing performance of variant S242Q compared to that of Ethyl and Xtra in the LAUNDER-O-METER under conditions of 85°C, for 30 min. at 0.01 ppm active protein. The desizing was performed in the presence of 0 or 5 ppm  $\text{CaCl}_2$ . See Example 14.

Figure 16 shows the effects of added  $\text{Ca}^{2+}$  on desizing performance of variant S242Q compared to that of Ethyl and Xtra in the LAUNDER-O-METER under conditions of 97°C, for 30 min. at 0.01 ppm active protein. The desizing was performed in the presence of 0 or 5 ppm  $\text{CaCl}_2$ . See Example 14.

## **DETAILED DISCLOSURE**

### **1. Definitions & Abbreviations**

In accordance with this disclosure, the following abbreviations and definitions apply. It should be noted that as used herein, the singular forms “a,” “an,” and “the” include plural referents unless the context clearly dictates otherwise. Thus, for example, reference to “a polypeptide” includes a plurality of such polypeptides and reference to “the formulation” includes reference to one or more formulations and equivalents thereof known to those skilled in the art, and so forth.

Unless defined otherwise, all technical and scientific terms used herein have the

same meaning as commonly understood by one of ordinary skill in the art. The following terms are provided below.

### 1.1. Abbreviations

The following abbreviations apply unless indicated otherwise:

5	AATCC	American Association of Textile Chemists and Colorists;
	ADW	automatic dish washing;
	AE	alcohol ethoxylate;
	AEO	alcohol ethoxylate;
	AEOS	alcohol ethoxysulfate;
10	AES	alcohol ethoxysulfate;
	AFAU	acid fungal alpha-amylase units;
	AGU	glucoamylase activity units;
	AOS	$\alpha$ -olefinsulfonate;
	AS	alcohol sulfate;
15	BAA	bacterial alpha-amylase;
	°C	degrees Centigrade;
	CCL	combinatorial charge library;
	cDNA	complementary DNA;
	CMC	carboxymethylcellulose;
20	dE	total color difference, as defined by the CIE-LAB color space;
	dH <sub>2</sub> O	deionized water;
	dIH <sub>2</sub> O	deionized water, Milli-Q filtration;
	DE	Dextrose Equivalent;
	DNA	deoxyribonucleic acid;
25	dNTP	deoxyribonucleotide triphosphates;
	DO	dissolved oxygen;
	DP3	degree of polymerization with three subunits;
	DP <sub>n</sub>	degree of polymerization with n subunits;
	DS (or ds)	dry solids content;
30	DSC	differential scanning calorimetry;

	DTMPA	diethyltriaminepentaacetic acid;
	EC	enzyme commission for enzyme classification;
	EDTA	ethylenediaminetetraacetic acid;
	EDTMPA	ethylenediaminetetramethylene phosphonic acid;
5	EO	ethylene oxide;
	eq	equivalents;
	ETOH	ethanol;
	F&HC	fabric and household care;
	FTU	“fitase” units, phytate hydrolyzing unit;
10	g (or gm)	grams;
	GAU	glucoamylase unit;
	gpg	grains per gallon;
	g/l	grams per liter;
	Genencor	Danisco US Inc, Genencor Division, Palo Alto, CA;
15	H <sub>2</sub> O	water;
	HDG	heavy duty granular detergent;
	HDL	heavy duty liquid detergent;
	HFCS	high-fructose corn syrup;
	HFSS	high-fructose starch-based syrup;
20	HPAEC-PAD	high performance anion exchange chromatography with pulsed amperometric detection;
	hr(s)	hour/hours;
	IKA	IKA Works Inc. 2635 North Chase Parkway SE, Wilmington, NC;
	IPTG	isopropyl β-D-thiogalactoside;
25	JPN	Japan;
	kg	kilograms;
	LA	Luria Agar;
	LAS	linear alkylbenzenesulfonate;
	LB	Luria Broth;
30	LU	Lipase Units;

	M	molar;
	MBD medium	MOPS-based defined medium;
	MES	2-( <i>N</i> -morpholino)ethanesulfonic acid;
	mg	milligrams;
5	min(s)	minute/minutes;
	mL (or ml)	milliliters;
	mm	millimeters;
	mM	millimolar;
	MOPS	3-( <i>N</i> -Morpholino)-propanesulfonic acid;
10	MW	molecular weight;
	NA	North America;
	Ncm	Newton centimeter;
	NEO	neomycin;
	ng	nanogram;
15	nm	nanometer;
	NOBS	nonanoyloxybenzenesulfonate;
	N	Normal;
	NTA	nitrilotriacetic acid;
	PAHBAH	p-hydroxybenzoic acid hydrazide;
20	PCR	polymerase chain reaction;
	PEG	polyethyleneglycol;
	pI	isoelectric point;
	ppm	parts per million;
	PVA	poly(vinyl alcohol);
25	PVP	poly(vinylpyrrolidone);
	RAU	Reference Amylase Units;
	RMS	root mean square;
	RNA	ribonucleic acid;
	rpm	revolutions per minute;
30	SAPU	spectrophotometric acid protease unit;

	SAS	secondary alkane sulfonates;
	1X SSC	0.15 M NaCl, 0.015 M sodium citrate, pH 7.0;
	sec	seconds;
	%SRI	percent stain removal index;
5	SSF	simultaneous saccharification and fermentation;
	TAED	tetraacetylenediamine;
	T <sub>m</sub>	thermal midpoint for a DSC curve, or melting temperature of a protein;
	TNBS	trinitrobenzenesulfonic acid;
10	μg	micrograms;
	μl, (μL)	microliters;
	μNm	microNewton meters;
	μm	micrometer;
	μM	micromolar;
15	U	units;
	V/V	volume to volume;
	WE	Western Europe;
	wt%	weight percent;
	w/v (or W/V)	weight/volume;
20	w/w(or W/w)	weight/weight;
	wt	wild-type.

## 1.2. Definitions

In some aspects, the present disclosure relies on routine techniques and methods used in the field of genetic engineering and molecular biology. The following resources include descriptions of general methodology useful in accordance with what is disclosed herein: Sambrook *et al.*, MOLECULAR CLONING: A LABORATORY MANUAL (2nd Ed., 1989); Kreigler, GENE TRANSFER AND EXPRESSION; A LABORATORY MANUAL (1990) and Ausubel *et al.*, Eds. CURRENT PROTOCOLS IN MOLECULAR BIOLOGY (1994).

30 These general references provide definitions and methods known to those in the



art. Unless defined otherwise herein, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which the disclosure pertains. Singleton, *et al.*, DICTIONARY OF MICROBIOLOGY AND MOLECULAR BIOLOGY, 2D ED., John Wiley and Sons, New York (1994) and Hale & Markham, THE HARPER COLLINS DICTIONARY OF BIOLOGY, Harper Perennial, NY (1991) provide one of skill with general dictionaries of many of the terms used in this disclosure.

“Isolated” means that the isolated substance, e.g. a compound or a sequence, is modified by the hand of man relative to that compound or sequence as found in nature. For example, an isolated sequence is at least partially free, or even substantially free, from at least one other component with which the sequence is naturally associated as found in nature.

“Purified” when used to describe a material or substance means that the material or substance is in a relatively pure state, e.g., at least about 90% pure, at least about 95% pure, at least about 98% pure, or at least about 99% pure.

As used herein, “starch” refers to any carbohydrate composition comprising complex polysaccharides, comprising amylose and/or amylopectin with the formula  $(C_6H_{10}O_5)_x$ , wherein “X” can be any number. Preferably, starch refers to any such carbohydrate that is naturally present in plants, including but not limited to grains, grasses, tubers, and roots, and more specifically from wheat, barley, corn, rye, rice, sorghum, cassava, millet, potato, sweet potato, and tapioca. Starch can also refer to synthetic starches or modified starches, such as chemically-modified starch for use as a detectable substrate for enzyme assays, or starches chemically- or enzymatically-modified to improve one or more properties for use.

As used herein, “phytic acid” (or inositol hexakisphosphate (IP6)), is the principle storage form of phosphorus in many plant tissues, such as bran, seeds, and the like. Phytic acid is also referred to as “phytate” herein, especially when in salt form. Various other inositol phosphates such as inositol penta- (IP5), tetra- (IP4), and triphosphate (IP3) are also referred to herein as phytates. Phytates are generally indigestible by man and most monogastric animals.

Enzymes that degrade phytates are referred to herein as “phytases” or “fytases” are generally myo-inositol-hexaphosphate phosphohydrolases. Phytase activity is defined as fytase units (FTU or U), where one FTU is defined as the quantity of enzyme that liberates 1 micromol of inorganic-P per minute from 0.0015 mol/l sodium phytate at pH 5.5, and 37 °C. This definition provides a useful measure of quantity of phytase activity and represents a simple bench mark measurement. Phytate-degrading enzymes of yeasts (e.g., *Schwanniomyces occidentalis*, *Pichia anomala*, *Arxula adeninivorans*), gram-negative bacteria (e.g., *Escherichia coli*, *Pseudomonas* spp., *Klebsiella* spp.), and gram-positive (e.g., *Bacillus* spp.) have been identified and characterized. Phytases from many plants, and from filamentous fungi such as *Penicillium* spp., *Aspergillus* spp., *Trichoderma* spp. *Mucor piriformis*, and *Cladosporium* spp., are also known. 3-phytases (EC 3.1.3.8) and 6-phytases (EC 3.1.3.26), depending on the site of initiation of hydrolysis, have been characterized. Also, phytase have been characterized, based on their pH “optima,” as either acid (pH optima around 5) or alkaline (pH optima around 9). A variety of commercial phytases are available, including ROVABIO (Genencor International).

“Amylase” refers to an enzyme that is capable of catalyzing the cleavage of a starch substrate, leading to a degradation or partial degradation of the starch. Amylases are generally hydrolases that cleave glycosidic linkages in starch. As used herein amylase includes any glucoamylase, alpha-amylase, beta-amylase, for example, the wild-type alpha-amylases of *Bacillus* spp., especially *B. licheniformis*. Generally, alpha-amylases (EC 3.2.1.1;  $\alpha$ -D-(1→4)-glucan glucohydrolase) are endo-acting enzymes defined as cleaving  $\alpha$ -D-(1→4) O-glycosidic linkages within the starch molecule in a random fashion. In contrast, the exo-acting amylolytic enzymes, such as beta-amylases (EC 3.2.1.2;  $\alpha$ -D-(1→4)-glucan maltohydrolase) and some product-specific amylases like maltogenic alpha-amylase (EC 3.2.1.133) cleave the substrate starch molecule from the non-reducing end. beta-Amylases, alpha-glucosidases (EC 3.2.1.20;  $\alpha$ -D-glucoside glucohydrolase), glucoamylase (EC 3.2.1.3;  $\alpha$ -D-(1→4)-glucan glucohydrolase), and product-specific amylases can produce malto-oligosaccharides of specific length from starch. Wild-type alpha-amylase from *Bacillus stearothermophilus* or “AmyS” amylase

is sometimes referred to herein as XTRA or SPEZYME XTRA, which are commercial AmyS products from Genencor International.

As used herein, "AmyS-like alpha-amylases" are useful as parent amylases herein. AmyS-like alpha-amylases constitute a class of alpha-amylases herein, based on the  
5 substantial homology found between them. "AmyS-like alpha-amylase" is intended to indicate the class of alpha-amylases, in particular *Bacillus* alpha-amylases, especially *Geobacillus* *stearothermophilus* alpha-amylases, which, at the amino acid level, exhibit a substantial identity to the alpha-amylase having the amino acid sequence shown in SEQ ID NO: 2, herein. Spezyme Xtra is commercially available from Danisco US Inc,  
10 Genencor Division. *Geobacillus stearothermophilus* has been referred to as *Bacillus stearothermophilus* in the literature and the two may be used interchangeably herein. All the alpha-amylases having the amino acid sequences provided herein as SEQ ID NOS: 1, 6, 7, 8, 9, 10, 11, 12, 15 and 16, respectively, are considered to be AmyS-like alpha-amylases and thus are suitable as parent alpha-amylases. AmyS-like alpha-amylases also  
15 include alpha-amylases i) having amino acid sequences with at least about 60% homology (identity), such as at least about 70%, at least about 75%, or at least about 80%, at least about 85%, at least about 90%, at least about 95%, at least about 96%, at least about 97%, at least about 98%, or at least about 99% identity, with at least one of the amino acid sequences shown in SEQ ID NOS: 1, 6, 7, 8, 9, 10, 11, 12, 15 and 16,  
20 and/or ii) that are encoded by a DNA sequence that hybridizes with a DNA sequence encoding any of the above-specified alpha-amylases, or those apparent from SEQ ID NOS: 9 (BAN), 5 (BSG), 3 (SP722), 1 (SP690), 7 (LAT), 11 (AA560) of WO 06/002643 or of the present specification, which encode any of the amino acid sequences shown in SEQ ID NOS: 1, 6, 7, 8, 9, 10, 11, 12, 15 and 16 herein, respectively. Still further  
25 homologous alpha-amylases useful as AmyS-like alpha-amylases and thus, as parent enzymes for producing variants herein, include the alpha-amylase produced by the *B. licheniformis* strain described in EP 0252666; (ATCC 27811), and the alpha-amylases identified in WO 91/00353 and WO 94/18314; commercial AmyS-like alpha-amylases are comprised in the products sold under the following tradenames: Spezyme® AA and  
30 ULTRAPHLOW (available from Danisco US Inc, Genencor Division), and Keistase™

(available from Daiwa) and LIQUEZYME SC (available from Novozymes, Denmark). Section 1.5 herein below provides further information regarding AmyS-like alpha-amylases. Table A therein provides a list of several useful AmyS-like alpha-amylases, as well as a convenient method of comparing amino acid sequence identities therebetween.

5 The skilled artisan will appreciate the similar tables can be constructed for other alpha-amylases to determine their suitability for use herein as apparent enzyme.

As used herein, "spectrophotometric acid protease unit" ("SAPU") is a unit of protease enzyme activity, wherein in 1 SAPU is the amount of protease enzyme activity that liberates one micromole of tyrosine per minute from a casein substrate under  
10 conditions of the assay.

"Glucoamylase unit" ("GAU"), is a measure of amylolytic activity defined as the amount of enzyme activity that will produce 1 g of reducing sugar, calculated as glucose, per hour from a soluble starch substrate at pH 4.2 and 60°C).

As used herein, the term "variant" may be used interchangeably with the term  
15 "mutant." "Variants" can refer to either polypeptides or nucleic acids. Variants include substitutions, insertions, deletions, truncations, transversions, and/or inversions, at one or more locations relative to a reference sequence. Variant nucleic acids include sequences that are complementary to sequences that are capable of hybridizing to the nucleotide sequences presented herein. For example, a variant nucleic acid sequence herein can be  
20 at least partially complementary to a sequence capable of hybridizing under stringent conditions (*e.g.*, 50°C and 0.2X SSC {1X SSC = 0.15 M NaCl, 0.015 M sodium citrate, pH 7.0}) to a nucleotide sequences presented herein. More preferably, the term variant encompasses sequences that are complementary to sequences that are capable of hybridizing under highly stringent conditions (*e.g.*, 65°C and 0.1X SSC) to the nucleotide  
25 sequences presented herein.

"Thermostable" when used to describe an enzyme means the enzyme is more thermostable than a reference enzyme. In the present application, an alpha-amylase variant is more thermostable than a wild-type *B. licheniformis* alpha-amylase if the variant has a relatively higher enzymatic activity after a specific interval of time under  
30 the same experimental conditions, *e.g.*, the same temperature, substrate concentration,

etc. Alternatively, a more thermostable enzyme has a higher heat capacity determined by differential scanning calorimetry, compared to a reference enzyme.

“Melting temperature” ( $T_m$ ) of a polypeptide is a temperature at which the conformation of the polypeptide undergoes a measurable temperature-dependent change.

5 Protein conformation and  $T_m$  can be analyzed, for example, by circular dichroism, one of the most general and basic tools to study protein folding. Circular dichroism spectroscopy measures the absorption of circularly polarized light. In proteins, structures such as alpha helices and beta sheets are generally chiral, and thus absorb circularly polarized light. The light absorption provides a measure of the degree of foldedness of  
10 the protein. Changes in this absorption as a function of temperature or concentration of a denaturant can be used to study equilibrium unfolding of the protein. This type of spectroscopy can also be combined with devices, such as stopped flow mixers, to measure kinetics of protein folding/unfolding.

“Calcium dependent” means that, a particular enzyme requires calcium to  
15 substantially exhibit catalytic activity. Generally as used herein, “calcium dependent” encompasses a property of any enzyme that has a strict requirement for a divalent metal ion to exhibit catalytic activity, and also includes enzymes whose catalytic activity is substantially (e.g. more than 20%) increased in the presence of calcium or another divalent cation.

20 As used herein, “pH stable” with respect to an enzyme can refer to the enzyme activity or the protein conformation of the enzyme. In the first sense, “pH stable” means the enzyme remains catalytically-active at a specified pH or across a specified pH range. In the second sense, an enzyme may be deemed “stable” at a pH wherein the protein is not irreversibly denatured. In such a case, the enzyme would become catalytically active  
25 when returned to a pH capable of supporting catalytic activity. pH stability may also be used in a relative or comparative manner, for example, with a reference enzyme. In the present application, an alpha-amylase variant can be more pH stable than a wild-type *B. licheniformis* alpha-amylase when the variant has a relatively higher activity than the wild-type, e.g., when held at a given pH or assayed under the same conditions, including  
30 pH. pH’s of most interest are typically either the conditions of actual use, or pH’s that

are at or near the boundaries or extremes of the enzyme's natural ability to remain catalytically active.

“pH range” means a range of pH values e.g., from more acid to more basic, or vice versa. With respect to an enzyme activity, a pH range indicates the upper and lower pH values at which the enzyme exhibits a specified level of activity- e.g. a minimum activity, a specified percentage of maximal activity, or a specified level of substrate conversion or product formation.

“Recombinant” when used in reference to a cell, nucleic acid, protein, or vector, indicates that the cell, nucleic acid, protein or vector, is the result of, or has been modified by, the introduction of a heterologous sequence or the alteration of a native sequence, or that the cell is derived from a cell so modified or altered. Thus, for example, recombinant cells may express genes that are not found within the native (non-recombinant) form of the cell or may express native genes that are otherwise differently expressed (e.g. under-expressed, or over-expressed), abnormally expressed, or not expressed at all.

As used herein, “nucleotide sequence” or “nucleic acid sequence” refers to any sequence of two or more nucleotides, ribonucleotides, or the like, or derivatives thereof. Nucleotide sequences include oligonucleotide and polynucleotide sequences, as well as variants, homologues, fragments and derivatives thereof. A nucleotide sequence may be single-, double-, or multi-stranded. The nucleotide sequence may be from any source or origin, e.g., genomic, synthetic, or recombinant, and includes genomic DNA, cDNA, synthetic DNA, and RNA, and the like as well as hybrids thereof. Nucleotide sequences may comprise one or more codons and may encode one or more polypeptides. Nucleotide sequences may preferentially assume one or more energetically preferred three-dimensional structures.

A “vector” refers to a nucleotide sequence frequently useful for experimental use *in vitro*, or for introduction of nucleic acids into one or more cell types. Vectors include cloning vectors, *in vivo* or *in vitro* expression vectors, shuttle vectors, plasmids, phagemids, cosmids, phage particles, cassettes and the like.

An “expression vector” as used herein means a DNA construct comprising a DNA

sequence which is operably-linked to a suitable control sequence capable of effecting expression of the DNA in a suitable host. Such control sequences may include a promoter to effect transcription, an optional operator sequence to control transcription, a sequence encoding suitable ribosome binding sites on the mRNA, enhancers and  
5 sequences which control termination of transcription and translation.

A polynucleotide or a polypeptide having a certain percent (*e.g.*, at least about 80%, 85%, 90%, 95%, or 99%) of sequence identity with another sequence means that, when aligned, that percentage of bases or amino acid residues are the same in comparing the two sequences. This alignment and the percent homology or identity can be  
10 determined using any suitable software program known in the art, for example those described in CURRENT PROTOCOLS IN MOLECULAR BIOLOGY (F. M. Ausubel *et al.* (eds) 1987, Supplement 30, section 7.7.18). Such programs may include the GCG Pileup program, FASTA (Pearson *et al.* (1988) *Proc. Natl. Acad. Sci USA* 85:2444–2448), and BLAST (BLAST Manual, Altschul *et al.*, Natl Cent. Biotechnol. Inf., Natl  
15 Lib. Med. (NCIB NLM NIH), Bethesda, Md., and Altschul *et al.*, (1997) *NAR* 25:3389–3402). Another alignment program is ALIGN Plus (Scientific and Educational Software, PA), using default parameters. Another sequence software program that finds use is the TFASTA Data Searching Program available in the Sequence Software Package Version 6.0 (Genetics Computer Group, University of Wisconsin, Madison, WI).

20 One skilled in the art will recognize that sequences encompassed by the disclosure are also defined by the ability to hybridize under stringent hybridization conditions with the exemplified *amyS* sequence (*e.g.*, SEQ ID NO:5 of WO 06/002643). A nucleic acid is hybridizable to another nucleic acid sequence when a single stranded form of the nucleic acid can anneal to the other nucleic acid under appropriate conditions of  
25 temperature and solution ionic strength. Hybridization and washing conditions are well known in the art (*see, e.g.*, Sambrook (1989) *supra*, particularly chapters 9 and 11). In some embodiments, stringent conditions correspond to a  $T_m$  of 65°C and 0.1×SSC, 0.1% SDS.

A “gene” refers to a DNA segment that is involved in producing a polypeptide  
30 and includes regions preceding and following the coding regions as well as intervening

sequences (introns) between individual coding segments (exons).

“Heterologous” with reference to a polynucleotide or protein refers to a polynucleotide or protein that does not naturally occur in a host cell. In some embodiments, the protein is a commercially important industrial protein. It is intended that the term encompass proteins that are encoded by naturally occurring genes, mutated genes, and/or synthetic genes.

“Endogenous” with reference to a polynucleotide or protein refers to a polynucleotide or protein that occurs naturally in the host cell.

As used herein, “transformed”, “stably transformed”, and “transgenic” used in reference to a cell means the cell comprises at least one non-native (*e.g.*, heterologous) nucleic acid sequence. A stably-transformed cell comprises at least one such nucleic acid sequence integrated into its genome, or in an episomal plasmid that is maintained through multiple generations.

As used herein, “expression” refers to the process by which a polypeptide is produced based on the nucleic acid sequence of a gene. The process includes both transcription and translation.

A “signal sequence” means a sequence of amino acids covalently-bound to the N-terminal portion of a protein, which facilitates the transport of the protein, *e.g.*, secretion of the mature form of the protein outside the cell. The definition of a signal sequence is functional. The mature form of the extracellular protein lacks the signal sequence which is cleaved off, *e.g.*, during the secretion process.

As used herein, the term “derived” encompasses the terms “originated from”, “obtained from” or “obtainable from”, and “isolated from”.

The terms “protein” and “polypeptide” are used interchangeably herein. The conventional one-letter or three-letter code for amino acid residues is used herein.

A “promoter” is a regulatory sequence that is involved in binding RNA polymerase to initiate transcription of a gene. The promoter may be an inducible promoter or a constitutive promoter. For example, *cbhl* from *Trichoderma reesei*, an inducible promoter, can be used herein.



“Operably-linked” refers to juxtaposition wherein elements are in an arrangement allowing them to be functionally related, even where not in close physical proximity. For example, a promoter is operably-linked to a coding sequence if it is capable of controlling the coding sequence and does control the transcription of the sequence under conditions  
5 permissive thereof, or conducive thereto.

“Selective marker” refers to a gene capable of expression in a host, and which allows selecting those hosts expressing the marker gene. Examples of selectable markers include but are not limited to gene that provide altered resistance to an antimicrobial agent (*e.g.*, hygromycin, bleomycin, or chloramphenicol) and/or genes that confer  
10 metabolic selectivity, for example, a nutritional advantage on the host cell, such as growth on a particular substrate as a sole source of carbohydrate.

“Introduced” in the context of inserting a nucleic acid sequence into a cell, means “transfection”, or “transformation” or “transduction” and includes reference to the incorporation of a nucleic acid sequence into a eukaryotic or prokaryotic cell wherein the  
15 nucleic acid sequence may be incorporated into the genome of the cell (*e.g.*, chromosome, plasmid, plastid, or mitochondrial DNA), converted into an autonomous replicon, or transiently expressed (*e.g.*, transfected mRNA).

“Host,” “host strain,” or “host cell” means a suitable cell in which to place an expression vector or DNA construct comprising a polynucleotide, *e.g.*, encoding a variant  
20 alpha-amylase. Host strains are preferably bacterial cells. In a preferred embodiment, “host cell” means cells and/or protoplasts created from the cells of a microbial strain, *e.g.*, a *Bacillus* spp.

The term “culturing” refers to growing a population of microbial cells under suitable conditions in a medium capable of supporting such growth. In one embodiment,  
25 culturing refers to fermentative bioconversion of a starch substrate containing granular starch to an end-product (typically in a vessel or reactor).

The term “enzymatic conversion” in general refers to the modification of a substrate by enzyme action. The term as used herein also refers to the modification of a starch substrate by the action of an enzyme.

As used herein the term “saccharification” refers to enzymatic conversion of starch to glucose.

The term “degree of polymerization (DP)” refers to the number (n) of anhydroglucopyranose units in a given saccharide. Examples of DP1 are the  
5 monosaccharides, such as glucose and fructose. Examples of DP2 are the disaccharides, such as maltose and sucrose. A DP>3 denotes polymers with a degree of polymerization of greater than 3. The skilled artisan will understand that compounds with greater DE are more polymeric.

“End-product” or “desired end-product” refer to any intended product of an  
10 enzymatic reaction, e.g. a starch-derived molecule that is enzymatically converted from the starch substrate.

The term “residual starch” refers to any remaining starch (soluble or insoluble) left in a composition after fermentation of a starch-containing substrate.

As used herein, “specific activity” means an enzyme unit defined as the number  
15 of moles of substrate converted to product by an enzyme preparation per unit time under specific conditions. Specific activity is expressed as units (U)/unit weight of protein, generally, U/mg protein.

“Yield” refers to the amount of end-product or desired end-products produced using the methods of the present disclosure. In some embodiments, the yield is greater  
20 than that produced using methods known in the art. In some embodiments, the term refers to the volume of the end product and in other embodiment the term refers to the concentration of the end product.

As used herein, “biologically-active” refers to a compound or sequence that has a measurable effect on a biological system, e.g., a cell, an organ, or an organism.

25 “ATCC” refers to American Type Culture Collection located at Manassas, VA 20108 (ATCC).

“NRRL” refers to the Agricultural Research Service Culture Collection, National Center for Agricultural Utilization Research (and previously known as USDA Northern Regional Research Laboratory), Peoria, Ill.

As used herein, "food" means any ingredient, component or composition that provides a nutritive value for an animal, including a human.

As used herein, by convention, when describing proteins and genes that encode them, the term for the gene is generally italicized, (*e.g.*, the gene that encodes amyL (*B. licheniformis* AA) may be denoted as *amyL*). The term for the protein is generally not  
5 italicized and the first letter is generally capitalized, (*e.g.*, the protein encoded by the *amyL* gene may be denoted as AmyL or amyL). Unless otherwise indicated, nucleic acids are written left to right in 5' to 3' orientation, and amino acid sequences are written left to right in amino to carboxy orientation, respectively.

As used herein the term "comprising" and its cognates are used in their inclusive  
10 sense; that is, equivalent to the term "including" and its corresponding cognates. Numeric ranges are inclusive of the numbers defining the range.

The headings provided herein are not limitations of the various aspects or embodiments of what is disclosed.

Although any methods and materials similar or equivalent to those described  
15 herein can be used in the practice or testing of that which is disclosed, certain presently preferred methods and materials are described with no intention to limit the practitioner to any particular methods, protocols, and reagents described, as these may be varied. All patents and publications, including all sequences disclosed within such patents and  
20 publications, referred to herein are expressly incorporated by reference.

## 2. Nomenclature

In the present description and claims, the conventional one-letter and three-letter  
25 codes for amino acid residues are used. For ease of reference, alpha-amylase variants are generally described by use of the following nomenclature:

Original amino acid(s): position(s): substituted amino acid(s)

According to this nomenclature, for instance the substitution of serine by an alanine in position 242 is shown as:

30 Ser242Ala or S242A

a deletion of alanine in position 30 is shown as:

Ala30\* or A30\* or  $\Delta$ A30

and insertion of an additional amino acid residue, such as lysine, is shown as:

Ala30AlaLys or A30AK

A deletion of a consecutive stretch of amino acid residues, such as amino acid  
5 residues 30-33, is indicated as (30-33)\* or  $\Delta$ (A30-N33).

Where a specific alpha-amylase contains a "deletion" in comparison with other  
alpha-amylases and an insertion is made in such a position this is indicated as:

\*36Asp or \*36D

for insertion of an aspartic acid in position 36.

10 Multiple mutations are separated by plus signs, i.e.:

Ala30Asp+Glu34Ser or A30N+E34S

representing mutations in positions 30 and 34 substituting alanine and glutamic acid for  
asparagine and serine, respectively.

When one or more alternative amino acid residues may be inserted in a given  
15 position it is indicated as

A30N,E or alternatively, A30N or A30E

Furthermore, when a position suitable for modification is identified herein  
without any specific modification being suggested, it is to be understood that any amino  
acid residue may be substituted for the amino acid residue present in the position. Thus,  
20 for instance, when a modification of an alanine in position 30 is mentioned, but not  
specified, it is to be understood that the alanine may be deleted or substituted for any  
other amino acid, i.e., any one of:

R, N, D, A, C, Q, E, G, H, I, L, K, M, F, P, S, T, W, Y, V.

Further, "A30X" means any one of the following substitutions: A30R, A30N,  
25 A30D, A30C, A30Q, A30E, A30G, A30H, A30I, A30L, A30K, A30M, A30F, A30P,  
A30S, A30T, A30W, A30Y, or A30V; or in short:

A30R,N,D,C,Q,E,G,H,I,L,K,M,F,P,S,T,W,Y,V.

If the parent enzyme--used for the numbering--already has the amino acid residue  
in question suggested for substitution in that position the following nomenclature is used:

"X30N" or "X30N,V" in the case where, for instance, one or N or V is present in the wild-type. This indicates that other corresponding parent enzymes are substituted to an "Asn" or "Val" in position 30.

5    **3. Characteristics of Amino Acid Residues**

*Charged amino acids:*

Asp, Glu, Arg, Lys, His

*Negatively charged amino acids (with the most negative residue first):*

Asp, Glu

10

*Positively charged amino acids (with the most positive residue first):*

Arg, Lys, His

*Neutral amino acids:*

Gly, Ala, Val, Leu, Ile, Phe, Tyr, Trp, Met, Cys, Asn, Gln, Ser, Thr, Pro

15

*Hydrophobic amino acid residues (with the most hydrophobic residue listed last):*

Gly, Ala, Val, Pro, Met, Leu, Ile, Tyr, Phe, Trp,

*Hydrophilic amino acids (with the most hydrophilic residue listed last):*

Thr, Ser, Cys, His, Glu, Gln, Asn, Asp, Lys, Arg

20    **4. Alpha-Amylases and AmyS-like Amylases**

**4.1 Amino Acid Identities of Various Alpha-Amylase**

A number of alpha-amylases produced by *Bacillus* spp. are highly homologous (identical) on the amino acid level and may be useful as parent enzymes herein. The percent identity (based on amino acid sequence) of a number of known *Bacillus* alpha-amylases, relative to each other can be found in the below Table A:

25

**TABLE A:** Amino acid sequence identity of several known *Bacillus* alpha-amylases

	<b>707</b>	<b>AP1378</b>	<b>BAN</b>	<b>BSG</b>	<b>SP690</b>	<b>SP722</b>	<b>AA560</b>	<b>LAT</b>
<b>707</b>	100.0	86.4	66.9	66.5	87.6	86.2	95.5	68.1
<b>AP1378</b>	86.4	100.0	67.1	68.1	95.1	86.6	86.0	69.4
<b>BAN</b>	66.9	67.1	100.0	65.6	67.1	68.8	66.9	80.7
<b>BSG</b>	66.5	68.1	65.6	100.0	67.9	67.1	66.3	65.4
<b>SP690</b>	87.6	95.1	67.1	67.9	100.0	87.2	87.0	69.2

	707	AP1378	BAN	BSG	SP690	SP722	AA560	LAT
SP722	86.2	86.6	68.8	67.1	87.2	100.0	86.8	70.8
AA560	95.5	86.0	66.9	66.3	87.0	86.8	100.0	68.3
LAT	68.1	69.4	80.7	65.4	69.2	70.8	68.3	100.0

The skilled artisan will appreciate that percent identities can be determined from the literature, or by any means disclosed herein or known in the art. For instance, the *B. licheniformis* alpha-amylase (LAT) (SEQ ID NO: 7) has been found to be about 81% homologous with the *B. amyloliquefaciens* alpha-amylase (SEQ ID NO: 9), and about 5 65% homologous with the *G. stearothermophilus* alpha-amylase (BSG) (SEQ ID NO: 1). Additional homologous alpha-amylases include SP690 and SP722 disclosed in WO 95/26397, and the #707 alpha-amylase derived from *Bacillus* spp. (SEQ ID NO: 6), described by Tsukamoto *et al.*, *Biochemical and Biophysical Research Communications*, 10 151 (1988), pp. 25-31. The KSM AP1378 alpha-amylase is disclosed in WO 97/00324 (from KAO Corporation).

#### 4.2 Parent Alpha-Amylases

AmyS-like alpha-amylases, as defined above, may be used as a parent alpha-amylase. In a preferred embodiment, the parent alpha-amylase is derived from *G. stearothermophilus*, e.g., one of those referred to above, such as the *G. stearothermophilus* alpha-amylase having the amino acid sequence shown in SEQ ID NO: 1 or 2.

#### 4.3 Parent Hybrid Alpha-Amylases

20 The parent alpha-amylase (i.e., backbone alpha-amylase) may also be a hybrid alpha-amylase, i.e., an alpha-amylase that comprises a combination of partial amino acid sequences derived from at least two alpha-amylases.

The parent hybrid alpha-amylase may be one, which on the basis of amino acid homology (identity) and/or DNA hybridization (as defined above), can be determined to belong to the AmyS-like alpha-amylase family described above. In such a case, the 25 hybrid alpha-amylase is typically composed of at least one part of a AmyS-like alpha-amylase and part(s) of one or more other alpha-amylases selected from AmyS-like alpha-

amylases or non-AmyS-like alpha-amylases of microbial (bacterial or fungal) and/or mammalian origin.

Thus, the parent hybrid alpha-amylase may comprise a combination of partial amino acid sequences deriving from at least two AmyS-like alpha-amylases, or from at least one AmyS-like and at least one non-AmyS-like bacterial alpha-amylase, or from at least one AmyS-like and at least one fungal alpha-amylase. The AmyS-like alpha-amylase from which a partial amino acid sequence derives, may be any of the specific AmyS-like alpha-amylase referred to herein.

For instance, the parent alpha-amylase may comprise a C-terminal part of an alpha-amylase derived from a strain of *B. licheniformis*, and an N-terminal part of an alpha-amylase derived from a strain of *G. stearothermophilus* or from a strain of *G. stearothermophilus* (BSG).

#### 5. Homology (Identity)

Homology may be determined as the degree of identity between two sequences indicating a relationship therebetween, e.g. a derivation of the first sequence from the second or vice versa. The homology may be determined by visual inspection or manual calculations, but more conveniently by means of computer programs known in the art, such as GAP, a program provided in the GCG program package (described above). Thus, Gap GCG v8 may be used, for example with the default scoring matrix for identity and the following default parameters: GAP creation penalty of 5.0 and GAP extension penalty of 0.3, respectively for nucleic acidic sequence comparison, and GAP creation penalty of 3.0 and GAP extension penalty of 0.1, respectively, for protein sequence comparison. GAP uses the method of Needleman and Wunsch, (1970), *J. Mol. Biol.* 48: 443-453, to make alignments and to calculate the identity.

A structural alignment between Spezyme Xtra (SEQ ID NO: 2) and, e.g., another alpha-amylase may be used to identify equivalent/corresponding positions in other AmyS-like alpha-amylases. One method of obtaining said structural alignment is to use the Pile Up program from the GCG package using default values of gap penalties, i.e., a gap creation penalty of 3.0 and gap extension penalty of 0.1. Other structural alignment

methods include the hydrophobic cluster analysis (Gaboriaud *et al.*, *FEBS Lett.* 224: 149-155, 1987) and reverse threading (Huber, T; Torda, AE, *Protein Sci.* 7(1) 142-149, 1998).

## 6. Hybridization

5 The oligonucleotide probe used in the characterization of the AmyS-like alpha-amylase above may suitably be prepared on the basis of the full or partial nucleotide or amino acid sequence of the alpha-amylase in question.

Suitable conditions for assessing hybridization involve pre-soaking in 5X SSC and pre-hybridizing for 1 hour at 40 °C in a solution of 20% formamide, 5X Denhardt's solution, 50 mM sodium phosphate, pH 6.8, and 50 mg of denatured sonicated calf  
10 thymus DNA, followed by hybridization in the same solution supplemented with 100 mM ATP for 18 hours at 40°C, followed by three times washing of the filter in 2X SSC, 0.2% SDS at 40°C for 30 minutes (low stringency), preferred at 50°C (medium stringency), more preferably at 65°C (high stringency), even more preferably at 75°C  
15 (very high stringency). More details about the hybridization method can be found in Sambrook *et al.*, *MOLECULAR CLONING: A LABORATORY MANUAL*, 2<sup>nd</sup> Ed., Cold Spring Harbor, 1989.

In the present context, "derived from" is intended not only to indicate an alpha-amylase produced or producible by a strain of the organism in question, but also an  
20 alpha-amylase encoded by a DNA sequence isolated from such strain and produced in a host organism transformed with said DNA sequence. Finally, the term is intended to indicate an alpha-amylase, which is encoded by a DNA sequence of synthetic and/or cDNA origin and which has the identifying characteristics of the alpha-amylase in question. The term is also intended to indicate that the parent alpha-amylase may be a  
25 variant of a naturally occurring alpha-amylase, i.e., a variant, which is the result of a modification (insertion, substitution, deletion) of one or more amino acid residues of the naturally occurring alpha-amylase.

## 7. General Mutations in Variant Alpha-Amylases

30 A variant described herein may, in one embodiment, comprise one or more



modifications in addition to those outlined above. Thus, it may be advantageous that one or more proline residues (Pro) present in the part of the alpha-amylase variant that is modified is/are replaced with a non-proline residue which may be any of the possible, naturally-occurring non-proline residues, and which preferably is an alanine, glycine, serine, threonine, valine or leucine.

Analogously, in one embodiment, one or more cysteine residues present in the parent alpha-amylase may be replaced with a non-cysteine residue such as serine, alanine, threonine, glycine, valine or leucine.

It is to be understood that this disclosure encompasses variants incorporating two or more of the above outlined modifications.

Furthermore, it may be advantageous to introduce mutations in one or more of the following positions (using SEQ ID NO: 7 for the numbering):

M15, V128, A111, H133, W138, T149, M197, N188, A209, A210, H405, T412, in particular the following single, double or triple or multi mutations:

M15X, in particular M15T,L;  
V128X, in particular V128E;  
H133X, in particular H133Y;  
N188X, in particular N188S,T,P;  
M197X, in particular M197T,L;  
A209X, in particular A209V;  
M197T/W138F; M197T/138Y; M15T/H133Y/N188S;  
M15N128E/H133Y/N188S; E119C/S130C; D124C/R127C;  
H133Y/T149I; and/or  
G475R, H133Y/S187D; H133Y/A209V.

In the case of the parent alpha-amylase having the amino acid sequence shown in SEQ ID No. 7, relevant amino acid residues which may be deleted or substituted with a view to improving the oxidation stability include the single cysteine residue (C363) and the methionine residues located in positions M8, M9, M96, M200, M206, M284, M307, M311, M316 and M438 in SEQ ID NO: 2.

With respect to increasing the thermal stability of an alpha-amylase variant relative to its parent alpha-amylase, it appears to be particularly desirable to delete at least one, and preferably two, or even three, of the following amino acid residues in the amino acid sequence shown in SEQ ID NO: 2: F178, R179, G180, I181, G182 and  
5 K183.

Particularly interesting pair-wise deletions of this type are R179\*+G180\*; and I181\*+G182\* (SEQ ID NOS: 16 or 15, respectively) (or equivalents of these pair-wise deletions in another alpha-amylase meeting the requirements of a parent alpha-amylase in the context of the present disclosure).

10 Other residues of interest include N193F and V416G in the amino acid sequence shown in SEQ ID NO: 2.

## **8. Altered Properties of Variants**

### **8.1 General**

The following section describes the relationship between mutations, which are  
15 present in a variant described herein, and desirable alterations in properties (relative to those of a parent AmyS-like alpha-amylase), which may result therefrom.

Described herein are AmyS-like alpha-amylases with altered properties. Parent alpha-amylases specifically contemplated herein are AmyS-like alpha-amylases and parent hybrid AmyS-like alpha-amylases.

20 In one embodiment, the *Geobacillus stearothermophilus* alpha-amylase (SEQ ID NO: 2) is used as the starting point, i.e., the parent amylase, but in other embodiments, the SP722, BLA, BAN, AA560, SP690, KSM AP1378, #707 and other *Bacillus* alpha-amylases may be used. Amino acid positions corresponding to positions in SEQ ID NO: 2 are readily determined in accordance herewith.

25 The skilled artisan will appreciate that while any parent alpha-amylase could be used as a reference amylase for the purpose of numbering/identifying the amino acid residues modified or to be modified in a particular variant, SEQ ID NO: 1 is presently a preferred sequence for such purpose, because it is the longest *B. stearothermophilus* sequence presently available herein.

In one aspect, this disclosure relates to variant with altered properties, e.g., as described above.

In one of its several aspects, this disclosure provides a variant of a parent *G. stearothermophilus* alpha-amylase, comprising an alteration at one or more positions  
5 (using e.g., SEQ ID NO: 1 for the amino acid numbering) selected from the group of:

P17, D19, T21, N28, S51, G72, V74, A82, Q86, Q89, A93, G95, Q97, W115,  
D117, P123, S124, D125, N127, I130, G132, Q135, P145, G146, G148, S153, Y159,  
W166, S169, K171, W187, P209, N224, S242, G256, D269, N271, T278, N281, G302,  
A304, R308, T321, Q358, P378, S382, K383, T398, H405, T417, E418, P420, G421,  
10 P432, W437, G446, G454, S457, T459, T461, S464, G474, R483,

wherein

(a) the alteration(s) are independently (i) an insertion of an amino acid  
downstream of the amino acid that occupies the position; (ii) a deletion of the amino acid  
that occupies the position; or (iii) a substitution of the amino acid that occupies the  
15 position with a different amino acid,

(b) the variant has alpha-amylase activity, and

(c) each position corresponds to a position of the amino acid sequence of the  
parent amylase, e.g., a *G. stearothermophilus* alpha-amylase, e.g., having the amino acid  
sequence shown in SEQ ID NO: 2, e.g., a truncated alpha-amylase that is available  
20 commercially as SPEZYME XTRA from Genencor.

Specifically contemplated herein are S242A, S242Q, S242N and S242E.

Additionally, residues R179, G180, I181, G182, K183 were chosen to explore the  
effect of mutations in the calcium-sodium binding region, and P245 was chosen because  
a proline in the middle of an alpha-helix is unusual.

25 Corresponding positions in other parent AmyS-like alpha-amylases can be found  
by alignment as described above, for example, as with those sequences shown in the  
alignment in Figure 4. Thus, variants of a parent AmyS-like alpha-amylase, comprising  
an alteration at one or more of the above enumerated positions (using, e.g., SEQ ID NO:  
1 for comparative amino acid numbering) is contemplated herein.

## 8.2 Altered Properties: Stability

In the context of the variants described herein, mutations (including amino acid substitutions and deletion) of importance with respect to achieving altered stability, in particular improved stability (i.e., higher or lower), at especially high temperatures (i.e., about 70-120°C) and/or extreme pH (i.e. low or high pH, i.e., pH 4-6 or pH 8-11, respectively), in particular at free (i.e., unbound, therefore in solution) calcium concentrations below 60 ppm, include any of the mutations listed in the "Altered Properties" section. The stability may be determined as described in the "Methods" section below.

## 8.3 Altered Properties: Ca<sup>2+</sup> Stability

Altered Ca<sup>2+</sup> stability means the stability of the enzyme under Ca<sup>2+</sup> depletion has been improved, i.e., higher or lower stability, relative to the parent enzyme. In the context of the presently described variants, mutations (including amino acid substitutions and deletions) of importance with respect to achieving altered Ca<sup>2+</sup> stability, in particular improved Ca<sup>2+</sup> stability, i.e., higher or lower stability, at especially high pH (i.e., pH 8-10.5) include any of the mutations listed in the "Altered Properties" section.

## 8.4 Altered Properties: Specific Activity

In a further aspect, important mutations (including amino acid substitutions and deletions) with respect to obtaining variants exhibiting altered specific activity, in particular increased or decreased specific activity, especially at temperatures from about 10-60°C, preferably about 20-50°C, especially about 0-40°C, include any of the mutations listed in the in "Altered Properties" section. The specific activity may be determined as described in the "Methods" section below.

## 8.5 Altered Properties: Oxidation Stability

The described variants may have altered oxidation stability, in particular higher oxidation stability, in comparison to the parent alpha-amylase. Increased oxidation stability is advantageous in, e.g., detergent compositions and decreased oxidation stability may be advantageous in compositions intended for starch liquefaction. Oxidation stability may be determined as described in the "Methods" section below.

### 8.6 Altered Properties: Altered pH Profile

Important positions and mutations with respect to obtaining variants with altered pH profile, in particular improved activity at especially high pH (i.e., pH 8-10.5) or low pH (i.e., pH 4-6) include mutations of amino residues located close to the active site residues.

Preferred specific mutations/substitutions include those listed above in the section "Altered Properties" for the positions in question. Suitable assays are described in the "Methods" section below.

### 8.7 Altered Properties: Wash Performance

Important positions and mutations with respect to obtaining variants with improved wash performance at especially high pH (i.e., pH 8.5-11) include the specific mutations/substitutions listed above in the section "Altered Properties" for the positions in question. The wash performance may be tested as described below in the "Methods" section.

## 9. Methods of Preparing $\alpha$ -Amylase Variants

Methods for introducing mutations into genes are known in the art, as are cloning methods for  $\alpha$ -amylase-encoding DNA sequences. Such methods including methods for generating mutations at specific sites within the  $\alpha$ -amylase-encoding sequence will be discussed below.

### 9.1 Cloning a DNA Sequence Encoding an $\alpha$ -Amylase

The DNA sequence encoding a parent  $\alpha$ -amylase may be isolated from any cell or microorganism producing the  $\alpha$ -amylase in question, using various methods well known in the art. First, a genomic DNA and/or cDNA library should be constructed using chromosomal DNA or messenger RNA from the organism that produces the  $\alpha$ -amylase to be studied. If the amino acid sequence of the  $\alpha$ -amylase is known, homologous, labeled oligonucleotide probes may be synthesized and used to identify  $\alpha$ -amylase-encoding clones from a genomic library prepared from the organism in question. Alternatively, a labeled oligonucleotide probe containing sequences homologous to a known  $\alpha$ -amylase

gene can be used as a probe to identify  $\alpha$ -amylase-encoding clones, e.g., using hybridization and washing conditions of lower stringency.

Another method for identifying  $\alpha$ -amylase-encoding clones is based on inserting fragments of genomic DNA into an expression vector, such as a plasmid, transforming  $\alpha$ -amylase-negative bacteria with the resulting genomic DNA library, and plating the  
5 transformed bacteria onto agar containing a substrate for  $\alpha$ -amylase, thereby allowing clones expressing the  $\alpha$ -amylase to be readily identified.

Alternatively, the DNA sequence encoding the enzyme may be prepared synthetically by established, standard methods, e.g. the phosphoramidite method described  
10 by S. L. Beaucage and M. H. Caruthers, *Tetrahedron Letters* 22: 1859-1869 (1981) or the method described by Matthes *et al.*, *EMBO J.* 3:801-895 (1984). In the phosphoramidite method, oligonucleotides are synthesized, e.g., in an automatic DNA synthesizer, purified, annealed, ligated, and cloned in appropriate vectors.

Finally, the DNA sequence may be of mixed origin comprising e.g., genomic and  
15 synthetic sequences, synthetic and cDNA sequences, or genomic and cDNA sequences, prepared by ligating fragments of synthetic, genomic, or cDNA origin (as appropriate, the fragments corresponding to various parts of the entire DNA sequence), in accordance with standard techniques. The DNA sequence may also be prepared by polymerase chain reaction (PCR) using specific primers, for instance as described in U.S. Pat. No.  
20 4,683,202 or R. K. Saiki *et al.* *EMBO J.* 3:801-895 (1988).

## 9.2 Site-directed Mutagenesis

Once an  $\alpha$ -amylase-encoding DNA sequence has been isolated, and desirable sites for mutation identified, mutations may be introduced using synthetic oligonucleotides. These oligonucleotides contain nucleotide sequences flanking the desired mutation sites;  
25 mutant nucleotides are inserted during oligonucleotide synthesis. In a specific method, a single-stranded gap of DNA, bridging the  $\alpha$ -amylase-encoding sequence, is created in a vector carrying the  $\alpha$ -amylase gene. Then the synthetic nucleotide, bearing the desired mutation, is annealed to a homologous portion of the single-stranded DNA. The remaining gap is then filled in with DNA polymerase I (Klenow fragment) and the  
30 construct is ligated using T4 ligase. A specific example of this method is described in

Morinaga *et al.* *Biotechnology* 2:636-639 (1984). U.S. Pat. No. 4,760,025 discloses the introduction of oligonucleotides encoding multiple mutations by performing minor alterations of the cassette. However, an even greater variety of mutations can be introduced at any one time by the Morinaga method, because a multitude of  
5 oligonucleotides, of various lengths, can be introduced.

Another method of introducing mutations into  $\alpha$ -amylase-encoding DNA sequences is described in Nelson and Long, *Analytical Biochem.*, 180: 147-151, 1989. It involves the 3-step generation of a PCR fragment containing the desired mutation introduced by using a chemically synthesized DNA strand as one of the primers in the  
10 PCR reactions. From the PCR-generated fragment, a DNA fragment carrying the mutation may be isolated by cleavage with restriction endonucleases and reinserted into an expression plasmid.

The skilled artisan will appreciate that many alternative methods are available for providing or obtaining variants herein. For example, gene shuffling, e.g., as described in  
15 WO 95/22625 (from Affymax Technologies N.V.) or in WO 96/00343 (from Novo Nordisk A/S), or other corresponding techniques resulting in hybrid enzymes comprising the mutation(s), e.g., substitution(s) and/or deletion(s), in question.

### 9.3 Expression of Alpha-Amylase Variants

A DNA sequence encoding the variant produced by methods described above, or  
20 by any alternative methods known in the art, can be expressed, in enzyme form, using an expression vector which typically includes control sequences encoding a promoter, operator, ribosome binding site, translation initiation signal, and, optionally, a repressor gene or various activator genes.

The recombinant expression vector carrying the DNA sequence encoding an  
25 alpha-amylase variant for use herein may be any vector, which may conveniently be subjected to recombinant DNA procedures, and the choice of vector will often depend on the host cell into which it is to be introduced. Thus, the vector may be an autonomously replicating vector, i.e., a vector that exists as an extrachromosomal entity, the replication of which is independent of chromosomal replication, e.g., a plasmid, a bacteriophage, an  
30 extrachromosomal element, a minichromosome, or an artificial chromosome.

Alternatively, the vector may be integrated into the host cell genome and replicated together with the chromosome(s) into which it has been integrated.

In the vector, the DNA sequence should be operably-connected to a suitable promoter sequence. The promoter may be any DNA sequence, which shows  
5 transcriptional activity in the host cell of choice and may be derived from genes encoding proteins either homologous or heterologous to the host cell. Examples of suitable promoters for directing the transcription of the DNA sequence encoding an alpha-amylase variant for use herein, especially in a bacterial host, are the promoter of the *lac* operon of *E. coli*, the *Streptomyces coelicolor* agarase gene *dagA* promoters, the  
10 promoters of the *Bacillus licheniformis* alpha-amylase gene (*amyL*), the promoters of the *Geobacillus stearothermophilus* maltogenic amylase gene (*amyM*), the promoters of the *Bacillus amyloliquefaciens* alpha-amylase (*amyQ*), the promoters of the *Bacillus subtilis* *xylA* and *xylB* genes etc. For transcription in a fungal host, examples of useful promoters are those derived from the gene encoding *A. oryzae* TAKA amylase, *Rhizomucor miehei*  
15 aspartic proteinase, *A. niger* neutral alpha-amylase, *A. niger* acid stable alpha-amylase, *A. niger* glucoamylase, *Rhizomucor miehei* lipase, *A. oryzae* alkaline protease, *A. oryzae* triose phosphate isomerase or *A. nidulans* acetamidase.

Expression vectors for use herein may also comprise a suitable transcription terminator and, in eukaryotes, polyadenylation sequences operably-connected to the  
20 DNA sequence encoding the alpha-amylase variant. Termination and polyadenylation sequences may suitably be derived from the same sources as the promoter.

The vector may further comprise a DNA sequence enabling the vector to replicate in the host cell in question. Examples of such sequences are the origins of replication of plasmids pUC19, pACYC177, pUB110, pE194, pAMB1 and pIJ702.

25 The vector may also comprise a selectable marker, e.g. a gene the product of which complements a defect in the host cell, such as the *dal* genes from *B. subtilis* or *B. licheniformis*, or one that confers antibiotic resistance such as ampicillin, kanamycin, chloramphenicol or tetracyclin resistance. Furthermore, the vector may comprise *Aspergillus* selection markers such as *amdS*, *argB*, *niaD* and *sC*, a marker giving rise to



hygromycin resistance, or the selection may be accomplished by co-transformation, e.g., as described in WO 91/17243.

While intracellular expression may be advantageous in some respects, e.g., when using certain bacteria as host cells, it is generally preferred that the expression is  
5 extracellular. In general, the *Bacillus* alpha-amylases mentioned herein comprise a pre-region permitting secretion of the expressed protease into the culture medium. If desirable, this pre-region may be replaced by a different pre-region or signal sequence, conveniently accomplished by substitution of the DNA sequences encoding the respective pre-regions.

10 The procedures used to ligate a DNA construct encoding an alpha-amylase variant, the promoter, terminator and other elements, respectively, and to insert them into suitable vectors containing the information necessary for replication, are well known to persons skilled in the art (cf., for instance, Sambrook *et al.*, MOLECULAR CLONING: A LABORATORY MANUAL, 2<sup>nd</sup> Ed., Cold Spring Harbor, 1989).

15 Cells for use herein, e.g. comprising a DNA construct or an expression vector as defined above, can be used as host cells in the recombinant production of an alpha-amylase variant. The cell may be transformed with a DNA construct encoding the variant, conveniently by integrating the DNA construct (in one or more copies) in the host chromosome. This integration is generally considered to be an advantage as the  
20 DNA sequence is more likely to be stably maintained in the cell. Integration of the DNA constructs into the host chromosome may be performed according to conventional methods, e.g., by homologous or heterologous recombination. Alternatively, the cell may be transformed with an expression vector as described above in connection with the different types of host cells.

25 Cells for use herein may be cells of a higher organism such as a mammal or an insect, but are preferably microbial cells, e.g., a bacterial or a fungal (including yeast) cell.

30 Examples of suitable bacteria are Gram-positive bacteria such as *Bacillus subtilis*, *Bacillus licheniformis*, *Bacillus lentus*, *Bacillus brevis*, *Geobacillus stearothermophilus*, *Bacillus alkalophilus*, *Bacillus amyloliquefaciens*, *Bacillus coagulans*, *Bacillus circulans*,

*Bacillus lautus*, *Bacillus megaterium*, *Bacillus thuringiensis*, or *Streptomyces lividans* or *Streptomyces murinus*, or gram-negative bacteria such as *E. coli*. The transformation of the bacteria may, for instance, be effected by protoplast transformation or by using competent cells in a manner known per se.

5           Where used for expression, a yeast may favorably be selected from a species of *Saccharomyces* or *Schizosaccharomyces*, e.g. *Saccharomyces cerevisiae*. A filamentous fungus may advantageously be selected from a species of *Aspergillus*, e.g., *Aspergillus oryzae* or *Aspergillus niger*. Fungal cells may be transformed by a process involving protoplast formation and transformation of the protoplasts followed by regeneration of  
10 the cell wall in a manner known per se. A suitable procedure for transformation of *Aspergillus* host cells is described in EP 238 023.

          In a yet further aspect, the disclosure relates to a method of producing an alpha-amylase variant, which method comprises cultivating a host cell as described above under conditions conducive to the production of the variant and recovering the variant from the  
15 cells and/or culture medium.

          The medium used to cultivate the cells may be any conventional medium suitable for growing the host cell in question and obtaining expression of the alpha-amylase variant. Suitable media are available from commercial suppliers or may be prepared according to published recipes (e.g., as described in catalogues of the ATCC).

20           The alpha-amylase variant secreted from the host cells may be recovered from the culture medium by known procedures, including separating the cells from the medium by centrifugation or filtration, and precipitating proteinaceous components of the medium by means of a salt such as ammonium sulphate, followed by the use of chromatographic procedures such as ion exchange chromatography, affinity chromatography, or the like.

## 25           **9.4 Methods for Characterizing and Screening Variants**

### **9.4.1 Filter Screening Assays**

          The below assays may be used to screening of AmyS-like alpha-amylase variants having altered stability at high or low pH and/or under Ca<sup>2+</sup> depleted conditions compared to the parent enzyme and AmyS-like alpha-amylase.

#### 9.4.2 High pH Filter Assay

*Bacillus* libraries are plated on a sandwich of cellulose acetate (OE 67, Schleicher & Schuell, Dassel, Germany)--and nitrocellulose filters (Protran-Ba 85, Schleicher & Schuell, Dassel, Germany) on TY agar plates with 10 µg/mL kanamycin at 37°C for at least 21 hours. The cellulose acetate layer is located on the TY agar plate.

Each filter sandwich is specifically marked with a needle after plating, but before incubation in order to be able to localize positive variants on the filter and the nitrocellulose filter with bound variants is transferred to a container with glycine-NaOH buffer, pH 8.6-10.6 and incubated at room temperature (can be altered from about 10-60°C) for 15 min. The cellulose acetate filters with colonies are stored on the TY-plates at room temperature until use. After incubation, residual activity is detected on plates containing 1% agarose, 0.2% starch in glycine-NaOH buffer, pH 8.6-10.6. The assay plates with nitrocellulose filters are marked the same way as the filter sandwich and incubated for 2 hours at room temperature. After removal of the filters the assay plates are stained with 10% Lugol solution. Starch degrading variants are detected as white spots on dark blue background and then identified on the storage plates. Positive variants are rescreened twice under the same conditions as the first screen.

#### 9.4.3 Low Calcium Filter Assay

*Bacillus* libraries are plated on a sandwich of cellulose acetate (OE 67, Schleicher & Schuell, Dassel, Germany)--and nitrocellulose filters (Protran-Ba 85, Schleicher & Schuell, Dassel, Germany) on TY agar plates with a relevant antibiotic, e.g., kanamycin or chloramphenicol, at 37°C for at least 21 hours. The cellulose-acetate layer is located on the TY agar plate.

Each filter sandwich is specifically marked with a needle after plating, but before incubation in order to be able to localize positive variants on the filter and the nitrocellulose filter with bound variants is transferred to a container with carbonate/bicarbonate buffer about pH 8.5-10 and with different EDTA concentrations (about 0.001 mM to about 100 mM). The filters are incubated at room temperature for 1 hour. The cellulose acetate filters with colonies are stored on the TY-plates at room temperature until use. After incubation, residual activity is detected on plates containing

1% agarose, 0.2% starch in carbonate/bicarbonate buffer about pH 8.5-10. The assay plates with nitrocellulose filters are marked the same way as the filter sandwich and incubated for about 2 hours at room temperature. After removal of the filters, the assay plates are stained with about 10% Lugol solution. Starch degrading variants are detected as white spots on dark blue background and then identified on the storage plates. Positive variants are rescreened twice under the same conditions as the first screen.

#### 9.4.4 Low pH Filter Assay

*Bacillus* libraries are plated on a sandwich of cellulose acetate (OE 67, Schleicher & Schuell, Dassel, Germany)--and nitrocellulose filters (Protran-Ba 85, Schleicher & Schuell, Dassel, Germany) on TY agar plates with 10 µg/mL chloramphenicol at 37°C for at least 21 hours. The cellulose acetate layer is located on the TY agar plate.

Each filter sandwich is specifically marked with a needle after plating, but before incubation in order to be able to localize positive variants on the filter, and the nitrocellulose filter with bound variants is transferred to a container with citrate buffer, pH 4.5 and incubated at 80°C for 20 minutes (when screening for variants in the wild-type backbone) or 85°C for 60 minutes (when screening for variants of the parent alpha-amylase). The cellulose acetate filters with colonies are stored on the TY-plates at room temperature until use. After incubation, residual activity is detected on assay plates containing 1% agarose, 0.2% starch in citrate buffer, pH 6.0. The assay plates with nitrocellulose filters are marked the same way as the filter sandwich and incubated for 2 hours at 50°C. After removal of the filters the assay plates are stained with 10% Lugol solution. Starch degrading variants are detected as white spots on dark blue background and then identified on the storage plates. Positive variants are re-screened twice under the same conditions as the first screen.

#### 9.4.5 Secondary Screening

Positive transformants after rescreening are picked from the storage plate and tested in a secondary plate assay. Positive transformants are grown for 22 hours at 37°C in 5 mL LB + chloramphenicol. The *Bacillus* culture of each positive transformant and as a control a clone expressing the corresponding backbone are incubated in citrate buffer, pH 4.5 at 90°C and samples are taken at 0, 10, 20, 30, 40, 60 and 80 minutes. A

3- $\mu$ L sample is spotted on an assay plate. The assay plate is stained with 10% Lugol solution. Improved variants are seen as variants with higher residual activity (detected as halos on the assay plate) than the backbone. The improved variants are determined by nucleotide sequencing.

#### 5           **9.4.6 Stability Assay of Unpurified Variants**

The stability of the variants may be assayed as follows: *Bacillus* cultures expressing the variants to be analyzed are grown for 21 hours at 37°C in 10 mL LB + chloramphenicol. 800  $\mu$ L culture is mixed with 200 microliters citrate buffer, pH 4.5. A number of 70  $\mu$ L aliquots corresponding to the number of sample time points are made in  
10 PCR tubes and incubated at 70°C or 90°C for various time points (typically 5, 10, 15, 20, 25 and 30 minutes) in a PCR machine. The 0 min sample is not incubated at high temperature. Activity in the sample is measured by transferring 20  $\mu$ L to 200  $\mu$ L of the alpha-amylase PNP-G<sub>7</sub> substrate MPR3 ((Boehringer Mannheim Cat. No. 1660730) as described below under "Assays for Alpha-Amylase Activity". Results are plotted as  
15 percentage activity (relative to the 0 time point) versus time, or stated as percentage residual activity after incubation for a certain period of time.

#### **9.4.7 Fermentation and Purification of Alpha-Amylase Variants**

*A. B. subtilis* strain harboring the relevant expression plasmid may be fermented and purified as follows: The strain is streaked on a LB-agar plate with 10  $\mu$ g/mL  
20 kanamycin from -80°C stock, and grown overnight at 37°C. The colonies are transferred to 100 mL PS-1 media supplemented with 10  $\mu$ g/mL chloramphenicol in a 500 mL shaking flask.

#### **Composition of PS-1 medium**

25	Pearl sugar	100 g/L
	Soy Bean Meal	40 g/L
	Na <sub>2</sub> HPO <sub>4</sub> , 12 H <sub>2</sub> O	10 g/L
	Pluronic™ PE 6100	0.1 g/L
30	CaCO <sub>3</sub>	5 g/L

The culture is shaken at 37°C at 270 rpm for 5 days.

Cells and cell debris are removed from the fermentation broth by centrifugation at 4500 rpm in 20-25 minutes. Afterwards the supernatant is filtered to obtain a completely clear solution. The filtrate is concentrated and washed on a UF-filter (10000 cut off membrane) and the buffer is changed to 20 mM Acetate at pH 5.5. The UF-filtrate is applied on an S-SEPHAROSE F.F (Pharmacia) and elution is carried out by step elution with 0.2 M NaCl in the same buffer. The eluate is dialyzed against 10 mM Tris, pH 9.0 and applied on a Q-SEPHAROSE F.F. and eluted with a linear gradient from 0-0.3M NaCl over 6 column volumes. The fractions that contain the activity (measured by the PHADEBAS assay) are pooled, pH was adjusted to pH 7.5, and remaining color was removed by treatment with 0.5% w/v active charcoal in 5 minutes.

#### **9.4.8 Specific Activity Determination**

The specific activity can be determined using the PHADEBAS® assay (Magle Life Sciences) as activity/mg enzyme. The manufactures instructions are followed (see also below under "Assay for Alpha-Amylase Activity").

#### **9.4.9 Determination of Isoelectric Point**

The pI can be determined by isoelectric focusing (e.g., using Pharmacia, Ampholine, pH 3.5-9.3).

#### **9.4.10 Stability Determination**

The amylase stability may be measured using the method as follows:

The enzyme is incubated under the relevant conditions. Samples are taken at various time points, e.g., after 0, 5, 10, 15 and 30 minutes and diluted 25 times (same dilution for all taken samples) in assay buffer (50 mM Britton buffer pH 7.3) and the activity is measured using the PHADEBAS assay (Magle Life Sciences) under standard conditions pH 7.3, 37 °C.

The activity measured before incubation (0 minutes) is used as reference (100%). The decline in percent is calculated as a function of the incubation time. The table shows the residual activity after, e.g., 30 minutes of incubation.

### 9.4.11 Assays for Alpha -Amylase Activity

#### 1. PHADEBAS Assay

Alpha-amylase activity is determined by a method employing PHADEBAS® tablets as substrate. PHADEBAS tablets (PHADEBAS® Amylase Test, supplied by  
5 Magle Life Sciences) contain a cross-linked insoluble blue-colored starch polymer, which has been mixed with bovine serum albumin and a buffer substance and tableted.

For every single measurement one tablet is suspended in a tube containing 5 mL 50 mM Britton-Robinson buffer (50 mM acetic acid, 50 mM phosphoric acid, 50 mM boric acid, 0.1 mM CaCl<sub>2</sub>, pH adjusted to the value of interest with NaOH). The test is  
10 performed in a water bath at the temperature of interest. The alpha-amylase to be tested is diluted in 50 mM Britton-Robinson buffer. One mL of this alpha-amylase solution is added to the 5 mL 50 mM Britton-Robinson buffer. The starch is hydrolyzed by the alpha-amylase giving soluble blue fragments. The absorbance of the resulting blue solution, measured spectrophotometrically at 620 nm, is a function of the alpha-amylase  
15 activity.

It is important that the measured 620 nm absorbance after 10 or 15 minutes of incubation (testing time) is in the range of 0.2 to 2.0 absorbance units at 620 nm. In this absorbance range there is linearity between activity and absorbance (Lambert-Beer law). The dilution of the enzyme must therefore be adjusted to fit this criterion. Under a  
20 specified set of conditions (temp., pH, reaction time, buffer conditions) 1 mg of a given alpha-amylase will hydrolyze a certain amount of substrate and a blue color will be produced. The color intensity is measured at 620 nm. The measured absorbance is directly proportional to the specific activity (activity/mg of pure alpha-amylase protein) of the alpha-amylase in question under the given set of conditions.

25

#### 2. Alternative Method

Alpha-amylase activity is determined by a method employing the PNP-G<sub>7</sub> substrate. PNP-G<sub>7</sub>, which is a abbreviation for p-nitrophenyl-alpha,D-maltoheptaoside, is a blocked oligosaccharide which can be cleaved by an endo-amylase. Following the  
30 cleavage, the alpha-glucosidase included in the kit digest the substrate to liberate a free

PNP molecule which has a yellow color and thus can be measured by visible spectrophotometry at  $\lambda=405$  nm (400-420 nm). Kits containing PNP-G<sub>7</sub> substrate and alpha-Glucosidase is manufactured by Boehringer-Mannheim (Cat. No. 1054635).

To prepare the reagent solution 10 mL of substrate/buffer solution is added to 50  
5 mL enzyme/buffer solution as recommended by the manufacturer. The assay is performed by transferring a 20  $\mu$ L sample to a 96 well microtitre plate and incubating at 25°C. 200  $\mu$ L reagent solution pre-equilibrated to 25°C is added. The solution is mixed and pre-incubated 1 minute and absorption is measured every 30 seconds over 4 minutes at OD 405 nm in an ELISA reader.

10 The slope of the time dependent absorption-curve is directly proportional to the activity of the alpha-amylase in question under the given set of conditions.

#### 9.4.12 Determination of LAS Sensitivity

The variant is incubated with different concentrations of LAS (linear alkyl benzene sulphonate; Nansa 1169/P) for 10 minutes at 40°C.

15 The residual activity is determined using the PHADEBAS® assay method or the alternative method employing the PNP-G<sub>7</sub> substrate.

LAS is diluted in 0.1 M phosphate buffer pH 7.5.

The following concentrations are used: 500 ppm, 250 ppm, 100 ppm, 50 ppm, 25 ppm, and 10 ppm on no LAS.

20 The variant is diluted in the different LAS buffers to concentration of 0.01-5 mg/l in a total volume of 10 mL and incubated for 10 minutes in a temperature controlled water bath. The incubation is stopped by transferring a small aliquot into cold assay buffer. It is important that during activity measurement the LAS concentration is below 1 ppm, in order not to affect the activity measurement. The residual activity is determined  
25 in duplicate using the above mentioned PHADEBAS® assay or alternative method. The activity is measured after subtraction of the blank. The activity with no LAS is 100%.

### 10. Methods of Using the Amylase Variants: Industrial Applications

30 The alpha-amylase variants presented herein possess valuable properties allowing for a variety of industrial applications in cleaning processes and stain removal. One or



more of the variant enzymes or compositions described herein may also be used in detergents, in particular laundry detergent compositions and dishwashing detergent compositions, hard surface cleaning compositions. The variants can also be used in compositions for desizing of textiles, fabrics or garments, for production of pulp and paper, beer making, ethanol production, and starch conversion processes as described  
5 above.

The variants herein may also be useful for desizing of textiles, fabrics, and garments (see, e.g., WO 95/21247, U.S. Pat. No. 4,643,736, and EP 119,920 hereby incorporated by reference), beer making or brewing, and in pulp and paper production or  
10 related processes.

### **10.1 Pulp and Paper Production**

The variant alkaline alpha-amylase may also be used in the production of lignocellulosic materials, such as pulp, paper and cardboard, from starch reinforced waste paper and cardboard, especially where re-pulping occurs at pH above about 7 and where  
15 amylases facilitate the disintegration of the waste material through degradation of the reinforcing starch. The alpha-amylase variants are especially useful in a process for producing a papermaking pulp from starch-coated printed-paper. The process may be performed as described in WO 95/14807, comprising the following steps:

- a) disintegrating the paper to produce a pulp,
- 20 b) treating with a starch-degrading enzyme before, during or after step a), and
- c) separating ink particles from the pulp after steps a) and b).

The alpha-amylases may also be very useful in modifying starch where enzymatically modified starch is used in papermaking together with alkaline fillers such as calcium carbonate, kaolin and clays. With the alkaline alpha-amylase variants it is  
25 possible to modify the starch in the presence of the filler thus allowing for a simpler integrated process.

### **10.2 Desizing of Textiles, Fabrics and Garments**

An alpha-amylase variant may also be very useful in textile, fabric or garment desizing. In the textile processing industry, alpha-amylases are traditionally used as  
30 auxiliaries in the desizing process to facilitate the removal of starch-containing size,

which has served as a protective coating on weft yarns during weaving. Complete removal of the size coating after weaving is important to ensure optimum results in the subsequent processes, in which the fabric is scoured, bleached and dyed. Enzymatic starch breakdown is preferred because it does not involve any harmful effect on the fiber material. In order to reduce processing cost and increase mill throughput, the desizing processing is sometimes combined with the scouring and bleaching steps. In such cases, non-enzymatic auxiliaries such as alkali or oxidation agents are typically used to break down the starch, because traditional alpha-amylases are not very compatible with high pH levels and bleaching agents. The non-enzymatic breakdown of the starch size does lead to some fiber damage because of the rather aggressive chemicals used. Accordingly, it would be desirable to use the alpha-amylase variants as they have an improved performance in alkaline solutions. The alpha-amylases may be used alone or in combination with a cellulase when desizing cellulose-containing fabric or textile.

Desizing and bleaching processes are well known in the art. For instance, such processes are described in WO 95/21247, U.S. Pat. No. 4,643,736, and EP 119,920, which are hereby incorporated by reference.

Commercially available products for desizing include OPTISIZE® FLEX from Genencor.

### 10.3 Cleaning Processes and Detergent Compositions

The variant alpha-amylases described herein may be added to and thus become a component of a detergent composition for various cleaning or washing processes, including laundry and dishwashing.

The detergent composition provided for herein may for example be formulated as a hand or machine laundry detergent composition including a laundry additive composition suitable for pretreatment of stained fabrics and a rinse added fabric softener composition or be formulated as a detergent composition for use in general household hard surface cleaning operations, or be formulated for hand or machine dishwashing operations.

In a specific aspect, there is provided for herein a detergent additive comprising a variant enzyme described herein. The detergent additive as well as the detergent

composition may comprise one or more other enzymes such as a protease, a lipase, a peroxidase, another amylolytic enzyme, e.g., another alpha-amylase, glucoamylase, maltogenic amylase, CGTase and/or a cellulase mannanase (such as MANNASTAR™ from Danisco US Inc., Genencor Division)), pectinase, pectin lyase, cutinase, and/or  
5 laccase.

In general the properties of the chosen enzyme(s) should be compatible with the selected detergent, (i.e., pH-optimum, compatibility with other enzymatic and non-enzymatic ingredients, etc.), and the enzyme(s) should be present in effective amounts.

*Proteases:* Suitable proteases include those of animal, vegetable or microbial  
10 origin. Microbial origin is preferred. Chemically modified or protein engineered mutants are included. The protease may be a serine protease or a metalloprotease, preferably an alkaline microbial protease or a trypsin-like protease. Examples of alkaline proteases are subtilisins, especially those derived from *Bacillus*, e.g., subtilisin Novo, subtilisin Carlsberg, subtilisin 309, subtilisin 147 and subtilisin 168 (described in WO 89/06279).  
15 Examples of trypsin-like pro-teases are trypsin (e.g., of porcine or bovine origin) and the *Fusarium* protease described in WO 89/06270 and WO 94/25583. Other examples of useful proteases may be found in WO98/23732, WO99/20770, WO 92/19729, WO 98/20115, WO 98/20116, and WO 98/34946.

Preferred commercially available protease enzymes include ALCALASE®,  
20 SAVINASE®, PRIMASE®, DURALASE®, ESPERASE®, and KANNASE® (from Novozymes A/S), MAXATASE®, MAXACAL, MAXAPEM®, PROPERASE®, PURAFECT®, PURAFECT OXP®, FN2®, FN3®, FN4® (Genencor International Inc.).

*Lipases:* Suitable lipases include those of bacterial or fungal origin. Chemically modified or protein engineered mutants are included. Examples of useful lipases include  
25 lipases from *Humicola* (synonym *Thermomyces*), e.g., from *H. lanuginosa* (*T. lanuginosus*) as described in EP 258 068 and EP 305 216 or from *H. insolens* as described in WO 96/13580, a *Pseudomonas* lipase, e.g., from *P. alcaligenes* or *P. pseudoalcaligenes* (EP 218 272), *P. cepacia* (EP 331 376), *P. stutzeri* (GB 1,372,034), *P. fluorescens*, *Pseudomonas* spp. strain SD 705 (WO 95/06720 and WO 96/27002), *P.*  
30 *wisconsinensis* (WO 96/12012), a *Bacillus* lipase, e.g., from *B. subtilis* (Dartois *et al.*

(1993), *Biochemica et Biophysica Acta*, 1131, 253-360), *B. stearothermophilus* (JP 64/744992) or *B. pumilus* (WO 91/16422). Other examples are lipase variants such as those described in WO 92/05249, WO 94/01541, EP 407 225, EP 260 105, WO 95/35381, WO 96/00292, WO 95/30744, WO 94/25578, WO 95/14783, WO 95/22615,  
5 WO 97/04079 and WO 97/07202.

Preferred commercially available lipase enzymes include LIPOLASE™ and LIPOLASE ULTRA™ (Novozymes A/S).

*Amylases*: One or more additional amylases may also be included. Suitable amylases (alpha and/or beta) include those of bacterial or fungal origin. Chemically  
10 modified or protein engineered mutants are included. Amylases include, for example, alpha-amylases obtained from *Bacillus*, e.g., a special strain of *B. licheniformis*, described in more detail in GB 1,296,839. Examples of useful alpha-amylases are the variants described in WO 94/18314, WO 96/39528, WO 94/02597, WO 94/18314, WO 96/23873, and WO 97/43424.

15 Commercially available alpha-amylases are DURAMYL™, LIQUEZYME™, TERMAMY™, NATALASE™, FUNGAMYL™ and BAN™ (Novozymes A/S), RAPIDASE™ and PURASTAR™ (from Genencor).

*Cellulases*: Suitable cellulases include those of bacterial or fungal origin. Chemically modified or protein engineered mutants are included. Suitable cellulases  
20 include cellulases from the genera *Bacillus*, *Pseudomonas*, *Trichoderma*, *Humicola*, *Fusarium*, *Thielavia*, *Acremonium*, e.g., the fungal cellulases produced from *Humicola insolens*, *Myceliophthora thermophila* and *Fusarium oxysporum* disclosed in U.S. Pat. No. 4,435,307, U.S. Pat. No. 5,648,263, U.S. Pat. No. 5,691,178, U.S. Pat. No. 5,776,757 and WO 89/09259. The *Trichoderma reesei* cellulases are disclosed in U.S. Pat. No.  
25 4,689,297, U.S. Pat. No. 5,814,501, U.S. Pat. No. 5,324,649, WO 92/06221 and WO 92/06165. *Bacillus* cellulases are disclosed in U.S. Pat. No. 6,562,612.

Commercially available cellulases include CELLUZYME®, and CAREZYME® (Novozymes A/S), CLAZINASE®, and PURADAX HA® (Genencor International Inc.), and KAC-500(B)® (Kao Corporation).

*Peroxidases/Oxidases*: Suitable peroxidases/oxidases include those of plant, bacterial or fungal origin. Chemically modified or protein engineered mutants are included. Examples of useful peroxidases include peroxidases from *Coprinus*, e.g., from *C. cinereus*, and variants thereof as those described in WO 93/24618, WO 95/10602, and  
5 WO 98/15257.

Commercially available peroxidases include GUARDZYME® (Novozymes A/S).

The detergent enzyme(s) may be included in a detergent composition by adding separate additives containing one or more enzymes, or by adding a combined additive comprising all of these enzymes. A detergent additive, e.g., a separate additive or a  
10 combined additive, can be formulated, e.g., granulate, a liquid, a slurry, etc. Preferred detergent additive formulations are granulates, in particular non-dusting granulates, liquids, in particular stabilized liquids, or slurries.

Non-dusting granulates may be produced, e.g., as disclosed in U.S. Pat. Nos. 4,106,991 and 4,661,452 and may optionally be coated by methods known in the art.  
15 Examples of waxy coating materials are poly(ethylene oxide) products (polyethyleneglycol, PEG) with mean molar weights of about 1000 to about 20000; ethoxylated nonyl-phenols having from 16 to 50 ethylene oxide units; ethoxylated fatty alcohols in which the alcohol contains from 12 to 20 carbon atoms and in which there are 15 to 80 ethylene oxide units; fatty alcohols; fatty acids; and mono- and di- and  
20 triglycerides of fatty acids. Examples of film-forming coating materials suitable for application by fluid bed techniques are given in GB 1483591. Liquid enzyme preparations may, for instance, be stabilized by adding a polyol such as propylene glycol, a sugar or sugar alcohol, lactic acid or boric acid according to established methods. Protected enzymes may be prepared according to the method disclosed in EP 238,216.

25 The detergent composition may be in any convenient form, e.g., a bar, a tablet, a powder, a granule, a paste or a liquid. A liquid detergent may be aqueous, typically containing up to about 70% water and 0 to about 30% organic solvent, or non-aqueous.

The detergent composition comprises one or more surfactants, which may be non-ionic, semi-polar, anionic, cationic, and/or zwitterionic. The surfactants are typically  
30 present at a level of from about 0.1% to about 60% by weight.

When included therein, the detergent will usually contain from about 1% to about 40% of an anionic surfactant such as linear alkylbenzenesulfonate, alpha-olefinsulfonate, alkyl sulfate (fatty alcohol sulfate), alcohol ethoxysulfate, secondary alkanesulfonate, alpha-sulfo fatty acid methyl ester, alkyl- or alkenylsuccinic acid or soap.

5           When included therein, the detergent will usually contain from about 0.2% to about 40% of a non-ionic surfactant such as alcohol ethoxylate, nonyl-phenol ethoxylate, alkylpolyglycoside, alkyl dimethylamine-oxide, ethoxylated fatty acid monoethanol- amide, fatty acid monoethanolamide, polyhydroxy alkyl fatty acid amide, or N-acyl N- alkyl derivatives of glucosamine ("glucamides").

10           The detergent may contain 0 to about 65% of a detergent builder or complexing agent such as zeolite, diphosphate, triphosphate, phosphonate, carbonate, citrate, nitrilotriacetic acid, ethylenediaminetetraacetic acid, diethylenetriaminepentaacetic acid, alkyl- or alkenylsuccinic acid, soluble silicates or layered silicates (e.g. SKS-6 from Hoechst).

15           The detergent may comprise one or more polymers. Examples are carboxymethylcellulose, poly(vinyl-pyrrolidone), poly(ethylene glycol), poly(vinyl alcohol), poly(vinylpyridine-N-oxide), poly(vinylimidazole), polycarboxylates such as polyacrylates, maleic/acrylic acid copolymers and lauryl methacrylate/acrylic acid co- polymers.

20           The detergent may contain a bleaching system, which may comprise a H<sub>2</sub>O<sub>2</sub> source such as perborate or percarbonate that may be combined with a peracid-forming bleach activator such as tetraacetythylenediamine or nonanoyloxybenzenesulfonate. Alternatively, the bleaching system may comprise peroxy acids of, e.g., the amide, imide, or sulfone type.

25           The enzyme(s) of the detergent composition may be stabilized using conventional stabilizing agents, e.g., a polyol such as propylene glycol or glycerol, a sugar or sugar alcohol, lactic acid, boric acid, or a boric acid derivative, e.g., an aromatic borate ester, or a phenyl boronic acid derivative, such as 4-formylphenyl boronic acid, and the composition may be formulated as described in, e.g., WO 92/19709 and WO 92/19708.

The detergent may also contain other conventional detergent ingredients such as e.g. fabric conditioners including clays, foam boosters, suds suppressors, anti-corrosion agents, soil-suspending agents, anti-soil re-deposition agents, dyes, bactericides, optical brighteners, hydrotropes, tarnish inhibitors, or perfumes.

5 It is at present contemplated that in the detergent compositions, any enzyme, in particular, one or more of the variant enzymes described herein, may be added, e.g., at about 0.01 mg to about 100 mg of enzyme protein per liter of wash liquor. In one embodiment, about 0.055 mg of enzyme protein per liter of wash liquor are used. In other embodiments, about 0.1 mg to about 1.0 mg of enzyme protein per liter of wash  
10 liquor are used.

One or more of the variant enzymes described herein may additionally be incorporated in the detergent formulations disclosed in WO 97/07202, which is hereby incorporated as reference.

#### 10.4 Dish Wash Detergent Compositions

15 The enzymes may also be used in dish wash detergent compositions, including the following:

##### 1) POWDER AUTOMATIC DISHWASHING COMPOSITION

	Nonionic surfactant	0.4-2.5%
	Sodium metasilicate	0-20%
20	Sodium disilicate	3-20%
	Sodium triphosphate	20-40%
	Sodium carbonate	0-20%
	Sodium perborate	2-9%
	Tetraacetyl ethylene diamine (TAED)	1-4%
25	Sodium sulphate	5-33%
	Enzymes	0.0001-0.1%

##### 2) POWDER AUTOMATIC DISHWASHING COMPOSITION

30	Nonionic surfactant (e.g. alcohol ethoxylate)	1-2%
	Sodium disilicate	2-30%
	Sodium carbonate	10-50%
	Sodium phosphonate	0-5%
35	Trisodium citrate dihydrate	9-30%
	Nitritotrisodium acetate (NTA)	0-20%
	Sodium perborate monohydrate	5-10%

	Tetraacetyl ethylene diamine (TAED)	1-2%
	Polyacrylate polymer (e.g. maleic acid/acrylic acid copolymer)	6-25%
5	Enzymes	0.0001-0.1%
	Perfume	0.1-0.5%
	Water	5-10 %

### 3) POWDER AUTOMATIC DISHWASHING COMPOSITION

	Nonionic surfactant	0.5-2.0%
10	Sodium disilicate	25-40%
	Sodium citrate	30-55%
	Sodium carbonate	0-29%
	Sodium bicarbonate	0-20%
	Sodium perborate monohydrate	0-15%
15	Tetraacetyl ethylene diamine (TAED)	0-6%
	Maleic acid/acrylic acid copolymer	0-5%
	Clay	1-3%
	Polyamino acids	0-20%
20	Sodium polyacrylate	0-8%
	Enzymes	0.0001-0.1%

### 4) POWDER AUTOMATIC DISHWASHING COMPOSITION

	Nonionic surfactant	1-2%
25	Zeolite MAP	15-42%
	Sodium disilicate	30-34%
	Sodium citrate	0-12%
	Sodium carbonate	0-20%
	Sodium perborate monohydrate	7-15%
30	Tetraacetyl ethylene diamine (TAED) Polymer	0-3% 0-4%
	Maleic acid/acrylic acid copolymer	0-5%
	Organic phosphonate	0-4%
	Clay	1-2%
35	Enzymes	0.0001-0.1%
	Sodium sulphate	Balance

### 5) POWDER AUTOMATIC DISHWASHING COMPOSITION

	Nonionic surfactant	1-7%
40	Sodium disilicate	18-30%
	Trisodium citrate	10-24%
	Sodium carbonate	12-20%
	Monopersulphate (2 KHSO <sub>5</sub> .KHSO <sub>4</sub> .K <sub>2</sub> SO <sub>4</sub> )	15-21%
45	Bleach stabilizer	0.1-2%



	Maleic acid/acrylic acid copolymer	0-6%
	Diethylene triamine pentaacetate, pentasodium salt	0-2.5%
	Enzymes	0.0001-0.1%
5	Sodium sulphate, water	Balance

#### 6) POWDER AND LIQUID DISHWASHING COMPOSITION WITH CLEANING SURFACTANT SYSTEM

	Nonionic surfactant	0-1.5%
10	Octadecyl dimethylamine N-oxide dihydrate	0-5%
	80:20 wt. C18/C16 blend of octadecyl dimethylamine N-oxide dihydrate and hexadecyldimethyl amine N- oxide dihydrate	0-4%
15	70:30 wt. C18/C16 blend of octadecyl bis (hydroxyethyl)amine N-oxide anhydrous and hexadecyl bis (hydroxyethyl)amine N-oxide anhydrous	0-5%
	C <sub>13</sub> -C <sub>15</sub> alkyl ethoxysulfate with an average degree of ethoxylation of 3	0-10%
20	C <sub>12</sub> -C <sub>15</sub> alkyl ethoxysulfate with an average degree of ethoxylation of 3	0-5%
	C <sub>13</sub> -C <sub>15</sub> ethoxylated alcohol with an average degree of ethoxylation of 12	0-5%
25	A blend of C <sub>12</sub> -C <sub>15</sub> ethoxylated alcohols with an average degree of ethoxylation of 9	0-6.5%
	A blend of C <sub>13</sub> -C <sub>15</sub> ethoxylated alcohols with an average degree of ethoxylation of 30	0-4%
	Sodium disilicate	0-33%
30	Sodium tripolyphosphate	0-46%
	Sodium citrate	0-28%
	Citric acid	0-29%
	Sodium carbonate	0-20%
	Sodium perborate monohydrate	0-11.5%
	Tetraacetyl ethylene diamine (TAED)	0-4%
35	Maleic acid/acrylic acid copolymer	0-7.5%
	Sodium sulphate	0-12.5%
	Enzymes	0.0001-0.1%

#### 7) NON-AQUEOUS LIQUID AUTOMATIC DISHWASHING COMPOSITION

40	Liquid nonionic surfactant (e.g. alcohol ethoxylates)	2.0-10.0%
	Alkali metal silicate	3.0-15.0%
	Alkali metal phosphate	20.0-40.0%
	Liquid carrier selected from higher glycols, polyglycols, polyoxides, glycolethers	25.0-45.0%
45	Stabilizer (e.g. a partial ester of phosphoric acid and a	0.5-7.0%

	C <sub>16</sub> -C <sub>18</sub> alkanol)	
	Foam suppressor (e.g. silicone)	0-1.5%
	Enzymes	0.0001-0.1%
5	8) NON-AQUEOUS LIQUID DISHWASHING COMPOSITION	
	Liquid nonionic surfactant (e.g. alcohol ethoxylates)	2.0-10.0%
	Sodium silicate	3.0-15.0%
	Alkali metal carbonate	7.0-20.0%
	Sodium citrate	0.0-1.5%
10	Stabilizing system (e.g. mixtures of finely divided silicone and low molecular weight dialkyl polyglycol ethers)	0.5-7.0%
	Low molecule weight polyacrylate polymer	5.0-15.0%
	Clay gel thickener (e.g. bentonite)	0.0-10.0%
	Hydroxypropyl cellulose polymer	0.0-0.6%
15	Enzymes	0.0001-0.1%
	Liquid carrier selected from higher glycols, polyglycols, polyoxides and glycol ethers	Balance
	9) THIXOTROPIC LIQUID AUTOMATIC DISHWASHING COMPOSITION	
20	C <sub>12</sub> -C <sub>14</sub> fatty acid	0-0.5%
	Block co-polymer surfactant	1.5-15.0%
	Sodium citrate	0-12%
	Sodium tripolyphosphate	0-15%
	Sodium carbonate	0-8%
25	Aluminum tristearate	0-0.1%
	Sodium cumene sulphonate	0-1.7%
	Polyacrylate thickener	1.32-2.5%
	Sodium polyacrylate	2.4-6.0%
	Boric acid	0-4.0%
30	Sodium formate	0-0.45%
	Calcium formate	0-0.2%
	Sodium n-decydiphenyl oxide disulphonate	0-4.0%
	Monoethanol amine (MEA)	0-1.86%
	Sodium hydroxide (50%)	1.9-9.3%
35	1,2-Propanediol	0-9.4%
	Enzymes	0.0001-0.1%
	Suds suppressor, dye, perfumes, water	Balance
	10) LIQUID AUTOMATIC DISHWASHING COMPOSITION	
40	Alcohol ethoxylate	0-20%
	Fatty acid ester sulphonate	0-30%
	Sodium dodecyl sulphate	0-20%
	Alkyl polyglycoside	0-21%
	Oleic acid	0-10%
45	Sodium disilicate monohydrate	18-33%

	Sodium citrate dihydrate	18-33%
	Sodium stearate	0-2.5%
	Sodium perborate monohydrate	0-13%
	Tetraacetyl ethylene diamine (TAED)	0-8%
5	Maleic acid/acrylic acid copolymer	4-8%
	Enzymes	0.0001-0.1%

#### 11) LIQUID AUTOMATIC DISHWASHING COMPOSITION CONTAINING PROTECTED BLEACH PARTICLES

10	Sodium silicate	5-10%
	Tetrapotassium pyrophosphate	15-25%
	Sodium triphosphate	0-2%
	Potassium carbonate	4-8%
	Protected bleach particles, e.g. chlorine	5-10%
15	Polymeric thickener	0.7-1.5%
	Potassium hydroxide	0-2%
	Enzymes	0.0001-0.1%
	Water	Balance

20 12) Automatic dishwashing compositions as described in 1), 2), 3), 4), 6) and 10), wherein perborate is replaced by percarbonate.

13) Automatic dishwashing compositions as described in 1)-6) which additionally contain a manganese catalyst. The manganese catalyst may, e.g., be one of the compounds described in "Efficient manganese catalysts for low-temperature bleaching",  
 25 *Nature* 369: 637-639 (1994).

#### 14) PREMIUM HDL LIQUID DETERGENT FORMULATIONS

	Bio-Soft S-101	Linear alkylbenzene sulfonic acid
	Steol CS-330	Sodium Laureth sulfate
	Bio-soft N25-7	Linear alkylethoxylate with 7 moles of EO
30	Stepanate SXS	Sodium xylene sulfonate

#### 15) ULTRA LIQUID DETERGENT FORMULATION

	Tionopal CBS-X	Fluorescent whitening agent
	Alpha-step MC-48	Sodium alpha-sulfomethylester
	Makon TD-6	Tridecylalcoholethoxylate

35

### 11. Compositions Comprising the Variant Alpha-Amylases

In one of its several aspects, this disclosure provides compositions comprising:

a) at least one variant alpha-amylase comprising an amino acid sequence at least 95% identical to that of a parent AmyS-like alpha-amylase, and having a substitution at an amino acid position corresponding to position 242 of a reference alpha-amylase, said variant having detectable alpha-amylase activity, and

b) at least one of an additional enzyme, a detergent, a surfactant, a chelator, an oxidizing agent, an acidulant, an alkalizing agent, a source of peroxide, a source of hardness, a salt, a detergent complexing agent, a polymer, a stabilizing agent, or a fabric conditioner.

In preferred embodiments the variant is altered, as compared to a parent AmyS-like alpha-amylase or a reference amylase, in one or more of a variety of properties that can alter its use or performance for certain applications, e.g., commercial processes described herein. The altered properties can include any property, for example, such as net charge, substrate specificity, substrate cleavage, substrate binding, thermal stability, activity at one or more pH's, stability at one or more pH's, stability in oxidizing conditions,  $\text{Ca}^{2+}$  requirements, specific activity, catalytic rate, catalytic efficiency, activity in the presence of a chelator, thermal or pH stability in the presence of a chelator, utility for desizing, or utility for a cleaning process, or amount of expression in a protein expression system. As the skilled artisan will appreciate, these altered properties preferably have utility to the end-user, or to the producer of the amylase, or both.

A number of amylases of known or readily-determined sequence can be used as the reference amylase. In various embodiments, the reference amylase is SEQ ID NOS: 1 or 2. The parent amylase and the reference amylase can be the same amylase in some embodiments.

The composition is, in certain embodiments, a component of a product for use in laundry, dish, or hard-surface cleaning, desizing, or fabric or stain treatment. For example the composition may be part of a dishwashing detergent for application as a liquid, semi-solid, solid, etc, or it can be a granular or liquid laundry detergent formulation. The composition comprises additional components as required for the

intended application. Examples of many such formulations are provided herein, and still others will be familiar to those of skill in the art.

In one embodiment, the composition comprises an additional enzyme that is a protease, a lipase, an amylase, a cellulase, a peroxidase, an oxidase, a pectinase, a lyase, a cutinase, or a laccase, or other useful enzyme. The skilled artisan will be familiar with these and other enzymes that may be useful in connection with the variant amylases provided herein. The amounts of enzyme that are useful can be determined empirically for a given application, however, guidelines are provided herein, e.g., in the examples.

In various embodiments, the composition comprises one or more surfactants. The surfactant is generally nonionic, anionic, cationic, or zwitterionic, or a combination thereof.

In one embodiment, the amylase variant is preferably a S242A, S242D, S242E, S242F, S242G, S242H, S242L, S242M, S242N, S242Q, or S242T variant. For certain uses, such as in washing and cleaning embodiments, stability to oxidation and stability to chelators or altered metal ion concentrations are useful. Accordingly in various embodiments, the variant amylase has altered stability to oxidation and the variant further includes deletion or substitution of one or more methionine residues including residues located at amino positions 8, 9, 96, 200, 206, 284, 307, 311, 316, and 438 of a parent amylase, where the reference amylase in SEQ ID NOS: 1 or 2. Variant amylases can further comprise an amino acid sequence modification at one or more amino acid positions corresponding to amino acid positions 97, 179, 180, 193, 319, 349, 358, 416, 428, or 443 of the reference amylase (which is preferably SEQ ID NOS: 1 or 2).

In various embodiments, the variant comprises or further comprises one or more of substitution at positions as follows: a cysteine at 349, a cysteine at 428, a glutamic acid at 97, an arginine at 97, a glutamic acid at 319, an arginine at 319, a glutamic acid at 358, an arginine at 358, a glutamic acid at 443, or an arginine at 443.

Moreover, the variant in one embodiment comprises a substitution of an N193 or a V416 or both, for example, a substitution that is N193F, or V416G, or both. Other embodiments include further modification such as deletion of one or more amino acids at positions F178, R179, G180, I181, G182 and K183. As described elsewhere herein, such

deletions may be even more useful when provided in pair-wise fashion or more.

Preferably in such embodiments, the variant has altered metal ion dependence, or altered stability or activity in the absence of added calcium, or in the presence of a chelator, or a combination thereof. Such variants may also have excellent utility in cleaning and

5 washing processes.

In one embodiment, the variant alpha-amylase has at least 95% homology to SEQ ID NO: 2 and comprises a substitution of amino acid 242 relative to numbering in a reference amylase comprising the amino acid sequence SEQ ID NO: 1. As with the other embodiments described herein, the variant preferably has detectable alpha-amylase

10 activity, particularly under the conditions of use.

In certain presently preferred embodiments, the parent alpha-amylase is SEQ ID NO: 1, 2, 6, 7, 8, 9, 10, 11, 12, 15, or 16, and the reference amylase is SEQ ID NO: 1 or 2.

In various embodiments, the variant amylase has improved performance in a wash process at very low and very high pH's. In one embodiment, the wash performance is

15 improved at  $\text{pH} \geq$  about 8, relative to the parent amylase. More preferred are those variants with improved wash performance above about pH 8.5 to about pH 11.

The variant in one embodiment comprises a set of substitutions of a) Q97E, Q319E, Q358E, Q443E; b) Q97E, Q319R, Q358E, Q443R; c) Q97E, Q319R, Q358E; d) Q97E, Q319R, Q443E; e) Q97E, Q319R, Q443R; f) Q97E, Q358R; g) Q97E, Q443E; h) Q319R, Q358E, Q443E; or i) Q319R, Q358R, Q443E relative to the reference amylase, e.g. a SEQ ID NO: 1 or 2 amylase sequence.

In another aspect of the disclosure, provided are detergent or cleaning formulations comprising at least one variant amylase comprising an amino acid sequence at least 95% identical to that of a parent AmyS-like alpha-amylase. The amylase variants

25 have a substitution at an amino acid position corresponding to position 242 of a reference alpha-amylase, and have detectable alpha-amylase activity. Preferably, the reference amylase is SEQ ID NOS: 1 or 2.

The detergent or cleaning formulation preferably comprises an amylase variant that an S242 variant comprising at least a S242A, S242D, S242E, S242F, S242G, S242H,

30 S242L, S242M, S242N, S242Q, or S242T substitution. As with the compositions

provided above, the variant can comprise any one or combination of the variant features and alterations disclosed herein.

In another of its several aspects, this disclosure provides kits. One embodiment of the kit comprises

5 a) one or more variant alpha-amylases comprising an amino acid sequence at least 95% identical to that of a parent AmyS-like alpha-amylase, and having a substitution at an amino acid position corresponding to position 242 of a reference alpha-amylase, said variant having detectable alpha-amylase activity, and

10 b) at least one of an additional enzyme, a detergent, a surfactant, a chelator, an oxidizing agent, an acidulant, an alkalizing agent, a source of peroxide, a source of hardness, a salt, a detergent complexing agent, a polymer, a stabilizing agent, or a fabric conditioner.

In one embodiment, the kits further comprise instructions for using the kit in a process for desizing a woven material or washing or cleaning one or more items soiled with a starch-containing substance.

The skilled artisan will also appreciate that kits for making the described alpha-amylases are also provided. The kits provide representative sequences e.g. amino acid sequences and/or nucleic acid derived therefrom, for use as parent alpha-amylases and reference amylases.

20

## **12. Using Amylase Variants in Desizing and Washing/Cleaning Processes**

In another aspect, this disclosure provides methods of using the variant alpha-amylases in desizing of fabrics or other woven material, and in washing or cleaning processes.

25 In aspect, this disclosure provides methods of desizing a woven material subsequent to a weaving process. The methods generally comprise contacting the woven material with a variant alpha-amylase under conditions and for a time effective for at least partially removing sizing from the woven material. The variant comprises an amino acid sequence at least 95% identical to that of a parent AmyS-like alpha-amylase, and has

a substitution at an amino acid position corresponding to position 242 of a reference alpha-amylase. The variant has detectable alpha-amylase activity.

The variant is preferably altered in one more of its physical or enzymatic properties, as compared to a parent AmyS-like alpha-amylase or a reference amylase. In various embodiments, the amylase is altered in one or more characteristics of: net charge, substrate specificity, substrate cleavage, substrate binding, thermal stability, activity at one or more pH's, stability at one or more pH's, stability in oxidizing conditions, Ca<sup>2+</sup> requirements, specific activity, catalytic rate, catalytic efficiency, activity in the presence of a chelator, thermal or pH stability in the presence of a chelator, effectiveness for desizing, or amount of expression in a protein expression system.

Reference amylases are discussed above, and in one embodiment of the method, the reference amylase is SEQ ID NOS: 1 or 2.

In one embodiment, the parent alpha-amylase is SEQ ID NOS: 1, 2, 6, 7, 8, 9, 10, 11, 12, 15, or 16, and the reference amylase is SEQ ID NOS: 1 or 2. In certain embodiments, the variant is a S242A, S242D, S242E, S242F, S242G, S242H, S242L, S242M, S242N, S242Q, or S242T variant.

The variant can further comprise one or more substitutions at positions as follows: a cysteine at 349, a cysteine at 428, a glutamic acid at 97, an arginine at 97, a glutamic acid at 319, an arginine at 319, a glutamic acid at 358, an arginine at 358, a glutamic acid at 443, or an arginine at 443, wherein the reference amylase is SEQ ID NO: 1 or 2.

In another aspect, provided herein are methods of washing or cleaning. While washing and cleaning operations can frequently benefit from the inclusion of one or more enzyme activities, the washing or cleaning processes can subject the enzymes, including amylases to extreme conditions and challenge the limits of the enzyme activity. Accordingly, the methods provided comprise contacting one or more items to be washed or cleaned with a composition comprising a variant alpha-amylase comprising an amino acid sequence at least 95% identical to that of a parent AmyS-like alpha-amylase, and having a substitution at an amino acid position corresponding to position 242 of a reference alpha-amylase. The variant preferably has detectable alpha-amylase activity, and the contacting step is under conditions and for a time effective for at least partially



washing or cleaning the one or more items. Preferably, at least one of the one or more items is soiled with at least one starch-containing material, the removal of which is aided by the variant amylase.

5 In one embodiment, the composition further comprises at least one component of a detergent composition or a cleaning formulation. For example, the composition comprises one or more of an additional enzyme, a detergent, a surfactant, a chelator, an oxidizing agent, an acidulant, an alkalizing agent, a source of peroxide, a source of hardness, a salt, a detergent complexing agent, a polymer, a stabilizing agent, or a fabric conditioner.

10 In one embodiment, the parent alpha-amylase can be any of SEQ ID NOS: 1, 2, 6, 7, 8, 9, 10, 11, 12, 15, or 16, and the reference amylase is SEQ ID NOS: 1 or 2. In certain embodiments, the parent alpha-amylase is conveniently SEQ ID NOS: 1, 2, 15, or 16, while in others, the parent alpha-amylase is SEQ ID NOS: 6, 7, 8, 9, 10, 11, or 12.

15 In presently preferred embodiments, the variant is a S242A, S242D, S242E, S242F, S242G, S242H, S242L, S242M, S242N, S242Q, or S242T variant. The reference amylase is SEQ ID NO: 1 or 2, and the variant is a S242A, S242D, S242E, S242F, S242G, S242H, S242L, S242M, S242N, S242Q, or S242T variant in certain embodiments.

20 In various embodiments, e.g., where the variant is a S242A, S242D, S242E, S242F, S242G, S242H, S242L, S242M, S242N, S242Q, or S242T variant, the variant further comprises a sequence modification at one or more amino acid positions corresponding to amino acid positions 97, 179, 180, 193, 319, 349, 358, 416, 428, or 443 of the reference amylase. More particularly, the variant comprises one or more of substitution at positions as follows: a cysteine at 349, a cysteine at 428, a glutamic acid at 25 97, an arginine at 97, a glutamic acid at 319, an arginine at 319, a glutamic acid at 358, an arginine at 358, a glutamic acid at 443, or an arginine at 443 in various embodiments. Substitution of an N193 or a V416 or both, such as a substitution of N193F or V416G, or both are also useful in certain variants.

30 In other embodiments, the variant comprises deletion of one or more amino acids at any of specific positions F178, R179, G180, I181, G182 and K183. In such

embodiments, the variant preferably has altered metal ion dependence or altered stability, or activity in the absence of added calcium or the presence of a chelator. As with the other modifications, the foregoing deletions of amino acids can also be used – alone or in combination with any of the foregoing alterations.

5           The variant generally has improved performance in a wash process relative to the parent amylase, for example under conditions such as  $\text{pH} \geq$  about 8.

          In one presently preferred embodiment, the method includes the use of a variant that comprises a set of substitutions of a) Q97E, Q319E, Q358E, Q443E; b) Q97E, Q319R, Q358E, Q443R; c) Q97E, Q319R, Q358E; d) Q97E, Q319R, Q443E; e) Q97E,  
10   Q319R, Q443R; f) Q97E, Q358R; g) Q97E, Q443E; h) Q319R, Q358E, Q443E; or i) Q319R, Q358R, Q443E.

          This disclosure includes further detail in the following examples, which are not in any way intended to limit the scope of what is claimed. The attached Figures are integral parts of the specification and description provided. All references cited are herein  
15   specifically incorporated by reference for all that is described therein. The following examples are thus offered to illustrate, but not to limit what is claimed.

## **EXAMPLES**

### **Example 1 - Construction of Variants**

20           The variants at position S242 of the mature sequence of AmyS were constructed using site directed mutagenesis. The template for mutagenesis was methylated pHPLT-AmyS (*see* Figure 2) using dam-Methylase from New England Biolabs (Massachusetts). Degenerate primers (S242F(forward) and S242R(reverse), given below) were synthesized and diluted to 10  $\mu\text{M}$  at Operon (Huntsville, AL) with complementary forward and  
25   reverse sequences both containing a 5' phosphate group for ligation in the reaction. The sequence of the parent alpha-amylase (SEQ ID NO: 2) is attached hereto. Libraries were created with the Stratagene Quik-Change™ Multi-site kit (Stratagene, La Jolla CA) using oligonucleotide primers randomized with NN(G/C) at the target position. The selected amino acid (i.e., S242) was randomly replaced with all 19 possible alternatives.

30

S242 primers for mutagenesis:

S242 F:

5'[Phos]GTCAAGCATATTAAGTTCNNSTTTTTTCCTGATTGGTTG 3' SEQ ID NO: 17

S242 R:

5'[Phos]CAACCAATCAGGAAAAAASNNGAACTTAATATGCTTGAC 3' SEQ ID NO: 18

The reaction was performed as follows:

**QUIK-CHANGE reaction:**

The reaction consisted of 18  $\mu\text{L}$  of sterile distilled  $\text{H}_2\text{O}$ , 2.5  $\mu\text{L}$  of 10x buffer from the kit, 1  $\mu\text{L}$  dNTPs from the kit, 1.25  $\mu\text{L}$  of the forward primers (of 10  $\mu\text{M}$  stock), 1.25  $\mu\text{L}$  of the reverse primers (of 10  $\mu\text{M}$  stock), 1  $\mu\text{L}$  of pHPLT-AmyS plasmid DNA as template (~70 ng), and 1  $\mu\text{L}$  of the enzyme blend from the kit for a total of 26.5  $\mu\text{L}$ .

**Cycling conditions:**

The cycling conditions were 95°C for 1 min once, then 95°C for 1 min, 55°C for 1 min, 65°C for 10 min for 25 cycles.

One microliter *DpnI* (10 U/ $\mu\text{L}$ ) was added to the Multi-site Quik-Change™ reaction mixture and incubated at 37°C for 18 hours and then another 0.5  $\mu\text{L}$  was added for an additional 3 hours.

One microliter of *DpnI* digested reaction was used as template for rolling circle amplification with the TEMPLIPHI amplification kit (Amersham Biosciences, Piscataway, NJ), and the reaction was performed according to the Amersham protocol.

One microliter of rolling circle DNA was transformed into 100  $\mu\text{L}$  of *Bacillus subtilis* competent cells (2 protease deleted *B. subtilis* strain ( $\Delta aprE$ ,  $\Delta nprE$ , *amyE::xylRPxylAcomK-phleo*)) and shaken at 37 °C for 1 hour. The entire transformation was next plated on LA + 10 ppm Neo + 1% insoluble starch plates (25  $\mu\text{L}$  one plate, 75  $\mu\text{L}$  on another plate) and incubated overnight at 37 °C. Ninety-six transformants were picked into 150  $\mu\text{L}$  of LB + 10 ppm Neo in a micro-titer plate and grown overnight at 37°C. The overnight plate was stamped onto a large LA + 10 ppm Neo + 1% insoluble starch plate with a 96 pin replicating tool and submitted to Quintara Biosciences (Berkeley, CA) for colony PCR and sequencing.

After variant sequences were determined, the variants were picked into a 96 well micro-titer plates containing 125  $\mu$ L of LB + 10 ppm Neo, arraying the variants into a quad format with controls. The arrayed micro-titer plate was grown for 6 hours at 37°C and 250 rpm. Using a replicating tool (EnzyScreen, Leiden, The Netherlands) the micro-titer culture plate was used to inoculate a new micro-titer plate (micro-titer plate and plate lids from EnzyScreen, Leiden, The Netherlands) containing 150  $\mu$ L of MBD medium for protein expression (G. Vogtentanz *et al.*, "A *Bacillus subtilis* fusion protein system to produce soybean Bowman-Birk protease inhibitor," *Prot. Expr. & Purif.*, 55: 40-52, 2007) and supplemented with 5 mM CaCl<sub>2</sub> for protein expression. Expression plates were grown for 64 hours at 37°C, 250 rpm, and 70% humidity. Expression cultures were next filtered through a micro-filter plate (0.22  $\mu$ m, Millipore, Billerica, MA) and screened for improved thermostability (*see* Example 3).

#### **Example 2 – Expression, Purification & Characterization of Variants**

Colonies were streaked from the microtiter plates from Example 1 onto starch plates with 10 ppm Neomycin. The plates were incubated overnight at 37°C, and single colonies were picked and used to inoculate shake flasks (250 mL with 25mL media) containing media (*see* below) and 20 ppm Neomycin. The cultures were grown at 37°C, 275 rpm, for about 8 hrs (until an OD (600 nm) of 2.0 was reached). The culture broths were mixed with 50% glycerol at 2:1 ratio, put into individually-labeled culture vials and frozen at -80°C. Subsequent production of the selected alpha-amylases were made from these glycerol stocks.

Fermentations for alpha-amylases were carried out in 500 mL shake flasks grown at 37°C for 60 hours in minimal MOPS culture medium (Neidhardt *et al.*, *J. Bacteriol.* 119(3):736-747, 1974) with 1% (w/v) Soytone. Enzymes were purified from the fermentation broth using hydrophobic interaction chromatography as follows: the broth was concentrated 10-fold then diluted back to its original volume with 50 mM MES, 2 mM CaCl<sub>2</sub>, pH 6.8 with 1 M ammonium sulfate, then sterile-filtered using a glass fiber filter. Samples were then loaded onto PHENYL SEPHAROSE FF high density column (20 x 95 mm; Amersham, GE Healthcare Bio-Sciences, Sweden) pre-equilibrated with

the same buffer. Non-amylase proteins were removed with 10 column volumes of the same buffer without ammonium sulfate followed by 5 column volumes of water. Enzymes of interest were eluted with 50 mM MES, 2 mM CaCl<sub>2</sub>, pH 6.8 containing 40% propylene glycol.

5 Protein concentrations were determined either with a standard quantitative SDS page gel densitometry method or using an activity assay using a standard amylase assay kit from Megazyme (Wicklow, Ireland). A standard curve generated using purified amylase (*Bacillus* 707 amylase; SEQ ID NO: 6) was used for comparing assay data.

### Example 3 - Determination of Altered Properties: Thermal stress

10 This example shows that the variants described herein may have an altered property relative to the parent alpha-amylase. A high-throughput thermal stability screen of *G. stearothermophilus* alpha-amylase (AmyS) variants was carried out.

After an initial investigation, heat-stress conditions were chosen such that the wild-type enzyme showed approximately 40% of its initial (pre-stress) activity after the  
15 heat stress (i.e., (activity after heat stress) / (activity before heat stress) was approximately 0.4). Libraries of mutants were screened in quadruplicate, and potential winners were identified as those that showed residual activity after heat stress that was at least two standard deviations more than the average residual activity of the wild-type enzyme.

20 Amylase expression was approximately 100 ppm in the culture supernatants of the expression plates. After 60-65 hours of growth at 37°C in a humidified shaker (250 rpm and 70% relative humidity), the culture supernatants were clarified to remove cellular material using filter plates. The clarified supernatants were diluted 10-fold into buffer containing 50 mM NaOAc / 2.6 mM CaCl<sub>2</sub> / 0.002% Tween-20, at pH 5.8., to a  
25 final concentration of approximately 10 ppm. One aliquot of each supernatant was further diluted to 0.02 ppm, for determination of activity of the enzyme variants as described below using a fluorescently-labeled corn starch substrate. A second aliquot of each supernatant was subjected to a 30 minute heat stress at 95°C in a thermocycler then diluted to 0.02 ppm in 50 mM NaOAc / 2.6 mM CaCl<sub>2</sub> / 0.002% Tween-20, at pH 5.8 and  
30 assayed for residual activity using the fluorescent substrate and assay described below.

Amylase activity was determined using the amylase ENZCHECK ULTRA AMYLASE assay kit essentially as described by the manufacturer (Invitrogen, San Diego CA). Final concentration of the amylase in the assay was approximately 0.02 ppm. Assay buffer was 50 mM NaOAc / 2.6 mM CaCl<sub>2</sub> / 0.002% Tween-20, pH 5.8. The substrate was BODIPY fluorescence dye conjugated 100 µg/mL DQ™ starch from corn (Invitrogen - Eugene, OR). Increased fluorescence, indicating amylase activity, was measured using a SpectraMAX M2 (Molecular Devices, Sunnyvale, CA). The reaction was monitored at room temperature for 5 minutes with the instrument recording in kinetic mode. Excitation wavelength was 485 nm; emission was monitored at 520 nm with a cutoff filter at 515 nm.

The wild-type AmyS (Xtra) showed 33-43% residual activity after being subject to thermal stress for 30 minutes at 95°C. AmyS variants, S242A and S242Q, retained 55-65% and 70-80% residual activities, respectively, following the same thermal stress conditions. See Figure 3 and Table 3-1. These residual activity measurements indicate the two variants are more thermostable than the wild-type alpha-amylase.

**Table 3-1:** Percent residual activities of each variant. Wild-type (SPEZYME XTRA). Each plate includes SPEZYME ETHYL and SPEZYME XTRA as controls as indicated.

Variant	% Residual Activity				Avg	Std. Dev
A	65.0	53.4	48.5	71.1	59.5	10.4
C	35.9	24.5	27.3	29.6	29.3	4.9
D	52.2	32.6	38.5	43.3	41.6	8.3
E	40.2	53.3	33.2	51.8	44.6	9.6
F	41.7	31.8	30.1	31.7	33.8	5.3
G	34.3	27.1	27.4	37.5	31.6	5.2
H	22.6	20.5	16.2	17.8	19.3	2.8
I	36.2	26.9	19.7	25.5	27.0	6.8
K	22.3	22.6	23.3	23.0	22.8	0.5
L	26.1	29.6	30.6	27.8	28.5	2.0
M	48.8	46.6	40.5	35.9	42.9	5.9
N	32.0	29.0	24.6	35.1	30.2	4.5
P	7.2	7.7	6.4	5.7	6.7	0.9
Q	61.0	65.7	49.1	69.3	61.3	8.8
R	14.5	14.3	11.7	11.7	13.0	1.5
wildtype	44.3	27.1	29.2	35.5	34.0	7.7
T	24.6	25.4	27.7	21.5	24.8	2.5
V	17.5	25.9	22.1	23.9	22.3	3.6

Variant	% Residual Activity				Avg	Std. Dev
	5.0	6.3	3.9	7.0		
W	5.0	6.3	3.9	7.0	5.6	1.4
Y	18.5	13.5	14.2	16.5	15.7	2.3
Ethyl	111.8	77.3	84.3	66.7	85.0	19.2
Xtra	27.1	36.1	40.7	25.2	32.3	7.4

#### Example 4 - Determination of Altered Properties: DSC

Spezyme Xtra, S242A, S242E, and S242Q were purified from shake flask fermentation broth (see Example 2) using hydrophobic interaction chromatography. The protein was eluted from the column in purified form using 50 mM MES, pH 6.8, containing 40% propylene glycol and 2 mM CaCl<sub>2</sub>.

Excessive heat capacity curves were measured using an  $\mu$ Ltrasensitive scanning high-throughput microcalorimeter, VP-CAP DSC (MicroCal, Inc., Northampton, MA). The standard procedure for DSC measurements and the theory of the technique has been published (E. Freire, "Differential Scanning Calorimetry," *Methods. Mol. Biol.* 41: 191-218, 1995). Approximately 500  $\mu$ L of 0.5 mg/mL wild-type *Bacillus stearothermophilus*  $\alpha$ -amylase or variant S242A, S242E, and S242Q (both in the absence and in the presence of 2 mM calcium chloride) were scanned over a 30-120°C temperature range. The same sample was then re-scanned to check the reversibility of the process. For  $\alpha$ -amylase, the thermal unfolding process was irreversible. The buffer used was 10 mM sodium acetate, pH 5.5. A 200°C/hr scan rate was used to minimize any artifacts that may have resulted from aggregation. The thermal midpoint ( $T_m$ ) of the DSC curves was used as an indicator of the thermal stability of the tested protein. Table 4-1 shows the thermal melting points for the amylase proteins tested. The thermal melting curves and the melting points for the wild-type and variant amylases are shown in Figure 5.

The thermal unfolding for the amylase variants S242A, S242E, and S242Q in the absence and presence of 2 mM calcium chloride show considerable increase in the melting points for the variants when compared to that for the wild-type. In the absence of added calcium chloride, the wild-type amylase has a thermal melting point of 100.8°C while the  $T_m$ 's for S242A, S242E, and S242Q are 106.5°C, 107.8°C, and 110.1°C, respectively. Thus, the substitution of S242 with A results in an increase in the  $T_m$  of

5.7°C; the substitution of S242 with E results in an increase in the  $T_m$  of 7.0°C; and the substitution of S242 with Q results in an increase in the  $T_m$  of 9.3°C.

In the presence of 2 mM calcium chloride, the wild-type amylase displayed a thermal melting point of 106.8°C while the  $T_m$ 's for S242A, S242E, and S242Q were 111.8°C, 112.2°C, and 113.8°C, respectively. Thus, relative to measurements in the absence of calcium, in the presence of 2 mM calcium chloride, all four proteins had increased  $T_m$  values. The increase in  $T_m$  for wild-type and the S242A variants in the presence of calcium was 6°C and 5.3°C, respectively. The increase in  $T_m$  for the S242E variant was 4.4°C. The increase in  $T_m$  for the S242Q variant was 3.7°C. This suggests that the S242Q variants is stabilized less by calcium, or the variant is less dependent on calcium for stability. The increase in the  $T_m$  of the S242A, S242E, and S242Q relative to wild-type in the presence of calcium chloride was 5°C, 5.4°C, and 3°C, respectively. This suggests that the thermodynamic properties of the variants differ from those of the wild-type, or Spezyme Xtra. This observation was consistent with its enhanced performance in application studies (*see* Example 5).

Table 4-1  $T_m$  (°C) for various amylases by DSC

	$T_m$ (No $Ca^{2+}$ )	$\Delta T$ (°C)	$T_m$ (w/ 2 mM $Ca^{2+}$ )	$\Delta T$ (°C)
Spezyme Xtra	100.8		106.8	
S242A	106.5	5.7	111.8	5.7
S242E	107.8	7.0	112.2	5.4
S242Q	110.1	9.3	113.8	7.0

#### Example 5 – Activity Profiles

This example shows that the tested variants have altered activity profiles relative not only to the parent alpha-amylase but also to an industry standard enzyme. Protein determinations were made on purified or plate samples. The variants and standard alpha-amylases were each assayed on the basis of equal protein concentration.

Either plate or purified variants were diluted to approximately 20 ppm using pH 5.6 malic acid buffer. The substrate consisted of 15% cornstarch in the same 50 mM Malic acid buffer, pH 5.6. Four hundred microliters of the starch suspension was



equilibrated to 70°C for 2.5 minutes. Seven (7)  $\mu\text{L}$  of the diluted enzyme was quickly added to the equilibrated starch at a final protein concentration of about 0.36 ppm. The reaction mix was then put into a pre-heated 85°C shaking heating block and mixed at 300 rpm. The reactions were quenched with 50  $\mu\text{L}$  of 125 mM NaOH at predetermined time intervals. The reaction tubes were spun and the supernatant was diluted 10 fold into 10 mM NaOH, for analysis of DP profile by HPAEC-PAD.

Reactions were set up for 4, 10 and 20 minutes. The 4 min reaction provides an indication of the enzyme initial conversion of product to substrate; the 10 minute reaction provides an indication of the enzyme's thermal activity, and the 20 minute reaction provides an indication of the enzyme's thermal stability.

Total area from DP2 to the end of the HPLC run was integrated, and divided by the total protein and reaction time. The results are provided in Figures 6 and 7.

#### Example 6 - Additional Methods

The following assays were used in the Examples. Deviations from the protocols provided below are generally indicated in the Examples. In these experiments, a spectrophotometer was used to measure the absorbance of the products formed during the reactions.

##### A. Protein Content Determination

**BCA (bicinchoninic acid) Assay.** BCA (Pierce) assay was used to determine the protein concentration in samples on microtiter plate (MTP) scale. The chemical and reagent solutions used were: BCA protein assay reagent, and Pierce dilution buffer (50 mM MES, pH 6.5, 2 mM  $\text{CaCl}_2$ , 0.005% TWEEN®-80). The equipment included a SpectraMAX (type 340; Molecular Devices) MTP reader. The MTPs were obtained from Costar (type 9017).

Two-hundred (200)  $\mu\text{L}$  BCA Reagent was pipetted into each well, followed by 20  $\mu\text{L}$  diluted protein. After thorough mixing, the MTPs were incubated for 30 minutes at 37°C. Air bubbles were removed before the optical density (OD) of the solution in the wells was read at 562 nm. To determine the protein concentration, the background reading was subtracted from the sample readings. The  $\text{OD}_{562}$  was plotted for protein

standards (purified enzyme) to produce a standard curve. The protein concentration of the samples were interpolated from the standard curve.

**Bradford Assay.** The Bradford dye reagent (Quick Start) assay was used to determine the protein concentration in samples on MTP scale. The chemical and reagent solutions used were: Quick Start Bradford Dye Reagent (BIO-RAD Catalog No. 500-0205), Dilution buffer (10 mM NaCl, 0.1 mM CaCl<sub>2</sub>, 0.005% TWEEN®-80). The equipment used was a Biomek FX Robot (Beckman) and a SpectraMAX (type 340) MTP reader. The MTPs were from Costar (type 9017).

Two-hundred (200) µL Bradford dye reagent was pipetted into each well, followed by 15 µL dilution buffer. Ten (10) µL of filtered culture broth were added to the wells. After thorough mixing, the MTPs were incubated for at least 10 minutes at room temperature. Air bubbles were blown away and the OD of each well was read at 595 nm. To determine the protein concentration, the background reading (*i.e.*, from uninoculated wells) was subtracted from the sample readings. The OD<sub>595</sub> values obtained provide a relative measure of the protein content in the samples.

#### **B. Microswatch Assay for Testing Enzyme Performance**

The detergents used in this assay did not contain enzymes or the enzymes present in commercial detergents had been destroyed through heat deactivation as described elsewhere in this document. The equipment used included an Eppendorf Thermomixer and a SpectraMAX (type 340) MTP reader. The MTPs were obtained from Costar (type 9017).

**Detergent Preparation** (AATCC HDL; US conditions). Milli-Q water was adjusted to 6 gpg water hardness (Ca/Mg=3/1), and 1.5 g/l AATCC 2003 standard reference liquid detergent without brightener was added. The detergent solution was vigorously stirred for at least 15 minutes. Then, 5 mM HEPES (free acid) was added and the pH adjusted to 8.0.

**Rice Starch Microswatch Assay for testing Amylase Performance.** Test detergents were prepared as described elsewhere in this document. The equipment used included a New Brunswick Innova 4230 shaker/incubator and a SpectraMAX (type 340) MTP reader. The MTPs were obtained from Corning (type 3641). Aged rice starch with

orange pigment swatches (CS-28) were obtained from Center for Test Materials (Vlaardingen, Netherlands). Before cutting 0.25-inch circular microswatches, the fabric was washed with water. Two microswatches were placed in each well of a 96-well microtiter plate. The test detergent was equilibrated at 20°C (North America) or 40°C (Western Europe). 190 µL of detergent solution were added to each well of the MTP, containing microswatches. To this mixture, 10 µL of the diluted enzyme solution was added. The MTP was sealed with adhesive foil and placed in the incubator for 1 hour with agitation at 750 rpm at the desired test temperature (typically 20°C or 40 C). Following incubation, 150 µL of the solution from each well were transferred into a fresh MTP and read at 488 nm using a SpectraMAX MTP reader to quantify cleaning. Blank controls, as well as controls containing microswatches and detergent, but no enzyme, were also included.

**Calculation of Enzyme Performance.** The obtained absorbance value was corrected for the blank value (*i.e.*, obtained after incubation of microswatches in the absence of enzyme). The resulting absorbance was a measure of the hydrolytic activity.

### C. Amylase Concentration Determination by Antibody Titration

Alpha-amylase concentration and specific activity was determined, in some cases, by titration with an inhibitory polyclonal antibody. Polyclonal antibodies raised to *Bacillus stearothermophilus* alpha-amylase (AmyS) were found to be strongly inhibitory of AmyS and the alpha-amylase from *Bacillus sp.* TS-23 (e.g., the binding is tight enough to produce a linear titration of activity loss). Therefore, this antibody can be used to measure enzyme concentration, which, in turn, is used to calculate specific activity.

Briefly, the amount of enzyme inhibition produced by several known concentrations of antibody is measured. From this information, the concentration of antibody required for complete inhibition is extrapolated, which is equivalent to the enzyme concentration in the sample. Alpha-amylase activity and inhibition was measured using the fluorogenic BODIPY-starch assay. The buffer was 50 mM MOPS, pH 7.0, containing 0.005% Tween-80.

A polyclonal antibody directed against purified AmyS was raised in a rabbit and purified by standard methods. An empirical “apparent concentration” value of an

antibody stock solution was determined by measuring the inhibition of a sample of AmyS of known specific activity. The antibody sample was used to determine the concentration and specific activity of AmyS and TS23t variants. These values were used to create normalized 96-well enzyme stock plates, in which all of the variants were diluted to a  
5 common concentration.

#### **D. Native Protein Gel Electrophoresis**

Electrophoretic mobility of variant protein samples was measured using the PHASTGEL system (GE Healthcare) on pre-cast native polyacrylamide gels (PHASTGEL Homogeneous) at either 7.5% or 12.5% concentration. Buffer strips  
10 (PHASTGEL Native) were used and consisted of pH 8.8 in 0.88 M L-Alanine, 0.25 M Tris buffer. Typical run conditions consisted of 400 V for 12.75 minutes with an anode-to-cathode distance of 3.7 cm.

Alternatively, electrophoretic mobility of variant protein samples was measured on 1 mm-thick 0.5-1.5% agarose gels at various pH values (i.e. 5.8, 8.0 and 10.0) through  
15 a choice of a suitable buffer system. The electrophoresis was carried out under non-denaturing conditions. The Cathode–Anode length was 13.9 cm. A sample of 1-2 µg protein was mixed with 5% glycerol + 0.05% bromophenol blue and loaded on each lane. Gels were run typically for 1 hour at 100 V.

Gels were stained with Louiseville blue dye dissolved in 10% acetic acid and  
20 destained with 10% methanol and 10% acidic acid-in-water. Between 12 and 20 protein variants were loaded simultaneously, depending on native gel system used. As a consequence, the electrophoretic mobility of a protein variant can be immediately assessed, relative to charge ladder standards loaded on the same gel.

#### **E. Detergent Heat Inactivation**

Heat inactivation of commercial detergent formulas serves to destroy the  
25 enzymatic activity of any protein components while retaining the properties of non-enzymatic components. Thus, this method was suitable for preparing commercially-purchased detergents for use in testing the enzyme variants. For North American (NA) and Western European (WE) heavy duty liquid laundry (HDL) detergents, heat  
30 inactivation was performed by placing pre-weighed liquid detergent (in a glass bottle) in

a water bath at 95°C for 2 hours. The incubation time for heat inactivation of North American (NA) and Japanese (JPN) heavy duty granular laundry (HDG) detergent was 8 hours and that for Western European (WE) HDG detergent was 5 hours. The incubation time for heat inactivation of NA and WE auto dishwashing (ADW) detergents was 8 hours. The detergents were purchased from local supermarket stores. Both un-heated and heated detergents were assayed within 5 minutes of dissolving the detergent to accurately determine percentage deactivated. Enzyme activity was tested by the suc-AAPF-pNA assay.

For testing of enzyme activity in heat-inactivated detergents, working solutions of detergents were made from the heat inactivated stocks. Appropriate amounts of water hardness (6 gpg or 12 gpg) and buffer were added to the detergent solutions to match the desired conditions (Table 6-1). The solutions were mixed by vortexing or inverting the bottles.

Region	Form	Dose	Detergent*	Buffer	Gpg	pH	T (°C)
Laundry (heavy duty liquid and granular)							
NA	HDL	0.78 g/l	P&G TIDE® 2X	5 mM HEPES	6	8.0	20
WE	HDL	5.0 g/L	Henkel Persil	5 mM HEPES	12	8.2	40
WE	HDG	8.0 g/L	P&G Ariel	2 mM Na <sub>2</sub> CO <sub>3</sub>	12	10.5	40
JPN	HDG	0.7 g/L	P&G TIDE®	2 mM Na <sub>2</sub> CO <sub>3</sub>	6	10.0	20
NA	HDG	1.0 g/L	P&G TIDE®	2 mM Na <sub>2</sub> CO <sub>3</sub>	6	10.0	20
Automatic Dish Washing							
WE	ADW	3.0 g/L	RB Calgonit	2 mM Na <sub>2</sub> CO <sub>3</sub>	21	10.0	40
NA	ADW	3.0 g/L	P&G Cascade	2 mM Na <sub>2</sub> CO <sub>3</sub>	9	10.0	40

\* Abbreviations: Procter & Gamble (P&G); and Reckitt Benckiser (RB).

15

#### **F. TERG-O-TOMETER Assay For Cleaning Performance Determination**

A standard protocol for assessing protein and carbohydrate soil cleaning was used whereby the soil level on a fabric swatch was measured before and after cleaning under standard conditions. The fabric swatches consisted of woven cotton fabric soiled with

20

either maize starch, rice starch or a blood, milk, and carbon black mixture. Swatches were purchased from Testfabrics, Inc. (West Pittston, PA). Maize Starch (EMPA 161) and Blood, Milk, Carbon Black (EMPA 116) technical soils were produced by EMPA Test materials AG (St. Gallen, Switzerland). Rice Starch (CFT CS-28) soils were  
5 produced by the Center for Testmaterials BV (Vlaardingen, Netherlands). Each stain was measured before and after treatment by optical reflectance using a Minolta Reflectometer CR-410 set to a D65 (6500°K) standard illuminant. The difference in the L, a, b values was converted to total color difference (dE), as defined by the CIE-LAB color space. Cleaning of the stains are expressed as percent stain removal index (%SRI) by taking a  
10 ratio between the color difference before and after washing and comparing it to the difference of unwashed soils (before wash) to unsoiled fabric.

Cleaning experiments were conducted in a TERG-O-TOMETER (United States Testing Co., Hoboken, NJ) equipped with 6 stainless steel 2 L pots fitted with overhead agitators. Each treatment was conducted in 1 L total volume consisting of either 6 grains  
15 per gallon 3:1 (calcium:magnesium) water hardness or 12 grains per gallon water hardness. Detergents used in the wash experiments were 1.5 g/L AATCC HDL WOB 2003 liquid detergent with 5 mM HEPES buffer at pH 8, 0.7g/L AATCC HDD WOB 1993 granular detergent, 8 g/L IEC A\* 60456 granular detergent with perborate and TAED bleach, or 5 g/L Persil Power Gel liquid detergent. Enzyme was added directly  
20 into the wash solution and reactions were then initiated by addition of either 40 g/L or 200 g/L of soiled and ballast fabric. The washing reactions were agitated at 100 rpm for 10, 15, or 40 minutes at 20°C, 25°C, 30°C, 40°C, or 50°C. Following cleaning, swatches were rinsed for 3 minutes in tap water, spun in a front-loading washing machine at 1000 rpm to remove excess water, and dried in a dryer at low heat on a permanent press cycle  
25 for approximately 45 minutes. Comparison of the extent of soil removal was assessed by reflectometry and expressed as the % soil removal index (%SRI). The control condition did not contain enzyme and the positive control consisted of various doses of benchmark commercial enzymes.

#### **G. BODIPY-Starch Assay for Determination of Amylase Activity**

30 The BODIPY-starch assay was performed using the EnzChek® Ultra Amylase

Assay Kit (E33651, Invitrogen). A 1 mg/mL stock solution of the DQ starch substrate was prepared by dissolving the contents of the vial containing the lyophilized substrate in 100  $\mu$ L of 50 mM sodium acetate buffer at pH 4.0. The vial was vortexed for about 20 seconds and left at room temperature, in the dark, with occasional mixing until dissolved. 5 900  $\mu$ L of assay buffer (50 mM sodium acetate with 2.6 mM  $\text{CaCl}_2$  at pH 5.8) was added, and the vial was mixed by vortex for about 20 seconds. The substrate solution was stored at room temperature, in the dark, until ready to use or at 4°C. For the assay, a 100  $\mu$ g/mL of working solution of the DQ substrate was prepared from the 1 mg/mL substrate solution in the assay buffer. 190  $\mu$ L of 100  $\mu$ g/mL substrate solution was added to each 10 well in a flat-bottom 96-well microtiter plate. 10  $\mu$ L of each enzyme sample was added to a well, mixed for 30 seconds using a thermomixer at 800 rpm. A blank sample containing buffer and substrate only (no-enzyme blank) was included in the assay. The rate of change of fluorescence intensity was measured (excitation: 485 nm, emission: 520 nm) in a fluorescence microtiter plate reader at 25°C for 5 minutes.

#### 15 H. Measurement of Enzyme Binding to Macromolecular Substrates

Binding assays were done to determine substrate binding of Amylase (AmyS) charge ladder variants (charge change = -12 to +12 relative to wild-type AmyS) to corn stover and bagasse. Substrates used included bagasse (sugarcane bagasse from Brazil, dilute-acid pre-treated by National Renewable Energy Laboratory, washed and buffered 20 at pH 5), AFEX (ammonia fiber expansion corn stover), and PCS (dilute sulfuric acid pre-treated corn stover, washed and adjusted to pH 5). All substrates were brought to the desired percentage solids prior to use.

*Amylase Binding:* Amylase charge ladder variants were purified and diluted to 200 ppm for testing. A 1% cellulose bagasse solution was prepared in borate buffer 25 (40mM, pH8.5, 0.016% Tween80). 150  $\mu$ L of the bagasse solution was added into each well in a microtiter filtration plate. 150  $\mu$ L of borate buffer was added into a set of separate wells, which served as controls. 10  $\mu$ L of amylase charge ladder variants was added into the filtration plate, each condition was in duplicates. The plate was incubated at room temperature for 2 hours. The filtrate was collected and amylase activity in the 30 supernatant was measured by BODIPY-starch assay.

*Measurement of Enzyme Binding to Microswatches:* Alpha-amylase variants were incubated with or without CS-28 rice starch microswatches under standard wash conditions for 30 min. The amount of free enzyme was measured by the BODIPY-starch assay. The fraction of enzyme bound to the microswatches was calculated as follows:

5 Fraction bound = (Activity of enzyme in absence of swatch - Activity of enzyme in presence of swatch) / (Activity of enzyme in absence of swatch).

### Example 7 - Amylase Production in *B. subtilis*

In this Example, production of a mutant truncated form of *Bacillus*  
 10 *stearothermophilus* amylase alpha-amylase (having a S242Q mutation and a 29 amino acid deletion from the C-terminus; also referred to herein as S242Q) and variants thereof in *B. subtilis* are described. Transformation was performed as known in the art (*see e.g.*, WO 02/14490). Briefly, the gene encoding the parent amylases was cloned into the pHPLT expression vector, which contains the LAT promoter (PLAT), a sequence  
 15 encoding the LAT signal peptide (preLAT), followed by *Pst*I and *Hpa*I restriction sites for cloning.

The coding region for the LAT signal peptide is shown below:

atgaaacaac aaaaacggct ttacgcccga ttgctgaecg tgttatttgc gctcatcttc  
 ttgctgcctc attctgcagc ttcagca (SEQ ID NO: 19).

20 The amino acid sequence of the LAT signal peptide is shown below:

MKQQKRLYAR LLTLLFALIF LLPHSAASA (SEQ ID NO: 20)

The amino acid sequence of the mature truncated S242Q amylase with the substituted amino acid shown in italics was used as the basis for making the variant libraries described herein:

25 AAFPNGTMMQ YFEWYLPDDG TLWTKVANEI NNLSSLGITA LWLPPAYKGT SRSDVGYGVY  
 DLYDLGEFNQ KGTVRTKYGT KAQYLQAIQA AHAAGMQVYA DVVFDHKGGA DGTEWVDAVE  
 VNPSDRNQEI SGTYQIQAWT KFDFPGRGNT YSSFKWRWYH FDGVDWDESR KLSRIYKFRG  
 IGKAWDWEVD TENGNYDYLM YADLMDHPE VVTELKNWGK WYVNTTNIDG FRLDAVKHIK  
 FQFFPDWLSY VRSQTGKPLF TVGEYWSYDI NKLHNYITKT NGTMSLFDAP LHNKFYTASK  
 30 SGGAFDMRTL MTNTLMKDQP TLAVTFVDNH DTEPGQALQS WVDPWFKPLA YAFILTRQEG  
 YPCVFYGDYY GIPQYNIPSL KSKIDPLLIA RRDYAYGTQH DYLDHSDIIG WTREGVTEKP



GSGLAALITD GPGGSKWMYV GKQHAGKVFY DLTGNRSDTV TINS DGWGEF KVNGGSVSVW  
VPRKTT (SEQ ID NO: 21).

The PCR products were purified using QIAQUIK columns from Qiagen, and resuspended in 50  $\mu$ L of deionized water. 50  $\mu$ L of the purified DNA was digested with  
5 *HpaI* (Roche) and *PstI* (Roche), and the resultant DNA resuspended in 30  $\mu$ L of deionized water. 10-20 ng/ $\mu$ L of the DNA was cloned into plasmid pHPLT using *PstI* and *HpaI* cloning sites. The ligation mixtures were directly transformed into competent  
10 *B. subtilis* cells (genotype:  $\Delta vpr$ ,  $\Delta wprA$ ,  $\Delta mpr-ybfJ$ ,  $\Delta nprB$ ). The *B. subtilis* cells have a competency gene (*comK*) which is placed under a xylose inducible promoter, so xylose was used to induce competency for DNA binding and uptake (*see* Hahn *et al.*, *Mol. Microbiol.*, 21: 763-775, 1996).

The elements of plasmid pHPLT-AmyS include: pUB110 = DNA fragment from plasmid pUB110 (McKenzie *et al.*, *Plasmid* 15: 93-103, 1986). Plasmid features include:  
15 ori-pUB110 = origin of replication from pUB110; neo = neomycin resistance gene from pUB110; Plat = transcriptional promoter from *B. licheniformis* amylase; Pre LAT = signal peptide from *B. licheniformis* amylase; SAMY 425ss = the coding region for truncated *AmyE* gene sequence (replaced by the coding regions for each truncated *AmyE* variant expressed in this study); and Terminator = transcriptional terminator from *B. licheniformis* amylase.

20

### Example 8 - Expression of Enzyme Variants

This Example describes the methods used to express various recombinant enzymes of the transformed *B. subtilis* of the preceding Examples.

*Alpha-Amylase Expression – 2 mL scale.* *B. subtilis* clones containing S242Q  
25 (or a variant thereof) expression vectors were replicated with a steel 96-well replicator from glycerol stocks into 96-well culture plates (BD, 353075) containing 150  $\mu$ L of LB media + 10  $\mu$ g/mL neomycin, grown overnight at 37°C, 220 rpm in a humidified enclosure. A 100  $\mu$ L aliquot from the overnight culture was used to inoculate 2000  $\mu$ L defined media + 10  $\mu$ g/mL neomycin in 5 mL plastic culture tubes. The cultivation  
30 media was an enriched semi-defined media based on MOPS buffer, with urea as major

nitrogen source, glucose as the main carbon source, and supplemented with 1% SOY-TONE and 5 mM calcium for robust cell growth. Culture tubes were incubated at 37°C, 250 rpm, for 72 hours. Following this incubation, the culture broths were centrifuged for 10 minutes at 3000 x g. The supernatant solution was decanted into 15 mL polypropylene conical tubes; 80 µL of each sample were aliquoted into 96 well plates for protein quantitation.

### Example 9 - Production of Enzyme Variants

This Example describes the production of enzyme charge ladders and combinatorial charge libraries.

**Enzyme Charge Ladders.** Multiple protein variants spanning a range of physical properties of interest are selected from existing libraries or are generated by site-directed mutagenesis techniques as known in the art (*see e.g.*, US Pat. Appln. Ser. Nos., 10/576,331, 11/581,102, and 11/583,334, assigned to Genencor International). This defined set of probe proteins is then assayed in a test of interest.

Exemplary amylase charge ladder variants are shown in the following tables and assayed as described herein. In these tables, the charge change is relative to the parent enzyme.

Table 9-1. AmyS-S242Q Charge Ladder	
AmyS-S242Q Variant	Δ Charge
Q97E-Q319E-Q358E-Q443E	-4
Q97E-Q319E-Q358E	-3
Q97E-Q319E	-2
Q97E	-1
Q97R-Q319E	0
Parent AmyS-S242Q	0
Q97R	+1
Q97R-Q319R	+2
Q97R-Q319R-Q358R	+3
Q97R-Q319R-Q358R	+4

### Enzyme Combinatorial Charge Libraries (CCL)

**Generation of *B. stearothermophilus* AmyS-S242Q CCL.** The AmyS-S242Q plasmid DNA was isolated from a transformed *B. subtilis* strain (genotype:  $\Delta aprE$ ,  $\Delta nprE$ , *amyE::xylRPxylAcomK-phleo*) and sent to DNA2.0 Inc. as the template for CCL construction. A request was made to DNA2.0 Inc. (Mountain View, CA) for the generation of positional libraries at each of the four sites in AmyS-S242Q (S242Q) amylase that are shown in Table 9-2. Variants were supplied as glycerol stocks in 96-well plates.

The AmyS S242Q combinatorial charge library was designed by identifying the following four residues: Gln97, Gln319, Gln358, and Gln 443. A four site, 81-member CCL was created by making all combinations of three possibilities at each site: wild-type, arginine, or aspartic acid.

**Table 9-2. S242Q CCL Variants**

Variant #	Q97	Q319	Q358	Q443	$\Delta$ Charge
1	Q97E	Q319E	Q358E	Q443E	-4
2	Q97E	Q319E	Q358E	Q443R	-2
3	Q97E	Q319E	Q358E	-	-3
4	Q97E	Q319E	Q358R	Q443E	-2
5	Q97E	Q319E	Q358R	Q443R	0
6	Q97E	Q319E	Q358R	-	-1
7	Q97E	Q319E	-	Q443E	-3
8	Q97E	Q319E	-	Q443R	-1
9	Q97E	Q319E	-	-	-2
10	Q97E	Q319R	Q358E	Q443E	-2
11	Q97E	Q319R	Q358E	Q443R	0
12	Q97E	Q319R	Q358E	-	-1
13	Q97E	Q319R	Q358R	Q443E	0
14	Q97E	Q319R	Q358R	Q443R	+2
15	Q97E	Q319R	Q358R	-	+1
16	Q97E	Q319R	-	Q443E	-1
17	Q97E	Q319R	-	Q443R	+1
18	Q97E	Q319R	-	-	0
19	Q97E	-	Q358E	Q443E	-3
20	Q97E	-	Q358E	Q443R	-1
21	Q97E	-	Q358E	-	-2

<b>Table 9-2. S242Q CCL Variants</b>					
<b>Variant #</b>	<b>Q97</b>	<b>Q319</b>	<b>Q358</b>	<b>Q443</b>	<b><math>\Delta</math> Charge</b>
22	Q97E	-	Q358R	Q443E	-1
23	Q97E	-	Q358R	Q443R	+1
24	Q97E	-	Q358R	-	0
25	Q97E	-	-	Q443E	-2
26	Q97E	-	-	Q443R	0
27	Q97E	-	-	-	-1
28	Q97R	Q319E	Q358E	Q443E	-2
29	Q97R	Q319E	Q358E	Q443R	0
30	Q97R	Q319E	Q358E	-	-1
31	Q97R	Q319E	Q358R	Q443E	0
32	Q97R	Q319E	Q358R	Q443R	+2
33	Q97R	Q319E	Q358R	-	+1
34	Q97R	Q319E	-	Q443E	-1
35	Q97R	Q319E	-	Q443R	+1
36	Q97R	Q319E	-	-	0
37	Q97R	Q319R	Q358E	Q443E	0
38	Q97R	Q319R	Q358E	Q443R	+2
39	Q97R	Q319R	Q358E	-	+1
40	Q97R	Q319R	Q358R	Q443E	+2
41	Q97R	Q319R	Q358R	Q443R	+4
42	Q97R	Q319R	Q358R	-	+3
43	Q97R	Q319R	-	Q443E	+1
44	Q97R	Q319R	-	Q443R	+3
45	Q97R	Q319R	-	-	+2
46	Q97R	-	Q358E	Q443E	-1
47	Q97R	-	Q358E	Q443R	+1
48	Q97R	-	Q358E	-	0
49	Q97R	-	Q358R	Q443E	+1
50	Q97R	-	Q358R	Q443R	+3
51	Q97R	-	Q358R	-	+2
52	Q97R	-	-	Q443E	0
53	Q97R	-	-	Q443R	+2
54	Q97R	-	-	-	+1
55	-	Q319E	Q358E	Q443E	-3
56	-	Q319E	Q358E	Q443R	-1
57	-	Q319E	Q358E	-	-2
58	-	Q319E	Q358R	Q443E	-1
59	-	Q319E	Q358R	Q443R	+1
60	-	Q319E	Q358R	-	0
61	-	Q319E	-	Q443E	-2

<b>Variant #</b>	<b>Q97</b>	<b>Q319</b>	<b>Q358</b>	<b>Q443</b>	<b>Δ Charge</b>
62	-	Q319E	-	Q443R	0
63	-	Q319E	-	-	-1
64	-	Q319R	Q358E	Q443E	-1
65	-	Q319R	Q358E	Q443R	+1
66	-	Q319R	Q358E	-	0
67	-	Q319R	Q358R	Q443E	+1
68	-	Q319R	Q358R	Q443R	+3
69	-	Q319R	Q358R	-	+2
70	-	Q319R	-	Q443E	0
71	-	Q319R	-	Q443R	+2
72	-	Q319R	-	-	+1
73	-	-	Q358E	Q443E	-2
74	-	-	Q358E	Q443R	0
75	-	-	Q358E	-	-1
76	-	-	Q358R	Q443E	0
77	-	-	Q358R	Q443R	+2
78	-	-	Q358R	-	+1
79	-	-	-	Q443E	-1
80	-	-	-	Q443R	+1
81 (parent)	Q97	Q319	Q358	Q443	0

### **Example 10 - Enzyme Wash Performance**

This Example describes the testing of S242Q variant in a microswatch assay 1.0 μg/mL in AATCC HDL detergent or 5 mM HEPES buffer under varying ionic strength.

- 5 The methods provided in Example 6 were used (*See*, “Rice Starch Microswatch Assay for testing Amylase Performance” and “Corn Four Hydrolysis”).

10 There is an optimal net charge change for cleaning performance for enzyme in AATCC HDL detergent. Performance is measured in terms of relative cleaning performance observed in a rice starch microswatch activity assay. A value of around 1.0 indicates top cleaning performance in this assay. This is an example of optimizing a protein physical property (*e.g.*, net charge) for improving a given outcome or benefit (*e.g.*, cleaning performance in a liquid laundry detergent). The charge optimum identified with this limited set of probe proteins coincides with the optimum charge

observed when measuring the entire charge combinatorial library. The use of probe proteins is therefore predictive of the behavior of the entire library.

According to the Debye-Hückel theory (Israelachvili, INTERMOLECULAR AND SURFACE FORCES, SECOND EDITION: WITH APPLICATIONS TO COLLOIDAL AND BIOLOGICAL SYSTEMS, Academic Press 2<sup>nd</sup> Ed. [1992]), electrostatic interactions are governed primarily by the strength of double-layer forces between interacting species at constant potential or constant charge (enzymes, substrates, fabric, and detergent), their size, and the dielectric constant of the surrounding medium. In order to characterize the electrostatic behavior of particles in a complex medium, such as a detergent formulation, their interaction in a reduced environment possessing the same Debye screening length is sufficient. This was accomplished by choosing a buffer of matching pH and conductivity to that of the detergent under wash conditions. An appropriate buffer for such testing is 5 mM HEPES buffer at pH 8.0 with varying amounts of indifferent electrolyte, such as NaCl. Addition of 2.5 mM NaCl to this buffer matches the pH and conductivity of typical North American wash conditions. Addition of a higher concentration of NaCl is representative of Japanese and European wash conditions, which typically are higher in ionic strength due to both increased water hardness and detergent concentrations.

Figure 10 shows that positive charge S242Q charge variants are superior for cleaning of rice starch microswatch under North American laundry conditions. Likewise, negative charge TS23t variants are superior for cleaning of rice starch microswatches in Western European laundry conditions (Figure 11).

Figure 12 demonstrates that positive S242Q variants exhibit higher specific activity for granular corn starch substrates hydrolysis.

#### 25 **Example 11 - Thermostability**

This Example describes determining the relationship between protein charge and thermal stability. Alpha-amylase assays were based on BODIPY starch hydrolysis before and after heating the culture supernatant. The same chemical and reagent solutions used are as described in Example 6.

**Thermal stability assay for alpha-amylases.** The filtered culture supernatants were serially diluted in 50 mM sodium acetate + 2 mM CaCl<sub>2</sub>, at pH 5.8 with 0.002% Tween. 10 μL of each diluted culture supernatant was assayed to determine the initial amylase activity by the BODIPY starch assay. 50 μL of each diluted culture supernatant was placed in a VWR low profile PCR 96 well plate. 30 μL of mineral oil was added to each well as a sealant. The plate was incubated in a BioRad DNA engine Peltier Thermal Cycler at 95°C for 30 or 60 minutes depending on the stability of the parent enzyme. Following incubation, the plate was cooled to 4°C for 5 min and then kept at room temperature. 10 μL of each sample was added to a fresh plate and assayed to determine the final amylase activity by the BODIPY starch assay as described in Example 1.

**Calculation of Thermostability.** The residual activity of a sample was expressed as the ratio of the final absorbance and the initial absorbance, both corrected for blanks. A higher index indicates a more thermally-stable variant. This is an example of optimizing a protein physical property, in this case net charge, for improving enzyme thermal stability for a liquid laundry application.

**Thermostability Assay.** Thermostability of the variants was assessed as described above. Thermostability winners are listed in Table 11-1. Winners were defined as those having a ratio of mutant residual activity to parent (i.e., S242Q) residual activity greater than 1.

**Table 11-1: S242Q CCL - thermal stability winners**

Variant #	97	319	358	443	Mut residual act./WT residual act.
2	Q97E	Q319E	Q358E	Q443R	1.12
10	Q97E	Q319R	Q358E	Q443E	1.12
13	Q97E	Q319R	Q358R	Q443E	1.36
14	Q97E	Q319R	Q358R	Q443R	1.16
15	Q97E	Q319R	Q358R		1.37
17	Q97E	Q319R		Q443R	1.29
18	Q97E	Q319R			1.11
27	Q97E				1.16
32	Q97R	Q319E	Q358R	Q443R	1.18
37	Q97R	Q319R	Q358E	Q443E	1.29
38	Q97R	Q319R	Q358E	Q443R	1.22
39	Q97R	Q319R	Q358E		1.21

Variant #	97	319	358	443	Mut residual act./WT residual act.
40	Q97R	Q319R	Q358R	Q443E	1.20
41	Q97R	Q319R	Q358R	Q443R	1.26
42	Q97R	Q319R	Q358R		1.48
43	Q97R	Q319R		Q443E	1.21
44	Q97R	Q319R		Q443R	1.21
45	Q97R	Q319R			1.14
50	Q97R		Q358R	Q443R	1.14
62		Q319E		Q443R	1.26
63		Q319E			1.18
64		Q319R	Q358E	Q443E	1.19
65		Q319R	Q358E	Q443R	1.28
68		Q319R	Q358R	Q443R	1.14
70		Q319R		Q443E	1.22
73			Q358E	Q443E	1.15
74			Q358E	Q443R	1.15
75			Q358E		1.18

### Example 12 - Enzyme Performance

This Example demonstrates that enzyme performance may be affected by charge.

- 5 Enzyme performance was assessed using heat inactivated detergents as described above in Example 6. Winners were defined as those having Performance Index (PI) a greater than 1. PI is the ratio of mutant residual activity to parent (i.e., S242Q) residual activity. Results are shown in Tables 12-1 and 12-2.

10 **Table 12-1:** S242Q CCL - CS-28 rice starch microswatch winners, Tide 2x (North American conditions as described in Example 6).

Variant #	97	319	358	443	rel charge	PI
13	Q97E	Q319R	Q358R	Q443E	0	1.44
14	Q97E	Q319R	Q358R	Q443R	2	1.32
15	Q97E	Q319R	Q358R		1	1.40
16	Q97E	Q319R		Q443E	-1	1.33
17	Q97E	Q319R		Q443R	1	1.40
18	Q97E	Q319R			0	1.41
20	Q97E		Q358E	Q443R	-1	1.15
23	Q97E		Q358R	Q443R	1	1.21
25	Q97E			Q443E	-2	1.18



Variant #	97	319	358	443	rel charge	PI
26	Q97E			Q443R	0	1.25
27	Q97E				-1	1.16
28	Q97R	Q319E	Q358E	Q443E	-2	2.32
29	Q97R	Q319E	Q358E	Q443R	0	2.54
30	Q97R	Q319E	Q358E		-1	2.93
31	Q97R	Q319E	Q358R	Q443E	0	2.27
32	Q97R	Q319E	Q358R	Q443R	2	2.28
33	Q97R	Q319E	Q358R		1	2.34
34	Q97R	Q319E		Q443E	-1	2.31
35	Q97R	Q319E		Q443R	1	2.31
36	Q97R	Q319E			0	2.14
37	Q97R	Q319R	Q358E	Q443E	0	1.93
38	Q97R	Q319R	Q358E	Q443R	2	1.85
39	Q97R	Q319R	Q358E		1	2.14
40	Q97R	Q319R	Q358R	Q443E	2	1.92
41	Q97R	Q319R	Q358R	Q443R	4	1.37
42	Q97R	Q319R	Q358R		3	1.61
43	Q97R	Q319R		Q443E	1	1.90
44	Q97R	Q319R		Q443R	3	1.64
45	Q97R	Q319R			2	1.99
46	Q97R		Q358E	Q443E	-1	1.40
47	Q97R		Q358E	Q443R	1	1.29
48	Q97R		Q358E		0	1.60
49	Q97R		Q358R	Q443E	1	1.57
50	Q97R		Q358R	Q443R	3	1.38
51	Q97R		Q358R		2	1.37
52	Q97R			Q443E	0	1.51
54	Q97R				1	1.51
55		Q319E	Q358E	Q443E	-3	1.14
56		Q319E	Q358E	Q443R	-1	1.38
57		Q319E	Q358E		-2	1.10
58		Q319E	Q358R	Q443E	-1	1.25
59		Q319E	Q358R	Q443R	1	1.41
60		Q319E	Q358R		0	1.49
61		Q319E		Q443E	-2	1.16
62		Q319E		Q443R	0	1.45
63		Q319E			-1	1.28
64		Q319R	Q358E	Q443E	-1	1.12
65		Q319R	Q358E	Q443R	1	1.19
66		Q319R	Q358E		0	1.36
67		Q319R	Q358R	Q443E	1	1.24

Variant #	97	319	358	443	rel charge	PI
69		Q319R	Q358R		2	1.19
70		Q319R		Q443E	0	1.29
76			Q358R	Q443E	0	1.22
78			Q358R		1	1.25
79				Q443E	-1	1.24
80				Q443R	1	1.17

**Table 12-2:** S242Q CCL - CS-28 rice starch microswatch winners, Persil (Western European conditions)

Variant #	97	319	358	443	rel charge	PI
2	Q97E	Q319E	Q358E	Q443R	-2	1.41
3	Q97E	Q319E	Q358E		-3	1.94
4	Q97E	Q319E	Q358R	Q443E	-2	1.61
5	Q97E	Q319E	Q358R	Q443R	0	1.39
6	Q97E	Q319E	Q358R		-1	2.04
7	Q97E	Q319E		Q443E	-3	2.05
8	Q97E	Q319E		Q443R	-1	1.84
9	Q97E	Q319E			-2	2.27
10	Q97E	Q319R	Q358E	Q443E	-2	1.35
13	Q97E	Q319R	Q358R	Q443E	0	1.45
14	Q97E	Q319R	Q358R	Q443R	2	1.17
15	Q97E	Q319R	Q358R		1	1.22
16	Q97E	Q319R		Q443E	-1	1.26
17	Q97E	Q319R		Q443R	1	1.29
18	Q97E	Q319R			0	1.76
26	Q97E			Q443R	0	1.36
27	Q97E				-1	1.31
28	Q97R	Q319E	Q358E	Q443E	-2	2.21
29	Q97R	Q319E	Q358E	Q443R	0	1.96
30	Q97R	Q319E	Q358E		-1	1.94
31	Q97R	Q319E	Q358R	Q443E	0	2.11
32	Q97R	Q319E	Q358R	Q443R	2	1.87
33	Q97R	Q319E	Q358R		1	2.41
34	Q97R	Q319E		Q443E	-1	2.20
35	Q97R	Q319E		Q443R	1	2.21
36	Q97R	Q319E			0	2.07
37	Q97R	Q319R	Q358E	Q443E	0	1.86
38	Q97R	Q319R	Q358E	Q443R	2	1.83
39	Q97R	Q319R	Q358E		1	1.99
40	Q97R	Q319R	Q358R	Q443E	2	1.85

Variant #	97	319	358	443	rel charge	PI
41	Q97R	Q319R	Q358R	Q443R	4	1.36
42	Q97R	Q319R	Q358R		3	1.90
43	Q97R	Q319R		Q443E	1	1.99
44	Q97R	Q319R		Q443R	3	1.94
45	Q97R	Q319R			2	1.75
46	Q97R		Q358E	Q443E	-1	1.71
47	Q97R		Q358E	Q443R	1	1.39
48	Q97R		Q358E		0	1.85
50	Q97R		Q358R	Q443R	3	1.24
51	Q97R		Q358R		2	1.36
52	Q97R			Q443E	0	1.25
54	Q97R				1	1.88
55		Q319E	Q358E	Q443E	-3	1.12
56		Q319E	Q358E	Q443R	-1	1.17
58		Q319E	Q358R	Q443E	-1	1.16
59		Q319E	Q358R	Q443R	1	1.25
60		Q319E	Q358R		0	1.50
63		Q319E			-1	1.36
64		Q319R	Q358E	Q443E	-1	1.10
65		Q319R	Q358E	Q443R	1	1.18
66		Q319R	Q358E		0	1.25
67		Q319R	Q358R	Q443E	1	1.29
70		Q319R		Q443E	0	1.15

Activity was also measured using the BODIPY starch hydrolysis assay as provided herein. The results are shown in Table 12-3. The relative specific activity on this starch substrate (a corn starch) greater than 1 indicates the variant has higher specific activity than the S242Q parent. Relative ppm is expression titers, greater than 1 indicates higher titers (in shake tubes) than the S242Q parent.

**Table 12-3:** S242Q CCL - titer and/or BODIPY-starch winners

Variant #	97	319	358	443	Charge	Rel ppm	Rel Sp act
1	Q97E	Q319E	Q358E	Q443E	-4	1.27	1.29
2	Q97E	Q319E	Q358E	Q443R	-2	1.19	1.31
3	Q97E	Q319E	Q358E		-3	1.00	1.43
4	Q97E	Q319E	Q358R	Q443E	-2	1.23	1.43
5	Q97E	Q319E	Q358R	Q443R	0	0.94	1.78
6	Q97E	Q319E	Q358R		-1	0.89	1.81
7	Q97E	Q319E		Q443E	-3	1.40	1.41

Variant #	97	319	358	443	Charge	Rel ppm	Rel Sp act
8	Q97E	Q319E		Q443R	-1	1.12	1.58
9	Q97E	Q319E			-2	1.09	1.56
10	Q97E	Q319R	Q358E	Q443E	-2	1.45	1.32
11	Q97E	Q319R	Q358E	Q443R	0	1.32	1.49
12	Q97E	Q319R	Q358E		-1	1.58	1.27
13	Q97E	Q319R	Q358R	Q443E	0	0.65	1.44
14	Q97E	Q319R	Q358R	Q443R	2	0.66	1.65
15	Q97E	Q319R	Q358R		1	0.80	1.64
16	Q97E	Q319R		Q443E	-1	1.09	1.51
17	Q97E	Q319R		Q443R	1	1.00	1.42
18	Q97E	Q319R			0	0.87	1.78
19	Q97E		Q358E	Q443E	-3	1.22	0.88
21	Q97E		Q358E		-2	1.12	0.88
22	Q97E		Q358R	Q443E	-1	0.91	1.16
23	Q97E		Q358R	Q443R	1	0.78	1.25
24	Q97E		Q358R		0	1.08	1.14
25	Q97E			Q443E	-2	1.12	1.00
28	Q97R	Q319E	Q358E	Q443E	-2	0.78	1.87
29	Q97R	Q319E	Q358E	Q443R	0	0.80	1.81
30	Q97R	Q319E	Q358E		-1	0.68	2.21
31	Q97R	Q319E	Q358R	Q443E	0	0.68	1.96
32	Q97R	Q319E	Q358R	Q443R	2	0.70	2.05
33	Q97R	Q319E	Q358R		1	0.60	2.27
34	Q97R	Q319E		Q443E	-1	0.65	2.25
35	Q97R	Q319E		Q443R	1	0.70	2.15
36	Q97R	Q319E			0	0.73	2.23
37	Q97R	Q319R	Q358E	Q443E	0	0.93	2.11
38	Q97R	Q319R	Q358E	Q443R	2	0.65	2.21
39	Q97R	Q319R	Q358E		1	0.82	2.22
40	Q97R	Q319R	Q358R	Q443E	2	0.74	2.28
41	Q97R	Q319R	Q358R	Q443R	4	0.55	2.09
42	Q97R	Q319R	Q358R		3	0.67	2.48
43	Q97R	Q319R		Q443E	1	0.84	2.35
44	Q97R	Q319R		Q443R	3	0.73	2.41
45	Q97R	Q319R			2	0.76	2.45
46	Q97R		Q358E	Q443E	-1	0.79	1.45
47	Q97R		Q358E	Q443R	1	0.75	1.42
48	Q97R		Q358E		0	0.82	1.46
49	Q97R		Q358R	Q443E	1	0.67	1.69
50	Q97R		Q358R	Q443R	3	0.60	1.60
51	Q97R		Q358R		2	0.64	1.29
52	Q97R			Q443E	0	0.83	1.43

Variant #	97	319	358	443	Charge	Rel ppm	Rel Sp act
54	Q97R				1	0.72	1.49
55		Q319E	Q358E	Q443E	-3	0.99	1.15
56		Q319E	Q358E	Q443R	-1	0.77	1.40
57		Q319E	Q358E		-2	0.83	1.34
58		Q319E	Q358R	Q443E	-1	0.73	1.49
59		Q319E	Q358R	Q443R	1	0.67	1.61
60		Q319E	Q358R		0	0.80	1.67
61		Q319E		Q443E	-2	0.91	1.39
62		Q319E		Q443R	0	0.73	1.45
63		Q319E			-1	0.75	1.41
64		Q319R	Q358E	Q443E	-1	1.05	1.28
65		Q319R	Q358E	Q443R	1	0.94	1.42
66		Q319R	Q358E		0	0.96	1.39
67		Q319R	Q358R	Q443E	1	1.02	1.50
68		Q319R	Q358R	Q443R	3	0.71	1.57
69		Q319R	Q358R		2	0.71	1.58
70		Q319R		Q443E	0	0.91	1.49
72		Q319R			1	0.95	1.56
77			Q358R	Q443R	2	0.67	1.22
78			Q358R		1	0.66	1.15

### Example 13 - Balancing Mutational Effects on Amylase Activity and Expression

This example illustrates that two separate enzyme properties can be simultaneously optimized by the introduction of multiple amino acid substitutions, even where the properties are negatively correlated due, for example, to oppositely linked to charge characteristics of the protein.

It was determined during experimentation that the median expression of AmyS-242Q decreased with increasing positive charge. However, specific BODIPY starch hydrolysis increased with increasing positive charge. Enhanced recombinant amylase expression and starch hydrolysis are desirable in an engineered variant of AmyS-242Q suitable for starch liquefaction in the fuel ethanol industry or cleaning in detergent applications for instance. These properties, however, are apparently conflicting properties. Using the methods provided herein, it is possible to produce a more highly expressed amylase variant without severely compromising starch hydrolysis by

selectively combining single mutations. The strategy described herein was successfully used to produce and select multiply-substituted AmyS-242Q variants having improvements in a first property (*e.g.*, expression as the primary property), while improving or not sacrificing a second property (*e.g.*, starch hydrolysis as the secondary property).

In addition, in converse to median expression of AmyS-242Q variants, corn starch microswatch cleaning increased with increasing positive charge. Enhanced recombinant amylase expression and cleaning performance are desirable in an engineered variant of AmyS-242Q. These properties, however, are also apparently conflicting properties.

Using the methods disclosed herein, it is possible to produce a more highly expressed amylase variant without severely compromising cleaning performance by selectively combining single mutations. The strategy described herein was successfully used to produce and select multiply-substituted AmyS-242Q variants having improvements in a first property (*e.g.*, expression as the primary property), while improving or not sacrificing a second property (*e.g.*, rice starch microswatch cleaning as the secondary property).

In particular, an eighty member AmyS-S242Q charge combinatorial library (CCL) comprising variants having combinations of from one to four substitutions of charged residues was tested for shake tube expression, BODIPY-starch hydrolysis, and rice starch cleaning activity. AmyS-S242Q winners are shown in Tables 13-1 and 13-2. Importantly, the multiply-substituted variants of Table 13-1 have equal or improved expression and equal or improved BODIPY-starch hydrolysis as compared to the parent enzyme. Similarly, the multiply-substituted variants of Table 13-2 have equal or improved expression and equal or improved rice starch cleaning activity as compared to the parent enzyme.

<b>Variant</b>	<b>97</b>	<b>319</b>	<b>358</b>	<b>443</b>	<b>Charge</b>	<b>Expression (PI)</b>	<b>BODIPY (PI)</b>
1	Q97E	Q319E	Q358E	Q443E	-4	1.27	1.29

**Table 13-1. AmyS-S242Q Expression and BODIPY-Starch Hydrolysis Winners**

<b>Variant</b>	<b>97</b>	<b>319</b>	<b>358</b>	<b>443</b>	<b>Charge</b>	<b>Expression (PI)</b>	<b>BODIPY (PI)</b>
2	Q97E	Q319E	Q358E	Q443R	-2	1.19	1.31
3	Q97E	Q319E	Q358E		-3	1.00	1.43
4	Q97E	Q319E	Q358R	Q443E	-2	1.23	1.43
7	Q97E	Q319E		Q443E	-3	1.40	1.41
8	Q97E	Q319E		Q443R	-1	1.12	1.58
9	Q97E	Q319E			-2	1.09	1.56
10	Q97E	Q319R	Q358E	Q443E	-2	1.45	1.32
11	Q97E	Q319R	Q358E	Q443R	0	1.32	1.49
12	Q97E	Q319R	Q358E		-1	1.58	1.27
16	Q97E	Q319R		Q443E	-1	1.09	1.51
17	Q97E	Q319R		Q443R	+1	1.00	1.42
24	Q97E		Q358R		0	1.08	1.14
25	Q97E			Q443E	-2	1.12	1.00
64		Q319R	Q358E	Q443E	-1	1.05	1.28
67		Q319R	Q358R	Q443E	+1	1.02	1.50

**Table 13-2. AmyS-S242Q Expression and Rice-Starch Hydrolysis Winners**

<b>Variant</b>	<b>97</b>	<b>319</b>	<b>358</b>	<b>443</b>	<b>Charge</b>	<b>Expression</b>	<b>CS-28</b>
1	Q97E	Q319E	Q358E	Q443E	-4	1.27	1.01
11	Q97E	Q319R	Q358E	Q443R	0	1.32	1.18
12	Q97E	Q319R	Q358E		-1	1.58	1.13
16	Q97E	Q319R		Q443E	-1	1.09	1.43
17	Q97E	Q319R		Q443R	+1	1.00	1.55
24	Q97E		Q358R		0	1.08	1.15
25	Q97E			Q443E	-2	1.12	1.09
64		Q319R	Q358E	Q443E	-1	1.05	1.18
67		Q319R	Q358R	Q443E	+1	1.02	1.15

In sum, because enzyme activity and enzyme production have different charge dependencies (*see* FIG. 13A, 13B, 14A, and 14B) they are negatively correlated (*see* FIG. 12A and 12B). However, there are a number of variants that are improved in both expression and activity, and analyzing the library in this manner allows them to be  
5 identified.

Although demonstrated with amylases, this method is applicable to other enzyme classes such as proteases, lipases, cellulases, transferases and pectinases. Moreover any combination of two or more properties can be analyzed simultaneously such as expression, activity, binding, thermal stability, stability in the presence of one or  
10 detergents, and chelant stability.

#### **Example 14 - Desizing performance of amylases**

In this example, the desizing performance of variant S242Q was compared against  
15 Ethyl and Xtra at 85°C and 97°C at several concentrations of calcium.

CaCl<sub>2</sub> concentration was varied from 0-20 ppm per test by adding various amounts of stock CaCl<sub>2</sub> solution to Milli Q water, pH ~6.5. Ethyl, Xtra and variant S242Q were used at 0.01 ppm active protein per test. The assay was performed in a LAUNDER-O-METER using a liquor ratio of 50:1. Performance tests were conducted  
20 on rice starch-stained fabric swatches with an indicator dye bound to the starch (TestFabrics Cat. No. CS-28; TestFabrics Inc.). Three CS-28 swatches (6 cm x 8 cm) and 4 greige print cloth swatches (Testfabrics, Style 400R; 3 inches x 4 inches) were used as substrates per experiment. The temperature of the LAUNDER-O-METER with Milli Q water/Ca was pre-adjusted to 85°C or 97°C, after which the enzymes and swatches  
25 were added. The reaction was carried out for 30 min, after which the swatches were rinsed in water and dried before reading.

Measurements are made by reflectometry using the CIE L\*a\*b\* color space. Every perceivable color can be represented by L\*a\*b\* coordinate in the color space. “L\*” represents the lightness or grey scale value on a scale of 0 to 100, pure black to pure  
30 white. “a\*” represents the magenta to green shift, wherein large positive values represent a very magenta hue and large negative values represent a very green hue. “b\*” represents



the yellow to blue shift where large positive values represent a very yellow hue and large negative values represent a very blue hue. When both  $a^*$  and  $b^*$  values are 0, there is an absence of color, leaving pure grey colors with their lightness defined by the  $L^*$  value.

A Minolta Chromameter CR 200 in the CIE Lab color space with a D 65 light source was utilized for measuring desizing performance. To quantify desizing performance, four CIE  $L^*$  readings (i.e., 2 readings each from the front and the back of the swatch) were taken from each CS-28 swatch following the amylase treatment. Higher CIE  $L^*$  values indicate better desizing performance.

As shown in Figures 15 and 16, the S242Q variant showed significantly lower calcium dependency for desizing performance compared to both Ethyl and Xtra under the conditions tested.

All publications and patents mentioned in the above specification are incorporated herein by reference. Although the disclosed methods and enzymes have in some instances been described in connection with specific or preferred embodiments, it should be understood what is covered by the appended claims is not limited to such specific or preferred embodiments. Indeed, various modifications and variations of the disclosed methods and enzymes will be apparent to those skilled in the art, and various modifications of the described modes for practicing what has been disclosed are included within the scope of the following claims.

Where the terms “comprise”, “comprises”, “comprised” or “comprising” are used in this specification, they are to be interpreted as specifying the presence of the stated features, integers, steps or components referred to, but not to preclude the presence or addition of one or more other feature, integer, step, component or group thereof.

Further, any prior art reference or statement provided in the specification is not to be taken as an admission that such art constitutes, or is to be understood as constituting, part of the common general knowledge in Australia.

The Claims defining the Invention are as follows:

1. A composition comprising:
  - a) at least one variant alpha-amylase comprising an amino acid sequence at least 95% identical to that of a parent AmyS-like alpha-amylase, and having a substitution at an amino acid position corresponding to position 242 of a reference alpha-amylase, wherein the substitution is a S242Q substitution, and said variant alpha-amylase having detectable alpha-amylase activity, wherein the reference alpha-amylase is SEQ ID NO:1 or 2; and
  - b) at least one of an additional enzyme, a detergent, a surfactant, a chelator, an oxidizing agent, an acidulant, an alkalizing agent, a source of peroxide, a source of hardness, a salt, a detergent complexing agent, a polymer, a stabilizing agent, or a fabric conditioner.
2. The composition of claim 1, wherein the variant alpha-amylase is altered, as compared to the parent AmyS-like alpha-amylase or the reference alpha-amylase, in one or more characteristics of: (a) net charge, (b) substrate specificity, (c) substrate cleavage, (d) substrate binding, (e) thermal stability, (f) activity at one or more pH's, (g) stability at one or more pH's, (h) stability in oxidizing conditions, (i) Ca<sup>2+</sup> requirements, (j) specific activity, (k) catalytic rate, (l) catalytic efficiency, (m) activity in the presence of a chelator, (n) thermal or pH stability in the presence of a chelator, (o) utility for desizing, or utility for a cleaning process, or (p) amount of expression in a protein expression system.
3. The composition of claim 1, which is a component of a product for use in laundry, dish, or hard-surface cleaning, desizing, or fabric or stain treatment.
4. The composition of any one of claims 1 to 3, wherein the additional enzyme is a protease, a lipase, an amylase, a cellulase, a peroxidase, an oxidase, a pectinase, a lyase, a cutinase, a laccase, or a combination thereof.
5. The composition of any one of claims 1 to 4, wherein the surfactant is nonionic, anionic, cationic, or zwitterionic.

6. The composition of claim 1, wherein the variant alpha-amylase has altered stability to oxidation and the variant alpha-amylase further includes deletion or substitution of one or more methionine residues including residues located at amino positions 8, 9, 96, 200, 206, 284, 307, 311, 316, and 438 of a parent AmyS-like alpha-amylase, where the reference alpha-amylase in SEQ ID NO:2.
7. The composition of any one of claims 1 to 6, wherein the variant alpha-amylase further comprises:
  - i) a sequence modification at one or more amino acid positions corresponding to amino acid positions 97, 179, 180, 193, 319, 349, 358, 416, 428, or 443 of the reference alpha-amylase, wherein the modification is optionally a substitution as follows: a cysteine at 349, a cysteine at 428, a glutamic acid at 97, an arginine at 97, a glutamic acid at 319, an arginine at 319, a glutamic acid at 358, an arginine at 358, a glutamic acid at 443, or an arginine at 443, whereby the variant alpha-amylase comprises a set of substitutions selected from the group consisting of a) Q97E, Q319E, Q358E, Q443E; b) Q97E, Q319R, Q358E, Q443R; c) Q97E, Q319R, Q358E; d) Q97E, Q319R, Q443E; e) Q97E, Q319R, Q443R; f) Q97E, Q358R; g) Q97E, Q443E; h) Q319R, Q358E, Q443E; and i) Q319R, Q358R, Q443E; and optionally
  - ii) substitution of an N193 or a V416 or both, optionally selected from N193F and V416G, or both; and optionally
  - iii) deletion of one or more amino acids at positions F178, R179, G180, I181, G182 and K183.
8. The composition of claim 7, wherein the variant alpha-amylase has altered metal ion dependence or altered stability or activity in an absence of added calcium or a presence of a chelator.
9. The composition of claim 1, wherein the variant alpha-amylase has at least 95% homology to SEQ ID NO:2 and the substitution of amino acid 242 relative to numbering

- in a reference alpha-amylase comprising SEQ ID NO:1, and wherein the variant alpha-amylase has alpha-amylase activity.
10. The composition of any one of claims 1 to 9, wherein the parent AmyS-like alpha-amylase is SEQ ID NO:1, 2, 6, 7, 8, 9, 10, 11, 12, 15, or 16.
  11. The composition of any one of claims 1 to 10, wherein the variant alpha-amylase has improved performance in a wash process at a pH  $\geq$  about 8, relative to the parent AmyS-like alpha-amylase.
  12. A detergent or cleaning formulation comprising at least one variant alpha-amylase comprising an amino acid sequence at least 95% identical to that of a parent AmyS-like alpha-amylase, and having a substitution at an amino acid position corresponding to position 242 of a reference alpha-amylase, wherein the substitution is a S242Q substitution, said variant alpha-amylase having detectable alpha-amylase activity; wherein the reference amylase is SEQ ID NO:1 or 2.
  13. A method of desizing a woven material subsequent to a weaving process comprising contacting the woven material with a variant alpha-amylase comprising an amino acid sequence at least 95% identical to that of a parent AmyS-like alpha-amylase, and having a substitution at an amino acid position corresponding to position 242 of a reference alpha-amylase, wherein the substitution is a S242Q substitution, said variant alpha-amylase having detectable alpha-amylase activity, under conditions and for a time effective for at least partially removing sizing from the woven material; wherein the reference amylase is SEQ ID NO: 1 or 2.
  14. The method of claim 13, wherein the variant alpha-amylase is altered, as compared to the parent AmyS-like alpha-amylase or the reference alpha-amylase, in one or more of: (a) net charge, (b) substrate specificity, (c) substrate cleavage, (d) substrate binding, (e) thermal stability, (f) activity at one or more pH's, (g) stability at one or more pH's, (h) stability in oxidizing conditions, (i) Ca<sup>2+</sup> requirements, (j) specific activity, (k) catalytic

rate, (l) catalytic efficiency, (m) activity in a presence of a chelator, (n) thermal or pH stability in the presence of a chelator, (o) effectiveness for desizing, or (p) amount of expression in a protein expression system.

15. The method of claim 13 or 14, wherein the parent AmyS-like alpha-amylase is SEQ ID NO:1, 2, 6, 7, 8, 9, 10, 11, 12, 15, or 16.
16. The method of any one of claims 13 to 15, wherein the variant alpha-amylase further comprises one or more of substitution at positions as follows: a cysteine at 349, a cysteine at 428, a glutamic acid at 97, an arginine at 97, a glutamic acid at 319, an arginine at 319, a glutamic acid at 358, an arginine at 358, a glutamic acid at 443, or an arginine at 443, wherein the reference alpha-amylase is SEQ ID NO: 1 or 2.
17. A method of washing or cleaning comprising contacting one or more items to be washed or cleaned with a detergent of claim 12 or a composition comprising a variant alpha-amylase comprising an amino acid sequence at least 95% identical to that of a parent AmyS-like alpha-amylase, and having a substitution at an amino acid position corresponding to position 242 of a reference alpha-amylase, wherein the substitution is a S242Q substitution, said variant alpha-amylase having detectable alpha-amylase activity, under conditions and for a time effective for at least partially washing or cleaning the one or more items; wherein the reference amylase is SEQ ID NO: 1 or 2.
18. The method of claim 17, wherein at least one item is soiled with at least one starch-containing material, removal of said starch soil is aided by the variant alpha-amylase.
19. The method of claim 17 or 18, wherein the composition further comprises one or more of an additional enzyme, a detergent, a surfactant, a chelator, an oxidizing agent, an acidulant, an alkalizing agent, a source of peroxide, a source of hardness, a salt, a detergent complexing agent, a polymer, a stabilizing agent, or a fabric conditioner.

20. The method of any one of claims 17 to 19, wherein the parent AmyS-like alpha-amylase is SEQ ID NO: 1, 2, 6, 7, 8, 9, 10, 11, 12, 15, or 16.
21. The method of any one of claims 17 to 20, wherein the variant alpha-amylase has improved performance in a wash process at a pH  $\geq$  about 8, relative to the parent AmyS-like alpha-amylase.
22. The method of any one of claims 17 to 21, wherein the variant alpha-amylase comprises:
  - i) one or more substitutions at positions as follows: a cysteine at 349, a cysteine at 428, a glutamic acid at 97, an arginine at 97, a glutamic acid at 319, an arginine at 319, a glutamic acid at 358, an arginine at 358, a glutamic acid at 443, or an arginine at 443, whereby the variant alpha-amylase optionally comprises a set of substitutions of a) Q97E, Q319E, Q358E, Q443E; b) Q97E, Q319R, Q358E, Q443R; c) Q97E, Q319R, Q358E; d) Q97E, Q319R, Q443E; e) Q97E, Q319R, Q443R; f) Q97E, Q358R; g) Q97E, Q443E; h) Q319R, Q358E, Q443E; or i) Q319R, Q358R, Q443E; and optionally
  - ii) a deletion of one or more amino acids at positions F178, R179, G180, I181, G182, or K183.
23. The method of claim 22, wherein the variant alpha-amylase has altered metal ion dependence or altered stability, or activity in an absence of added calcium or the presence of a chelator.
24. A composition according to any one of claims 1 to 11, substantially as hereinbefore described.

		1		50
SEQID No 1	(1)	-AAPFNGTMMQYFEWYLPDDGTLWTKVANEANNLSSLGITALWLPPAYKG		
SEQID No 2	(1)	-AAPFNGTMMQYFEWYLPDDGTLWTKVANEANNLSSLGITALWLPPAYKG		
SEQID No 3	(1)	-AAPFNGTMMQYFEWYLPDDGTLWTKVANEANNLSSLGITALWLPPAYKG		
SEQID No 4	(1)	-AAPFNGTMMQYFEWYLPDDGTLWTKVANEANNLSSLGITALWLPPAYKG		
SEQID No 5	(1)	-AAPFNGTMMQYFEWYLPDDGTLWTKVANEANNLSSLGITALWLPPAYKG		
SEQID No 6	(1)	HHNGTNGTMMQYFEWYLPNDGNHWNRLNSDASNLSKSGITAVWI PPAYKG		
SEQID No 7	(1)	--ANLNGTLMQYFEWYMPNDGQHWKRLQNDSAYLAEHGITAVWI PPAYKG		
SEQID No 8	(1)	---ANLNGTLMQYFEWYMPNDGQHWKRLQNDSAYLAEHGITAVWI PPAYKG		
SEQID No 9	(1)	----VNGTLMQYFEWYTPNDGQHWKRLQNDAEHLSDIGITAVWI PPAYKG		
SEQID No 10	(1)	HHNGTNGTMMQYFEWYLPNDGNHWNRLRSDASNLSKSGISAVWI PPAYKG		
SEQID No 11	(1)	HHNGTNGTMMQYFEWHL PNDGNHWNRLRDDASNLRNRGITAIWI PPAYKG		
SEQID No 12	(1)	HHNGTNGTMMQYFEWHL PNDGNHWNRLRDDAANLSKSGITAVWI PPAYKG		
SEQID No 13	(1)	--DGLNGTMMQYFEWHL PNDGNHWNRLHDDAAALS DAGITAIWI PPAYKG		
SEQID No 14	(1)	--DGLNGTMMQYFEWHL PNDGNHWNRLHDDAEALS NAGITAIWI PPAYKG		
SEQID No 15	(1)	-AAPFNGTMMQYFEWYLPDDGTLWTKVANEANNLSSLGITALWLPPAYKG		
Consensus 1	(1)	A NGTMMQYFEWYLPNDGQHW RL NDA NLSS GITALWI PPAYKG		
		51		100
SEQID No 1	(50)	TSRSDVGYGVYDLYDLGFEFNQKGTVRTKYGTKAQYLQAIQAAHAAGMQVY		
SEQID No 2	(50)	TSRSDVGYGVYDLYDLGFEFNQKGTVRTKYGTKAQYLQAIQAAHAAGMQVY		
SEQID No 3	(50)	TSRSDVGYGVYDLYDLGFEFNQKGTVRTKYGTKAQYLQAIQAAHAAGMQVY		
SEQID No 4	(50)	TSRSDVGYGVYDLYDLGFEFNQKGTVRTKYGTKAQYLQAIQAAHAAGMQVY		
SEQID No 5	(50)	TSRSDVGYGVYDLYDLGFEFNQKGTVRTKYGTKAQYLQAIQAAHAAGMQVY		
SEQID No 6	(51)	ASQNDVGYGAYDLYDLGFEFNQKGTVRTKYGTRS QLA AVTSLKNNGIQVY		
SEQID No 7	(49)	TSQADVGYGAYDLYDLGFEFHQKGTVRTKYGT K GELQSAIKSLHSRDINVY		
SEQID No 8	(49)	TSQADVGYGAYDLYDLGFEFHQKGTVRTKYGT K GELQSAIKSLHSRDINVY		
SEQID No 9	(47)	LSQSDNGYGPYDLYDLGFEFQQKGTVRTKYGT K SELQDAIGSLHSRNVQVY		
SEQID No 10	(51)	ASQNDVGYGAYDLYDLGFEFNQKGT IRTKYGTRNQLAAVNALKSNGIQVY		
SEQID No 11	(51)	TSQNDVGYGAYDLYDLGFEFNQKGTVRTKYGTR SQLES AIHALKNNGVQVY		
SEQID No 12	(51)	TSQNDVGYGAYDLYDLGFEFNQKGTVRTKYGTR SQLGAVTSLKNNGIQVY		
SEQID No 13	(49)	NSQADVGYGAYDLYDLGFEFNQKGTVRTKYGT KAQLERAIGSLKSN DINVY		
SEQID No 14	(49)	NSQADVGYGAYDLYDLGFEFNQKGTVRTKYGT KAQLERAIGSLKSN DINVY		
SEQID No 15	(50)	TSRSDVGYGVYDLYDLGFEFNQKGTVRTKYGT KAQYLQAIQAAHAAGMQVY		
Consensus 1	(51)	TSQSDVGYGAYDLYDLGFEFNQKGTVRTKYGT KAQL AI ALHA GIQVY		
		101		150
SEQID No 1	(100)	ADVVFDPHKKGGADGTEWVDAVEVNPSDRNQEISGTYQIQAWTKFDFPGRGN		
SEQID No 2	(100)	ADVVFDPHKKGGADGTEWVDAVEVNPSDRNQEISGTYQIQAWTKFDFPGRGN		
SEQID No 3	(100)	ADVVFDPHKKGGADGTEWVDAVEVNPSDRNQEISGTYQIQAWTKFDFPGRGN		
SEQID No 4	(100)	ADVVFDPHKKGGADGTEWVDAVEVNPSDRNQEISGTYQIQAWTKFDFPGRGN		
SEQID No 5	(100)	ADVVFDPHKKGGADGTEWVDAVEVNPSDRNQEISGTYQIQAWTKFDFPGRGN		
SEQID No 6	(101)	GDVVMNHKGGADATEMVR AVEVNPNNRNQEVTGEY TIEAWTRFDFPGRGN		
SEQID No 7	(99)	GDVVINHKGGADATEDVTAVEVDPADRNRVISGEHLIKAWTHFHFPGRGS		
SEQID No 8	(99)	GDVVINHKGGADATEDVTAVEVDPADRNRVISGEHLIKAWTHFHFPGRGS		
SEQID No 9	(97)	GDVVLNKHKAGADATEDVTAVEVNPANRNQETSEEYQIKAWTDFRFPGRGN		
SEQID No 10	(101)	GDVVMNHKGGADATEMVR AVEVNPNNRNQEVSGEY TIEAWTKFDFPGRGN		
SEQID No 11	(101)	GDVVMNHKGGADATENVL AVEVNPNNRNQEISGDY TIEAWTKFDFPGRGN		
SEQID No 12	(101)	GDVVMNHKGGADGTEMVNAVEVNRNQRNQEISGEY TIEAWTKFDFPGRGN		
SEQID No 13	(99)	GDVVMNHKMGADFT EAVQAVQVNPTNRWQDISGAY TIDAWTGDFDFPGRGN		
SEQID No 14	(99)	GDVVMNHKLGADFT EAVQAVQVNPSNRWQDISGVY TIDAWTGDFDFPGRGN		
SEQID No 15	(100)	ADVVFDPHKKGGADGTEWVDAVEVNPSDRNQEISGTYQIQAWTKFDFPGRGN		
Consensus 1	(101)	GDVVMNHKGGADGTE V AVEVNPNNRNQEISG Y I AWTKFDFPGRGN		

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		151		200
SEQID No 1	(150)	TYSSFKRWYHFDGVDWDESRLS-RIYKFRGIGKAWDWEVDTENGNIDY		
SEQID No 2	(150)	TYSSFKRWYHFDGVDWDESRLS-RIYKFRGIGKAWDWEVDTENGNIDY		
SEQID No 3	(150)	TYSSFKRWYHFDGVDWDESRLS-RIYKFRGIGKAWDWEVDTENGNIDY		
SEQID No 4	(150)	TYSSFKRWYHFDGVDWDESRLS-RIYKFRGIGKAWDWEVDTENGNIDY		
SEQID No 5	(150)	TYSSFKRWYHFDGVDWDESRLS-RIYKFRGIGKAWDWEVDTENGNIDY		
SEQID No 6	(151)	THSSFKRWYHFDGVDWQSRRLNNRIYKFRGHGKAWDWEVDTENGNIDY		
SEQID No 7	(149)	TYSDFKWHWYHFDGTDWDESRLN-RIYKFQ--KAWDWEVSNENGNIDY		
SEQID No 8	(149)	TYSDFKWHWYHFDGTDWDESRLN-RIYKFQ--KAWDWEVSNENGNIDY		
SEQID No 9	(147)	TYSDFKWHWYHFDGADWDESRLS-RIYKFRGEGKAWDWEVSSENGNIDY		
SEQID No 10	(151)	THSNFKRWYHFDGVDWQSRRLNNRIYKFRGDGKAWDWEVDTENGNIDY		
SEQID No 11	(151)	TYSDFKRWYHFDGVDWQSRQFQNRRIYKFRGDGKAWDWEVDSENGNIDY		
SEQID No 12	(151)	THSNFKRWYHFDGTDWQSRQLQNKIYKFRGTGKAWDWEVDIENGNIDY		
SEQID No 13	(149)	AYSDFKRWRFHFNVDWQRYQEN-HIFRFAN--TNWNWRVDEENGNIDY		
SEQID No 14	(149)	AYSDFKRWRFHFNVDWQRYQEN-HLFRFAN--TNWNWRVDEENGNIDY		
SEQID No 15	(150)	TYSSFKRWYHFDGVDWDESRLS-RIYKFRG--KAWDWEVDFEFGNIDY		
Consensus 1	(151)	TYS FKRWYHFDGVDWDESRLN RIYKFRG GKAWDWEVDTENGNIDY		
		201		250
SEQID No 1	(199)	LMYADLMDHPEVVTELKNWGKQYVNTTNIIDGFRLDVAVKHIFSFPPDWL		
SEQID No 2	(199)	LMYADLMDHPEVVTELKNWGKQYVNTTNIIDGFRLDVAVKHIFSFPPDWL		
SEQID No 3	(199)	LMYADLMDHPEVVTELKNWGKQYVNTTNIIDGFRLDVAVKHIFAFFPPDWL		
SEQID No 4	(199)	LMYADLMDHPEVVTELKNWGKQYVNTTNIIDGFRLDVAVKHIFQFFPPDWL		
SEQID No 5	(199)	LMYADLMDHPEVVTELKNWGKQYVNTTNIIDGFRLDVAVKHIFEFFPPDWL		
SEQID No 6	(201)	LMYADIDMDHPEVVNELRNWGWYVNTLGLDGFRLDAVKHIFYSFTRDWI		
SEQID No 7	(196)	LMYADIDYDHPDVAEIKRWGTWYANELQLDGFRLDAVKHIFSFRLRDWV		
SEQID No 8	(196)	LMYADIDYDHPDVAEIKRWGTWYANELQLDGFRLDAVKHIFSFRLRDWV		
SEQID No 9	(196)	LMYADVVDYDHPDVVAETKKWGIWYANESLDGFRLDAVKHIFSFRLRDWV		
SEQID No 10	(201)	LMYADIDMDHPEVVNELRNWGWYVNTLGLDGFRLDAVKHIFYSFTRDWI		
SEQID No 11	(201)	LMYADVMDHPEVVNELRRWGEWYVNTLNLGLDGFRLDAVKHIFYSFTRDWL		
SEQID No 12	(201)	LMYADIDMDHPEVINELRNWGWYVNTLNLGLDGFRLDAVKHIFYSYTRDWL		
SEQID No 13	(196)	LLGSNIDFSHPEVQDELKDWGSWFTDELGLDGYRLDAIKHIFWYTSDWV		
SEQID No 14	(196)	LLGSNIDFSHPEVQDELKDWGSWFTDELGLDGYRLDAIKHIFWYTSDWV		
SEQID No 15	(197)	LMYADLMDHPEVVTELKNWGKQYVNTTNIIDGFRLDVAVKHIFSFPPDWL		
Consensus 1	(201)	LMYADIDMDHPEVV ELKNWG WY NTLNLGLDGFRLDAVKHIFSF DWL		
		251		300
SEQID No 1	(249)	SYVRSQTGKPLFTVGEYWSYDINKLHNYITKTNGTMSLFDAPLHNKFYTA		
SEQID No 2	(249)	SYVRSQTGKPLFTVGEYWSYDINKLHNYITKTNGTMSLFDAPLHNKFYTA		
SEQID No 3	(249)	SYVRSQTGKPLFTVGEYWSYDINKLHNYITKTNGTMSLFDAPLHNKFYTA		
SEQID No 4	(249)	SYVRSQTGKPLFTVGEYWSYDINKLHNYITKTNGTMSLFDAPLHNKFYTA		
SEQID No 5	(249)	SYVRSQTGKPLFTVGEYWSYDINKLHNYITKTNGTMSLFDAPLHNKFYTA		
SEQID No 6	(251)	NHVSATGKNMFVAEAFWKNLGALENYLNKTNWNHVSFVPLHYNLNA		
SEQID No 7	(246)	NHVREKTGKEMFTVAEYQNDLGALENYLNKTNFNHVSFVPLHYQFHAA		
SEQID No 8	(246)	NHVREKTGKEMFTVAEYQNDLGALENYLNKTNFNHVSFVPLHYQFHAA		
SEQID No 9	(246)	QAVRQATGKEMFTVAEYQNNAGKLENYLNKTSFNQSVFVPLHFNLQAA		
SEQID No 10	(251)	NHVSATGKNMFVAEAFWKNLGALENYLNKTNWNHVSFVPLHYNLNA		
SEQID No 11	(251)	THVRNATGKEMFVAEAFWKNLGALENYLNKTNWNHVSFVPLHYNLNA		
SEQID No 12	(251)	THVRNTTGKPMFVAEAFWKNLAAIENYLNKTSWNHVSFVPLHYNLNA		
SEQID No 13	(246)	RHORNEADQDLFVVGEYWKDDVGALEFYLDENWEMSLFDVPLNYNFYRA		
SEQID No 14	(246)	RHORSEADQDLFVVGEYWKDDVGALEFYLDENWEMSLFDVPLNYNFYRA		
SEQID No 15	(247)	SYVRSQTGKPLFTVGEYWSYDINKLHNYITKTNGTMSLFDAPLHNKFYTA		
Consensus 1	(251)	SHVRS TGK LFTVGEYW DIGALENYL KTNW MSLFDVPLHYNFY A		

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301
SEQID No 1 (299) SKSGGAFDMRTLMTNTLMKDQPTLAVTFVDNHDTEPGQALQSWVDPWFKP
SEQID No 2 (299) SKSGGAFDMRTLMTNTLMKDQPTLAVTFVDNHDTEPGQALQSWVDPWFKP
SEQID No 3 (299) SKSGGAFDMRTLMTNTLMKDQPTLAVTFVDNHDTEPGQALQSWVDPWFKP
SEQID No 4 (299) SKSGGAFDMRTLMTNTLMKDQPTLAVTFVDNHDTEPGQALQSWVDPWFKP
SEQID No 5 (299) SKSGGAFDMRTLMTNTLMKDQPTLAVTFVDNHDTEPGQALQSWVDPWFKP
SEQID No 6 (301) SKSGGNYDMRNI FNGTVVQRHPSHAVTFVDNHDSPPEEALESFVEEWFKP
SEQID No 7 (296) STQGGGYDMRKLNGTVVSKHPLKSVTFVDNHDTPGGQSLSTVQTFWFKP
SEQID No 8 (296) STQGGGYDMRKLNGTVVSKHPLKSVTFVDNHDTPGGQSLSTVQTFWFKP
SEQID No 9 (296) SSQGGGYDMRLLDGTVVSRRHPEKAVTFVENHDTQPGQSLSTVQTFWFKP
SEQID No 10 (301) SKSGGNYDMRQIFNGTVVQRHPMHAFTVDNHDSPPEEALESFVEEWFKP
SEQID No 11 (301) SNSGGNYDMAKLLNGTVVQKHPMHAFTVDNHDSPGESLESFVQEWFKP
SEQID No 12 (301) SNSGGYFDMRNILNGSVVQKHPHIAVTFVDNHDSPGEALESFVQSWFKP
SEQID No 13 (296) SQGGGSYDMRNILRGLSVEAHPMHAFTVDNHDTPGESLESVWADWFKP
SEQID No 14 (296) SKQGGSYDMRNILRGLSVEAHPHIAVTFVDNHDTPGESLESVWADWFKP
SEQID No 15 (297) SKSGGAFDMRTLMTNTLMKDQPTLAVTFVDNHDTEPGQALQSWVDPWFKP
Consensus 1 (301) SKSGGAYDMR LL GTLV HP AVTFVDNHDTPGGQALESVWD WFKP
351
SEQID No 1 (349) LAYAFILTRQEGYPCVFYGDYYGIPQYN---IPSLKSKIDPLLIARRDYA
SEQID No 2 (349) LAYAFILTRQEGYPCVFYGDYYGIPQYN---IPSLKSKIDPLLIARRDYA
SEQID No 3 (349) LAYAFILTRQEGYPCVFYGDYYGIPQYN---IPSLKSKIDPLLIARRDYA
SEQID No 4 (349) LAYAFILTRQEGYPCVFYGDYYGIPQYN---IPSLKSKIDPLLIARRDYA
SEQID No 5 (349) LAYAFILTRQEGYPCVFYGDYYGIPQYN---IPSLKSKIDPLLIARRDYA
SEQID No 6 (351) LAYALTLTREQGYPSVFYGDYYGIPTHG---VPAMRSKIDPILEARQKYA
SEQID No 7 (346) LAYAFILTRESGYPQVFYGDYMGYGTGDSQREIPALKHKIEPILKARKQYA
SEQID No 8 (346) LAYAFILTRESGYPQVFYGDYMGYGTGDSQREIPALKHKIEPILKARKQYA
SEQID No 9 (346) LAYAFILTRESGYPQVFYGDYMGYGTGTSPEKIPSLKDNIEPILKARKEYA
SEQID No 10 (351) LAYALTLTREQGYPSVFYGDYYGIPTHG---VPAMKSKIDPILEARQKYA
SEQID No 11 (351) LAYALILTREQGYPSVFYGDYYGIPTHS---VPAMKAKIDPILEARQNF
SEQID No 12 (351) LAYALILTREQGYPSVFYGDYYGIPTHG---VPSMKSIDPLLQARQTYA
SEQID No 13 (346) LAYATILTREGGYPNVFYGDYYGIPNDN---ISAKKDMIDELLDARQNYA
SEQID No 14 (346) LAYATILTREGGYPNVFYGDYYGIPNDN---ISAKKDMIDELLDARQNYA
SEQID No 15 (347) LAYAFILTRQEGYPCVFYGDYYGIPQYN---IPSLKSKIDPLLIARRDYA
Consensus 1 (351) LAYAFILTRE GYP VFYGDYYGIPQYN IPSLKSKIDPLL ARR YA
401
SEQID No 1 (396) YGTQHDYLDHSDIIGWTREGVTEKPGSGLAALITDGPGGSKWMYVGKQHA
SEQID No 2 (396) YGTQHDYLDHSDIIGWTREGVTEKPGSGLAALITDGPGGSKWMYVGKQHA
SEQID No 3 (396) YGTQHDYLDHSDIIGWTREGVTEKPGSGLAALITDGPGGSKWMYVGKQHA
SEQID No 4 (396) YGTQHDYLDHSDIIGWTREGVTEKPGSGLAALITDGPGGSKWMYVGKQHA
SEQID No 5 (396) YGTQHDYLDHSDIIGWTREGVTEKPGSGLAALITDGPGGSKWMYVGKQHA
SEQID No 6 (398) YGKQNDYLDHSDIIGWTREGNTHPNSGLATIMSDGAGGSKWMFVGRNKA
SEQID No 7 (396) YGAQHDFDHHDIVGWTREGDSSVANSGLAALITDGPGGAKRMYVGRQNA
SEQID No 8 (396) YGAQHDFDHHDIVGWTREGDSSVANSGLAALITDGPGGAKRMYVGRQNA
SEQID No 9 (396) YGPQHDFIDHPDIVGWTREGDSSAAKSGLAALITDGPGGSKRMYAGLKNA
SEQID No 10 (398) YGRQNDYLDHSDIIGWTREGNTHPNSGLATIMSDGAGGNKWMFVGRNKA
SEQID No 11 (398) YGTQHDYFDHSDIIGWTREGNTHPNSGLATIMSDGPGGKEMMYVGQNK
SEQID No 12 (398) YGTQHDYFDHSDIIGWTREGDSSHPNSGLATIMSDGPGGNKWMYVGKHA
SEQID No 13 (393) YGTQHDYFDHWDVVGWTREGSSSRPNSGLATIMSNPGGSKWMYVGRQNA
SEQID No 14 (393) YGTQHDYFDHWDIVGWTREGTSSSRPNSGLATIMSNPGGSKWMYVGQQA
SEQID No 15 (394) YGTQHDYLDHSDIIGWTREGGTEKPGSGLAALITDGPGGSKWMYVGKQHA
Consensus 1 (401) YGTQHDYLDH DIIGWTREG TSKPNSGLAALITDGPGGSKWMYVGKQ A

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Figure 1 (3 of 4)

		451	500
SEQID No 1	(446)	GKVFYDLTGNRSDTVTINSDGWGEFKVNGGSVSVVWVPRKTTVSTIARPIT	
SEQID No 2	(446)	GKVFYDLTGNRSDTVTINSDGWGEFKVNGGSVSVVWVPRKTT-----	
SEQID No 3	(446)	GKVFYDLTGNRSDTVTINSDGWGEFKVNGGSVSVVWVPRKTTVSTIARPIT	
SEQID No 4	(446)	GKVFYDLTGNRSDTVTINSDGWGEFKVNGGSVSVVWVPRKTTVSTIARPIT	
SEQID No 5	(446)	GKVFYDLTGNRSDTVTINSDGWGEFKVNGGSVSVVWVPRKTTVSTIARPIT	
SEQID No 6	(448)	GQVWSDITGNRTGTVTINADGWGNFSVNGGSVSIWVNK-----	
SEQID No 7	(446)	GETWHDITGNRSEPVVINSEGWGEFHVNGGSVSIYVQR-----	
SEQID No 8	(446)	GETWHDITGNRSEPVVINSEGWGEFHVNGGSVSIYVQR-----	
SEQID No 9	(446)	GETWYDITGNRSDTVKIGSDGWGEFHVNDGSVSIYVQK-----	
SEQID No 10	(448)	GQVWTDITGNRAGTVTINADGWGNFSVNGGSVSIWVNK-----	
SEQID No 11	(448)	GQVWHDITGNKPGTVTINADGWANFSVNGGSVSIWVKR-----	
SEQID No 12	(448)	GQVWRDITGNRSGTVTINADGWGNFTVNGGAVSVVVKQ-----	
SEQID No 13	(443)	GQTWDLTGNGASVTINGDGWGEFFTNNGGSVSVYVNO-----	
SEQID No 14	(443)	GQTWDLTGNGAASVTINGDGWGEFFTNNGGSVSVYVNO-----	
SEQID No 15	(444)	GKVFYDLTGNRSDTVTINSDGWGEFKVNGGSVSVVWVPRKTTVS-----	
Consensus 1	(451)	G VWYDLTGNRSDTVTINSDGWGEF VNGGSVSVVW R	
		501	520
SEQID No 1	(496)	TRPWTGEFVRWTEPRLVAVP	
SEQID No 2	(487)	-----	
SEQID No 3	(496)	TRPWTGEFVRWTEPRLVAVP	
SEQID No 4	(496)	TRPWTGEFVRWTEPRLVAVP	
SEQID No 5	(496)	TRPWTGEFVRWTEPRLVAVP	
SEQID No 6	(486)	-----	
SEQID No 7	(484)	-----	
SEQID No 8	(484)	-----	
SEQID No 9	(484)	-----	
SEQID No 10	(486)	-----	
SEQID No 11	(486)	-----	
SEQID No 12	(486)	-----	
SEQID No 13	(481)	-----	
SEQID No 14	(481)	-----	
SEQID No 15	(487)	-----	
Consensus 1	(501)		

Figure 1 (4 of 4)

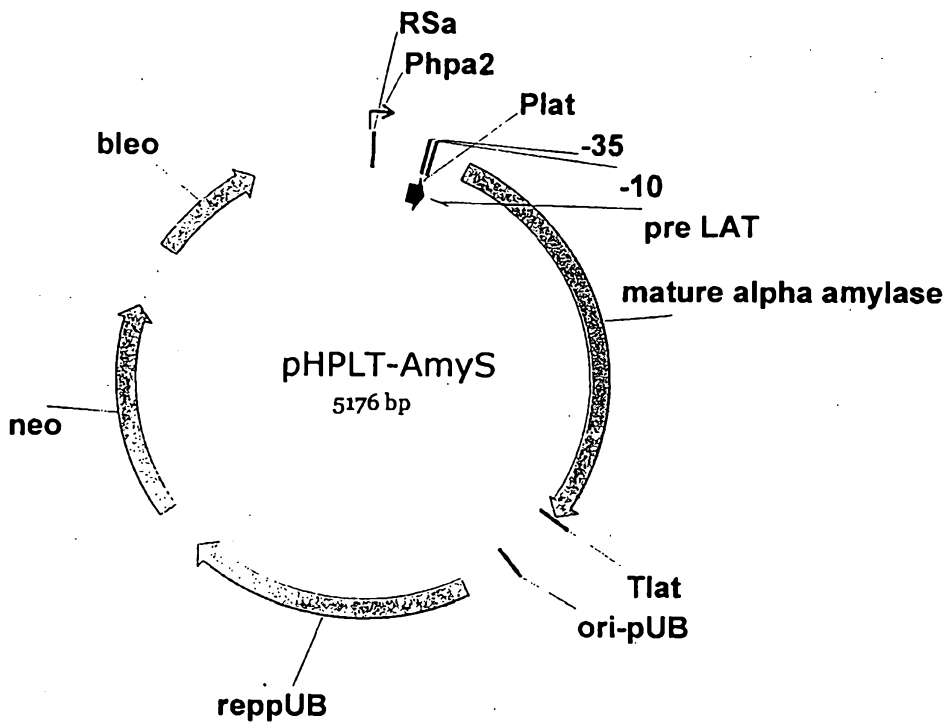


Figure 2

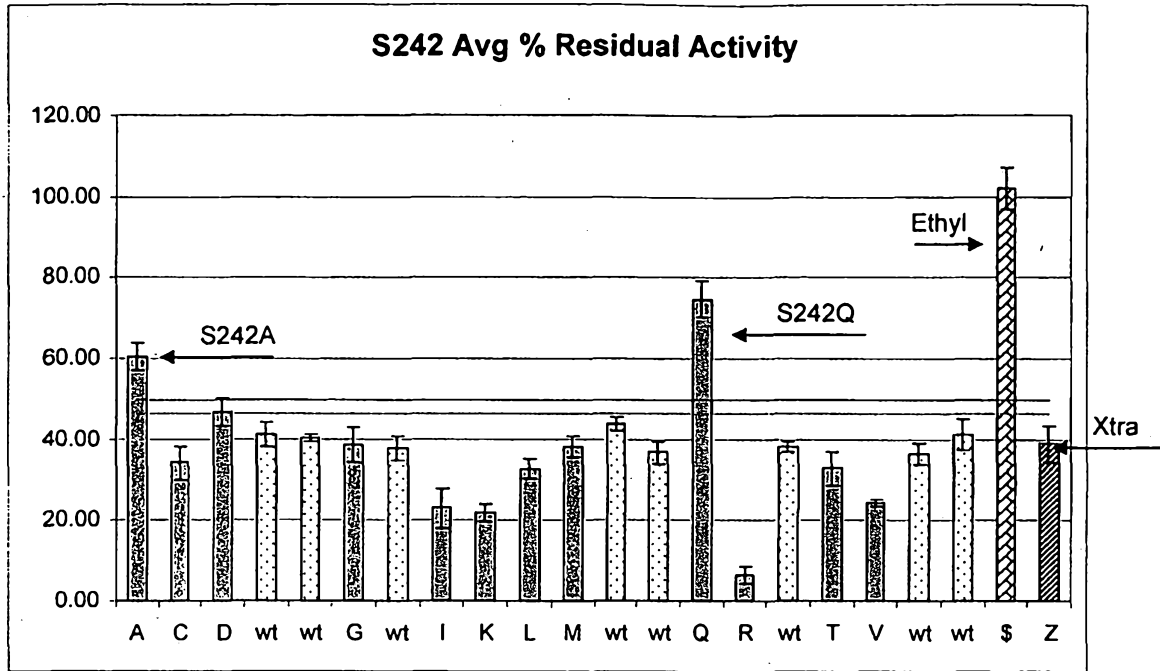


Figure 3

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1
50
SEQID No 1 (1) -AAPFNGTMMQYFEWYLPDDGTLWTKVANEANNLSSLGITALWLPPAYKG
SEQID No 6 (1) HHNGTNGTMMQYFEWYLPNDGNHWNRLNSDASNLKSKGITAVWIPPAWKG
Consensus 2 (1) NGTMMQYFEWYLP DG W KL DA NL S GITALWIPPAWKG
51
100
SEQID No 1 (50) TSRSDVGYGVYDLYDLGFEFNQKGTVRTKYGTAKQYLQAIQAAHAAGMQVY
SEQID No 6 (51) ASQNDVGYGAYDLYDLGFEFNQKGTVRTKYGTRSOLQAAVTSLKNNGIQVY
Consensus 2 (51) S DVGYG YDLYDLGFEFNQKGTVRTKYGTAKQ AI A GIQVY
101
150
SEQID No 1 (100) ADVVFDHKGADGTEWVDAVEVNPSPDRNQEISGTYQIQAWTKFDFPGRGN
SEQID No 6 (101) GDVVMNHKGGADATEMVRAVEVNPNNRNQEVTEGYTIEAWTRFDFPGRGN
Consensus 2 (101) ADVV HKGGADATE V AVEVNP RNQEISG Y I AWTKFDFPGRGN
151
200
SEQID No 1 (150) TYSSFKWRWYHFDGVDWDESRKLS-RIYKFRGIGKAWDWEVDTEGNNDY
SEQID No 6 (151) THSSFKWRWYHFDGVDWQSRRLNNRIYKFRGHGKAWDWEVDTEGNNDY
Consensus 2 (151) THSSFKWRWYHFDGVDW SRKL RIYKFRG GKAWDWEVDTEGNNDY
201
250
SEQID No 1 (199) LMYADLMDHPEVVTELKNWGKQYVNTTNIDGFRIDAVKHIFSFPPDWL
SEQID No 6 (201) LMYADIDMDHPEVVNELRNWGVWYNTNLGLDGFRIDAVKHIFSFTRDWI
Consensus 2 (201) LMYADIDMDHPEVV ELKNWG WY NT IDGFRIDAVKHIFSF DWI
251
300
SEQID No 1 (249) SYVRSQTGKPLFTVGEYWSYDINKLHNYITKNGTMSLFDAPLHNKFYTA
SEQID No 6 (251) NHVRSATGKNMFAVAEFWKNLGAIEENYLQKTNNWHSVFDVPLHYNLYNA
Consensus 2 (251) HVRS TGK LF VAEFW DI I NYI KTN SLFD PLH Y A
301
350
SEQID No 1 (299) SKSGGAFDMRMLTNTLMKDQPTLAVTFVDNHDTEPGQALQSWVDPWFKP
SEQID No 6 (301) SKSGGNYDMRNIFNGTVVQRHPSHAVTFVDNHDSPQEEALESFVEEWFKP
Consensus 2 (301) SKSGG FDMR I TLM PS AVTFVDNHD S P AL SFVD WFKP
351
400
SEQID No 1 (349) LAYAFILTRQEGYPCVFYGDYYGIPQYNIPSLKSKIDPLLIARRDYAYGT
SEQID No 6 (351) LAYALTLTREQGYPSVFYGDYYGIPTHGVPAMRSKIDPILEARQKYAYGK
Consensus 2 (351) LAYA LTR GYP VFYGDYYGIP H IPALKSKIDPIL AR YAYG
401
450
SEQID No 1 (399) QHDYLDHSDIIGWTREGVTEKPGSGLAALITDGGGSKWMYVYGKQKAGKV
SEQID No 6 (401) QNDYLDHNNIIGWTREGNTAHPNSGLATIMSDGAGGSKWMFVGRNKAGQV
Consensus 2 (401) Q DYLDH IIGWTREG T P SGLA IISDG GSKWMFVGRN AG V
451
500
SEQID No 1 (449) FYDLTGNRSDTVTINS DGWGEFKVNGGSVSVVWVPRKTTVSTIARPIITRP
SEQID No 6 (451) WSDITGNRTGTVTINADGWGNFSVNGGSVSIWVK-----
Consensus 2 (451) F DITGNRS TVTINADGWG F VNGGSVSIWV K
501 517
SEQID No 1 (499) WTGEFVRWTEPRLVAWP
SEQID No 6 (486) -----
Consensus 2 (501)

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Figure 4A

		1		50
SEQID No 1	(1)	AAPFNGTMMQYFEWYLPDDGTLWTKVANEANNLSSLGITALWLPPAYKGT		
SEQID No 8	(1)	-ANLNGTLMQYFEWYMPNDGQHWRRRLQND SAYLAEHGITAVWI PPAYKGT		
Consensus 3	(1)	A NGTLMQYFEWYLP DG W KL NDA LA GITALWIPPAYKGT		
		51		100
SEQID No 1	(51)	SRSDVGYGVYDLYDLGFEFNQKGTVRTKYGTKAQYLQAIQAAHAAGMQVYA		
SEQID No 8	(50)	SQADVGYGAYDLYDLGFEFHQKGTVRTKYGTGELQSAIKSLHSRDINVYG		
Consensus 3	(51)	S ADVGYG YDLYDLGEF QKGTVRTKYGTKA AI A HA INVYA		
		101		150
SEQID No 1	(101)	DVVFDPHKGADGTEWVDAVEVNPSPDRNQEISGTYQIQAWTKFDFPGRGNT		
SEQID No 8	(100)	DVVINHKGGADATEDVTAVEVDPADRNRVISGEHLIKAWTHFHFPGRGST		
Consensus 3	(101)	DVV HKGGADATE V AVEV PADRN ISG H I AWT F FPGRG T		
		151		200
SEQID No 1	(151)	YSSFKWRWYHFDGVDWDESRKLSRIYKFRGIGKAWDWEVDTENGNVDYLM		
SEQID No 8	(150)	YSDFKWHWYHFDGTDWDESRKLNRIYKFQ--GKAWDWEVSNENGNVDYLM		
Consensus 3	(151)	YS FKW WYHFDG DWDESRKL RIYKF GKAWDWEV ENGNVDYLM		
		201		250
SEQID No 1	(201)	YADLMDHPEVVTELKNWGKWYVNTTNIDGFRLDAVKHIKFSFFPDWLSY		
SEQID No 8	(198)	YADIDYDHPDVAAEIKRWGTWYANELQLDGFRLDAVKHIKFSFLRDWVNH		
Consensus 3	(201)	YADID DHPDV EIK WG WY N NIDGFRLDAVKHIKFSF DWL H		
		251		300
SEQID No 1	(251)	VRVQTKGKPLFTVGEYWSYDINKLHNYITKTNGTMSLFDAPLHNKFYTASK		
SEQID No 8	(248)	VREKTGKEMFTVAEYWQNDLGALENYLNKTNFNHSVFDVPLHYQFHAAS		
Consensus 3	(251)	VR TKG LFTVAEYW DI L NYI KTN SLFD PLH FH AS		
		301		350
SEQID No 1	(301)	SGGAFDMRMLTNTLTKDQPTLAVTFVDNHDTEPGQALQSWDPEWFKPLA		
SEQID No 8	(298)	QGGGYDMRKLNGTVVSKHPLKSVTFVDNHDTPGGQSLESTVQTFWFKPLA		
Consensus 3	(301)	GGAFDMR LL TLM P AVTFVDNHDTE PGQAL S V WFKPLA		
		351		400
SEQID No 1	(351)	YAFILTRQEGYPCVFYGDYYGIP---QYNIPSLKSKIDPLLIARRDYAYG		
SEQID No 8	(348)	YAFILTRESGYPQVFYGDYGTGKDSQREIPALKKHKIEPILKARKQYAYG		
Consensus 3	(351)	YAFILTR GYP VFYGD YG Q IPALK KIDPIL ARK YAYG		
		401		450
SEQID No 1	(398)	TQHDYLDHSDIIGWTREGVTEKPGSGLAALITDGPGGSKWYVVGKQHAGK		
SEQID No 8	(398)	AQHDFDHHDIVGWTRREGDSSVANSGLAALITDGPGGAKRMYVGRQAGE		
Consensus 3	(401)	QH DY DH DIIGWTREG S SGLAALITDGPGGAK MYVGKQ AG		
		451		500
SEQID No 1	(448)	VFYDLTGNRSDTVTINSDGWGEFKVNGGSVSVVWVPRKTTVSTIARPITTR		
SEQID No 8	(448)	TWHDITGNRSEPVVINSEGWGEFHVNGGSVSIYVQR-----		
Consensus 3	(451)	FHDITGNRSD V INSDGWGEF VNGGSVSIWV R		
		501		518
SEQID No 1	(498)	PWTGEFVRWTEPRLVAVP		
SEQID No 8	(484)	-----		
Consensus 3	(501)			

Figure 4B

		1		50
SEQID No 1	(1)	AAPFNGTMMQYFEWYLPDDGTLWTKVANEANNLSSLGITALWLPPAYKGT		
SEQID No 9	(1)	---VNGTLMQYFEWYTPNDGQHWKRLQNDAEHLSDIGITAVWIPPAYKGL		
Consensus 4	(1)	NGTLMQYFEWY P DG W KL NDA LS IGITALWIPPAYKG		
		51		100
SEQID No 1	(51)	SRSDVGYGVYDLYDLGFEFNQKGTVRTKYGTKAQYLQAIQAAHAAGMQVYA		
SEQID No 9	(48)	SQSDNGYGPYDLYDLGFEFQQKGTVRTKYGTKSELQDAIGSLHSRNVQVYG		
Consensus 4	(51)	S SD GYG YDLYDLGFEFNQKGTVRTKYGTKA AI A HA MQVYA		
		101		150
SEQID No 1	(101)	DVVFHDKGGADGTEWVDAVEVNPSDRNQEISGTYQIQAWTKFDFPGRGNT		
SEQID No 9	(98)	DVVLNHKAGADATEDVTAVEVNPANRNQETSEEYQIKAWTDFRFPGRGNT		
Consensus 4	(101)	DVV HKAGADATE V AVEVNPA RNQE S YQI AWT F FPGRGNT		
		151		200
SEQID No 1	(151)	YSSFKWRWYHFDGVDWDESRLKSRIYKFRGIGKAWDWEVDTENGYDYL		
SEQID No 9	(148)	YSDFKWHWYHFDGADWDESRLKISRIFKFRGEGKAWDWEVSSENGYDYL		
Consensus 4	(151)	YS FKW WYHFDG DWDESRLKISRIFKFRG GKAWDWEV SENGNYDYL		
		201		250
SEQID No 1	(201)	YADLMDHPEVVTELKNWGKWYVNTTNIDGFRLDAVKHIKFSFFPDWLSY		
SEQID No 9	(198)	YADVYDHPDVVAETKKWGIWYANELSLDGFRIDAAKHIKFSFLRDWVQA		
Consensus 4	(201)	YADLD DHPDVV E K WG WY N IDGFRIDA KHIKFSF DWL		
		251		300
SEQID No 1	(251)	VRSQTGKPLFTVGEYWSYDINKLHNYITKTNGTMSLFDAPLHNKFYTASK		
SEQID No 9	(248)	VRQATGKEMFTVAEYWQNNAGKLENYLNKTSFNQSVFDVPLHFNLQAASS		
Consensus 4	(251)	VR TGK LFTVAEYW KL NYI KT SLFD PLH AS		
		301		350
SEQID No 1	(301)	SGGAFDMRTLMTNTLMKDQPTLAVTFVDNHDTEPGQALQSWVDPWFKPLA		
SEQID No 9	(298)	QGGGYDMRRLLDGTVVSRRHEKAVTFVENHDTQPGQSLESTVQTFWKPLA		
Consensus 4	(301)	GGAFDMR LL TLM P AVTFVDNHDTE PGQAL S V WFKPLA		
		351		400
SEQID No 1	(351)	YAFILTRQEGYPCVFYGDYYGIPQYN---IPSLKSKIDPLLIARRDYAYG		
SEQID No 9	(348)	YAFILTRESGYPQVFYGDYMGTKGTSPKEIPSLKDNIEPILKARKEYAYG		
Consensus 4	(351)	YAFILTR GYP VFYGD YG IPSLK IDPIL ARKDYAYG		
		401		450
SEQID No 1	(398)	TQHDYLDHSDIIGWTREGVTEKPGSGLAALITDGPGGSKWMYVKGQHAGK		
SEQID No 9	(398)	PQHDYIDHPDVIGWTREGDSSAAKSGLAALITDGPGGSKRMYAGLKNAGE		
Consensus 4	(401)	QHDYIDH DIIGWTREG S SGLAALITDGPGGSK MY G AG		
		451		500
SEQID No 1	(448)	VFYDLTGNRSDTVINSDBGWGEFVKVNGGSVSVVWVPRKTTVSTIARPIPTR		
SEQID No 9	(448)	TWYDITGNRSDTVKIGSDGWGEFHVNDGSVSIYVQK-----		
Consensus 4	(451)	FYDITGNRSDTV I SDGWGEF VN GSVSIWV K		
		501		518
SEQID No 1	(498)	PWTGEFVRWTEPRLVAWP		
SEQID No 9	(484)	-----		
Consensus 4	(501)			

Figure 4C

		1	50
SEQID No 1	(1)	-AAPFNGTMMQYFEWYLPDDGTLWTKVANEANNLSSLGITALWLPPAYKG	
SEQID No 10	(1)	HHNGTNGTMMQYFEWYLPNDGNHWNRLRSASNLKDKGISAVWIPPAWKG	
Consensus 5	(1)	NGTMMQYFEWYLP DG W KL DA NL GISALWIPPAWKG	
		51	100
SEQID No 1	(50)	TSRSDVGYGVYDLYDLGFEFNQKGTVRTKYGTAKQYLQAIQAAHAAGMQVY	
SEQID No 10	(51)	ASQNDVGYGAYDLYDLGFEFNQKGTIRTQYGTNRNQLQAAVNALKSNGIQVY	
Consensus 5	(51)	S DVGYG YDLYDLGFEFNQKGTIRTQYGTK Q AINA A GIQVY	
		101	150
SEQID No 1	(100)	ADVVFHKGKGGADGTEWVDAVEVNPSPDRNQEISGTYQIQAWTKFDFPGRGN	
SEQID No 10	(101)	GDVVMNHKGGADATEMVRAVEVNPNNRNQEVSGEYIEAWTKFDFPGRGN	
Consensus 5	(101)	ADVV HKGGADATE V AVEVNP RNQEISG Y I AWTKFDFPGRGN	
		151	200
SEQID No 1	(150)	TYSSFKWRWYHFDGVDWDESRKLS-RIYKFRGIGKAWDWEVDTENGNYDY	
SEQID No 10	(151)	THSNFKWRWYHFDGVDWQSRKLNRIYKFRGDGKGDWEVDTENGNYDY	
Consensus 5	(151)	THS FKWRWYHFDGVDW SRKL RIYKFRG GKAWDWEVDTENGNYDY	
		201	250
SEQID No 1	(199)	LMYADLMDHPEVVTELKNWGKQYVNTTNIDGFRIDAVKHKIFSFDPDWL	
SEQID No 10	(201)	LMYADIDMDHPEVVNELRNWGVWYNTNLGLDGFRIDAVKHKIFSFTRDWI	
Consensus 5	(201)	LMYADIDMDHPEVV ELKNWG WY NT IDGFRIDAVKHKIFSF DWI	
		251	300
SEQID No 1	(249)	SYVRSQTGKPLFTVGEYWSYDINKLHNYITKTNGTMSLFDAPLHNKFYTA	
SEQID No 10	(251)	NHVRSATGKNMFAVAEFWKNDLGAIEYLNKTNWNHVSFVPLHYNLNA	
Consensus 5	(251)	HVRS TGK LF VAEFW DI I NYI KTN SLFD PLH Y A	
		301	350
SEQID No 1	(299)	SKSGGAFDMRTLMTNTLMKDQPTLAVTFVDNHDTEPGQALQSWVDPWFKP	
SEQID No 10	(301)	SKSGGNYDMRQIFNGTVVQRHPMHAVTFVDNHDSPQEEALESFVEEWFKP	
Consensus 5	(301)	SKSGG FDMR I TLM P AVTFVDNHDSP AL SFVD WFKP	
		351	400
SEQID No 1	(349)	LAYAFILTRQEGYPCVFGDYDGIPQYNIPSLKSKIDPLLIARRDYAYGT	
SEQID No 10	(351)	LAYALTLTREQGYPSVFGDYDGIPHGVPAKSKIDPILEARQKYAYGR	
Consensus 5	(351)	LAYA LTR GYP VFGDYDGIP H IPALKSKIDPIL AR YAYG	
		401	450
SEQID No 1	(399)	QHDYLDHSDIIGWTREGVTEKPGSGLAALITDGPGGSKWYVVGKQHAGKV	
SEQID No 10	(401)	QNDYLDHSDIIGWTREGNTAHPNSGLATIMSDGAGGNKWMFVGRNKAGQV	
Consensus 5	(401)	Q DYLDH IIGWTREG T P SGLA IISDG GG KWMFVGKN AG V	
		451	500
SEQID No 1	(449)	FYDLTGNRSDTVTINSDGWGEFKVNGGSVSVVWVPRKTTVSTIARPIITRP	
SEQID No 10	(451)	WTDITGNRAGTVTINADGWGNFSVNGGSVSIWVNK-----	
Consensus 5	(451)	F DITGNRA TVTINADGWG F VNGGSVSIWV K	
		501	517
SEQID No 1	(499)	WTGEFVRWTEPRLVAVP	
SEQID No 10	(486)	-----	
Consensus 5	(501)		

Figure 4D



		1		50
SEQID No 1	(1)	-AAPFNGTMMQYFEWYLPDDGTLWTKVANEANNLSSLGITALWLPPAYKG		
SEQID No 11	(1)	HHNGTNGTMMQYFEWHLPN DGNHWNRLRDDASNLNRGITA IWI PPAWKG		
Consensus 6	(1)	NGTMMQYFEWHL P DG W KL DA NL GITAIWI PPAWKG		
		51		100
SEQID No 1	(50)	TSRSDVGYGVYDLYDLGFEFNQKGTVRTKYGTKAQYLQAIQAAHAAGMQVY		
SEQID No 11	(51)	TSQNDVGYGAYDLYDLGFEFNQKGTVRTKYGTRSQLES A I H A L K N N G V Q V Y		
Consensus 6	(51)	TS DVG YG Y D L Y D L G F E F N Q K G T V R T K Y G T K A Q A I A G M Q V Y		
		101		150
SEQID No 1	(100)	ADV V F D H K G G A D G T E W D A V E V N P S D R N Q E I S G T Y Q I Q A W T K F D F P G R G N		
SEQID No 11	(101)	G D V V M N H K G G A D A T E N V L A V E V N P N N R N Q E I S G D Y T I E A W T K F D F P G R G N		
Consensus 6	(101)	ADV V H K G G A D A T E V A V E V N P R N Q E I S G Y I A W T K F D F P G R G N		
		151		200
SEQID No 1	(150)	TYSSFKWRWYHFDGVDWDESR-KLSRIYKFRGIGKAWDWEVDTENGN YDY		
SEQID No 11	(151)	TYSDFKWRWYHFDGVDWQSRQFQNR IYKFRGDGKAWDWEVDS ENGN YDY		
Consensus 6	(151)	TYS FKWRWYHFDGVDWD SR R IYKFRG GKAWDWEVDS ENGN YDY		
		201		250
SEQID No 1	(199)	LMYADLMDHPEVVTELKNWGK WYVNTT NIDGFR L D A V K H I K F S F F P D W L		
SEQID No 11	(201)	LMYADVDMDHPEVVNELRRWGEWY T N T L N L D G F R I D A V K H I K Y S F T R D W L		
Consensus 6	(201)	LMYADLMDHPEVV ELK WG WY NT NIDGFR I D A V K H I K F S F D W L		
		251		300
SEQID No 1	(249)	SYVRSQTGKPLFTVGEYWSYDINKLHNYITKTNGTMSLFDAPLHNKFYTA		
SEQID No 11	(251)	THVRNATGKEMFAVAEFWKNDLGALENYLNKTNWNH SVFDVPLHYNLYNA		
Consensus 6	(251)	SHVR TGK LF VAEFW DI L NYI KTN SLFD PLH Y A		
		301		350
SEQID No 1	(299)	SKSGGAFDMRTLMTNTL MKDQPTLAVTFVDNHDTEPGQALQSWVDPWFKP		
SEQID No 11	(301)	SNSGGNYDMAKLLNGTVVQKHPMAVTFVDNHD S Q P G E S L E S F V Q E W F K P		
Consensus 6	(301)	S SGG FDM LL TLM P AVTFVDNHD S PG AL SFV WFKP		
		351		400
SEQID No 1	(349)	LAYAFILTRQEGYPCVFYGDYYGIPQYNI PSLKSKIDPLLIARRDYAYGT		
SEQID No 11	(351)	LAYALILTR EQGYP S V F Y G D Y Y G I P T H S V P A M K A K I D P I L E A R Q N F A Y G T		
Consensus 6	(351)	LAYA ILTR GYP VFYGDYYGIP H IPALKAKIDPIL AR FAYGT		
		401		450
SEQID No 1	(399)	QHDYLDHSDIIGWTREGVTEKPGSGLAALITDGP GSKWMYVGQKHAGKV		
SEQID No 11	(401)	QHDYFDH H N I I G W T R E G N T T H P N S G L A T I M S D G P G G E K W M Y V G Q N K A G Q V		
Consensus 6	(401)	QHDY DH IIGWTREG T P SGLA IISDGP G K W M Y V G N A G V		
		451		500
SEQID No 1	(449)	FYDLTGNRSDT V T I N S D G W G E F K V N G G S V S V W V P R K T T V S T I A R P I T T R P		
SEQID No 11	(451)	WHDITGNKPGTVTINADGWANFSVNGG S V S I W V K R -----		
Consensus 6	(451)	FHDITGNK TVTINADGWA F VNGG S V S I W V R		
		501		517
SEQID No 1	(499)	WTGEFVRWTEPRLVAWP		
SEQID No 11	(486)	-----		
Consensus 6	(501)			

Figure 4E

```

1 50
SEQID No 1 (1) -AAPFNGTMMQYFEWYLPDDGTLWTKVANEANNLSSLGITALWLPPAYKG
SEQID No 12 (1) HHNGTNGTMMQYFEWHLPNLNDGNHWNRLRDDAANLKSNGITAVWIPPAWK
Consensus 7 (1) NGTMMQYFEWHL DP W KL DA NL S GITALWIPPAWK
51 100
SEQID No 1 (50) TSRSDVGYGVYDLYDLGFEFNQKGTVRTKYGTKAQYLQAIQAAHAAGMQVY
SEQID No 12 (51) TSQNDVGYGAYDLYDLGFEFNQKGTVRTKYGTRSOLQGAVTSLKNNGIQVY
Consensus 7 (51) TS DVGYG YDLYDLGFEFNQKGTVRTKYGTKAQ AI A GIQVY
101 150
SEQID No 1 (100) ADVVFDHKGADGTEWVDAVEVNPSDRNQEISGTYQIQAWTKFDFPGRGN
SEQID No 12 (101) GDVVMNHKGGADGTEMVNAVEVNRNRNQEISGEYTIEAWTKFDFPGRGN
Consensus 7 (101) ADVV HKGGADGTE V AVEVN S RNQEISG Y I AWTKFDFPGRGN
151 200
SEQID No 1 (150) TYSSFKWRWYHFDGVDWDESR-KLSRIYKFRGIGKAWDWEVDTEGNYDY
SEQID No 12 (151) THSNFKWRWYHFDGTDWDQSRQLQNKIYKFRGTGKAWDWEVDIENGNYDY
Consensus 7 (151) THS FKWRWYHFDG DWD SR KIYKFRG GKAWDWEVD ENGN DY
201 250
SEQID No 1 (199) LMYADLMDHPEVVTTELKNWGKWYVNTTIDGFRDLAVKHKIFSFFPDWL
SEQID No 12 (201) LMYADIDMDHPEVINELRNWGVWYNTLNLGDGFRIDAVKHKISYTRDWL
Consensus 7 (201) LMYADIDMDHPEVI ELKNWG WY NT NIDGFRIDAVKHKIFS DWL
251 300
SEQID No 1 (249) SYVRSQTGKPLFTVGEYWSYDINKLHNYITKTNGTMSLFDAPLHNKFYTA
SEQID No 12 (251) THVRNTTGKPMFAVAEFWKNDLAAIENYLNKTSWNHVSFVPLHYNLNA
Consensus 7 (251) SHVR TGKPLF VAEFW DI I NYI KT SLFD PLH Y A
301 350
SEQID No 1 (299) SKSGGAFDMRTLMTNTLMKDQPTLAVTFVDNHDTEPGQALQSWVDPWFKP
SEQID No 12 (301) SNSGGYFDMRNILNGSVVQKHPHIAVTFVDNHDSQPGEALESFVQSWFKP
Consensus 7 (301) S SGG FDMR IL SLM P AVTFVDNHDS PG AL SFV WFKP
351 400
SEQID No 1 (349) LAYAFILTRQEGYPCVFGDYGGIPQYNIPSLKSKIDPLLIARROYAYGT
SEQID No 12 (351) LAYALILTREQGYPSVFGDYGGIPHTGVPSMKSKIDPLLQARQTYAYGT
Consensus 7 (351) LAYA ILTR GYP VFGDYGGIP H IPSLKSKIDPLL AR YAYGT
401 450
SEQID No 1 (399) QHDYLDHSDIIGWTREGVTEKPGSGLAALITDGPGGSKWMYVGKQHAGKV
SEQID No 12 (401) QHDYFDHDDIIGWTREGDSSHPNSGLATIMSDGPGGNKMYVGKHKAGQV
Consensus 7 (401) QHDY DH DIIGWTREG S P SGLA IISDGP GG KMYVGK AG V
451 500
SEQID No 1 (449) FYDLTGNRSDTVTINSWGGEFKVNGGSVSVVWVPRKTTVSTIARPIITRP
SEQID No 12 (451) WRDITGNRSGTVTINADGWGNFTVNGGAVSVVWKQ-----
Consensus 7 (451) F DITGNRS TVTINADGWG F VNGGAVSVVW
501 517
SEQID No 1 (499) WTGEFVRWTEPRLVAVP
SEQID No 12 (486) -----
Consensus 7 (501)

```

Figure 4F

		1		50
SEQID No 1	(1)	AAPFNGTMMQYFEWYLPDDGTLWTKVANEANNLSSLGITALWLPPAYKGT		
SEQID No 13	(1)	-DGLNGTMMQYYEWHLENDGQHWNRHLHDDAAALSDAGITAIWIPPAYKGN		
Consensus 8	(1)	NGTMMQYFEWHL DG W KL DA LS GITAIWIPPAYKG		
		51		100
SEQID No 1	(51)	SRSDVGYGVYDLYDLGFEFNQKGTVRTKYGTAKQYLQAIQAAHAAGMQVYA		
SEQID No 13	(50)	SQADVGYGAYDLYDLGFEFNQKGTVRTKYGTAKQLERAIGSLKSNDINVYG		
Consensus 8	(51)	S ADVGYG YDLYDLGFEFNQKGTVRTKYGTAKQ AI A A INVYA		150
		101		150
SEQID No 1	(101)	DVVFHDHKGADGTEWVDAVEVNPSDRNQEISGTYQIQAWTKFDFPGRGNT		
SEQID No 13	(100)	DVVMNHKMGADTFEAVQAVQVNPTNRWQDISGAYTIDAWTGDFDFSGRNNNA		
Consensus 8	(101)	DVV HK GAD TE V AV VNPS R QDISG Y I AWT FDF GR N		200
		151		200
SEQID No 1	(151)	YSSFKWRWYHFDGVDWDESRKLSRIYKFRGIGKAWDWEVDTENGN DYLM		
SEQID No 13	(150)	YSDFKWRWFHFNQVDWDQRYQENHIFRFANTN--WNWRVDEENGN DYLL		
Consensus 8	(151)	YS FKWRWFHF GVDWD IFKF W W VD ENGN DYLL		250
		201		250
SEQID No 1	(201)	YADLMDHPEVVTELKNWGKQYVNTTNIDGFRDLAVKHKFSFFPDWLSY		
SEQID No 13	(198)	GSNIDFSHPEVQDELKDWGSWFTDELDDLGYRLDAIKHIPFYTSDWVRH		
Consensus 8	(201)	A ID HPEV ELK WG WF IDGFRDLAIKHI F F DWL H		300
		251		300
SEQID No 1	(251)	VRSQTGKPLFTVGEYWSYDINKLHNYITKTNGTMSLFDAPLHNKFYTASK		
SEQID No 13	(248)	QRNEADQDLFVVGGEYWKDDVGALEFYLDENWEMSLFDVPLNRYFYRASQ		
Consensus 8	(251)	R LF VGEYW DI L YI N MSLFD PL FY AS		350
		301		350
SEQID No 1	(301)	SGGAFDMRTLMTNTLMKDQPTLAVTFVDNHDTPEGQALQSWVDPWFKPLA		
SEQID No 13	(298)	QGGSYDMRNILRGLSVEAHPMHAVTFVDNHDTQPGESLESWVADWFKPLA		
Consensus 8	(301)	GGAFDMR IL SLM P AVTFVDNHDT PG AL SWV WFKPLA		400
		351		400
SEQID No 1	(351)	YAFILTRQEGYPCVFGDYDGIPQYNIPSLKSKIDPLLIARRDYAYGTQH		
SEQID No 13	(348)	YATILTRREGGYPNVFGDYDGIPNDNISAKKDMIDELLDARQNYAYGTQH		
Consensus 8	(351)	YA ILTR GYP VFYGDYDGIPN NI A K ID LL AR YAYGTQH		450
		401		450
SEQID No 1	(401)	DYLDHSDIIGWTREGVTEKPGSGLAALITDGPGGSKWMYVVGKQHAGKV FY		
SEQID No 13	(398)	DYFDHWDVVGWWTREGSSSRPNSGLATIMSNGPGGSKWMYVGRQAGQWT		
Consensus 8	(401)	DY DH DIIGWTREG S KP SGLA IIS GPGGSKWMYVVGKQ AG F		500
		451		500
SEQID No 1	(451)	DLTGNRSDTV TINS DGWGEFKVNGGSVSVVWVPRKTTVSTIARPI TRPWT		
SEQID No 13	(448)	DLTGNGASVTINGDGWGEFFTNGGSVSVVYNQ-----		
Consensus 8	(451)	DLTGN SVTIN DGWGEF NGGSVSVVW		515
		501		515
SEQID No 1	(501)	GEFVRWTEPRLVAMP		
SEQID No 13	(481)	-----		
Consensus 8	(501)			

Figure 4G

```

1
50
SEQID No 1 (1) AAPFNGTMMQYFEWYLPDDGTLWTKVANEANNLSSLGITALWLPPAYKGT
SEQID No 14 (1) -DGLNGTMMQYYEWHLENDGQHWNRHLHDDAEALS NAGITAIWI PPAYKGN
Consensus 9 (1) NGTMMQYFEWHL DG W KL DA LS GITAIWI PPAYKG
51
100
SEQID No 1 (51) SRSDVGYGVYDLYDLGEFNOQKGTVRTKYGTKAQYLQAIQAAHAAGMQVYA
SEQID No 14 (50) SQADVGYGAYDLYDLGEFNOQKGTVRTKYGTKAQLERAIGSLKSNNDINVYG
Consensus 9 (51) S ADVGYG YDLYDLGEFNOQKGTVRTKYGTKAQ AI A A INVYA
101
150
SEQID No 1 (101) DVVFDHKGGADGTEWVDAVEVNPSDRNQEISGTYQIQAWTKFDFPGRGNT
SEQID No 14 (100) DVVMNHKLGADTFEAVQAVQVNPSNRWQDISGVYTIDAWTGFDFPGRRNA
Consensus 9 (101) DVV HK GAD TE V AV VNPS R QDISG Y I AWT FDFPGR N
151
200
SEQID No 1 (151) YSSFKWRWYHFDGVDWDESRKLSRIYKFRGIGKAWDWEVDTENGNVDYLM
SEQID No 14 (150) YSDFKWRWFHFNVDWDQRYQENHLFRFANTN--WNWRVDEENGNVDYLL
Consensus 9 (151) YS FKWRWFHF GVDWD IFKF W W VD ENGNVDYLL
201
250
SEQID No 1 (201) YADLMDHPEVVTELKNWGKWYVNTNIDGFRLDVAVKHIFSFPPDWLSY
SEQID No 14 (198) GSNIDFSHPEVQEELKDWGSWFTDELDDLGYRLDAIKHIPFWYTSDWVRH
Consensus 9 (201) A ID HPEV ELK WG WF IDGFRLDVAVKHIF F F DWL H
251
300
SEQID No 1 (251) VRSQTGKPLFTVGEYWSYDINKLHNYITKTNGTMSLFDAPLHNKFYTASK
SEQID No 14 (248) QRSEADQDLFVVGEYWKDDVGALEFYLDENMWEMSLFDVPLNYNFYRASK
Consensus 9 (251) RS LF VGEYW DI L YI N MSLFD PL FY ASK
301
350
SEQID No 1 (301) SGGAFDMRTLMTNTLMKDQPTLAVTFVDNHDTPEGQALQSWVDPWFKPLA
SEQID No 14 (298) QGGSYDMRNILRGLSVEAHPIHAVTFVDNHDTQPGESLESWVADWFKPLA
Consensus 9 (301) GGAFDMR IL SLM P AVTFVDNHDT PG AL SWV WFKPLA
351
400
SEQID No 1 (351) YAFILTRQEGYPCVFGDYYGIPQYNIPSLKSKIDPLLIARRDYAYGTQH
SEQID No 14 (348) YATILTREGGYPNVFYGDYYGIPNDNISAKKDMIDELLDARQNYAYGTQH
Consensus 9 (351) YA ILTR GYP VFYGDYYGIPN NI A K ID LL AR YAYGTQH
401
450
SEQID No 1 (401) DYLDHSDIIGWTREGVTEKPGSGLAALITDGPGGSKWMYVGKQHAGKVYF
SEQID No 14 (398) DYFDHWDIVGWTRREGTSSRPNSGLATIMSNGPGGSKWMYVGOQHAGQWT
Consensus 9 (401) DY DH DIIGWTREG S KP SGLA IIS GPGGSKWMYVG QHAG F
451
500
SEQID No 1 (451) DLTGNRSDTVTINSDGWGEFKNVGGSVSVVWVPRKTTVSTIARPIITRPWT
SEQID No 14 (448) DLTGNHAASVTINGDGWGEFFTNNGGSVSVYVNQ-----
Consensus 9 (451) DLTGN A SVTIN DGWGEF NNGGSVSVVW
501
515
SEQID No 1 (501) GEFVRWTEPRLVAVP
SEQID No 14 (481) -----
Consensus 9 (501)

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Figure 4H

		1		50
SEQID No 1	(1)	AAPFNGTMMQYFEWYLPDDGTLWTKVANEANNLSSLGITALWLPPAYKGT		
SEQID No 15	(1)	AAPFNGTMMQYFEWYLPDDGTLWTKVANEANNLSSLGITALWLPPAYKGT		
Consensus 10	(1)	AAPFNGTMMQYFEWYLPDDGTLWTKVANEANNLSSLGITALWLPPAYKGT		
		51		100
SEQID No 1	(51)	SRSDVGYGVYDLYDLGFEFNQKGTVRTKYGTAKAYLQAIQAAHAAGMQVYA		
SEQID No 15	(51)	SRSDVGYGVYDLYDLGFEFNQKGTVRTKYGTAKAYLQAIQAAHAAGMQVYA		
Consensus 10	(51)	SRSDVGYGVYDLYDLGFEFNQKGTVRTKYGTAKAYLQAIQAAHAAGMQVYA		
		101		150
SEQID No 1	(101)	DVVFDHKGGADGTEWVDAVEVNPSDRNQEISGTYQIQAWTKFDFPGRGNT		
SEQID No 15	(101)	DVVFDHKGGADGTEWVDAVEVNPSDRNQEISGTYQIQAWTKFDFPGRGNT		
Consensus 10	(101)	DVVFDHKGGADGTEWVDAVEVNPSDRNQEISGTYQIQAWTKFDFPGRGNT		
		151		200
SEQID No 1	(151)	YSSFKWRWYHFDGVDWDESRLKSRIYKFRGIGKAWDWEVDTEGNYDYLM		
SEQID No 15	(151)	YSSFKWRWYHFDGVDWDESRLKSRIYKFR--GKAWDWEVDTEFGNYDYLM		
Consensus 10	(151)	YSSFKWRWYHFDGVDWDESRLKSRIYKFR GKAWDWEVDTE GNYDYLM		
		201		250
SEQID No 1	(201)	YADLMDHPEVVTTELKNWGKQYVNTTNIDGFRLDAVKHIKFSFFPDWLSY		
SEQID No 15	(199)	YADLMDHPEVVTTELKNWGKQYVNTTNIDGFRLDAVKHIKFSFFPDWLSY		
Consensus 10	(201)	YADLMDHPEVVTTELKNWGKQYVNTTNIDGFRLDAVKHIKFSFFPDWLSY		
		251		300
SEQID No 1	(251)	VRSQTGKPLFTVGEYWSYDINKLHNYITKTNGTMSLFDAPLHNKFTYTASK		
SEQID No 15	(249)	VRSQTGKPLFTVGEYWSYDINKLHNYITKTNGTMSLFDAPLHNKFTYTASK		
Consensus 10	(251)	VRSQTGKPLFTVGEYWSYDINKLHNYITKTNGTMSLFDAPLHNKFTYTASK		
		301		350
SEQID No 1	(301)	SGGAFDMRTLMTNTLMKDQPTLAVTFVDNHDEPGQALQSWVDPWFKPLA		
SEQID No 15	(299)	SGGAFDMRTLMTNTLMKDQPTLAVTFVDNHDEPGQALQSWVDPWFKPLA		
Consensus 10	(301)	SGGAFDMRTLMTNTLMKDQPTLAVTFVDNHDEPGQALQSWVDPWFKPLA		
		351		400
SEQID No 1	(351)	YAFILTRQEGYPCVFGDYGGIPQYNI PSLKSKIDPLLIARRDYAYGTQH		
SEQID No 15	(349)	YAFILTRQEGYPCVFGDYGGIPQYNI PSLKSKIDPLLIARRDYAYGTQH		
Consensus 10	(351)	YAFILTRQEGYPCVFGDYGGIPQYNI PSLKSKIDPLLIARRDYAYGTQH		
		401		450
SEQID No 1	(401)	DYLDHSDIIGWTREGVTEKPGSGLAALITDGPGGSKWYVVGKQHAGKVFY		
SEQID No 15	(399)	DYLDHSDIIGWTREGTEKPGSGLAALITDGPGGSKWYVVGKQHAGKVFY		
Consensus 10	(401)	DYLDHSDIIGWTREG TEKPGSGLAALITDGPGGSKWYVVGKQHAGKVFY		
		451		500
SEQID No 1	(451)	DLTGNRSDTVTINSDGWGEFKVNGGSVSVVWVPRKTTVSTIARPIITRPWT		
SEQID No 15	(449)	DLTGNRSDTVTINSDGWGEFKVNGGSVSVVWVPRKTTVS-----		
Consensus 10	(451)	DLTGNRSDTVTINSDGWGEFKVNGGSVSVVWVPRKTTVS		
		501		515
SEQID No 1	(501)	GEFVRWTEPRLVAWP		
SEQID No 15	(487)	-----		
Consensus 10	(501)			

Figure 4I

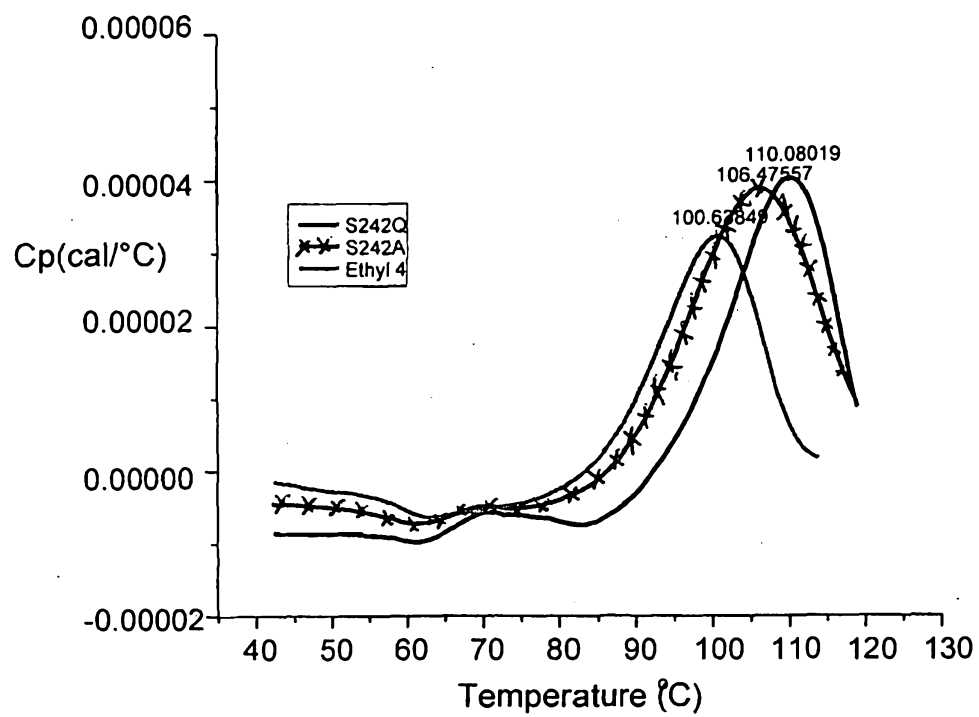


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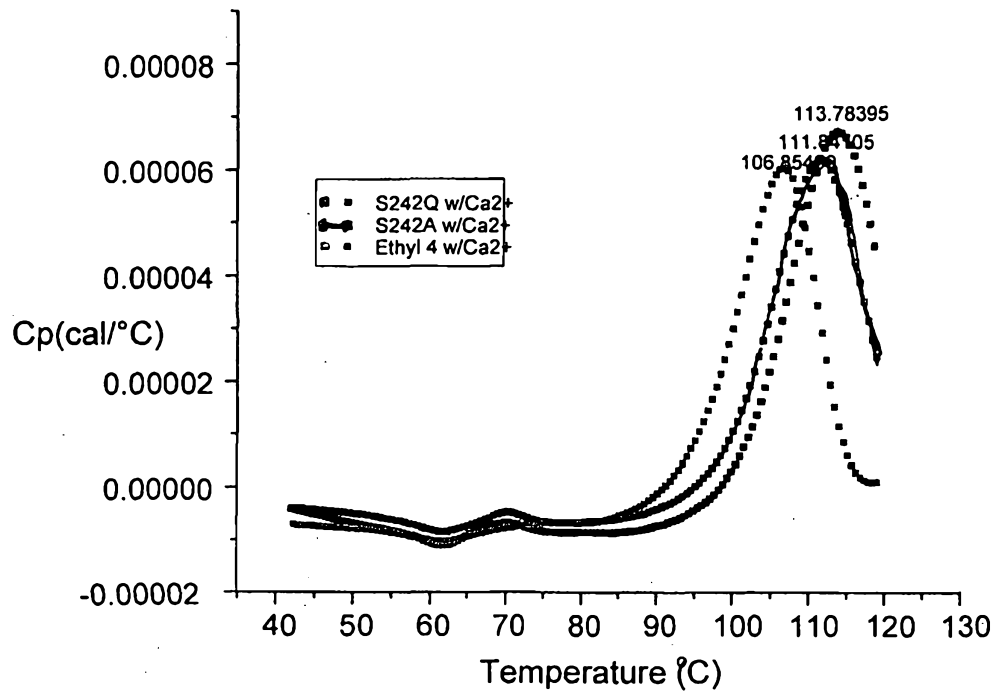


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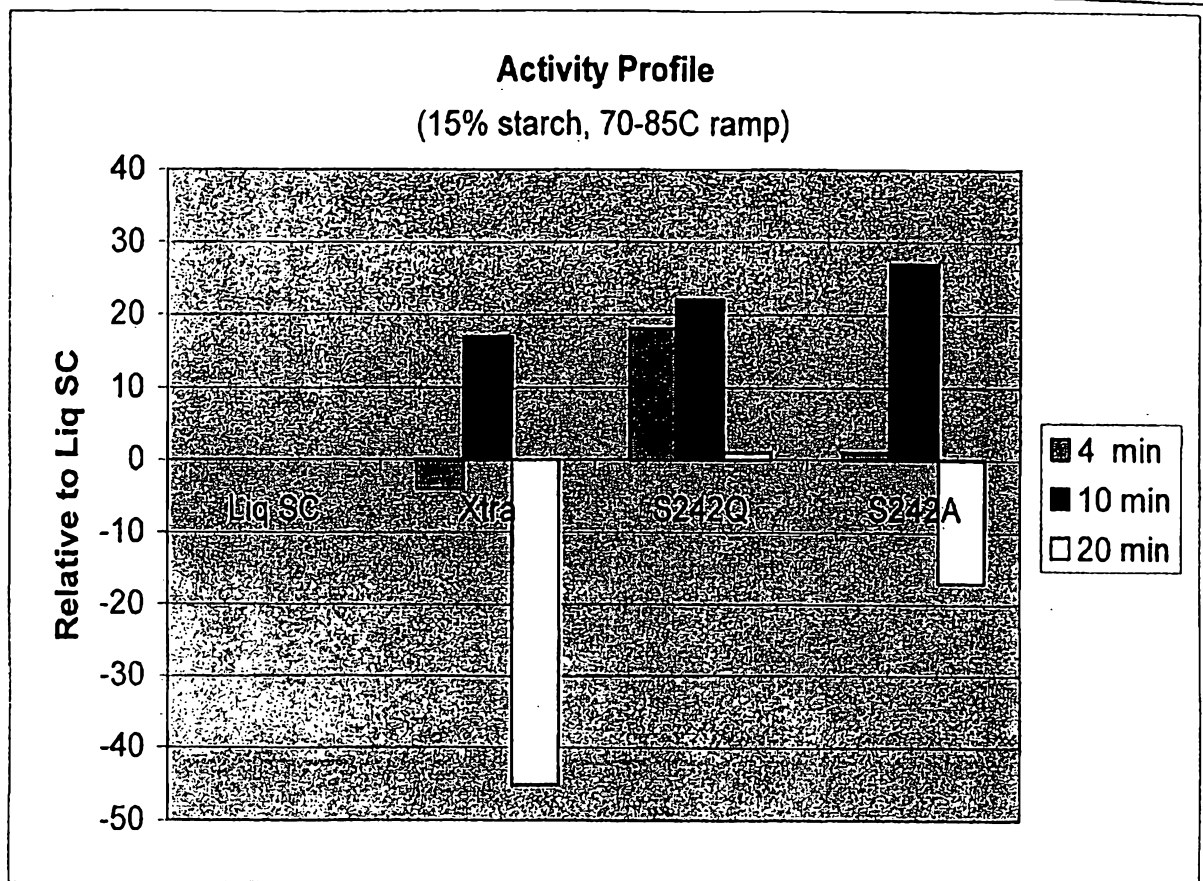


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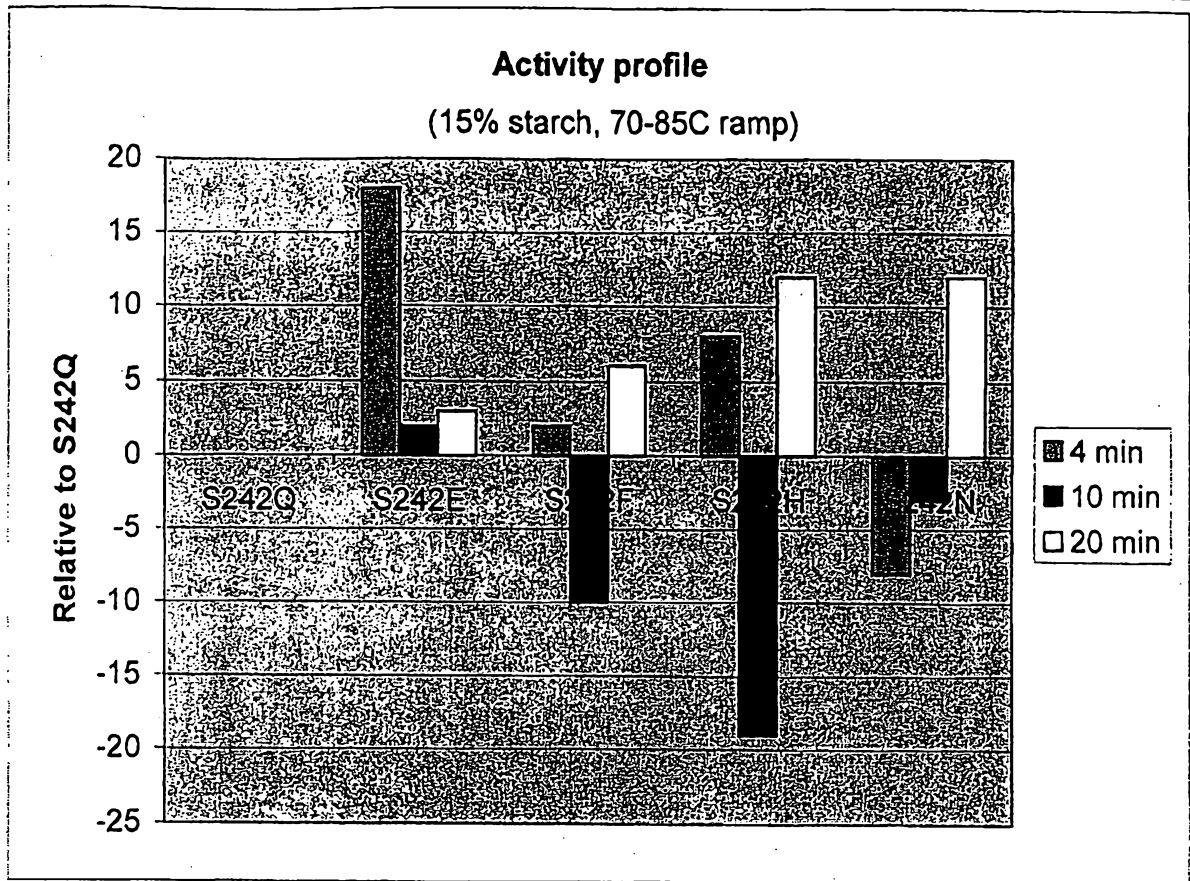


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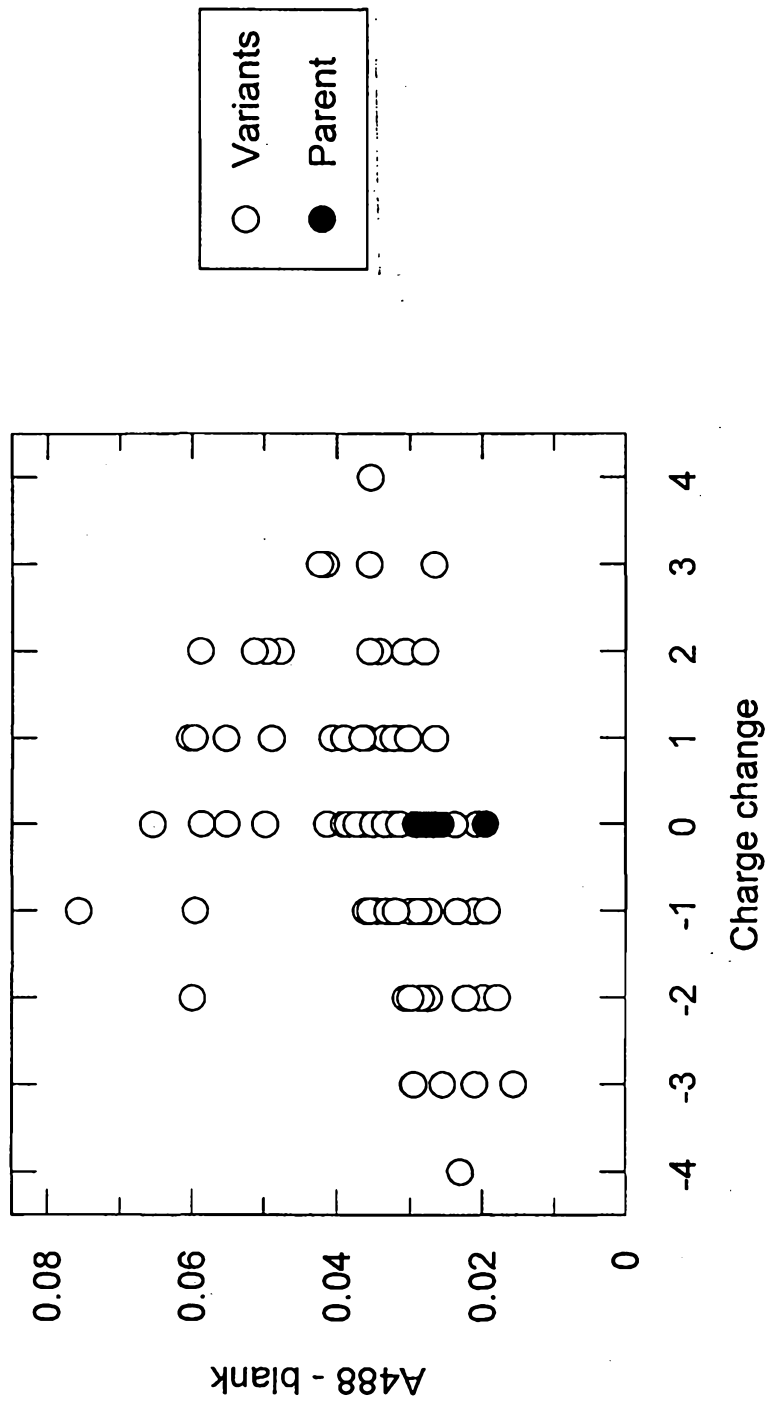


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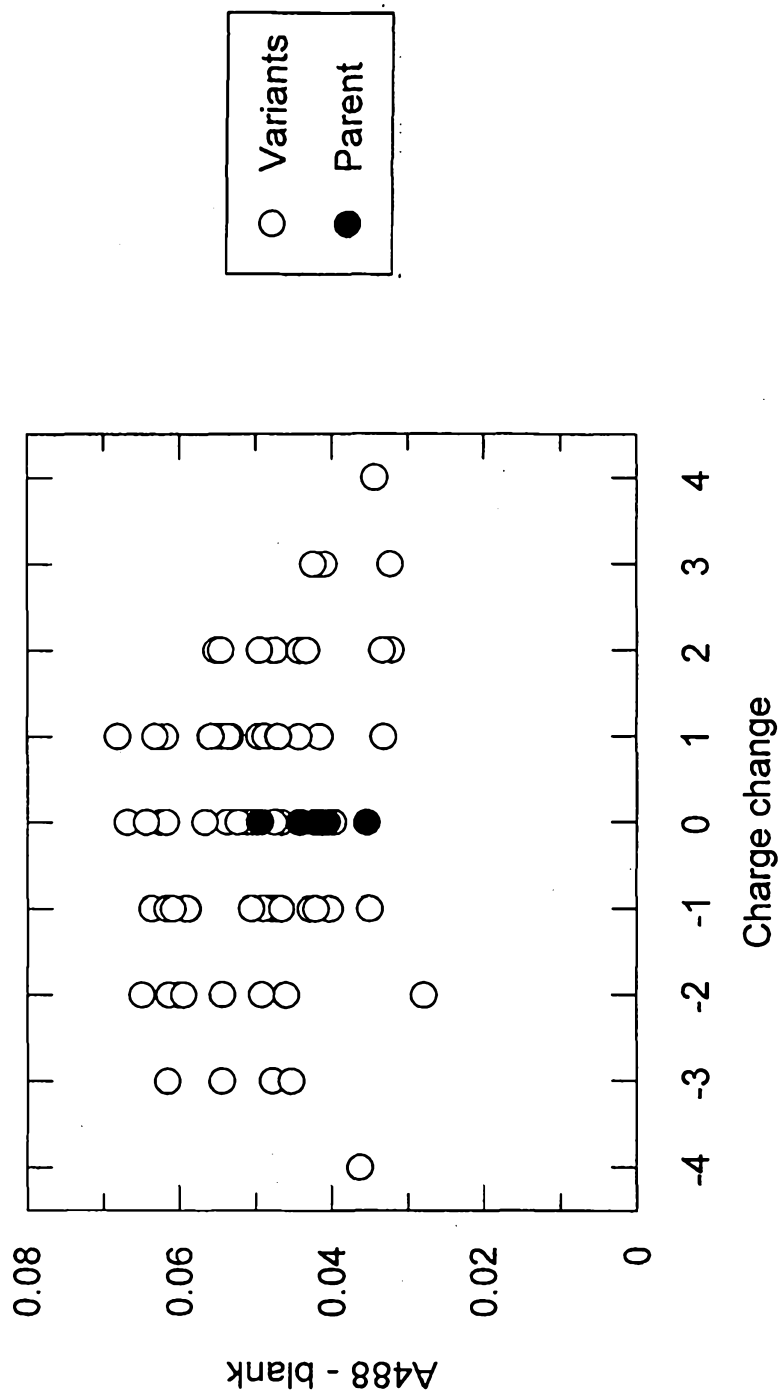


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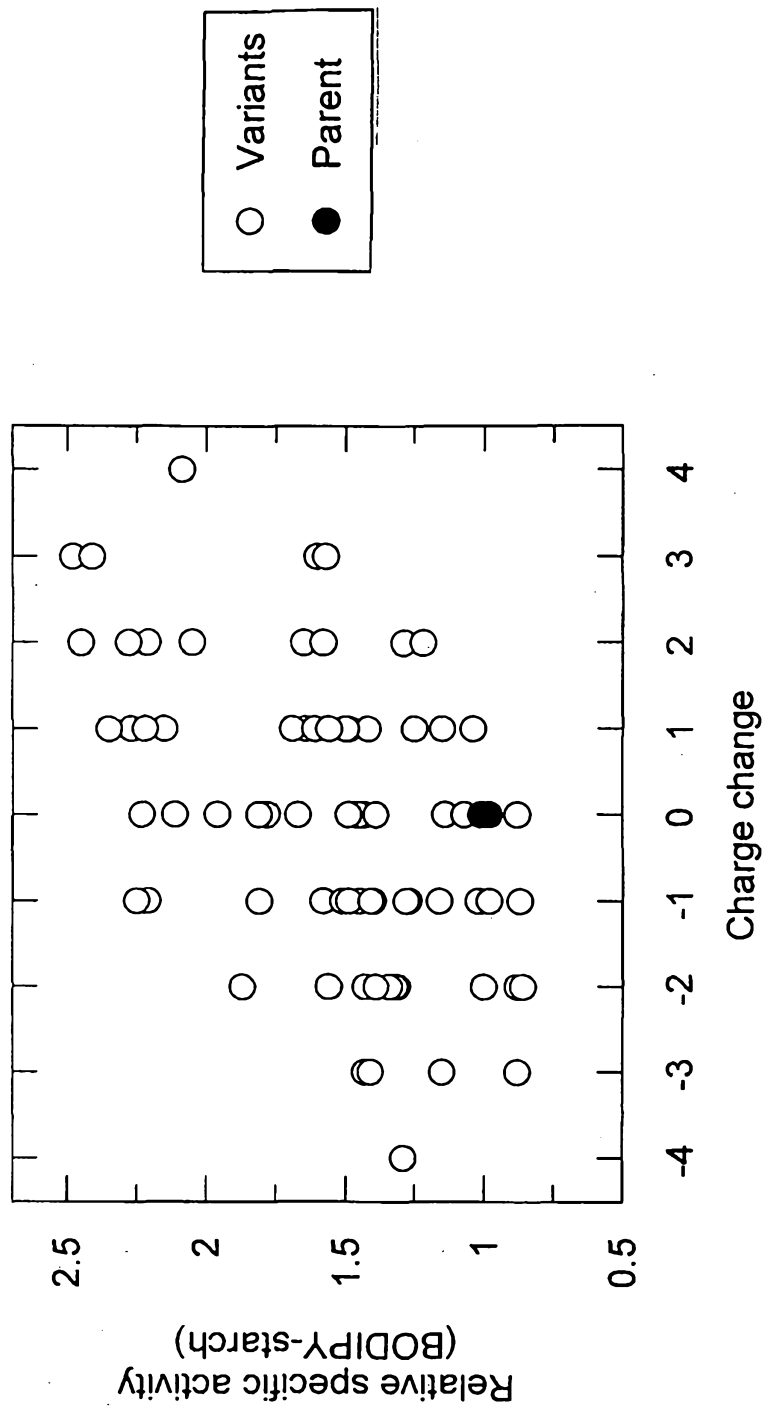


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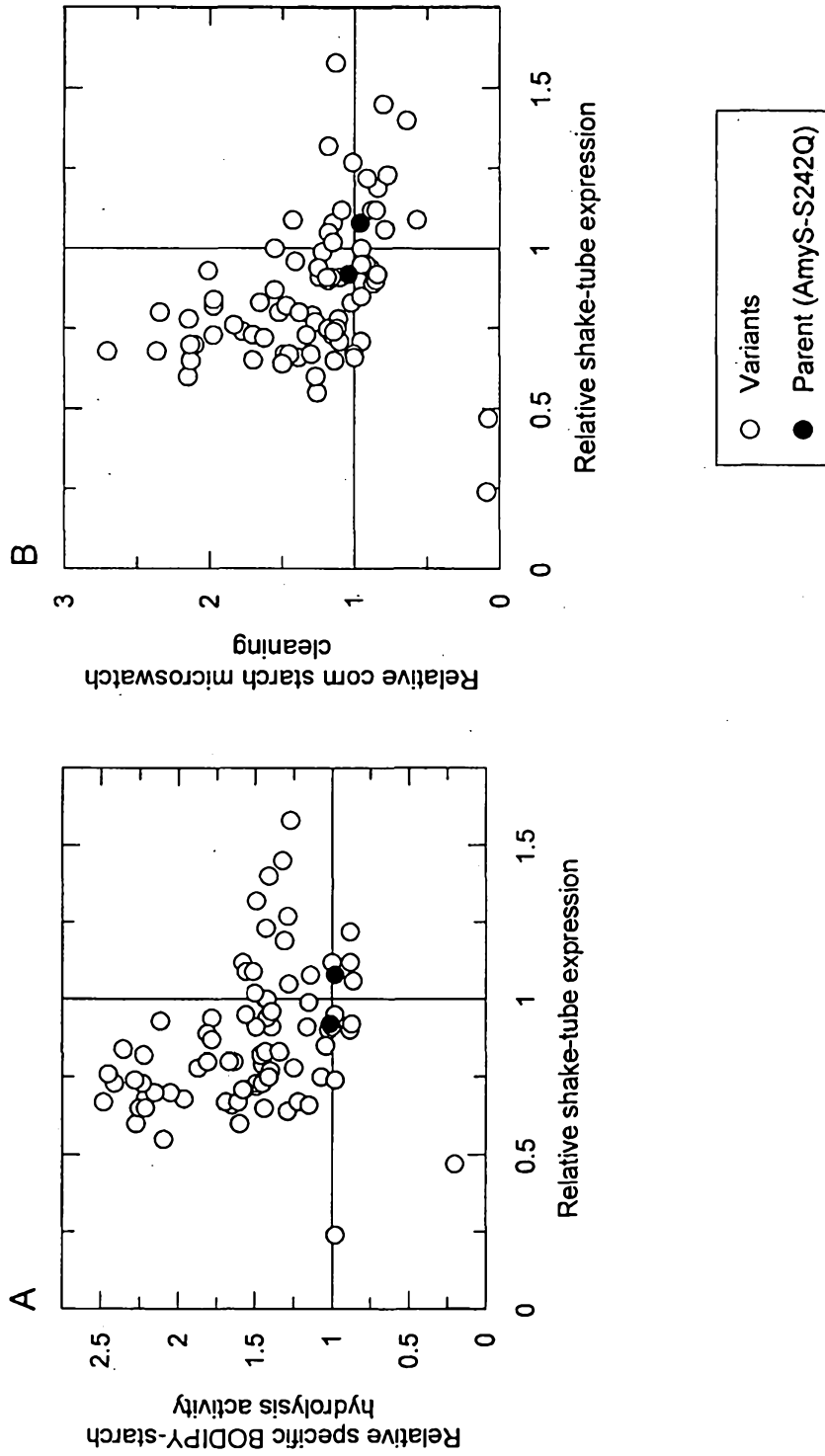


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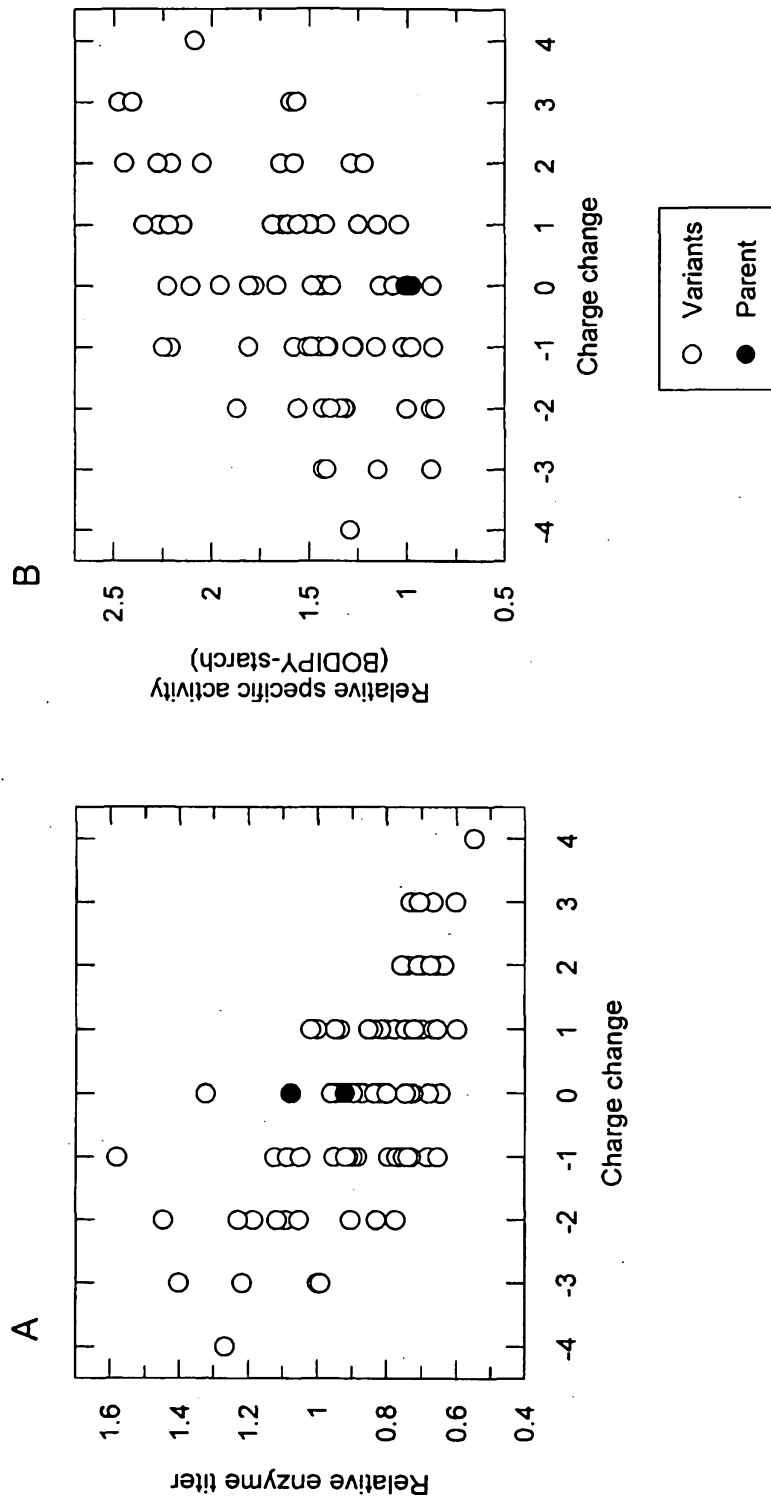


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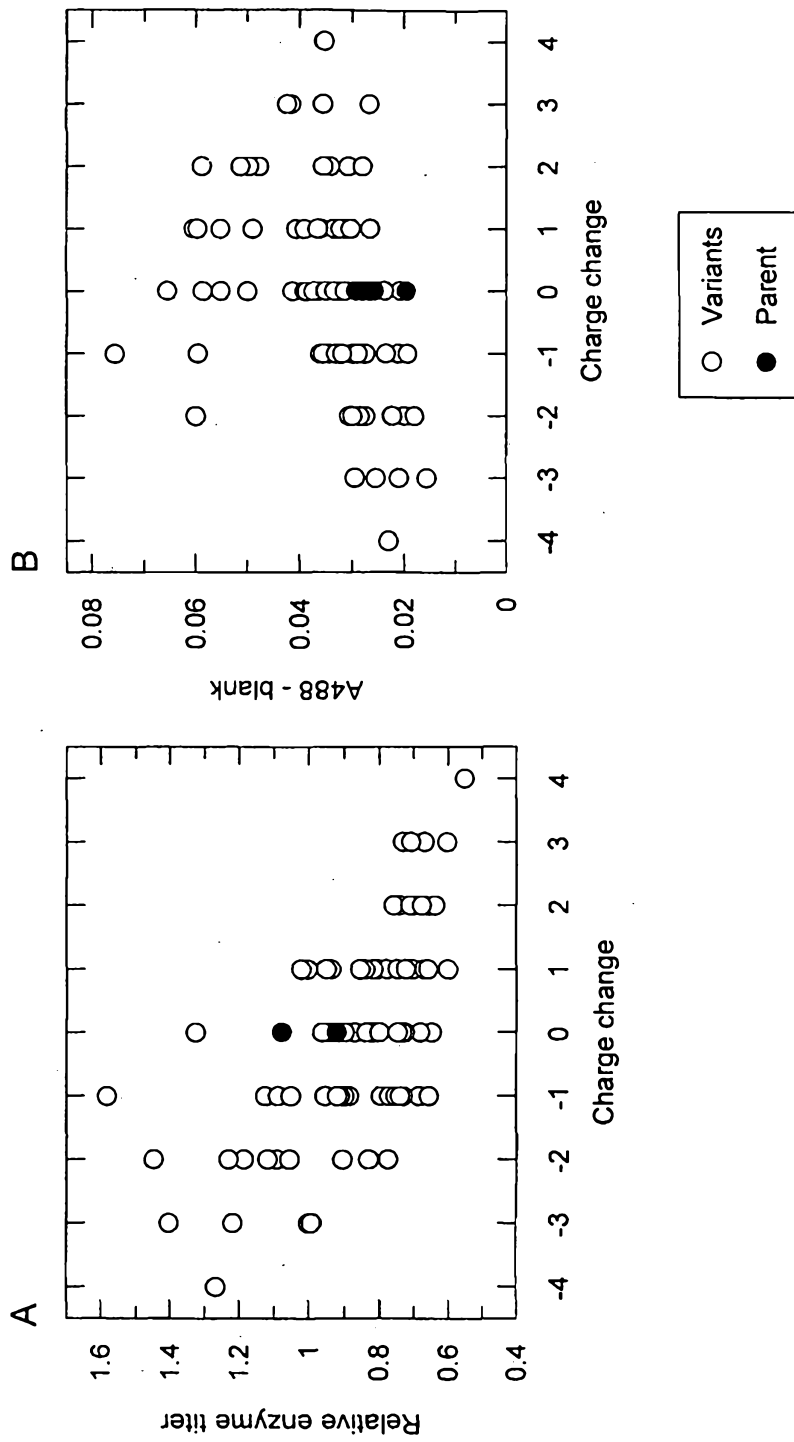


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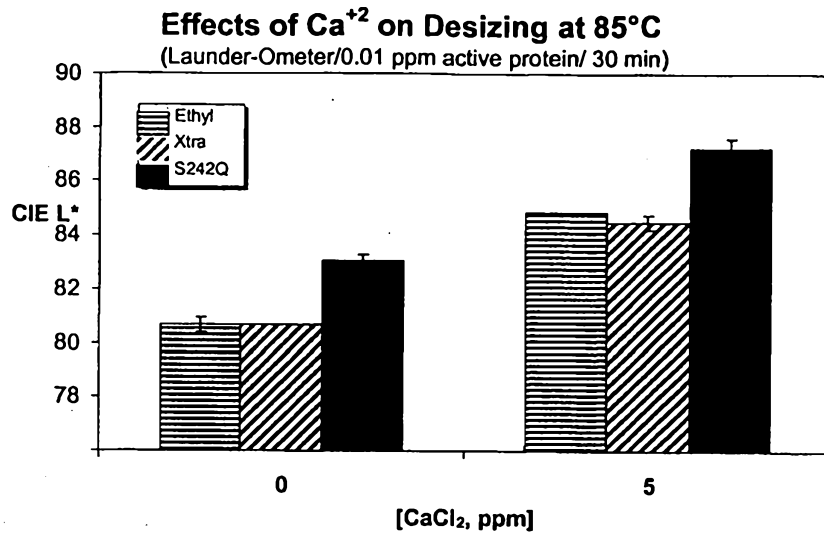


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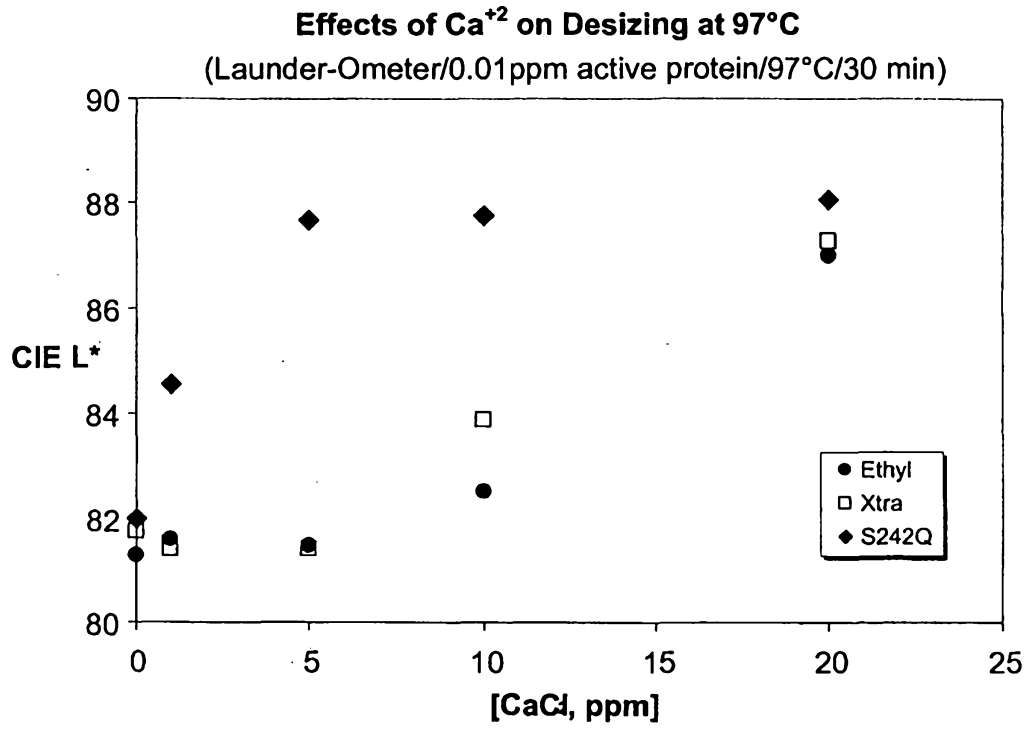


Figure 16

## SEQUENCE LISTING

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<151> 2008-06-06

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<150> 61/026,579

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Pro Glu Val Val Thr Glu Leu Lys Asn Trp Gly Lys Trp Tyr Val Asn  
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Leu His Asn Tyr Ile Thr Lys Thr Asn Gly Thr Met Ser Leu Phe Asp  
275 280 285

Ala Pro Leu His Asn Lys Phe Tyr Thr Ala Ser Lys Ser Gly Gly Ala  
290 295 300

Phe Asp Met Arg Thr Leu Met Thr Asn Thr Leu Met Lys Asp Gln Pro  
305 310 315 320

Thr Leu Ala Val Thr Phe Val Asp Asn His Asp Thr Glu Pro Gly Gln

3/251

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Ala Ala Pro Phe Asn Gly Thr Met Met Gln Tyr Phe Glu Trp Tyr Leu  
1 5 10 15

Pro Asp Asp Gly Thr Leu Trp Thr Lys Val Ala Asn Glu Ala Asn Asn  
20 25 30

Leu Ser Ser Leu Gly Ile Thr Ala Leu Trp Leu Pro Pro Ala Tyr Lys  
35 40 45

Gly Thr Ser Arg Ser Asp Val Gly Tyr Gly Val Tyr Asp Leu Tyr Asp  
50 55 60

Leu Gly Glu Phe Asn Gln Lys Gly Thr Val Arg Thr Lys Tyr Gly Thr  
65 70 75 80

Lys Ala Gln Tyr Leu Gln Ala Ile Gln Ala Ala His Ala Ala Gly Met  
85 90 95

Gln Val Tyr Ala Asp Val Val Phe Asp His Lys Gly Gly Ala Asp Gly  
100 105 110

Thr Glu Trp Val Asp Ala Val Glu Val Asn Pro Ser Asp Arg Asn Gln  
115 120 125

Glu Ile Ser Gly Thr Tyr Gln Ile Gln Ala Trp Thr Lys Phe Asp Phe  
130 135 140

Pro Gly Arg Gly Asn Thr Tyr Ser Ser Phe Lys Trp Arg Trp Tyr His  
145 150 155 160

Phe Asp Gly Val Asp Trp Asp Glu Ser Arg Lys Leu Ser Arg Ile Tyr  
165 170 175

Lys Phe Arg Gly Ile Gly Lys Ala Trp Asp Trp Glu Val Asp Thr Glu  
180 185 190

Asn Gly Asn Tyr Asp Tyr Leu Met Tyr Ala Asp Leu Asp Met Asp His  
195 200 205

Pro Glu Val Val Thr Glu Leu Lys Asn Trp Gly Lys Trp Tyr Val Asn  
210 215 220

Thr Thr Asn Ile Asp Gly Phe Arg Leu Asp Ala Val Lys His Ile Lys

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225					230					235					240
Phe	Ser	Phe	Phe	Pro	Asp	Trp	Leu	Ser	Tyr	Val	Arg	Ser	Gln	Thr	Gly
				245					250					255	
Lys	Pro	Leu	Phe	Thr	Val	Gly	Glu	Tyr	Trp	Ser	Tyr	Asp	Ile	Asn	Lys
			260					265					270		
Leu	His	Asn	Tyr	Ile	Thr	Lys	Thr	Asn	Gly	Thr	Met	Ser	Leu	Phe	Asp
		275					280					285			
Ala	Pro	Leu	His	Asn	Lys	Phe	Tyr	Thr	Ala	Ser	Lys	Ser	Gly	Gly	Ala
	290					295					300				
Phe	Asp	Met	Arg	Thr	Leu	Met	Thr	Asn	Thr	Leu	Met	Lys	Asp	Gln	Pro
305					310					315					320
Thr	Leu	Ala	Val	Thr	Phe	Val	Asp	Asn	His	Asp	Thr	Glu	Pro	Gly	Gln
				325					330					335	
Ala	Leu	Gln	Ser	Trp	Val	Asp	Pro	Trp	Phe	Lys	Pro	Leu	Ala	Tyr	Ala
			340					345					350		
Phe	Ile	Leu	Thr	Arg	Gln	Glu	Gly	Tyr	Pro	Cys	Val	Phe	Tyr	Gly	Asp
		355					360					365			
Tyr	Tyr	Gly	Ile	Pro	Gln	Tyr	Asn	Ile	Pro	Ser	Leu	Lys	Ser	Lys	Ile
	370					375					380				
Asp	Pro	Leu	Leu	Ile	Ala	Arg	Arg	Asp	Tyr	Ala	Tyr	Gly	Thr	Gln	His
385					390					395					400
Asp	Tyr	Leu	Asp	His	Ser	Asp	Ile	Ile	Gly	Trp	Thr	Arg	Glu	Gly	Val
				405					410					415	
Thr	Glu	Lys	Pro	Gly	Ser	Gly	Leu	Ala	Ala	Leu	Ile	Thr	Asp	Gly	Pro
			420					425					430		
Gly	Gly	Ser	Lys	Trp	Met	Tyr	Val	Gly	Lys	Gln	His	Ala	Gly	Lys	Val
		435					440					445			
Phe	Tyr	Asp	Leu	Thr	Gly	Asn	Arg	Ser	Asp	Thr	Val	Thr	Ile	Asn	Ser
	450					455					460				

Asp Gly Trp Gly Glu Phe Lys Val Asn Gly Gly Ser Val Ser Val Trp  
 465 470 475 480

Val Pro Arg Lys Thr Thr  
 485

<210> 3

<211> 515

<212> PRT

<213> Geobacillus stearothermophilus

<400> 3

Ala Ala Pro Phe Asn Gly Thr Met Met Gln Tyr Phe Glu Trp Tyr Leu  
 1 5 10 15

Pro Asp Asp Gly Thr Leu Trp Thr Lys Val Ala Asn Glu Ala Asn Asn  
 20 25 30

Leu Ser Ser Leu Gly Ile Thr Ala Leu Trp Leu Pro Pro Ala Tyr Lys  
 35 40 45

Gly Thr Ser Arg Ser Asp Val Gly Tyr Gly Val Tyr Asp Leu Tyr Asp  
 50 55 60

Leu Gly Glu Phe Asn Gln Lys Gly Thr Val Arg Thr Lys Tyr Gly Thr  
 65 70 75 80

Lys Ala Gln Tyr Leu Gln Ala Ile Gln Ala Ala His Ala Ala Gly Met  
 85 90 95

Gln Val Tyr Ala Asp Val Val Phe Asp His Lys Gly Gly Ala Asp Gly  
 100 105 110

Thr Glu Trp Val Asp Ala Val Glu Val Asn Pro Ser Asp Arg Asn Gln  
 115 120 125

Glu Ile Ser Gly Thr Tyr Gln Ile Gln Ala Trp Thr Lys Phe Asp Phe  
 130 135 140

Pro Gly Arg Gly Asn Thr Tyr Ser Ser Phe Lys Trp Arg Trp Tyr His  
 145 150 155 160

Phe Asp Gly Val Asp Trp Asp Glu Ser Arg Lys Leu Ser Arg Ile Tyr

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				165						170						175
Lys	Phe	Arg	Gly	Ile	Gly	Lys	Ala	Trp	Asp	Trp	Glu	Val	Asp	Thr	Glu	
			180					185					190			
Asn	Gly	Asn	Tyr	Asp	Tyr	Leu	Met	Tyr	Ala	Asp	Leu	Asp	Met	Asp	His	
		195					200					205				
Pro	Glu	Val	Val	Thr	Glu	Leu	Lys	Asn	Trp	Gly	Lys	Trp	Tyr	Val	Asn	
	210					215					220					
Thr	Thr	Asn	Ile	Asp	Gly	Phe	Arg	Leu	Asp	Ala	Val	Lys	His	Ile	Lys	
225					230					235					240	
Phe	Ala	Phe	Phe	Pro	Asp	Trp	Leu	Ser	Tyr	Val	Arg	Ser	Gln	Thr	Gly	
				245					250					255		
Lys	Pro	Leu	Phe	Thr	Val	Gly	Glu	Tyr	Trp	Ser	Tyr	Asp	Ile	Asn	Lys	
			260					265					270			
Leu	His	Asn	Tyr	Ile	Thr	Lys	Thr	Asn	Gly	Thr	Met	Ser	Leu	Phe	Asp	
		275					280					285				
Ala	Pro	Leu	His	Asn	Lys	Phe	Tyr	Thr	Ala	Ser	Lys	Ser	Gly	Gly	Ala	
	290					295					300					
Phe	Asp	Met	Arg	Thr	Leu	Met	Thr	Asn	Thr	Leu	Met	Lys	Asp	Gln	Pro	
305					310					315					320	
Thr	Leu	Ala	Val	Thr	Phe	Val	Asp	Asn	His	Asp	Thr	Glu	Pro	Gly	Gln	
				325					330					335		
Ala	Leu	Gln	Ser	Trp	Val	Asp	Pro	Trp	Phe	Lys	Pro	Leu	Ala	Tyr	Ala	
			340					345					350			
Phe	Ile	Leu	Thr	Arg	Gln	Glu	Gly	Tyr	Pro	Cys	Val	Phe	Tyr	Gly	Asp	
		355					360					365				
Tyr	Tyr	Gly	Ile	Pro	Gln	Tyr	Asn	Ile	Pro	Ser	Leu	Lys	Ser	Lys	Ile	
	370					375					380					
Asp	Pro	Leu	Leu	Ile	Ala	Arg	Arg	Asp	Tyr	Ala	Tyr	Gly	Thr	Gln	His	
385					390					395					400	



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Asp Tyr Leu Asp His Ser Asp Ile Ile Gly Trp Thr Arg Glu Gly Val  
405 410 415

Thr Glu Lys Pro Gly Ser Gly Leu Ala Ala Leu Ile Thr Asp Gly Pro  
420 425 430

Gly Gly Ser Lys Trp Met Tyr Val Gly Lys Gln His Ala Gly Lys Val  
435 440 445

Phe Tyr Asp Leu Thr Gly Asn Arg Ser Asp Thr Val Thr Ile Asn Ser  
450 455 460

Asp Gly Trp Gly Glu Phe Lys Val Asn Gly Gly Ser Val Ser Val Trp  
465 470 475 480

Val Pro Arg Lys Thr Thr Val Ser Thr Ile Ala Arg Pro Ile Thr Thr  
485 490 495

Arg Pro Trp Thr Gly Glu Phe Val Arg Trp Thr Glu Pro Arg Leu Val  
500 505 510

Ala Trp Pro  
515

<210> 4

<211> 515

<212> PRT

<213> Geobacillus stearothermophilus

<400> 4

Ala Ala Pro Phe Asn Gly Thr Met Met Gln Tyr Phe Glu Trp Tyr Leu  
1 5 10 15

Pro Asp Asp Gly Thr Leu Trp Thr Lys Val Ala Asn Glu Ala Asn Asn  
20 25 30

Leu Ser Ser Leu Gly Ile Thr Ala Leu Trp Leu Pro Pro Ala Tyr Lys  
35 40 45

Gly Thr Ser Arg Ser Asp Val Gly Tyr Gly Val Tyr Asp Leu Tyr Asp  
50 55 60

Leu Gly Glu Phe Asn Gln Lys Gly Thr Val Arg Thr Lys Tyr Gly Thr

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65					70					75					80
Lys	Ala	Gln	Tyr	Leu	Gln	Ala	Ile	Gln	Ala	Ala	His	Ala	Ala	Gly	Met
				85					90					95	
Gln	Val	Tyr	Ala	Asp	Val	Val	Phe	Asp	His	Lys	Gly	Gly	Ala	Asp	Gly
			100					105					110		
Thr	Glu	Trp	Val	Asp	Ala	Val	Glu	Val	Asn	Pro	Ser	Asp	Arg	Asn	Gln
		115					120					125			
Glu	Ile	Ser	Gly	Thr	Tyr	Gln	Ile	Gln	Ala	Trp	Thr	Lys	Phe	Asp	Phe
	130					135					140				
Pro	Gly	Arg	Gly	Asn	Thr	Tyr	Ser	Ser	Phe	Lys	Trp	Arg	Trp	Tyr	His
145					150					155					160
Phe	Asp	Gly	Val	Asp	Trp	Asp	Glu	Ser	Arg	Lys	Leu	Ser	Arg	Ile	Tyr
				165					170					175	
Lys	Phe	Arg	Gly	Ile	Gly	Lys	Ala	Trp	Asp	Trp	Glu	Val	Asp	Thr	Glu
			180					185					190		
Asn	Gly	Asn	Tyr	Asp	Tyr	Leu	Met	Tyr	Ala	Asp	Leu	Asp	Met	Asp	His
		195					200					205			
Pro	Glu	Val	Val	Thr	Glu	Leu	Lys	Asn	Trp	Gly	Lys	Trp	Tyr	Val	Asn
	210					215					220				
Thr	Thr	Asn	Ile	Asp	Gly	Phe	Arg	Leu	Asp	Ala	Val	Lys	His	Ile	Lys
225					230					235					240
Phe	Gln	Phe	Phe	Pro	Asp	Trp	Leu	Ser	Tyr	Val	Arg	Ser	Gln	Thr	Gly
				245					250					255	
Lys	Pro	Leu	Phe	Thr	Val	Gly	Glu	Tyr	Trp	Ser	Tyr	Asp	Ile	Asn	Lys
			260					265					270		
Leu	His	Asn	Tyr	Ile	Thr	Lys	Thr	Asn	Gly	Thr	Met	Ser	Leu	Phe	Asp
		275					280					285			
Ala	Pro	Leu	His	Asn	Lys	Phe	Tyr	Thr	Ala	Ser	Lys	Ser	Gly	Gly	Ala
	290					295					300				

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Phe Asp Met Arg Thr Leu Met Thr Asn Thr Leu Met Lys Asp Gln Pro  
305 310 315 320

Thr Leu Ala Val Thr Phe Val Asp Asn His Asp Thr Glu Pro Gly Gln  
325 330 335

Ala Leu Gln Ser Trp Val Asp Pro Trp Phe Lys Pro Leu Ala Tyr Ala  
340 345 350

Phe Ile Leu Thr Arg Gln Glu Gly Tyr Pro Cys Val Phe Tyr Gly Asp  
355 360 365

Tyr Tyr Gly Ile Pro Gln Tyr Asn Ile Pro Ser Leu Lys Ser Lys Ile  
370 375 380

Asp Pro Leu Leu Ile Ala Arg Arg Asp Tyr Ala Tyr Gly Thr Gln His  
385 390 395 400

Asp Tyr Leu Asp His Ser Asp Ile Ile Gly Trp Thr Arg Glu Gly Val  
405 410 415

Thr Glu Lys Pro Gly Ser Gly Leu Ala Ala Leu Ile Thr Asp Gly Pro  
420 425 430

Gly Gly Ser Lys Trp Met Tyr Val Gly Lys Gln His Ala Gly Lys Val  
435 440 445

Phe Tyr Asp Leu Thr Gly Asn Arg Ser Asp Thr Val Thr Ile Asn Ser  
450 455 460

Asp Gly Trp Gly Glu Phe Lys Val Asn Gly Gly Ser Val Ser Val Trp  
465 470 475 480

Val Pro Arg Lys Thr Thr Val Ser Thr Ile Ala Arg Pro Ile Thr Thr  
485 490 495

Arg Pro Trp Thr Gly Glu Phe Val Arg Trp Thr Glu Pro Arg Leu Val  
500 505 510

Ala Trp Pro  
515

11/251

<210> 5  
 <211> 515  
 <212> PRT  
 <213> Geobacillus stearothermophilus

<400> 5  
 Ala Ala Pro Phe Asn Gly Thr Met Met Gln Tyr Phe Glu Trp Tyr Leu  
 1 5 10 15

Pro Asp Asp Gly Thr Leu Trp Thr Lys Val Ala Asn Glu Ala Asn Asn  
 20 25 30

Leu Ser Ser Leu Gly Ile Thr Ala Leu Trp Leu Pro Pro Ala Tyr Lys  
 35 40 45

Gly Thr Ser Arg Ser Asp Val Gly Tyr Gly Val Tyr Asp Leu Tyr Asp  
 50 55 60

Leu Gly Glu Phe Asn Gln Lys Gly Thr Val Arg Thr Lys Tyr Gly Thr  
 65 70 75 80

Lys Ala Gln Tyr Leu Gln Ala Ile Gln Ala Ala His Ala Ala Gly Met  
 85 90 95

Gln Val Tyr Ala Asp Val Val Phe Asp His Lys Gly Gly Ala Asp Gly  
 100 105 110

Thr Glu Trp Val Asp Ala Val Glu Val Asn Pro Ser Asp Arg Asn Gln  
 115 120 125

Glu Ile Ser Gly Thr Tyr Gln Ile Gln Ala Trp Thr Lys Phe Asp Phe  
 130 135 140

Pro Gly Arg Gly Asn Thr Tyr Ser Ser Phe Lys Trp Arg Trp Tyr His  
 145 150 155 160

Phe Asp Gly Val Asp Trp Asp Glu Ser Arg Lys Leu Ser Arg Ile Tyr  
 165 170 175

Lys Phe Arg Gly Ile Gly Lys Ala Trp Asp Trp Glu Val Asp Thr Glu  
 180 185 190

Asn Gly Asn Tyr Asp Tyr Leu Met Tyr Ala Asp Leu Asp Met Asp His  
 195 200 205

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Pro Glu Val Val Thr Glu Leu Lys Asn Trp Gly Lys Trp Tyr Val Asn  
210 215 220

Thr Thr Asn Ile Asp Gly Phe Arg Leu Asp Ala Val Lys His Ile Lys  
225 230 235 240

Phe Glu Phe Phe Pro Asp Trp Leu Ser Tyr Val Arg Ser Gln Thr Gly  
245 250 255

Lys Pro Leu Phe Thr Val Gly Glu Tyr Trp Ser Tyr Asp Ile Asn Lys  
260 265 270

Leu His Asn Tyr Ile Thr Lys Thr Asn Gly Thr Met Ser Leu Phe Asp  
275 280 285

Ala Pro Leu His Asn Lys Phe Tyr Thr Ala Ser Lys Ser Gly Gly Ala  
290 295 300

Phe Asp Met Arg Thr Leu Met Thr Asn Thr Leu Met Lys Asp Gln Pro  
305 310 315 320

Thr Leu Ala Val Thr Phe Val Asp Asn His Asp Thr Glu Pro Gly Gln  
325 330 335

Ala Leu Gln Ser Trp Val Asp Pro Trp Phe Lys Pro Leu Ala Tyr Ala  
340 345 350

Phe Ile Leu Thr Arg Gln Glu Gly Tyr Pro Cys Val Phe Tyr Gly Asp  
355 360 365

Tyr Tyr Gly Ile Pro Gln Tyr Asn Ile Pro Ser Leu Lys Ser Lys Ile  
370 375 380

Asp Pro Leu Leu Ile Ala Arg Arg Asp Tyr Ala Tyr Gly Thr Gln His  
385 390 395 400

Asp Tyr Leu Asp His Ser Asp Ile Ile Gly Trp Thr Arg Glu Gly Val  
405 410 415

Thr Glu Lys Pro Gly Ser Gly Leu Ala Ala Leu Ile Thr Asp Gly Pro  
420 425 430

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Gly Gly Ser Lys Trp Met Tyr Val Gly Lys Gln His Ala Gly Lys Val  
435 440 445

Phe Tyr Asp Leu Thr Gly Asn Arg Ser Asp Thr Val Thr Ile Asn Ser  
450 455 460

Asp Gly Trp Gly Glu Phe Lys Val Asn Gly Gly Ser Val Ser Val Trp  
465 470 475 480

Val Pro Arg Lys Thr Thr Val Ser Thr Ile Ala Arg Pro Ile Thr Thr  
485 490 495

Arg Pro Trp Thr Gly Glu Phe Val Arg Trp Thr Glu Pro Arg Leu Val  
500 505 510

Ala Trp Pro  
515

<210> 6

<211> 485

<212> PRT

<213> Bacillus sp. 707

<400> 6

His His Asn Gly Thr Asn Gly Thr Met Met Gln Tyr Phe Glu Trp Tyr  
1 5 10 15

Leu Pro Asn Asp Gly Asn His Trp Asn Arg Leu Asn Ser Asp Ala Ser  
20 25 30

Asn Leu Lys Ser Lys Gly Ile Thr Ala Val Trp Ile Pro Pro Ala Trp  
35 40 45

Lys Gly Ala Ser Gln Asn Asp Val Gly Tyr Gly Ala Tyr Asp Leu Tyr  
50 55 60

Asp Leu Gly Glu Phe Asn Gln Lys Gly Thr Val Arg Thr Lys Tyr Gly  
65 70 75 80

Thr Arg Ser Gln Leu Gln Ala Ala Val Thr Ser Leu Lys Asn Asn Gly  
85 90 95

Ile Gln Val Tyr Gly Asp Val Val Met Asn His Lys Gly Gly Ala Asp  
100 105 110

Ala Thr Glu Met Val Arg Ala Val Glu Val Asn Pro Asn Asn Arg Asn  
115 120 125

Gln Glu Val Thr Gly Glu Tyr Thr Ile Glu Ala Trp Thr Arg Phe Asp  
130 135 140

Phe Pro Gly Arg Gly Asn Thr His Ser Ser Phe Lys Trp Arg Trp Tyr  
145 150 155 160

His Phe Asp Gly Val Asp Trp Asp Gln Ser Arg Arg Leu Asn Asn Arg  
165 170 175

Ile Tyr Lys Phe Arg Gly His Gly Lys Ala Trp Asp Trp Glu Val Asp  
180 185 190

Thr Glu Asn Gly Asn Tyr Asp Tyr Leu Met Tyr Ala Asp Ile Asp Met  
195 200 205

Asp His Pro Glu Val Val Asn Glu Leu Arg Asn Trp Gly Val Trp Tyr  
210 215 220

Thr Asn Thr Leu Gly Leu Asp Gly Phe Arg Ile Asp Ala Val Lys His  
225 230 235 240

Ile Lys Tyr Ser Phe Thr Arg Asp Trp Ile Asn His Val Arg Ser Ala  
245 250 255

Thr Gly Lys Asn Met Phe Ala Val Ala Glu Phe Trp Lys Asn Asp Leu  
260 265 270

Gly Ala Ile Glu Asn Tyr Leu Gln Lys Thr Asn Trp Asn His Ser Val  
275 280 285

Phe Asp Val Pro Leu His Tyr Asn Leu Tyr Asn Ala Ser Lys Ser Gly  
290 295 300

Gly Asn Tyr Asp Met Arg Asn Ile Phe Asn Gly Thr Val Val Gln Arg  
305 310 315 320

His Pro Ser His Ala Val Thr Phe Val Asp Asn His Asp Ser Gln Pro  
325 330 335

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Glu Glu Ala Leu Glu Ser Phe Val Glu Glu Trp Phe Lys Pro Leu Ala  
340 345 350

Tyr Ala Leu Thr Leu Thr Arg Glu Gln Gly Tyr Pro Ser Val Phe Tyr  
355 360 365

Gly Asp Tyr Tyr Gly Ile Pro Thr His Gly Val Pro Ala Met Arg Ser  
370 375 380

Lys Ile Asp Pro Ile Leu Glu Ala Arg Gln Lys Tyr Ala Tyr Gly Lys  
385 390 395 400

Gln Asn Asp Tyr Leu Asp His His Asn Ile Ile Gly Trp Thr Arg Glu  
405 410 415

Gly Asn Thr Ala His Pro Asn Ser Gly Leu Ala Thr Ile Met Ser Asp  
420 425 430

Gly Ala Gly Gly Ser Lys Trp Met Phe Val Gly Arg Asn Lys Ala Gly  
435 440 445

Gln Val Trp Ser Asp Ile Thr Gly Asn Arg Thr Gly Thr Val Thr Ile  
450 455 460

Asn Ala Asp Gly Trp Gly Asn Phe Ser Val Asn Gly Gly Ser Val Ser  
465 470 475 480

Ile Trp Val Asn Lys  
485

<210> 7

<211> 483

<212> PRT

<213> Bacillus sp. 707

<400> 7

Ala Asn Leu Asn Gly Thr Leu Met Gln Tyr Phe Glu Trp Tyr Met Pro  
1 5 10 15

Asn Asp Gly Gln His Trp Lys Arg Leu Gln Asn Asp Ser Ala Tyr Leu  
20 25 30

Ala Glu His Gly Ile Thr Ala Val Trp Ile Pro Pro Ala Tyr Lys Gly  
35 40 45



Thr Ser Gln Ala Asp Val Gly Tyr Gly Ala Tyr Asp Leu Tyr Asp Leu  
 50 55 60

Gly Glu Phe His Gln Lys Gly Thr Val Arg Thr Lys Tyr Gly Thr Lys  
 65 70 75 80

Gly Glu Leu Gln Ser Ala Ile Lys Ser Leu His Ser Arg Asp Ile Asn  
 85 90 95

Val Tyr Gly Asp Val Val Ile Asn His Lys Gly Gly Ala Asp Ala Thr  
 100 105 110

Glu Asp Val Thr Ala Val Glu Val Asp Pro Ala Asp Arg Asn Arg Val  
 115 120 125

Ile Ser Gly Glu His Leu Ile Lys Ala Trp Thr His Phe His Phe Pro  
 130 135 140

Gly Arg Gly Ser Thr Tyr Ser Asp Phe Lys Trp His Trp Tyr His Phe  
 145 150 155 160

Asp Gly Thr Asp Trp Asp Glu Ser Arg Lys Leu Asn Arg Ile Tyr Lys  
 165 170 175

Phe Gln Gly Lys Ala Trp Asp Trp Glu Val Ser Asn Glu Asn Gly Asn  
 180 185 190

Tyr Asp Tyr Leu Met Tyr Ala Asp Ile Asp Tyr Asp His Pro Asp Val  
 195 200 205

Ala Ala Glu Ile Lys Arg Trp Gly Thr Trp Tyr Ala Asn Glu Leu Gln  
 210 215 220

Leu Asp Gly Phe Arg Leu Asp Ala Val Lys His Ile Lys Phe Ser Phe  
 225 230 235 240

Leu Arg Asp Trp Val Asn His Val Arg Glu Lys Thr Gly Lys Glu Met  
 245 250 255

Phe Thr Val Ala Glu Tyr Trp Gln Asn Asp Leu Gly Ala Leu Glu Asn  
 260 265 270

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Tyr Leu Asn Lys Thr Asn Phe Asn His Ser Val Phe Asp Val Pro Leu  
275 280 285

His Tyr Gln Phe His Ala Ala Ser Thr Gln Gly Gly Tyr Asp Met  
290 295 300

Arg Lys Leu Leu Asn Gly Thr Val Val Ser Lys His Pro Leu Lys Ser  
305 310 315 320

Val Thr Phe Val Asp Asn His Asp Thr Gln Pro Gly Gln Ser Leu Glu  
325 330 335

Ser Thr Val Gln Thr Trp Phe Lys Pro Leu Ala Tyr Ala Phe Ile Leu  
340 345 350

Thr Arg Glu Ser Gly Tyr Pro Gln Val Phe Tyr Gly Asp Met Tyr Gly  
355 360 365

Thr Lys Gly Asp Ser Gln Arg Glu Ile Pro Ala Leu Lys His Lys Ile  
370 375 380

Glu Pro Ile Leu Lys Ala Arg Lys Gln Tyr Ala Tyr Gly Ala Gln His  
385 390 395 400

Asp Tyr Phe Asp His His Asp Ile Val Gly Trp Thr Arg Glu Gly Asp  
405 410 415

Ser Ser Val Ala Asn Ser Gly Leu Ala Ala Leu Ile Thr Asp Gly Pro  
420 425 430

Gly Gly Ala Lys Arg Met Tyr Val Gly Arg Gln Asn Ala Gly Glu Thr  
435 440 445

Trp His Asp Ile Thr Gly Asn Arg Ser Glu Pro Val Val Ile Asn Ser  
450 455 460

Glu Gly Trp Gly Glu Phe His Val Asn Gly Gly Ser Val Ser Ile Tyr  
465 470 475 480

Val Gln Arg

<210> 8  
 <211> 483  
 <212> PRT  
 <213> Bacillus licheniformis

<400> 8  
 Ala Asn Leu Asn Gly Thr Leu Met Gln Tyr Phe Glu Trp Tyr Met Pro  
 1 5 10 15

Asn Asp Gly Gln His Trp Arg Arg Leu Gln Asn Asp Ser Ala Tyr Leu  
 20 25 30

Ala Glu His Gly Ile Thr Ala Val Trp Ile Pro Pro Ala Tyr Lys Gly  
 35 40 45

Thr Ser Gln Ala Asp Val Gly Tyr Gly Ala Tyr Asp Leu Tyr Asp Leu  
 50 55 60

Gly Glu Phe His Gln Lys Gly Thr Val Arg Thr Lys Tyr Gly Thr Lys  
 65 70 75 80

Gly Glu Leu Gln Ser Ala Ile Lys Ser Leu His Ser Arg Asp Ile Asn  
 85 90 95

Val Tyr Gly Asp Val Val Ile Asn His Lys Gly Gly Ala Asp Ala Thr  
 100 105 110

Glu Asp Val Thr Ala Val Glu Val Asp Pro Ala Asp Arg Asn Arg Val  
 115 120 125

Ile Ser Gly Glu His Leu Ile Lys Ala Trp Thr His Phe His Phe Pro  
 130 135 140

Gly Arg Gly Ser Thr Tyr Ser Asp Phe Lys Trp His Trp Tyr His Phe  
 145 150 155 160

Asp Gly Thr Asp Trp Asp Glu Ser Arg Lys Leu Asn Arg Ile Tyr Lys  
 165 170 175

Phe Gln Gly Lys Ala Trp Asp Trp Glu Val Ser Asn Glu Asn Gly Asn  
 180 185 190

Tyr Asp Tyr Leu Met Tyr Ala Asp Ile Asp Tyr Asp His Pro Asp Val  
 195 200 205

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Ala Ala Glu Ile Lys Arg Trp Gly Thr Trp Tyr Ala Asn Glu Leu Gln  
210 215 220

Leu Asp Gly Phe Arg Leu Asp Ala Val Lys His Ile Lys Phe Ser Phe  
225 230 235 240

Leu Arg Asp Trp Val Asn His Val Arg Glu Lys Thr Gly Lys Glu Met  
245 250 255

Phe Thr Val Ala Glu Tyr Trp Gln Asn Asp Leu Gly Ala Leu Glu Asn  
260 265 270

Tyr Leu Asn Lys Thr Asn Phe Asn His Ser Val Phe Asp Val Pro Leu  
275 280 285

His Tyr Gln Phe His Ala Ala Ser Thr Gln Gly Gly Gly Tyr Asp Met  
290 295 300

Arg Lys Leu Leu Asn Gly Thr Val Val Ser Lys His Pro Leu Lys Ser  
305 310 315 320

Val Thr Phe Val Asp Asn His Asp Thr Gln Pro Gly Gln Ser Leu Glu  
325 330 335

Ser Thr Val Gln Thr Trp Phe Lys Pro Leu Ala Tyr Ala Phe Ile Leu  
340 345 350

Thr Arg Glu Ser Gly Tyr Pro Gln Val Phe Tyr Gly Asp Met Tyr Gly  
355 360 365

Thr Lys Gly Asp Ser Gln Arg Glu Ile Pro Ala Leu Lys His Lys Ile  
370 375 380

Glu Pro Ile Leu Lys Ala Arg Lys Gln Tyr Ala Tyr Gly Ala Gln His  
385 390 395 400

Asp Tyr Phe Asp His His Asp Ile Val Gly Trp Thr Arg Glu Gly Asp  
405 410 415

Ser Ser Val Ala Asn Ser Gly Leu Ala Ala Leu Ile Thr Asp Gly Pro  
420 425 430

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Gly Gly Ala Lys Arg Met Tyr Val Gly Arg Gln Asn Ala Gly Glu Thr  
435 440 445

Trp His Asp Ile Thr Gly Asn Arg Ser Glu Pro Val Val Ile Asn Ser  
450 455 460

Glu Gly Trp Gly Glu Phe His Val Asn Gly Gly Ser Val Ser Ile Tyr  
465 470 475 480

Val Gln Arg

<210> 9

<211> 483

<212> PRT

<213> Bacillus amyloliquefaciens

<400> 9

Val Asn Gly Thr Leu Met Gln Tyr Phe Glu Trp Tyr Thr Pro Asn Asp  
1 5 10 15

Gly Gln His Trp Lys Arg Leu Gln Asn Asp Ala Glu His Leu Ser Asp  
20 25 30

Ile Gly Ile Thr Ala Val Trp Ile Pro Pro Ala Tyr Lys Gly Leu Ser  
35 40 45

Gln Ser Asp Asn Gly Tyr Gly Pro Tyr Asp Leu Tyr Asp Leu Gly Glu  
50 55 60

Phe Gln Gln Lys Gly Thr Val Arg Thr Lys Tyr Gly Thr Lys Ser Glu  
65 70 75 80

Leu Gln Asp Ala Ile Gly Ser Leu His Ser Arg Asn Val Gln Val Tyr  
85 90 95

Gly Asp Val Val Leu Asn His Lys Ala Gly Ala Asp Ala Thr Glu Asp  
100 105 110

Val Thr Ala Val Glu Val Asn Pro Ala Asn Arg Asn Gln Glu Thr Ser  
115 120 125

Glu Glu Tyr Gln Ile Lys Ala Trp Thr Asp Phe Arg Phe Pro Gly Arg  
130 135 140

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Gly Asn Thr Tyr Ser Asp Phe Lys Trp His Trp Tyr His Phe Asp Gly  
145 150 155 160

Ala Asp Trp Asp Glu Ser Arg Lys Ile Ser Arg Ile Phe Lys Phe Arg  
165 170 175

Gly Glu Gly Lys Ala Trp Asp Trp Glu Val Ser Ser Glu Asn Gly Asn  
180 185 190

Tyr Asp Tyr Leu Met Tyr Ala Asp Val Asp Tyr Asp His Pro Asp Val  
195 200 205

Val Ala Glu Thr Lys Lys Trp Gly Ile Trp Tyr Ala Asn Glu Leu Ser  
210 215 220

Leu Asp Gly Phe Arg Ile Asp Ala Ala Lys His Ile Lys Phe Ser Phe  
225 230 235 240

Leu Arg Asp Trp Val Gln Ala Val Arg Gln Ala Thr Gly Lys Glu Met  
245 250 255

Phe Thr Val Ala Glu Tyr Trp Gln Asn Asn Ala Gly Lys Leu Glu Asn  
260 265 270

Tyr Leu Asn Lys Thr Ser Phe Asn Gln Ser Val Phe Asp Val Pro Leu  
275 280 285

His Phe Asn Leu Gln Ala Ala Ser Ser Gln Gly Gly Gly Tyr Asp Met  
290 295 300

Arg Arg Leu Leu Asp Gly Thr Val Val Ser Arg His Pro Glu Lys Ala  
305 310 315 320

Val Thr Phe Val Glu Asn His Asp Thr Gln Pro Gly Gln Ser Leu Glu  
325 330 335

Ser Thr Val Gln Thr Trp Phe Lys Pro Leu Ala Tyr Ala Phe Ile Leu  
340 345 350

Thr Arg Glu Ser Gly Tyr Pro Gln Val Phe Tyr Gly Asp Met Tyr Gly  
355 360 365

Thr Lys Gly Thr Ser Pro Lys Glu Ile Pro Ser Leu Lys Asp Asn Ile  
 370 375 380

Glu Pro Ile Leu Lys Ala Arg Lys Glu Tyr Ala Tyr Gly Pro Gln His  
 385 390 395 400

Asp Tyr Ile Asp His Pro Asp Val Ile Gly Trp Thr Arg Glu Gly Asp  
 405 410 415

Ser Ser Ala Ala Lys Ser Gly Leu Ala Ala Leu Ile Thr Asp Gly Pro  
 420 425 430

Gly Gly Ser Lys Arg Met Tyr Ala Gly Leu Lys Asn Ala Gly Glu Thr  
 435 440 445

Trp Tyr Asp Ile Thr Gly Asn Arg Ser Asp Thr Val Lys Ile Gly Ser  
 450 455 460

Asp Gly Trp Gly Glu Phe His Val Asn Asp Gly Ser Val Ser Ile Tyr  
 465 470 475 480

Val Gln Lys

<210> 10

<211> 485

<212> PRT

<213> Bacillus sp. 707

<400> 10

His His Asn Gly Thr Asn Gly Thr Met Met Gln Tyr Phe Glu Trp Tyr  
 1 5 10 15

Leu Pro Asn Asp Gly Asn His Trp Asn Arg Leu Arg Ser Asp Ala Ser  
 20 25 30

Asn Leu Lys Asp Lys Gly Ile Ser Ala Val Trp Ile Pro Pro Ala Trp  
 35 40 45

Lys Gly Ala Ser Gln Asn Asp Val Gly Tyr Gly Ala Tyr Asp Leu Tyr  
 50 55 60

Asp Leu Gly Glu Phe Asn Gln Lys Gly Thr Ile Arg Thr Lys Tyr Gly  
 65 70 75 80

Thr Arg Asn Gln Leu Gln Ala Ala Val Asn Ala Leu Lys Ser Asn Gly  
 85 90 95

Ile Gln Val Tyr Gly Asp Val Val Met Asn His Lys Gly Gly Ala Asp  
 100 105 110

Ala Thr Glu Met Val Arg Ala Val Glu Val Asn Pro Asn Asn Arg Asn  
 115 120 125

Gln Glu Val Ser Gly Glu Tyr Thr Ile Glu Ala Trp Thr Lys Phe Asp  
 130 135 140

Phe Pro Gly Arg Gly Asn Thr His Ser Asn Phe Lys Trp Arg Trp Tyr  
 145 150 155 160

His Phe Asp Gly Val Asp Trp Asp Gln Ser Arg Lys Leu Asn Asn Arg  
 165 170 175

Ile Tyr Lys Phe Arg Gly Asp Gly Lys Gly Trp Asp Trp Glu Val Asp  
 180 185 190

Thr Glu Asn Gly Asn Tyr Asp Tyr Leu Met Tyr Ala Asp Ile Asp Met  
 195 200 205

Asp His Pro Glu Val Val Asn Glu Leu Arg Asn Trp Gly Val Trp Tyr  
 210 215 220

Thr Asn Thr Leu Gly Leu Asp Gly Phe Arg Ile Asp Ala Val Lys His  
 225 230 235 240

Ile Lys Tyr Ser Phe Thr Arg Asp Trp Ile Asn His Val Arg Ser Ala  
 245 250 255

Thr Gly Lys Asn Met Phe Ala Val Ala Glu Phe Trp Lys Asn Asp Leu  
 260 265 270

Gly Ala Ile Glu Asn Tyr Leu Asn Lys Thr Asn Trp Asn His Ser Val  
 275 280 285

Phe Asp Val Pro Leu His Tyr Asn Leu Tyr Asn Ala Ser Lys Ser Gly  
 290 295 300



Gly Asn Tyr Asp Met Arg Gln Ile Phe Asn Gly Thr Val Val Gln Arg  
 305 310 315 320

His Pro Met His Ala Val Thr Phe Val Asp Asn His Asp Ser Gln Pro  
 325 330 335

Glu Glu Ala Leu Glu Ser Phe Val Glu Glu Trp Phe Lys Pro Leu Ala  
 340 345 350

Tyr Ala Leu Thr Leu Thr Arg Glu Gln Gly Tyr Pro Ser Val Phe Tyr  
 355 360 365

Gly Asp Tyr Tyr Gly Ile Pro Thr His Gly Val Pro Ala Met Lys Ser  
 370 375 380

Lys Ile Asp Pro Ile Leu Glu Ala Arg Gln Lys Tyr Ala Tyr Gly Arg  
 385 390 395 400

Gln Asn Asp Tyr Leu Asp His His Asn Ile Ile Gly Trp Thr Arg Glu  
 405 410 415

Gly Asn Thr Ala His Pro Asn Ser Gly Leu Ala Thr Ile Met Ser Asp  
 420 425 430

Gly Ala Gly Gly Asn Lys Trp Met Phe Val Gly Arg Asn Lys Ala Gly  
 435 440 445

Gln Val Trp Thr Asp Ile Thr Gly Asn Arg Ala Gly Thr Val Thr Ile  
 450 455 460

Asn Ala Asp Gly Trp Gly Asn Phe Ser Val Asn Gly Gly Ser Val Ser  
 465 470 475 480

Ile Trp Val Asn Lys  
 485

<210> 11  
 <211> 485  
 <212> PRT  
 <213> Bacillus halmapalus

<400> 11  
 His His Asn Gly Thr Asn Gly Thr Met Met Gln Tyr Phe Glu Trp His  
 1 5 10 15

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Leu Pro Asn Asp Gly Asn His Trp Asn Arg Leu Arg Asp Asp Ala Ser  
20 25 30

Asn Leu Arg Asn Arg Gly Ile Thr Ala Ile Trp Ile Pro Pro Ala Trp  
35 40 45

Lys Gly Thr Ser Gln Asn Asp Val Gly Tyr Gly Ala Tyr Asp Leu Tyr  
50 55 60

Asp Leu Gly Glu Phe Asn Gln Lys Gly Thr Val Arg Thr Lys Tyr Gly  
65 70 75 80

Thr Arg Ser Gln Leu Glu Ser Ala Ile His Ala Leu Lys Asn Asn Gly  
85 90 95

Val Gln Val Tyr Gly Asp Val Val Met Asn His Lys Gly Gly Ala Asp  
100 105 110

Ala Thr Glu Asn Val Leu Ala Val Glu Val Asn Pro Asn Asn Arg Asn  
115 120 125

Gln Glu Ile Ser Gly Asp Tyr Thr Ile Glu Ala Trp Thr Lys Phe Asp  
130 135 140

Phe Pro Gly Arg Gly Asn Thr Tyr Ser Asp Phe Lys Trp Arg Trp Tyr  
145 150 155 160

His Phe Asp Gly Val Asp Trp Asp Gln Ser Arg Gln Phe Gln Asn Arg  
165 170 175

Ile Tyr Lys Phe Arg Gly Asp Gly Lys Ala Trp Asp Trp Glu Val Asp  
180 185 190

Ser Glu Asn Gly Asn Tyr Asp Tyr Leu Met Tyr Ala Asp Val Asp Met  
195 200 205

Asp His Pro Glu Val Val Asn Glu Leu Arg Arg Trp Gly Glu Trp Tyr  
210 215 220

Thr Asn Thr Leu Asn Leu Asp Gly Phe Arg Ile Asp Ala Val Lys His  
225 230 235 240

Ile Lys Tyr Ser Phe Thr Arg Asp Trp Leu Thr His Val Arg Asn Ala  
 245 250 255

Thr Gly Lys Glu Met Phe Ala Val Ala Glu Phe Trp Lys Asn Asp Leu  
 260 265 270

Gly Ala Leu Glu Asn Tyr Leu Asn Lys Thr Asn Trp Asn His Ser Val  
 275 280 285

Phe Asp Val Pro Leu His Tyr Asn Leu Tyr Asn Ala Ser Asn Ser Gly  
 290 295 300

Gly Asn Tyr Asp Met Ala Lys Leu Leu Asn Gly Thr Val Val Gln Lys  
 305 310 315 320

His Pro Met His Ala Val Thr Phe Val Asp Asn His Asp Ser Gln Pro  
 325 330 335

Gly Glu Ser Leu Glu Ser Phe Val Gln Glu Trp Phe Lys Pro Leu Ala  
 340 345 350

Tyr Ala Leu Ile Leu Thr Arg Glu Gln Gly Tyr Pro Ser Val Phe Tyr  
 355 360 365

Gly Asp Tyr Tyr Gly Ile Pro Thr His Ser Val Pro Ala Met Lys Ala  
 370 375 380

Lys Ile Asp Pro Ile Leu Glu Ala Arg Gln Asn Phe Ala Tyr Gly Thr  
 385 390 395 400

Gln His Asp Tyr Phe Asp His His Asn Ile Ile Gly Trp Thr Arg Glu  
 405 410 415

Gly Asn Thr Thr His Pro Asn Ser Gly Leu Ala Thr Ile Met Ser Asp  
 420 425 430

Gly Pro Gly Gly Glu Lys Trp Met Tyr Val Gly Gln Asn Lys Ala Gly  
 435 440 445

Gln Val Trp His Asp Ile Thr Gly Asn Lys Pro Gly Thr Val Thr Ile  
 450 455 460

Asn Ala Asp Gly Trp Ala Asn Phe Ser Val Asn Gly Gly Ser Val Ser

465 470 475 480

Ile Trp Val Lys Arg  
485

<210> 12  
<211> 485  
<212> PRT  
<213> Bacillus sp.

<400> 12  
His His Asn Gly Thr Asn Gly Thr Met Met Gln Tyr Phe Glu Trp His  
1 5 10 15

Leu Pro Asn Asp Gly Asn His Trp Asn Arg Leu Arg Asp Asp Ala Ala  
20 25 30

Asn Leu Lys Ser Lys Gly Ile Thr Ala Val Trp Ile Pro Pro Ala Trp  
35 40 45

Lys Gly Thr Ser Gln Asn Asp Val Gly Tyr Gly Ala Tyr Asp Leu Tyr  
50 55 60

Asp Leu Gly Glu Phe Asn Gln Lys Gly Thr Val Arg Thr Lys Tyr Gly  
65 70 75 80

Thr Arg Ser Gln Leu Gln Gly Ala Val Thr Ser Leu Lys Asn Asn Gly  
85 90 95

Ile Gln Val Tyr Gly Asp Val Val Met Asn His Lys Gly Gly Ala Asp  
100 105 110

Gly Thr Glu Met Val Asn Ala Val Glu Val Asn Arg Ser Asn Arg Asn  
115 120 125

Gln Glu Ile Ser Gly Glu Tyr Thr Ile Glu Ala Trp Thr Lys Phe Asp  
130 135 140

Phe Pro Gly Arg Gly Asn Thr His Ser Asn Phe Lys Trp Arg Trp Tyr  
145 150 155 160

His Phe Asp Gly Thr Asp Trp Asp Gln Ser Arg Gln Leu Gln Asn Lys  
165 170 175

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Ile Tyr Lys Phe Arg Gly Thr Gly Lys Ala Trp Asp Trp Glu Val Asp  
180 185 190

Ile Glu Asn Gly Asn Tyr Asp Tyr Leu Met Tyr Ala Asp Ile Asp Met  
195 200 205

Asp His Pro Glu Val Ile Asn Glu Leu Arg Asn Trp Gly Val Trp Tyr  
210 215 220

Thr Asn Thr Leu Asn Leu Asp Gly Phe Arg Ile Asp Ala Val Lys His  
225 230 235 240

Ile Lys Tyr Ser Tyr Thr Arg Asp Trp Leu Thr His Val Arg Asn Thr  
245 250 255

Thr Gly Lys Pro Met Phe Ala Val Ala Glu Phe Trp Lys Asn Asp Leu  
260 265 270

Ala Ala Ile Glu Asn Tyr Leu Asn Lys Thr Ser Trp Asn His Ser Val  
275 280 285

Phe Asp Val Pro Leu His Tyr Asn Leu Tyr Asn Ala Ser Asn Ser Gly  
290 295 300

Gly Tyr Phe Asp Met Arg Asn Ile Leu Asn Gly Ser Val Val Gln Lys  
305 310 315 320

His Pro Ile His Ala Val Thr Phe Val Asp Asn His Asp Ser Gln Pro  
325 330 335

Gly Glu Ala Leu Glu Ser Phe Val Gln Ser Trp Phe Lys Pro Leu Ala  
340 345 350

Tyr Ala Leu Ile Leu Thr Arg Glu Gln Gly Tyr Pro Ser Val Phe Tyr  
355 360 365

Gly Asp Tyr Tyr Gly Ile Pro Thr His Gly Val Pro Ser Met Lys Ser  
370 375 380

Lys Ile Asp Pro Leu Leu Gln Ala Arg Gln Thr Tyr Ala Tyr Gly Thr  
385 390 395 400

Gln His Asp Tyr Phe Asp His His Asp Ile Ile Gly Trp Thr Arg Glu

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405  
 Gly Asp Ser Ser His Pro Asn Ser Gly Leu Ala Thr Ile Met Ser Asp  
 420 425 430

Gly Pro Gly Gly Asn Lys Trp Met Tyr Val Gly Lys His Lys Ala Gly  
 435 440 445

Gln Val Trp Arg Asp Ile Thr Gly Asn Arg Ser Gly Thr Val Thr Ile  
 450 455 460

Asn Ala Asp Gly Trp Gly Asn Phe Thr Val Asn Gly Gly Ala Val Ser  
 465 470 475 480

Val Trp Val Lys Gln  
 485

<210> 13  
 <211> 480  
 <212> PRT  
 <213> Bacillus sp.

<400> 13  
 Asp Gly Leu Asn Gly Thr Met Met Gln Tyr Tyr Glu Trp His Leu Glu  
 1 5 10 15

Asn Asp Gly Gln His Trp Asn Arg Leu His Asp Asp Ala Ala Ala Leu  
 20 25 30

Ser Asp Ala Gly Ile Thr Ala Ile Trp Ile Pro Pro Ala Tyr Lys Gly  
 35 40 45

Asn Ser Gln Ala Asp Val Gly Tyr Gly Ala Tyr Asp Leu Tyr Asp Leu  
 50 55 60

Gly Glu Phe Asn Gln Lys Gly Thr Val Arg Thr Lys Tyr Gly Thr Lys  
 65 70 75 80

Ala Gln Leu Glu Arg Ala Ile Gly Ser Leu Lys Ser Asn Asp Ile Asn  
 85 90 95

Val Tyr Gly Asp Val Val Met Asn His Lys Met Gly Ala Asp Phe Thr  
 100 105 110

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Glu Ala Val Gln Ala Val Gln Val Asn Pro Thr Asn Arg Trp Gln Asp  
115 120 125

Ile Ser Gly Ala Tyr Thr Ile Asp Ala Trp Thr Gly Phe Asp Phe Ser  
130 135 140

Gly Arg Asn Asn Ala Tyr Ser Asp Phe Lys Trp Arg Trp Phe His Phe  
145 150 155 160

Asn Gly Val Asp Trp Asp Gln Arg Tyr Gln Glu Asn His Ile Phe Arg  
165 170 175

Phe Ala Asn Thr Asn Trp Asn Trp Arg Val Asp Glu Glu Asn Gly Asn  
180 185 190

Tyr Asp Tyr Leu Leu Gly Ser Asn Ile Asp Phe Ser His Pro Glu Val  
195 200 205

Gln Asp Glu Leu Lys Asp Trp Gly Ser Trp Phe Thr Asp Glu Leu Asp  
210 215 220

Leu Asp Gly Tyr Arg Leu Asp Ala Ile Lys His Ile Pro Phe Trp Tyr  
225 230 235 240

Thr Ser Asp Trp Val Arg His Gln Arg Asn Glu Ala Asp Gln Asp Leu  
245 250 255

Phe Val Val Gly Glu Tyr Trp Lys Asp Asp Val Gly Ala Leu Glu Phe  
260 265 270

Tyr Leu Asp Glu Met Asn Trp Glu Met Ser Leu Phe Asp Val Pro Leu  
275 280 285

Asn Tyr Asn Phe Tyr Arg Ala Ser Gln Gln Gly Gly Ser Tyr Asp Met  
290 295 300

Arg Asn Ile Leu Arg Gly Ser Leu Val Glu Ala His Pro Met His Ala  
305 310 315 320

Val Thr Phe Val Asp Asn His Asp Thr Gln Pro Gly Glu Ser Leu Glu  
325 330 335

Ser Trp Val Ala Asp Trp Phe Lys Pro Leu Ala Tyr Ala Thr Ile Leu

340 345 350  
 Thr Arg Glu Gly Gly Tyr Pro Asn Val Phe Tyr Gly Asp Tyr Tyr Gly  
 355 360 365  
 Ile Pro Asn Asp Asn Ile Ser Ala Lys Lys Asp Met Ile Asp Glu Leu  
 370 375 380  
 Leu Asp Ala Arg Gln Asn Tyr Ala Tyr Gly Thr Gln His Asp Tyr Phe  
 385 390 395 400  
 Asp His Trp Asp Val Val Gly Trp Thr Arg Glu Gly Ser Ser Ser Arg  
 405 410 415  
 Pro Asn Ser Gly Leu Ala Thr Ile Met Ser Asn Gly Pro Gly Gly Ser  
 420 425 430  
 Lys Trp Met Tyr Val Gly Arg Gln Asn Ala Gly Gln Thr Trp Thr Asp  
 435 440 445  
 Leu Thr Gly Asn Asn Gly Ala Ser Val Thr Ile Asn Gly Asp Gly Trp  
 450 455 460  
 Gly Glu Phe Phe Thr Asn Gly Gly Ser Val Ser Val Tyr Val Asn Gln  
 465 470 475 480  
 <210> 14  
 <211> 480  
 <212> PRT  
 <213> Bacillus sp.  
 <400> 14  
 Asp Gly Leu Asn Gly Thr Met Met Gln Tyr Tyr Glu Trp His Leu Glu  
 1 5 10 15  
 Asn Asp Gly Gln His Trp Asn Arg Leu His Asp Asp Ala Glu Ala Leu  
 20 25 30  
 Ser Asn Ala Gly Ile Thr Ala Ile Trp Ile Pro Pro Ala Tyr Lys Gly  
 35 40 45  
 Asn Ser Gln Ala Asp Val Gly Tyr Gly Ala Tyr Asp Leu Tyr Asp Leu  
 50 55 60



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Gly Glu Phe Asn Gln Lys Gly Thr Val Arg Thr Lys Tyr Gly Thr Lys  
65 70 75 80

Ala Gln Leu Glu Arg Ala Ile Gly Ser Leu Lys Ser Asn Asp Ile Asn  
85 90 95

Val Tyr Gly Asp Val Val Met Asn His Lys Leu Gly Ala Asp Phe Thr  
100 105 110

Glu Ala Val Gln Ala Val Gln Val Asn Pro Ser Asn Arg Trp Gln Asp  
115 120 125

Ile Ser Gly Val Tyr Thr Ile Asp Ala Trp Thr Gly Phe Asp Phe Pro  
130 135 140

Gly Arg Asn Asn Ala Tyr Ser Asp Phe Lys Trp Arg Trp Phe His Phe  
145 150 155 160

Asn Gly Val Asp Trp Asp Gln Arg Tyr Gln Glu Asn His Leu Phe Arg  
165 170 175

Phe Ala Asn Thr Asn Trp Asn Trp Arg Val Asp Glu Glu Asn Gly Asn  
180 185 190

Tyr Asp Tyr Leu Leu Gly Ser Asn Ile Asp Phe Ser His Pro Glu Val  
195 200 205

Gln Glu Glu Leu Lys Asp Trp Gly Ser Trp Phe Thr Asp Glu Leu Asp  
210 215 220

Leu Asp Gly Tyr Arg Leu Asp Ala Ile Lys His Ile Pro Phe Trp Tyr  
225 230 235 240

Thr Ser Asp Trp Val Arg His Gln Arg Ser Glu Ala Asp Gln Asp Leu  
245 250 255

Phe Val Val Gly Glu Tyr Trp Lys Asp Asp Val Gly Ala Leu Glu Phe  
260 265 270

Tyr Leu Asp Glu Met Asn Trp Glu Met Ser Leu Phe Asp Val Pro Leu  
275 280 285

Asn Tyr Asn Phe Tyr Arg Ala Ser Lys Gln Gly Gly Ser Tyr Asp Met

290 295 300  
 Arg Asn Ile Leu Arg Gly Ser Leu Val Glu Ala His Pro Ile His Ala  
 305 310 315 320  
 Val Thr Phe Val Asp Asn His Asp Thr Gln Pro Gly Glu Ser Leu Glu  
 325 330 335  
 Ser Trp Val Ala Asp Trp Phe Lys Pro Leu Ala Tyr Ala Thr Ile Leu  
 340 345 350  
 Thr Arg Glu Gly Gly Tyr Pro Asn Val Phe Tyr Gly Asp Tyr Tyr Gly  
 355 360 365  
 Ile Pro Asn Asp Asn Ile Ser Ala Lys Lys Asp Met Ile Asp Glu Leu  
 370 375 380  
 Leu Asp Ala Arg Gln Asn Tyr Ala Tyr Gly Thr Gln His Asp Tyr Phe  
 385 390 395 400  
 Asp His Trp Asp Ile Val Gly Trp Thr Arg Glu Gly Thr Ser Ser Arg  
 405 410 415  
 Pro Asn Ser Gly Leu Ala Thr Ile Met Ser Asn Gly Pro Gly Gly Ser  
 420 425 430  
 Lys Trp Met Tyr Val Gly Gln Gln His Ala Gly Gln Thr Trp Thr Asp  
 435 440 445  
 Leu Thr Gly Asn His Ala Ala Ser Val Thr Ile Asn Gly Asp Gly Trp  
 450 455 460  
 Gly Glu Phe Phe Thr Asn Gly Gly Ser Val Ser Val Tyr Val Asn Gln  
 465 470 475 480

<210> 15  
 <211> 486  
 <212> PRT  
 <213> Geobacillus stearothermophilus

<400> 15  
 Ala Ala Pro Phe Asn Gly Thr Met Met Gln Tyr Phe Glu Trp Tyr Leu  
 1 5 10 15

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Pro Asp Asp Gly Thr Leu Trp Thr Lys Val Ala Asn Glu Ala Asn Asn  
20 25 30

Leu Ser Ser Leu Gly Ile Thr Ala Leu Trp Leu Pro Pro Ala Tyr Lys  
35 40 45

Gly Thr Ser Arg Ser Asp Val Gly Tyr Gly Val Tyr Asp Leu Tyr Asp  
50 55 60

Leu Gly Glu Phe Asn Gln Lys Gly Thr Val Arg Thr Lys Tyr Gly Thr  
65 70 75 80

Lys Ala Gln Tyr Leu Gln Ala Ile Gln Ala Ala His Ala Ala Gly Met  
85 90 95

Gln Val Tyr Ala Asp Val Val Phe Asp His Lys Gly Gly Ala Asp Gly  
100 105 110

Thr Glu Trp Val Asp Ala Val Glu Val Asn Pro Ser Asp Arg Asn Gln  
115 120 125

Glu Ile Ser Gly Thr Tyr Gln Ile Gln Ala Trp Thr Lys Phe Asp Phe  
130 135 140

Pro Gly Arg Gly Asn Thr Tyr Ser Ser Phe Lys Trp Arg Trp Tyr His  
145 150 155 160

Phe Asp Gly Val Asp Trp Asp Glu Ser Arg Lys Leu Ser Arg Ile Tyr  
165 170 175

Lys Phe Arg Gly Lys Ala Trp Asp Trp Glu Val Asp Thr Glu Phe Gly  
180 185 190

Asn Tyr Asp Tyr Leu Met Tyr Ala Asp Leu Asp Met Asp His Pro Glu  
195 200 205

Val Val Thr Glu Leu Lys Asn Trp Gly Lys Trp Tyr Val Asn Thr Thr  
210 215 220

Asn Ile Asp Gly Phe Arg Leu Asp Ala Val Lys His Ile Lys Phe Ser  
225 230 235 240

Phe Phe Pro Asp Trp Leu Ser Tyr Val Arg Ser Gln Thr Gly Lys Pro

				245						250						255
Leu	Phe	Thr	Val	Gly	Glu	Tyr	Trp	Ser	Tyr	Asp	Ile	Asn	Lys	Leu	His	
			260					265					270			
Asn	Tyr	Ile	Thr	Lys	Thr	Asn	Gly	Thr	Met	Ser	Leu	Phe	Asp	Ala	Pro	
		275					280					285				
Leu	His	Asn	Lys	Phe	Tyr	Thr	Ala	Ser	Lys	Ser	Gly	Gly	Ala	Phe	Asp	
	290					295					300					
Met	Arg	Thr	Leu	Met	Thr	Asn	Thr	Leu	Met	Lys	Asp	Gln	Pro	Thr	Leu	
305					310					315					320	
Ala	Val	Thr	Phe	Val	Asp	Asn	His	Asp	Thr	Glu	Pro	Gly	Gln	Ala	Leu	
				325					330					335		
Gln	Ser	Trp	Val	Asp	Pro	Trp	Phe	Lys	Pro	Leu	Ala	Tyr	Ala	Phe	Ile	
			340					345					350			
Leu	Thr	Arg	Gln	Glu	Gly	Tyr	Pro	Cys	Val	Phe	Tyr	Gly	Asp	Tyr	Tyr	
		355					360					365				
Gly	Ile	Pro	Gln	Tyr	Asn	Ile	Pro	Ser	Leu	Lys	Ser	Lys	Ile	Asp	Pro	
	370					375					380					
Leu	Leu	Ile	Ala	Arg	Arg	Asp	Tyr	Ala	Tyr	Gly	Thr	Gln	His	Asp	Tyr	
385					390					395					400	
Leu	Asp	His	Ser	Asp	Ile	Ile	Gly	Trp	Thr	Arg	Glu	Gly	Gly	Thr	Glu	
				405					410					415		
Lys	Pro	Gly	Ser	Gly	Leu	Ala	Ala	Leu	Ile	Thr	Asp	Gly	Pro	Gly	Gly	
			420					425					430			
Ser	Lys	Trp	Met	Tyr	Val	Gly	Lys	Gln	His	Ala	Gly	Lys	Val	Phe	Tyr	
		435					440					445				
Asp	Leu	Thr	Gly	Asn	Arg	Ser	Asp	Thr	Val	Thr	Ile	Asn	Ser	Asp	Gly	
	450					455					460					
Trp	Gly	Glu	Phe	Lys	Val	Asn	Gly	Gly	Ser	Val	Ser	Val	Trp	Val	Pro	
465					470					475					480	

Arg Lys Thr Thr Val Ser  
485

<210> 16  
<211> 520  
<212> PRT  
<213> Artificial sequence

<220>  
<221> source  
<223> /note="Description of artificial sequence: Synthetic  
consensus sequence"

<220>  
<221> VARIANT  
<222> (1)..(1)  
<223> /replace=" "

<220>  
<221> VARIANT  
<222> (2)..(2)  
<223> /replace="His" or " "

<220>  
<221> misc\_feature  
<222> (2)..(2)  
<223> /note="Residues given in the sequence have no preference  
with respect to those in the annotation for said positions"

<220>  
<221> VARIANT  
<222> (4)..(4)  
<223> /replace="Gly" or "Asn" or " "

<220>  
<221> VARIANT  
<222> (5)..(5)  
<223> /replace="Thr" or "Leu" or "Val"

<220>  
<221> misc\_feature  
<222> (4)..(5)  
<223> /note="Residues given in the sequence have no preference  
with respect to those in the annotation for said positions"

<220>  
<221> VARIANT  
<222> (25)..(25)  
<223> /replace="Asn" or "Lys" or "Arg"

<220>  
<221> misc\_feature  
<222> (25)..(25)

<223> /note="Residue given in the sequence has no preference with respect to those in the annotation for said position"

<220>

<221> VARIANT

<222> (28)..(28)

<223> /replace="Asn" or "Gln" or "Arg" or "His"

<220>

<221> misc\_feature

<222> (28)..(28)

<223> /note="Residue given in the sequence has no preference with respect to those in the annotation for said position"

<220>

<221> VARIANT

<222> (32)..(32)

<223> /replace="Ser" or "Ala" or "Glu"

<220>

<221> misc\_feature

<222> (32)..(32)

<223> /note="Residue given in the sequence has no preference with respect to those in the annotation for said position"

<220>

<221> VARIANT

<222> (37)..(37)

<223> /replace="Lys" or "His" or "Ile" or "Arg" or "Ala"

<220>

<221> misc\_feature

<222> (37)..(37)

<223> /note="Residue given in the sequence has no preference with respect to those in the annotation for said position"

<220>

<221> VARIANT

<222> (86)..(86)

<223> /replace="Gln" or "Glu"

<220>

<221> VARIANT

<222> (87)..(87)

<223> /replace="Ala" or "Ser" or "Asp" or "Gly" or "Arg"

<220>

<221> misc\_feature

<222> (86)..(87)

<223> /note="Residues given in the sequence have no preference with respect to those in the annotation for said positions"

<220>

<221> VARIANT

<222> (90)..(90)

<223> /replace="Thr" or "Lys" or "Gly" or "Asn" or "His"

<220>  
<221> misc\_feature  
<222> (90)..(90)  
<223> /note="Residue given in the sequence has no preference  
with respect to those in the annotation for said position"

<220>  
<221> VARIANT  
<222> (95)..(95)  
<223> /replace="Asn" or "Arg"

<220>  
<221> misc\_feature  
<222> (95)..(95)  
<223> /note="Residue given in the sequence has no preference  
with respect to those in the annotation for said position"

<220>  
<221> VARIANT  
<222> (116)..(116)  
<223> /replace="Met" or "Asp" or "Asn" or "Ala"

<220>  
<221> misc\_feature  
<222> (116)..(116)  
<223> /note="Residue given in the sequence has no preference  
with respect to those in the annotation for said position"

<220>  
<221> VARIANT  
<222> (118)..(118)  
<223> /replace="Arg" or "Thr" or "Leu" or "Asn" or "Gln"

<220>  
<221> misc\_feature  
<222> (118)..(118)  
<223> /note="Residue given in the sequence has no preference  
with respect to those in the annotation for said position"

<220>  
<221> VARIANT  
<222> (134)..(134)  
<223> /replace="Glu" or "Asp" or "Ala" or "Val"

<220>  
<221> misc\_feature  
<222> (134)..(134)  
<223> /note="Residue given in the sequence has no preference  
with respect to those in the annotation for said position"

<220>  
<221> VARIANT  
<222> (136)..(136)  
<223> /replace="Thr" or "Leu"

<220>  
<221> misc\_feature

```
<222> (136)..(136)
<223> /note="Residue given in the sequence has no preference
with respect to those in the annotation for said position"

<220>
<221> VARIANT
<222> (138)..(138)
<223> /replace="Glu" or "Lys" or "Asp"

<220>
<221> misc_feature
<222> (138)..(138)
<223> /note="Residue given in the sequence has no preference
with respect to those in the annotation for said position"

<220>
<221> VARIANT
<222> (154)..(154)
<223> /replace="Asp" or "Asn"

<220>
<221> misc_feature
<222> (154)..(154)
<223> /note="Residue given in the sequence has no preference
with respect to those in the annotation for said position"

<220>
<221> VARIANT
<222> (175)..(175)
<223> /replace=" "

<220>
<221> VARIANT
<222> (183)..(183)
<223> /replace="His" or "Glu" or "Asp" or "Thr" or " "

<220>
<221> misc_feature
<222> (183)..(183)
<223> /note="Residue given in the sequence has no preference
with respect to those in the annotation for said position"

<220>
<221> VARIANT
<222> (215)..(215)
<223> /replace="Asn" or "Ala" or "Asp" or "Glu"

<220>
<221> misc_feature
<222> (215)..(215)
<223> /note="Residue given in the sequence has no preference
with respect to those in the annotation for said position"

<220>
<221> VARIANT
<222> (222)..(222)
<223> /replace="Val" or "Thr" or "Ile" or "Glu" or "Ser"
```



<220>  
<221> misc\_feature  
<222> (222)..(222)  
<223> /note="Residue given in the sequence has no preference  
with respect to those in the annotation for said position"

<220>  
<221> VARIANT  
<222> (225)..(225)  
<223> /replace="Thr" or "Ala"

<220>  
<221> misc\_feature  
<222> (225)..(225)  
<223> /note="Residue given in the sequence has no preference  
with respect to those in the annotation for said position"

<220>  
<221> VARIANT  
<222> (246)..(246)  
<223> /replace="Thr" or "Leu"

<220>  
<221> misc\_feature  
<222> (246)..(246)  
<223> /note="Residues given in the sequence have no preference  
with respect to those in the annotation for said positions"

<220>  
<221> VARIANT  
<222> (247)..(247)  
<223> /replace="Pro" or "Ser"

<220>  
<221> VARIANT  
<222> (256)..(256)  
<223> /replace="Ala" or "Lys" or "Thr" or "Glu"

<220>  
<221> misc\_feature  
<222> (256)..(256)  
<223> /note="Residue given in the sequence has no preference  
with respect to those in the annotation for said position"

<220>  
<221> VARIANT  
<222> (260)..(260)  
<223> /replace="Asn" or "Glu" or "Asp"

<220>  
<221> misc\_feature  
<222> (260)..(260)  
<223> /note="Residue given in the sequence has no preference  
with respect to those in the annotation for said position"

<220>

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<221> VARIANT
<222> (269)..(269)
<223> /replace="Lys" or "Gln"

<220>
<221> misc_feature
<222> (269)..(269)
<223> /note="Residue given in the sequence has no preference
with respect to those in the annotation for said position"

<220>
<221> VARIANT
<222> (270)..(270)
<223> /replace="Tyr" or "Asp"

<220>
<221> VARIANT
<222> (280)..(280)
<223> /replace="Gln" or "Asn" or "Asp"

<220>
<221> misc_feature
<222> (280)..(280)
<223> /note="Residue given in the sequence has no preference
with respect to those in the annotation for said position"

<220>
<221> VARIANT
<222> (285)..(285)
<223> /replace="Thr" or "Glu"

<220>
<221> VARIANT
<222> (299)..(299)
<223> /replace="Asn" or "Ala" or "Arg"

<220>
<221> misc_feature
<222> (299)..(299)
<223> /note="Residue given in the sequence has no preference
with respect to those in the annotation for said position"

<220>
<221> VARIANT
<222> (311)..(311)
<223> /replace="Asn" or "Lys" or "Arg" or "Gln"

<220>
<221> misc_feature
<222> (311)..(311)
<223> /note="Residue given in the sequence has no preference
with respect to those in the annotation for said position"

<220>
<221> VARIANT
<222> (314)..(314)
<223> /replace="Asn" or "Asp" or "Arg"
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<220>
<221> misc_feature
<222> (314)..(314)
<223> /note="Residue given in the sequence has no preference
with respect to those in the annotation for said position"

<220>
<221> VARIANT
<222> (319)..(319)
<223> /replace="Gln" or "Ser" or "Glu"

<220>
<221> VARIANT
<222> (320)..(320)
<223> /replace="Arg" or "Lys" or "Ala"

<220>
<221> misc_feature
<222> (319)..(320)
<223> /note="Residues given in the sequence have no preference
with respect to those in the annotation for said positions"

<220>
<221> VARIANT
<222> (323)..(323)
<223> /replace="Ser" or "Leu" or "Glu" or "Met" or "Ile"

<220>
<221> VARIANT
<222> (324)..(324)
<223> /replace="His" or "Lys"

<220>
<221> misc_feature
<222> (323)..(324)
<223> /note="Residues given in the sequence have no preference
with respect to those in the annotation for said positions"

<220>
<221> VARIANT
<222> (346)..(346)
<223> /replace="Glu" or "Thr" or "Ser" or "Asp"

<220>
<221> misc_feature
<222> (346)..(346)
<223> /note="Residue given in the sequence has no preference
with respect to those in the annotation for said position"

<220>
<221> VARIANT
<222> (361)..(361)
<223> /replace="Gln" or "Ser" or "Gly"

<220>
<221> misc_feature
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<222> (361)..(361)
<223> /note="Residue given in the sequence has no preference
with respect to those in the annotation for said position"

<220>
<221> VARIANT
<222> (365)..(365)
<223> /replace="Ser" or "Gln" or "Asn"

<220>
<221> misc_feature
<222> (365)..(365)
<223> /note="Residue given in the sequence has no preference
with respect to those in the annotation for said position"

<220>
<221> VARIANT
<222> (379)..(379)
<223> /replace="Pro" or " "

<220>
<221> VARIANT
<222> (380)..(380)
<223> /replace="Lys" or " "

<220>
<221> VARIANT
<222> (381)..(381)
<223> /replace=" "

<220>
<221> VARIANT
<222> (394)..(394)
<223> /replace="Glu" or "Lys" or "Gln" or "Asp"

<220>
<221> misc_feature
<222> (394)..(394)
<223> /note="Residue given in the sequence has no preference
with respect to those in the annotation for said position"

<220>
<221> VARIANT
<222> (398)..(398)
<223> /replace="Lys" or "Gln" or "Glu" or "Asn" or "Thr"

<220>
<221> misc_feature
<222> (398)..(398)
<223> /note="Residue given in the sequence has no preference
with respect to those in the annotation for said position"

<220>
<221> VARIANT
<222> (411)..(411)
<223> /replace="His" or "Pro" or "Trp"
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<220>  
<221> misc\_feature  
<222> (411)..(411)  
<223> /note="Residue given in the sequence has no preference  
with respect to those in the annotation for said position"

<220>  
<221> VARIANT  
<222> (421)..(421)  
<223> /replace="Asn" or "Asp" or "Ser" or "Thr" or "Gly"

<220>  
<221> misc\_feature  
<222> (421)..(421)  
<223> /note="Residue given in the sequence has no preference  
with respect to those in the annotation for said position"

<220>  
<221> VARIANT  
<222> (449)..(449)  
<223> /replace="Lys" or "Asn"

<220>  
<221> misc\_feature  
<222> (449)..(449)  
<223> /note="Residue given in the sequence has no preference  
with respect to those in the annotation for said position"

<220>  
<221> VARIANT  
<222> (452)..(452)  
<223> /replace="Gln" or "Glu"

<220>  
<221> misc\_feature  
<222> (452)..(452)  
<223> /note="Residue given in the sequence has no preference  
with respect to those in the annotation for said position"

<220>  
<221> VARIANT  
<222> (476)..(476)  
<223> /replace="Ser" or "His" or "Thr" or "Phe"

<220>  
<221> misc\_feature  
<222> (476)..(476)  
<223> /note="Residue given in the sequence has no preference  
with respect to those in the annotation for said position"

<220>  
<221> VARIANT  
<222> (487)..(487)  
<223> /replace="Asn" or "Gln" or "Lys"

<220>  
<221> misc\_feature

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<222> (487)..(487)
<223> /note="Residue given in the sequence has no preference
with respect to those in the annotation for said position"

<220>
<221> VARIANT
<222> (489)..(489)
<223> /replace=" "

<220>
<221> VARIANT
<222> (490)..(491)
<223> /replace=" "

<220>
<221> VARIANT
<222> (492)..(492)
<223> /replace=" "

<220>
<221> VARIANT
<222> (493)..(493)
<223> /replace=" "

<220>
<221> VARIANT
<222> (494)..(494)
<223> /replace=" "

<220>
<221> VARIANT
<222> (495)..(495)
<223> /replace=" "

<220>
<221> VARIANT
<222> (496)..(496)
<223> /replace=" "

<220>
<221> VARIANT
<222> (497)..(497)
<223> /replace=" "

<220>
<221> VARIANT
<222> (498)..(498)
<223> /replace=" "

<220>
<221> VARIANT
<222> (499)..(499)
<223> /replace=" "

<220>
<221> VARIANT
<222> (500)..(501)
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<223> /replace=" "

<220>  
<221> VARIANT  
<222> (502)..(502)  
<223> /replace=" "

<220>  
<221> VARIANT  
<222> (503)..(503)  
<223> /replace=" "

<220>  
<221> VARIANT  
<222> (504)..(504)  
<223> /replace=" "

<220>  
<221> VARIANT  
<222> (505)..(505)  
<223> /replace=" "

<220>  
<221> VARIANT  
<222> (506)..(506)  
<223> /replace=" "

<220>  
<221> VARIANT  
<222> (507)..(507)  
<223> /replace=" "

<220>  
<221> VARIANT  
<222> (508)..(508)  
<223> /replace=" "

<220>  
<221> VARIANT  
<222> (509)..(509)  
<223> /replace=" "

<220>  
<221> VARIANT  
<222> (510)..(510)  
<223> /replace=" "

<220>  
<221> VARIANT  
<222> (511)..(511)  
<223> /replace=" "

<220>  
<221> VARIANT  
<222> (512)..(512)  
<223> /replace=" "

<220>  
 <221> VARIANT  
 <222> (513)..(513)  
 <223> /replace=" "

<220>  
 <221> VARIANT  
 <222> (514)..(514)  
 <223> /replace=" "

<220>  
 <221> VARIANT  
 <222> (515)..(515)  
 <223> /replace=" "

<220>  
 <221> VARIANT  
 <222> (516)..(516)  
 <223> /replace=" "

<220>  
 <221> VARIANT  
 <222> (517)..(517)  
 <223> /replace=" "

<220>  
 <221> VARIANT  
 <222> (518)..(518)  
 <223> /replace=" "

<220>  
 <221> VARIANT  
 <222> (519)..(519)  
 <223> /replace=" "

<220>  
 <221> VARIANT  
 <222> (520)..(520)  
 <223> /replace=" "

<220>  
 <221> misc\_feature  
 <222> (489)..(520)  
 <223> /note="Residues given in the sequence have no preference  
 with respect to those in the annotation for said positions"

<400> 16  
 His Ala Ala Pro Phe Asn Gly Thr Met Met Gln Tyr Phe Glu Trp Tyr  
 1 5 10 15

Leu Pro Asn Asp Gly Gln His Trp Thr Arg Leu Ala Asn Asp Ala Asn  
 20 25 30

Asn Leu Ser Ser Leu Gly Ile Thr Ala Leu Trp Ile Pro Pro Ala Tyr  
 35 40 45



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Lys Gly Thr Ser Gln Ser Asp Val Gly Tyr Gly Ala Tyr Asp Leu Tyr  
50 55 60

Asp Leu Gly Glu Phe Asn Gln Lys Gly Thr Val Arg Thr Lys Tyr Gly  
65 70 75 80

Thr Lys Ala Gln Leu Leu Gln Ala Ile Gln Ala Leu His Ala Ala Gly  
85 90 95

Ile Gln Val Tyr Gly Asp Val Val Met Asn His Lys Gly Gly Ala Asp  
100 105 110

Gly Thr Glu Trp Val Asp Ala Val Glu Val Asn Pro Ser Asp Arg Asn  
115 120 125

Gln Glu Ile Ser Gly Thr Tyr Gln Ile Gln Ala Trp Thr Lys Phe Asp  
130 135 140

Phe Pro Gly Arg Gly Asn Thr Tyr Ser Ser Phe Lys Trp Arg Trp Tyr  
145 150 155 160

His Phe Asp Gly Val Asp Trp Asp Glu Ser Arg Lys Leu Asn Asn Arg  
165 170 175

Ile Tyr Lys Phe Arg Gly Ile Gly Lys Ala Trp Asp Trp Glu Val Asp  
180 185 190

Thr Glu Asn Gly Asn Tyr Asp Tyr Leu Met Tyr Ala Asp Ile Asp Met  
195 200 205

Asp His Pro Glu Val Val Thr Glu Leu Lys Asn Trp Gly Lys Trp Tyr  
210 215 220

Val Asn Thr Leu Asn Leu Asp Gly Phe Arg Leu Asp Ala Val Lys His  
225 230 235 240

Ile Lys Phe Ser Phe Phe Arg Asp Trp Leu Ser His Val Arg Ser Gln  
245 250 255

Thr Gly Lys Pro Leu Phe Thr Val Gly Glu Tyr Trp Ser Asn Asp Ile  
260 265 270

Gly Ala Leu Glu Asn Tyr Leu Thr Lys Thr Asn Trp Asn Met Ser Leu  
275 280 285

Phe Asp Val Pro Leu His Tyr Asn Phe Tyr Thr Ala Ser Lys Ser Gly  
290 295 300

Gly Ala Tyr Asp Met Arg Thr Leu Leu Thr Gly Thr Leu Val Lys Asp  
305 310 315 320

His Pro Thr Leu Ala Val Thr Phe Val Asp Asn His Asp Thr Gln Pro  
325 330 335

Gly Gln Ala Leu Glu Ser Trp Val Asp Pro Trp Phe Lys Pro Leu Ala  
340 345 350

Tyr Ala Phe Ile Leu Thr Arg Glu Glu Gly Tyr Pro Cys Val Phe Tyr  
355 360 365

Gly Asp Tyr Tyr Gly Ile Pro Gln Tyr Asn Gln Arg Glu Ile Pro Ser  
370 375 380

Leu Lys Ser Lys Ile Asp Pro Leu Leu Ile Ala Arg Arg Asp Tyr Ala  
385 390 395 400

Tyr Gly Thr Gln His Asp Tyr Leu Asp His Ser Asp Ile Ile Gly Trp  
405 410 415

Thr Arg Glu Gly Val Thr Ser Lys Pro Asn Ser Gly Leu Ala Ala Leu  
420 425 430

Ile Thr Asp Gly Pro Gly Gly Ser Lys Trp Met Tyr Val Gly Lys Gln  
435 440 445

His Ala Gly Lys Val Trp Tyr Asp Leu Thr Gly Asn Arg Ser Asp Thr  
450 455 460

Val Thr Ile Asn Ser Asp Gly Trp Gly Glu Phe Lys Val Asn Gly Gly  
465 470 475 480

Ser Val Ser Val Trp Val Pro Arg Lys Thr Thr Val Ser Thr Ile Ala  
485 490 495

Arg Pro Ile Thr Thr Arg Pro Trp Thr Gly Glu Phe Val Arg Trp Thr  
 500 505 510

Glu Pro Arg Leu Val Ala Trp Pro  
 515 520

<210> 17  
 <211> 39  
 <212> DNA  
 <213> Artificial sequence

<220>  
 <221> source  
 <223> /note="Description of artificial sequence: Synthetic oligonucleotide"

<220>  
 <221> modified\_base  
 <222> (19)..(20)  
 <223> a, c, t, g, unknown or other

<400> 17  
 gtcaagcata ttaagttcnn sttttttcoct gattggttg 39

<210> 18  
 <211> 39  
 <212> DNA  
 <213> Artificial sequence

<220>  
 <221> source  
 <223> /note="Description of artificial sequence: Synthetic oligonucleotide"

<220>  
 <221> modified\_base  
 <222> (20)..(21)  
 <223> a, c, t, g, unknown or other

<400> 18  
 caaccaatca ggaaaaaasn ngaacttaat atgcttgac 39

<210> 19  
 <211> 87  
 <212> DNA  
 <213> Artificial sequence

<220>  
 <221> source  
 <223> /note="Description of artificial sequence: Synthetic oligonucleotide"

<400> 19  
atgaaacaac aaaaacggct ttacgcccga ttgctgacgc tgttatttgc gctcatcttc 60

ttgctgcctc attctgcagc ttcagca 87

<210> 20  
<211> 29  
<212> PRT  
<213> Artificial sequence

<220>  
<221> source  
<223> /note="Description of artificial sequence: Synthetic peptide"

<400> 20  
Met Lys Gln Gln Lys Arg Leu Tyr Ala Arg Leu Leu Thr Leu Leu Phe  
1 5 10 15

Ala Leu Ile Phe Leu Leu Pro His Ser Ala Ala Ser Ala  
20 25

<210> 21  
<211> 486  
<212> PRT  
<213> Artificial sequence

<220>  
<221> source  
<223> /note="Description of artificial sequence: Synthetic polypeptide"

<400> 21  
Ala Ala Pro Phe Asn Gly Thr Met Met Gln Tyr Phe Glu Trp Tyr Leu  
1 5 10 15

Pro Asp Asp Gly Thr Leu Trp Thr Lys Val Ala Asn Glu Ala Asn Asn  
20 25 30

Leu Ser Ser Leu Gly Ile Thr Ala Leu Trp Leu Pro Pro Ala Tyr Lys  
35 40 45

Gly Thr Ser Arg Ser Asp Val Gly Tyr Gly Val Tyr Asp Leu Tyr Asp  
50 55 60

Leu Gly Glu Phe Asn Gln Lys Gly Thr Val Arg Thr Lys Tyr Gly Thr  
65 70 75 80

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Lys Ala Gln Tyr Leu Gln Ala Ile Gln Ala Ala His Ala Ala Gly Met  
85 90 95

Gln Val Tyr Ala Asp Val Val Phe Asp His Lys Gly Gly Ala Asp Gly  
100 105 110

Thr Glu Trp Val Asp Ala Val Glu Val Asn Pro Ser Asp Arg Asn Gln  
115 120 125

Glu Ile Ser Gly Thr Tyr Gln Ile Gln Ala Trp Thr Lys Phe Asp Phe  
130 135 140

Pro Gly Arg Gly Asn Thr Tyr Ser Ser Phe Lys Trp Arg Trp Tyr His  
145 150 155 160

Phe Asp Gly Val Asp Trp Asp Glu Ser Arg Lys Leu Ser Arg Ile Tyr  
165 170 175

Lys Phe Arg Gly Ile Gly Lys Ala Trp Asp Trp Glu Val Asp Thr Glu  
180 185 190

Asn Gly Asn Tyr Asp Tyr Leu Met Tyr Ala Asp Leu Asp Met Asp His  
195 200 205

Pro Glu Val Val Thr Glu Leu Lys Asn Trp Gly Lys Trp Tyr Val Asn  
210 215 220

Thr Thr Asn Ile Asp Gly Phe Arg Leu Asp Ala Val Lys His Ile Lys  
225 230 235 240

Phe Gln Phe Phe Pro Asp Trp Leu Ser Tyr Val Arg Ser Gln Thr Gly  
245 250 255

Lys Pro Leu Phe Thr Val Gly Glu Tyr Trp Ser Tyr Asp Ile Asn Lys  
260 265 270

Leu His Asn Tyr Ile Thr Lys Thr Asn Gly Thr Met Ser Leu Phe Asp  
275 280 285

Ala Pro Leu His Asn Lys Phe Tyr Thr Ala Ser Lys Ser Gly Gly Ala  
290 295 300

Phe Asp Met Arg Thr Leu Met Thr Asn Thr Leu Met Lys Asp Gln Pro



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<220>
<221> VARIANT
<222> (1)..(1)
<223> /replace=" "

<220>
<221> VARIANT
<222> (2)..(2)
<223> /replace="His"

<220>
<221> VARIANT
<222> (3)..(3)
<223> /replace="Asn"

<220>
<221> VARIANT
<222> (4)..(4)
<223> /replace="Gly"

<220>
<221> VARIANT
<222> (5)..(5)
<223> /replace="Thr"

<220>
<221> misc_feature
<222> (1)..(5)
<223> /note="Residues given in the sequence have no preference
with respect to those in the annotation for said positions"

<220>
<221> VARIANT
<222> (19)..(19)
<223> /replace="Asn"

<220>
<221> misc_feature
<222> (19)..(19)
<223> /note="Residues given in the sequence have no preference
with respect to those in the annotation for said positions"

<220>
<221> VARIANT
<222> (22)..(22)
<223> /replace="Asn"

<220>
<221> VARIANT
<222> (23)..(23)
<223> /replace="His"

<220>
<221> misc_feature
<222> (22)..(23)
<223> /note="Residues given in the sequence have no preference"
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```
with respect to those in the annotation for said positions"
<220>
<221> VARIANT
<222> (25)..(25)
<223> /replace="Asn"

<220>
<221> misc_feature
<222> (25)..(25)
<223> /note="Residue given in the sequence has no preference
with respect to those in the annotation for said position"

<220>
<221> VARIANT
<222> (28)..(28)
<223> /replace="Asn"

<220>
<221> VARIANT
<222> (29)..(29)
<223> /replace="Ser"

<220>
<221> misc_feature
<222> (28)..(29)
<223> /note="Residues given in the sequence have no preference
with respect to those in the annotation for said positions"

<220>
<221> VARIANT
<222> (32)..(32)
<223> /replace="Ser"

<220>
<221> misc_feature
<222> (32)..(32)
<223> /note="Residue given in the sequence has no preference
with respect to those in the annotation for said position"

<220>
<221> VARIANT
<222> (35)..(35)
<223> /replace="Lys"

<220>
<221> misc_feature
<222> (35)..(35)
<223> /note="Residue given in the sequence has no preference
with respect to those in the annotation for said position"

<220>
<221> VARIANT
<222> (37)..(37)
<223> /replace="Lys"

<220>
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<221> misc_feature
<222> (37)..(37)
<223> /note="Residue given in the sequence has no preference
with respect to those in the annotation for said position"

<220>
<221> VARIANT
<222> (51)..(51)
<223> /replace="Ala"

<220>
<221> misc_feature
<222> (51)..(51)
<223> /note="Residue given in the sequence has no preference
with respect to those in the annotation for said position"

<220>
<221> VARIANT
<222> (53)..(53)
<223> /replace="Gln"

<220>
<221> VARIANT
<222> (54)..(54)
<223> /replace="Asn"

<220>
<221> misc_feature
<222> (53)..(54)
<223> /note="Residues given in the sequence have no preference
with respect to those in the annotation for said positions"

<220>
<221> VARIANT
<222> (60)..(60)
<223> /replace="Ala"

<220>
<221> misc_feature
<222> (60)..(60)
<223> /note="Residue given in the sequence has no preference
with respect to those in the annotation for said position"

<220>
<221> VARIANT
<222> (85)..(85)
<223> /replace="Leu"

<220>
<221> VARIANT
<222> (86)..(86)
<223> /replace="Gln"

<220>
<221> VARIANT
<222> (87)..(87)
<223> /replace="Ala"
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<220>  
<221> misc\_feature  
<222> (85)..(87)  
<223> /note="Residues given in the sequence have no preference  
with respect to those in the annotation for said positions"

<220>  
<221> VARIANT  
<222> (90)..(90)  
<223> /replace="Thr"

<220>  
<221> misc\_feature  
<222> (90)..(90)  
<223> /note="Residue given in the sequence has no preference  
with respect to those in the annotation for said position"

<220>  
<221> VARIANT  
<222> (92)..(92)  
<223> /replace="Leu"

<220>  
<221> VARIANT  
<222> (93)..(93)  
<223> /replace="Lys"

<220>  
<221> VARIANT  
<222> (94)..(95)  
<223> /replace="Asn"

<220>  
<221> misc\_feature  
<222> (92)..(95)  
<223> /note="Residues given in the sequence have no preference  
with respect to those in the annotation for said positions"

<220>  
<221> VARIANT  
<222> (105)..(105)  
<223> /replace="Met"

<220>  
<221> VARIANT  
<222> (106)..(106)  
<223> /replace="Asn"

<220>  
<221> misc\_feature  
<222> (105)..(106)  
<223> /note="Residues given in the sequence have no preference  
with respect to those in the annotation for said positions"

<220>  
<221> VARIANT

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<222> (116)..(116)
<223> /replace="Met "

<220>
<221> misc_feature
<222> (116)..(116)
<223> /note="Residue given in the sequence has no preference
with respect to those in the annotation for said position"

<220>
<221> VARIANT
<222> (118)..(118)
<223> /replace="Arg"

<220>
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<222> (118)..(118)
<223> /note="Residue given in the sequence has no preference
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<220>
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<222> (125)..(125)
<223> /replace="Asn"

<220>
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<223> /replace="Asn"

<220>
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<220>
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<223> /replace="Glu"

<220>
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<220>
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<223> /replace="Thr"

<220>
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<222> (136)..(136)
<223> /note="Residue given in the sequence has no preference
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<220>  
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<220>  
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<220>  
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<223> /replace="Asn"

<220>  
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<222> (175)..(175)  
<223> /replace=" "

<220>  
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<222> (174)..(175)  
<223> /note="Residues given in the sequence have no preference  
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<220>  
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<223> /replace="His"

<220>  
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<222> (183)..(183)  
<223> /note="Residue given in the sequence has no preference  
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<220>  
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<223> /replace="Asn"

<220>  
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<220>
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<222> (222)..(222)
<223> /replace="Val"

<220>
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<220>
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<220>
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<220>
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<220>
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<222> (229)..(229)
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<223> /replace="Thr"

<220>
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<222> (247)..(247)
<223> /replace="Arg"

<220>
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<222> (246)..(247)
<223> /note="Residues given in the sequence have no preference
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<223> /replace="Asn"

<220>  
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<222> (251)..(251)  
<223> /note="Residue given in the sequence has no preference  
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<220>  
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<223> /replace="Ala"

<220>  
<221> misc\_feature  
<222> (256)..(256)  
<223> /note="Residue given in the sequence has no preference  
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<222> (260)..(260)  
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<223> /replace="Ala"

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<223> /replace="Lys"

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<222> (270)..(270)  
<223> /replace="Asn"

<220>  
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<223> /replace="Gly"

<220>
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<222> (274)..(274)
<223> /replace="Ala"

<220>
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<223> /replace="Glu"

<220>
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<222> (276)..(276)
<223> /note="Residue given in the sequence has no preference
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<222> (280)..(280)
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<220>
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<222> (284)..(284)
<223> /replace="Trp"

<220>
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<222> (285)..(285)
<223> /replace="Asn"

<220>
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<222> (286)..(286)
<223> /replace="His"
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<220>  
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<223> /note="Residues given in the sequence have no preference  
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<220>  
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<223> /replace="Val"

<220>  
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<222> (291)..(291)  
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<220>  
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<222> (295)..(295)  
<223> /replace="Tyr"

<220>  
<221> VARIANT  
<222> (296)..(296)  
<223> /replace="Asn"

<220>  
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<222> (297)..(297)  
<223> /replace="Leu"

<220>  
<221> misc\_feature  
<222> (295)..(297)  
<223> /note="Residues given in the sequence have no preference  
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<220>  
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<223> /replace="Asn"

<220>  
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<222> (299)..(299)  
<223> /note="Residue given in the sequence has no preference  
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<223> /replace="Asn"

<220>  
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<222> (306)..(306)



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<223> /note="Residue given in the sequence has no preference  
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<222> (311)..(311)

<223> /replace="Asn"

<220>

<221> misc\_feature

<222> (311)..(311)

<223> /note="Residue given in the sequence has no preference  
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<222> (313)..(313)

<223> /replace="Phe"

<220>

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<222> (314)..(314)

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<222> (315)..(315)

<223> /replace="Gly"

<220>

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<222> (313)..(315)

<223> /note="Residues given in the sequence have no preference  
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<222> (319)..(319)

<223> /replace="Gln"

<220>

<221> VARIANT

<222> (320)..(320)

<223> /replace="Arg"

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<222> (321)..(321)

<223> /replace="His"

<220>

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<222> (319)..(321)

<223> /note="Residues given in the sequence have no preference  
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<220>  
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<223> /replace="Gln"

<220>  
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<222> (335)..(335)  
<223> /note="Residue given in the sequence has no preference  
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<220>  
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<223> /replace="Glu"

<220>  
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<222> (338)..(338)  
<223> /replace="Glu"

<220>  
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<222> (337)..(338)  
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<223> /replace="Glu"

<220>  
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<220>  
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<223> /replace="Glu"

<220>  
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<222> (346)..(346)  
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with respect to those in the annotation for said position"
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<223> /replace="Leu"

<220>
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<222> (356)..(356)
<223> /replace="Thr"

<220>
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<222> (355)..(356)
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<220>
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<223> /replace="Glu"

<220>
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<222> (361)..(361)
<223> /replace="Gln"

<220>
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<222> (365)..(365)
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<222> (376)..(376)
<223> /replace="Thr"

<220>
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<222> (376)..(376)
<223> /note="Residue given in the sequence has no preference
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<220>  
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<223> /replace="Glu"

<220>  
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<222> (391)..(391)  
<223> /note="Residue given in the sequence has no preference  
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<222> (394)..(394)  
<223> /replace="Gln"

<220>  
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<222> (395)..(395)  
<223> /replace="Lys"

<220>  
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<222> (394)..(395)  
<223> /note="Residues given in the sequence have no preference  
with respect to those in the annotation for said positions"

<220>  
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<223> /replace="Lys"

<220>  
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<222> (400)..(400)  
<223> /note="Residue given in the sequence has no preference  
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<220>  
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<223> /replace="Asn"

<220>  
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<223> /note="Residue given in the sequence has no preference"

with respect to those in the annotation for said position"

<220>  
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<223> /replace="His"

<220>  
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<222> (409)..(409)  
<223> /replace="Asn"

<220>  
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<222> (408)..(409)  
<223> /note="Residues given in the sequence have no preference  
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<220>  
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<223> /replace="Asn"

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<223> /replace="Ala"

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<223> /replace="His"

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with respect to those in the annotation for said positions"

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<223> /replace="Asn"

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<223> /note="Residue given in the sequence has no preference  
with respect to those in the annotation for said position"

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<220>
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<223> /replace="Ala"

<220>
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with respect to those in the annotation for said position"

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<223> /replace="Lys"

<220>
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<223> /replace="Gln"

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<223> /note="Residue given in the sequence has no preference
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<223> /replace="Ser"

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<222> (452)..(452)
<223> /note="Residue given in the sequence has no preference
with respect to those in the annotation for said position"

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<222> (460)..(460)
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<223> /replace="Gly"
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<223> /note="Residue given in the sequence has no preference
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<220>
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<222> (473)..(473)
<223> /replace="Ser"

<220>
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<222> (473)..(473)
<223> /note="Residue given in the sequence has no preference
with respect to those in the annotation for said position"

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<222> (484)..(484)
<223> /replace="Asn"

<220>
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<222> (484)..(484)
<223> /note="Residue given in the sequence has no preference
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<222> (486)..(486)
<223> /replace=" "

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<223> /replace=" "

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<223> /replace=" "

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<223> /replace=" "

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<223> /replace=" "

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<222> (512)..(512)

<223> /replace=" "

<220>

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<222> (513)..(513)

<223> /replace=" "

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<223> /replace=" "

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<223> /replace=" "

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His Ala Ala Pro Phe Asn Gly Thr Met Met Gln Tyr Phe Glu Trp Tyr
1          5          10          15

Leu Pro Asp Asp Gly Thr Leu Trp Thr Lys Leu Ala Asn Asp Ala Asn
20          25          30

Asn Leu Ser Ser Leu Gly Ile Thr Ala Leu Trp Ile Pro Pro Ala Trp
35          40          45

Lys Gly Thr Ser Arg Ser Asp Val Gly Tyr Gly Val Tyr Asp Leu Tyr
50          55          60

Asp Leu Gly Glu Phe Asn Gln Lys Gly Thr Val Arg Thr Lys Tyr Gly
65          70          75

Thr Lys Ala Gln Tyr Leu Gln Ala Ile Gln Ala Ala His Ala Ala Gly
85          90          95

Ile Gln Val Tyr Ala Asp Val Val Phe Asp His Lys Gly Gly Ala Asp
100         105         110

Ala Thr Glu Trp Val Asp Ala Val Glu Val Asn Pro Ser Asp Arg Asn
115         120         125

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Gln Glu Ile Ser Gly Thr Tyr Gln Ile Gln Ala Trp Thr Lys Phe Asp  
130 135 140

Phe Pro Gly Arg Gly Asn Thr His Ser Ser Phe Lys Trp Arg Trp Tyr  
145 150 155 160

His Phe Asp Gly Val Asp Trp Asp Glu Ser Arg Lys Leu Ser Asn Arg  
165 170 175

Ile Tyr Lys Phe Arg Gly Ile Gly Lys Ala Trp Asp Trp Glu Val Asp  
180 185 190

Thr Glu Asn Gly Asn Tyr Asp Tyr Leu Met Tyr Ala Asp Ile Asp Met  
195 200 205

Asp His Pro Glu Val Val Thr Glu Leu Lys Asn Trp Gly Lys Trp Tyr  
210 215 220

Val Asn Thr Thr Asn Ile Asp Gly Phe Arg Ile Asp Ala Val Lys His  
225 230 235 240

Ile Lys Phe Ser Phe Phe Pro Asp Trp Ile Ser His Val Arg Ser Gln  
245 250 255

Thr Gly Lys Pro Leu Phe Thr Val Ala Glu Phe Trp Ser Tyr Asp Ile  
260 265 270

Asn Lys Ile His Asn Tyr Ile Thr Lys Thr Asn Gly Thr Met Ser Leu  
275 280 285

Phe Asp Ala Pro Leu His Asn Lys Phe Tyr Thr Ala Ser Lys Ser Gly  
290 295 300

Gly Ala Phe Asp Met Arg Thr Ile Met Thr Asn Thr Leu Met Lys Asp  
305 310 315 320

Gln Pro Ser Leu Ala Val Thr Phe Val Asp Asn His Asp Ser Glu Pro  
325 330 335

Gly Gln Ala Leu Gln Ser Phe Val Asp Pro Trp Phe Lys Pro Leu Ala  
340 345 350

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Tyr Ala Phe Ile Leu Thr Arg Gln Glu Gly Tyr Pro Cys Val Phe Tyr  
355 360 365

Gly Asp Tyr Tyr Gly Ile Pro Gln His Asn Ile Pro Ala Leu Lys Ser  
370 375 380

Lys Ile Asp Pro Ile Leu Ile Ala Arg Arg Asp Tyr Ala Tyr Gly Thr  
385 390 395 400

Gln His Asp Tyr Leu Asp His Ser Asp Ile Ile Gly Trp Thr Arg Glu  
405 410 415

Gly Val Thr Glu Lys Pro Gly Ser Gly Leu Ala Ala Ile Ile Ser Asp  
420 425 430

Gly Pro Gly Gly Ser Lys Trp Met Phe Val Gly Lys Asn His Ala Gly  
435 440 445

Lys Val Phe Tyr Asp Ile Thr Gly Asn Arg Ser Asp Thr Val Thr Ile  
450 455 460

Asn Ala Asp Gly Trp Gly Glu Phe Lys Val Asn Gly Gly Ser Val Ser  
465 470 475 480

Ile Trp Val Pro Lys Lys Thr Thr Val Ser Thr Ile Ala Arg Pro Ile  
485 490 495

Thr Thr Arg Pro Trp Thr Gly Glu Phe Val Arg Trp Thr Glu Pro Arg  
500 505 510

Leu Val Ala Trp Pro  
515

<210> 23

<211> 518

<212> PRT

<213> Artificial sequence

<220>

<221> source

<223> /note="Description of artificial sequence: Synthetic  
consensus sequence"

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<222> (283)..(283)
<223> /replace="Asn"

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<222> (284)..(284)
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<223> /replace="Ala"

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<222> (301)..(301)
<223> /replace="Gln"

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<223> /replace="Ser"
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<223> /replace="Ser"

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<223> /replace="Met "

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<222> (373)..(373)
<223> /replace="Lys"

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<223> /replace=" "

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<223> /replace=" "

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<223> /replace=" "

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<223> /replace="Arg"
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<223> /replace="Glu"

<220>  
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<223> /replace="His"

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<223> /replace="Lys"

<220>  
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<222> (392)..(392)  
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<223> /replace="Gln"

<220>  
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<222> (396)..(396)  
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<223> /replace="Ala"

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<223> /replace="His"

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<222> (421)..(421)
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<220>
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<222> (422)..(422)
<223> /replace="Val"

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<223> /replace="Ala"

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<223> /replace="Asn"

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<223> /replace="Arg"

<220>  
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<223> /replace="Asn"

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<223> /replace=" "

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<220>  
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Pro Asp Asp Gly Thr Leu Trp Thr Lys Leu Ala Asn Asp Ala Asn Asn  
20 25 30

Leu Ala Ser Leu Gly Ile Thr Ala Leu Trp Ile Pro Pro Ala Tyr Lys  
35 40 45

Gly Thr Ser Arg Ala Asp Val Gly Tyr Gly Val Tyr Asp Leu Tyr Asp  
50 55 60

Leu Gly Glu Phe Asn Gln Lys Gly Thr Val Arg Thr Lys Tyr Gly Thr  
65 70 75 80

Lys Ala Gln Tyr Leu Gln Ala Ile Gln Ala Ala His Ala Ala Gly Ile  
85 90 95

Asn Val Tyr Ala Asp Val Val Phe Asp His Lys Gly Gly Ala Asp Ala  
100 105 110

Thr Glu Trp Val Asp Ala Val Glu Val Asn Pro Ala Asp Arg Asn Gln  
115 120 125

Glu Ile Ser Gly Thr His Gln Ile Gln Ala Trp Thr Lys Phe Asp Phe  
130 135 140

Pro Gly Arg Gly Asn Thr Tyr Ser Ser Phe Lys Trp Arg Trp Tyr His  
145 150 155 160

Phe Asp Gly Val Asp Trp Asp Glu Ser Arg Lys Leu Ser Arg Ile Tyr  
165 170 175

Lys Phe Arg Gly Ile Gly Lys Ala Trp Asp Trp Glu Val Asp Thr Glu

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180 185 190

Asn Gly Asn Tyr Asp Tyr Leu Met Tyr Ala Asp Ile Asp Met Asp His  
195 200 205

Pro Asp Val Val Thr Glu Ile Lys Asn Trp Gly Lys Trp Tyr Val Asn  
210 215 220

Thr Thr Asn Ile Asp Gly Phe Arg Leu Asp Ala Val Lys His Ile Lys  
225 230 235 240

Phe Ser Phe Phe Pro Asp Trp Leu Ser His Val Arg Ser Gln Thr Gly  
245 250 255

Lys Pro Leu Phe Thr Val Ala Glu Tyr Trp Ser Tyr Asp Ile Asn Lys  
260 265 270

Leu His Asn Tyr Ile Thr Lys Thr Asn Gly Thr Met Ser Leu Phe Asp  
275 280 285

Ala Pro Leu His Asn Lys Phe His Thr Ala Ser Lys Ser Gly Gly Ala  
290 295 300

Phe Asp Met Arg Thr Leu Leu Thr Asn Thr Leu Met Lys Asp Gln Pro  
305 310 315 320

Thr Leu Ala Val Thr Phe Val Asp Asn His Asp Thr Glu Pro Gly Gln  
325 330 335

Ala Leu Gln Ser Trp Val Asp Pro Trp Phe Lys Pro Leu Ala Tyr Ala  
340 345 350

Phe Ile Leu Thr Arg Gln Glu Gly Tyr Pro Cys Val Phe Tyr Gly Asp  
355 360 365

Tyr Tyr Gly Ile Pro Gly Asp Ser Gln Tyr Asn Ile Pro Ala Leu Lys  
370 375 380

Ser Lys Ile Asp Pro Ile Leu Ile Ala Arg Lys Asp Tyr Ala Tyr Gly  
385 390 395 400

Thr Gln His Asp Tyr Leu Asp His Ser Asp Ile Ile Gly Trp Thr Arg  
405 410 415

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Glu Gly Val Ser Glu Lys Pro Gly Ser Gly Leu Ala Ala Leu Ile Thr  
420 425 430

Asp Gly Pro Gly Gly Ala Lys Trp Met Tyr Val Gly Lys Gln His Ala  
435 440 445

Gly Lys Val Phe His Asp Ile Thr Gly Asn Arg Ser Asp Thr Val Thr  
450 455 460

Ile Asn Ser Asp Gly Trp Gly Glu Phe Lys Val Asn Gly Gly Ser Val  
465 470 475 480

Ser Ile Trp Val Pro Arg Lys Thr Thr Val Ser Thr Ile Ala Arg Pro  
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Ile Thr Thr Arg Pro Trp Thr Gly Glu Phe Val Arg Trp Thr Glu Pro  
500 505 510

Arg Leu Val Ala Trp Pro  
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<221> misc\_feature  
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<220>  
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<223> /replace="Thr"

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<220>  
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<223> /replace="His"

<220>  
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<222> (21)..(22)  
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<223> /replace="Lys"

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<220>  
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<222> (27)..(27)  
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<222> (27)..(27)  
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<223> /replace="Glu"

<220>  
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<223> /replace="His"

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<223> /replace="Asp"

<220>  
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<223> /replace="Glu"

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<222> (84)..(84)  
<223> /replace="Leu"

<220>  
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<223> /replace="Gln"

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<223> /replace="Asp"

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<220>  
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<223> /note="Residue given in the sequence has no preference  
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<223> /replace="Leu"

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<222> (91)..(91)

<223> /note="Residue given in the sequence has no preference  
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<223> /replace="Arg"

<220>

<221> VARIANT

<222> (95)..(95)

<223> /replace="Asn"

<220>

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<222> (94)..(95)

<223> /note="Residues given in the sequence have no preference  
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<222> (104)..(104)

<223> /replace="Leu"

<220>

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<222> (105)..(105)

<223> /replace="Asn"

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<222> (104)..(105)

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<223> /replace="Asp"

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<222> (115)..(115)

<223> /note="Residue given in the sequence has no preference  
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<223> /replace="Thr"

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<223> /replace="Asn"

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<223> /replace="Thr"

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<223> /replace="Glu"

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<223> /replace="Glu"

<220>  
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<222> (132)..(133)  
<223> /note="Residues given in the sequence have no preference  
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<223> /replace="Lys"

<220>  
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<222> (137)..(137)

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<223> /note="Residue given in the sequence has no preference  
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<222> (141)..(141)

<223> /replace="Asp"

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<223> /replace="Arg"

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<222> (143)..(143)

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<223> /replace="Asp"

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<222> (153)..(153)

<223> /note="Residue given in the sequence has no preference  
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<222> (157)..(157)

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<222> (157)..(157)

<223> /note="Residue given in the sequence has no preference  
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<222> (164)..(164)

<223> /replace="Ala"

<220>

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<222> (164)..(164)

<223> /note="Residue given in the sequence has no preference  
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<223> /replace="Glu"

<220>  
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<222> (181)..(181)  
<223> /note="Residue given in the sequence has no preference  
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<223> /replace="Ser"

<220>  
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<222> (190)..(190)  
<223> /note="Residue given in the sequence has no preference  
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<223> /replace="Tyr"

<220>  
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<222> (206)..(206)  
<223> /note="Residue given in the sequence has no preference  
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<223> /replace="Ala"

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<223> /replace="Thr"

<220>  
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<220>  
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<222> (217)..(217)  
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<223> /replace="Ile"

<220>  
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<222> (220)..(220)  
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with respect to those in the annotation for said position"

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<222> (223)..(223)  
<223> /replace="Ala"

<220>  
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<222> (223)..(223)  
<223> /note="Residue given in the sequence has no preference  
with respect to those in the annotation for said position"

<220>  
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<222> (225)..(225)  
<223> /replace="Glu"

<220>  
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<222> (226)..(226)  
<223> /replace="Leu"

<220>  
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<223> /replace="Ser"

<220>  
<221> misc\_feature  
<222> (225)..(227)  
<223> /note="Residues given in the sequence have no preference  
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<220>
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<222> (236)..(236)
<223> /note="Residue given in the sequence has no preference
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<220>
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<222> (244)..(244)
<223> /replace="Leu"

<220>
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<222> (245)..(245)
<223> /replace="Arg"

<220>
<221> misc_feature
<222> (244)..(245)
<223> /note="Residues given in the sequence have no preference
with respect to those in the annotation for said positions"

<220>
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<223> /replace="Gln"

<220>
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<222> (250)..(250)
<223> /replace="Ala"

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<220>
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<223> /replace="Gln"

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<223> /replace="Ala"

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<222> (253)..(254)
<223> /note="Residues given in the sequence have no preference
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<222> (258)..(258)  
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<220>  
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<222> (258)..(258)  
<223> /note="Residue given in the sequence has no preference  
with respect to those in the annotation for said position"

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<223> /replace="Gln"

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<222> (268)..(268)  
<223> /replace="Asn"

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<222> (269)..(269)  
<223> /replace="Asn"

<220>  
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<222> (270)..(270)  
<223> /replace="Ala"

<220>  
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<222> (271)..(271)  
<223> /replace="Gly"

<220>  
<221> misc\_feature  
<222> (267)..(271)  
<223> /note="Residues given in the sequence have no preference  
with respect to those in the annotation for said positions"

<220>  
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<223> /replace="Glu"

<220>  
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<223> /note="Residue given in the sequence has no preference  
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<223> /replace="Ser"

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<222> (282)..(282)
<223> /replace="Phe"

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<223> /replace="Asn"

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<223> /replace="Gln"

<220>
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<222> (281)..(284)
<223> /note="Residues given in the sequence have no preference
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<222> (289)..(289)
<223> /replace="Val"

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<222> (289)..(289)
<223> /note="Residue given in the sequence has no preference
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<222> (293)..(293)
<223> /replace="Phe"

<220>
<221> VARIANT
<222> (294)..(294)
<223> /replace="Asn"

<220>
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<222> (295)..(295)
<223> /replace="Leu"
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<223> /replace="Gln"

<220>  
<221> VARIANT  
<222> (297)..(297)  
<223> /replace="Ala"

<220>  
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<222> (293)..(297)  
<223> /note="Residues given in the sequence have no preference  
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<220>  
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<223> /replace="Ser"

<220>  
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<223> /replace="Gln"

<220>  
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with respect to those in the annotation for said positions"

<220>  
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<223> /replace="Arg"

<220>  
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<222> (309)..(309)  
<223> /note="Residue given in the sequence has no preference  
with respect to those in the annotation for said position"

<220>  
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<223> /replace="Asp"

<220>  
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<222> (313)..(313)  
<223> /replace="Gly"

<220>  
<221> misc\_feature  
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<223> /note="Residues given in the sequence have no preference"



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with respect to those in the annotation for said positions"

<220>  
<221> VARIANT  
<222> (317)..(317)  
<223> /replace="Ser"

<220>  
<221> VARIANT  
<222> (318)..(318)  
<223> /replace="Arg"

<220>  
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<222> (319)..(319)  
<223> /replace="His"

<220>  
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<220>  
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<223> /replace="Glu"

<220>  
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<223> /replace="Lys"

<220>  
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<222> (321)..(322)  
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<220>  
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<222> (333)..(333)  
<223> /replace="Gln"

<220>  
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<222> (333)..(333)  
<223> /note="Residue given in the sequence has no preference  
with respect to those in the annotation for said position"

<220>  
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<222> (339)..(339)  
<223> /replace="Glu"

<220>  
<221> misc\_feature

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<222> (339)..(339)
<223> /note="Residue given in the sequence has no preference
with respect to those in the annotation for said position"

<220>
<221> VARIANT
<222> (341)..(341)
<223> /replace="Thr"

<220>
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<222> (341)..(341)
<223> /note="Residue given in the sequence has no preference
with respect to those in the annotation for said position"

<220>
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<222> (343)..(343)
<223> /replace="Gln"

<220>
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<222> (344)..(344)
<223> /replace="Thr"

<220>
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<222> (343)..(344)
<223> /note="Residues given in the sequence have no preference
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<220>
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<223> /replace="Glu"

<220>
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<222> (359)..(359)
<223> /replace="Ser"

<220>
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<222> (358)..(359)
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with respect to those in the annotation for said positions"

<220>
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<223> /replace="Gln"

<220>
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<222> (363)..(363)
<223> /note="Residue given in the sequence has no preference
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<220>  
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<222> (369)..(369)  
<223> /replace="Met "

<220>  
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<222> (369)..(369)  
<223> /note="Residue given in the sequence has no preference  
with respect to those in the annotation for said position"

<220>  
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<222> (372)..(372)  
<223> /replace="Thr "

<220>  
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<222> (373)..(373)  
<223> /replace="Lys "

<220>  
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<222> (374)..(374)  
<223> /replace="Gly "

<220>  
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<222> (375)..(375)  
<223> /replace="Thr "

<220>  
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<222> (376)..(376)  
<223> /replace="Ser "

<220>  
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<222> (377)..(377)  
<223> /replace=" "

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<223> /replace=" "

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<222> (379)..(379)  
<223> /replace=" "

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<222> (372)..(379)  
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with respect to those in the annotation for said positions"

<220>  
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<222> (385)..(385)  
<223> /replace="Asp"

<220>  
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<222> (386)..(386)  
<223> /replace="Asn"

<220>  
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<222> (385)..(386)  
<223> /note="Residues given in the sequence have no preference  
with respect to those in the annotation for said positions"

<220>  
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<222> (392)..(392)  
<223> /replace="Lys"

<220>  
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<222> (392)..(392)  
<223> /note="Residue given in the sequence has no preference  
with respect to those in the annotation for said position"

<220>  
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<222> (401)..(401)  
<223> /replace="Pro"

<220>  
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<222> (401)..(401)  
<223> /note="Residue given in the sequence has no preference  
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<220>  
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<222> (409)..(409)  
<223> /replace="Pro"

<220>  
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<222> (409)..(409)  
<223> /note="Residue given in the sequence has no preference  
with respect to those in the annotation for said position"

<220>  
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<223> /replace="Asp"

<220>  
<221> misc\_feature

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<222> (419)..(419)
<223> /note="Residue given in the sequence has no preference
with respect to those in the annotation for said position"

<220>
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<222> (421)..(421)
<223> /replace="Ser"

<220>
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<222> (422)..(422)
<223> /replace="Ala"

<220>
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<222> (423)..(423)
<223> /replace="Ala"

<220>
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<222> (424)..(424)
<223> /replace="Lys"

<220>
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<222> (421)..(424)
<223> /note="Residues given in the sequence have no preference
with respect to those in the annotation for said positions"

<220>
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<222> (440)..(440)
<223> /replace="Arg"

<220>
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<222> (440)..(440)
<223> /note="Residue given in the sequence has no preference
with respect to those in the annotation for said position"

<220>
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<222> (443)..(443)
<223> /replace="Ala"

<220>
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<222> (443)..(443)
<223> /note="Residue given in the sequence has no preference
with respect to those in the annotation for said position"

<220>
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<222> (445)..(445)
<223> /replace="Leu"
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<220>  
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<222> (446)..(446)  
<223> /replace="Lys"

<220>  
<221> VARIANT  
<222> (447)..(447)  
<223> /replace="Asn"

<220>  
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<222> (445)..(447)  
<223> /note="Residues given in the sequence have no preference  
with respect to those in the annotation for said positions"

<220>  
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<222> (450)..(450)  
<223> /replace="Glu"

<220>  
<221> VARIANT  
<222> (451)..(451)  
<223> /replace="Thr"

<220>  
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<222> (450)..(451)  
<223> /note="Residues given in the sequence have no preference  
with respect to those in the annotation for said positions"

<220>  
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<222> (464)..(464)  
<223> /replace="Lys"

<220>  
<221> misc\_feature  
<222> (464)..(464)  
<223> /note="Residue given in the sequence has no preference  
with respect to those in the annotation for said position"

<220>  
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<222> (466)..(466)  
<223> /replace="Gly"

<220>  
<221> misc\_feature  
<222> (466)..(466)  
<223> /note="Residue given in the sequence has no preference  
with respect to those in the annotation for said position"

<220>  
<221> VARIANT  
<222> (474)..(474)

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<223> /replace="His"
<220>
<221> misc_feature
<222> (474)..(474)
<223> /note="Residue given in the sequence has no preference
with respect to those in the annotation for said position"

<220>
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<222> (477)..(477)
<223> /replace="Asp"

<220>
<221> misc_feature
<222> (477)..(477)
<223> /note="Residue given in the sequence has no preference
with respect to those in the annotation for said position"

<220>
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<222> (485)..(485)
<223> /replace="Gln"

<220>
<221> misc_feature
<222> (485)..(485)
<223> /note="Residue given in the sequence has no preference
with respect to those in the annotation for said position"

<220>
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<222> (487)..(487)
<223> /replace=" "

<220>
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<222> (488)..(489)
<223> /replace=" "

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<223> /replace=" "

<220>
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<222> (491)..(491)
<223> /replace=" "

<220>
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<222> (492)..(492)
<223> /replace=" "

<220>
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<223> /replace=" "

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<222> (495)..(495)  
<223> /replace=" "

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<223> /replace=" "

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<223> /replace=" "

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<223> /replace=" "

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<223> /replace=" "

<220>  
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<222> (501)..(501)  
<223> /replace=" "

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<222> (502)..(502)  
<223> /replace=" "

<220>  
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<222> (503)..(503)  
<223> /replace=" "

<220>  
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<222> (504)..(504)  
<223> /replace=" "

<220>  
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<222> (505)..(505)  
<223> /replace=" "



<220>  
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<222> (506)..(506)  
<223> /replace=" "

<220>  
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<222> (507)..(507)  
<223> /replace=" "

<220>  
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<222> (508)..(508)  
<223> /replace=" "

<220>  
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<222> (509)..(509)  
<223> /replace=" "

<220>  
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<223> /replace=" "

<220>  
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<222> (511)..(511)  
<223> /replace=" "

<220>  
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<222> (512)..(512)  
<223> /replace=" "

<220>  
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<222> (513)..(513)  
<223> /replace=" "

<220>  
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<222> (514)..(514)  
<223> /replace=" "

<220>  
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<223> /replace=" "

<220>  
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<223> /replace=" "

<220>

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<221> VARIANT  
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 <223> /replace=" "

<220>  
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 <222> (518)..(518)  
 <223> /replace=" "

<220>  
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 <222> (487)..(518)  
 <223> /note="Residues given in the sequence have no preference  
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<400> 24  
 Ala Ala Pro Phe Asn Gly Thr Leu Met Gln Tyr Phe Glu Trp Tyr Leu  
 1 5 10 15

Pro Asp Asp Gly Thr Leu Trp Thr Lys Leu Ala Asn Asp Ala Asn Asn  
 20 25 30

Leu Ser Ser Ile Gly Ile Thr Ala Leu Trp Ile Pro Pro Ala Tyr Lys  
 35 40 45

Gly Thr Ser Arg Ser Asp Val Gly Tyr Gly Val Tyr Asp Leu Tyr Asp  
 50 55 60

Leu Gly Glu Phe Asn Gln Lys Gly Thr Val Arg Thr Lys Tyr Gly Thr  
 65 70 75 80

Lys Ala Gln Tyr Leu Gln Ala Ile Gln Ala Ala His Ala Ala Gly Met  
 85 90 95

Gln Val Tyr Ala Asp Val Val Phe Asp His Lys Ala Gly Ala Asp Ala  
 100 105 110

Thr Glu Trp Val Asp Ala Val Glu Val Asn Pro Ala Asp Arg Asn Gln  
 115 120 125

Glu Ile Ser Gly Thr Tyr Gln Ile Gln Ala Trp Thr Lys Phe Asp Phe  
 130 135 140

Pro Gly Arg Gly Asn Thr Tyr Ser Ser Phe Lys Trp Arg Trp Tyr His  
 145 150 155 160

Phe Asp Gly Val Asp Trp Asp Glu Ser Arg Lys Ile Ser Arg Ile Phe

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				165						170						175
Lys	Phe	Arg	Gly	Ile	Gly	Lys	Ala	Trp	Asp	Trp	Glu	Val	Asp	Ser	Glu	
			180					185					190			
Asn	Gly	Asn	Tyr	Asp	Tyr	Leu	Met	Tyr	Ala	Asp	Leu	Asp	Met	Asp	His	
		195					200					205				
Pro	Asp	Val	Val	Thr	Glu	Leu	Lys	Asn	Trp	Gly	Lys	Trp	Tyr	Val	Asn	
	210					215					220					
Thr	Thr	Asn	Ile	Asp	Gly	Phe	Arg	Ile	Asp	Ala	Val	Lys	His	Ile	Lys	
225					230					235					240	
Phe	Ser	Phe	Phe	Pro	Asp	Trp	Leu	Ser	Tyr	Val	Arg	Ser	Gln	Thr	Gly	
				245					250					255		
Lys	Pro	Leu	Phe	Thr	Val	Ala	Glu	Tyr	Trp	Ser	Tyr	Asp	Ile	Asn	Lys	
			260					265					270			
Leu	His	Asn	Tyr	Ile	Thr	Lys	Thr	Asn	Gly	Thr	Met	Ser	Leu	Phe	Asp	
		275					280					285				
Ala	Pro	Leu	His	Asn	Lys	Phe	Tyr	Thr	Ala	Ser	Lys	Ser	Gly	Gly	Ala	
	290					295					300					
Phe	Asp	Met	Arg	Thr	Leu	Leu	Thr	Asn	Thr	Leu	Met	Lys	Asp	Gln	Pro	
305					310					315					320	
Thr	Leu	Ala	Val	Thr	Phe	Val	Asp	Asn	His	Asp	Thr	Glu	Pro	Gly	Gln	
				325					330					335		
Ala	Leu	Gln	Ser	Trp	Val	Asp	Pro	Trp	Phe	Lys	Pro	Leu	Ala	Tyr	Ala	
			340					345					350			
Phe	Ile	Leu	Thr	Arg	Gln	Glu	Gly	Tyr	Pro	Cys	Val	Phe	Tyr	Gly	Asp	
		355					360					365				
Tyr	Tyr	Gly	Ile	Pro	Gln	Tyr	Asn	Pro	Lys	Glu	Ile	Pro	Ser	Leu	Lys	
	370					375					380					
Ser	Lys	Ile	Asp	Pro	Ile	Leu	Ile	Ala	Arg	Lys	Asp	Tyr	Ala	Tyr	Gly	
385					390					395					400	

Thr Gln His Asp Tyr Ile Asp His Ser Asp Ile Ile Gly Trp Thr Arg  
 405 410 415

Glu Gly Val Ser Glu Lys Pro Gly Ser Gly Leu Ala Ala Leu Ile Thr  
 420 425 430

Asp Gly Pro Gly Gly Ser Lys Trp Met Tyr Val Gly Lys Gln His Ala  
 435 440 445

Gly Lys Val Phe Tyr Asp Ile Thr Gly Asn Arg Ser Asp Thr Val Thr  
 450 455 460

Ile Asn Ser Asp Gly Trp Gly Glu Phe Lys Val Asn Gly Gly Ser Val  
 465 470 475 480

Ser Ile Trp Val Pro Lys Lys Thr Thr Val Ser Thr Ile Ala Arg Pro  
 485 490 495

Ile Thr Thr Arg Pro Trp Thr Gly Glu Phe Val Arg Trp Thr Glu Pro  
 500 505 510

Arg Leu Val Ala Trp Pro  
 515

<210> 25  
 <211> 517  
 <212> PRT  
 <213> Artificial sequence

<220>  
 <221> source  
 <223> /note="Description of artificial sequence: Synthetic  
 consensus sequence"

<220>  
 <221> VARIANT  
 <222> (1)..(1)  
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 <223> /replace="His"

<220>  
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<222> (3)..(3)  
<223> /replace="Asn"

<220>  
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<222> (4)..(4)  
<223> /replace="Gly"

<220>  
<221> VARIANT  
<222> (5)..(5)  
<223> /replace="Thr"

<220>  
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<222> (1)..(5)  
<223> /note="Residues given in the sequence have no preference  
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<220>  
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<223> /replace="Asn"

<220>  
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<222> (19)..(19)  
<223> /note="Residue given in the sequence has no preference  
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<220>  
<221> VARIANT  
<222> (22)..(22)  
<223> /replace="Asn"

<220>  
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<222> (23)..(23)  
<223> /replace="His"

<220>  
<221> misc\_feature  
<222> (22)..(23)  
<223> /note="Residues given in the sequence have no preference  
with respect to those in the annotation for said positions"

<220>  
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<222> (25)..(25)  
<223> /replace="Asn"

<220>  
<221> misc\_feature  
<222> (25)..(25)  
<223> /note="Residue given in the sequence has no preference  
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<220>  
<221> VARIANT  
<222> (28)..(28)  
<223> /replace="Arg"

<220>  
<221> VARIANT  
<222> (29)..(29)  
<223> /replace="Ser"

<220>  
<221> misc\_feature  
<222> (28)..(29)  
<223> /note="Residues given in the sequence have no preference  
with respect to those in the annotation for said positions"

<220>  
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<222> (32)..(32)  
<223> /replace="Ser"

<220>  
<221> misc\_feature  
<222> (32)..(32)  
<223> /note="Residue given in the sequence has no preference  
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<220>  
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<222> (35)..(35)  
<223> /replace="Lys"

<220>  
<221> VARIANT  
<222> (36)..(36)  
<223> /replace="Asp"

<220>  
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<222> (37)..(37)  
<223> /replace="Lys"

<220>  
<221> misc\_feature  
<222> (35)..(37)  
<223> /note="Residues given in the sequence have no preference  
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<220>  
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<222> (51)..(51)  
<223> /replace="Ala"

<220>  
<221> misc\_feature  
<222> (51)..(51)  
<223> /note="Residue given in the sequence has no preference"

with respect to those in the annotation for said position"

<220>  
<221> VARIANT  
<222> (53)..(53)  
<223> /replace="Gln"

<220>  
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<222> (54)..(54)  
<223> /replace="Asn"

<220>  
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<222> (53)..(54)  
<223> /note="Residues given in the sequence have no preference  
with respect to those in the annotation for said positions"

<220>  
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<222> (60)..(60)  
<223> /replace="Ala"

<220>  
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<222> (60)..(60)  
<223> /note="Residue given in the sequence has no preference  
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<220>  
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<222> (83)..(83)  
<223> /replace="Asn"

<220>  
<221> misc\_feature  
<222> (83)..(83)  
<223> /note="Residue given in the sequence has no preference  
with respect to those in the annotation for said position"

<220>  
<221> VARIANT  
<222> (85)..(85)  
<223> /replace="Leu"

<220>  
<221> VARIANT  
<222> (86)..(86)  
<223> /replace="Gln"

<220>  
<221> VARIANT  
<222> (87)..(87)  
<223> /replace="Ala"

<220>  
<221> misc\_feature

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<222> (85)..(87)
<223> /note="Residues given in the sequence have no preference
with respect to those in the annotation for said positions"

<220>
<221> VARIANT
<222> (92)..(92)
<223> /replace="Leu"

<220>
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<222> (93)..(93)
<223> /replace="Lys"

<220>
<221> misc_feature
<222> (92)..(93)
<223> /note="Residues given in the sequence have no preference
with respect to those in the annotation for said positions"

<220>
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<222> (95)..(95)
<223> /replace="Asn"

<220>
<221> misc_feature
<222> (95)..(95)
<223> /note="Residue given in the sequence has no preference
with respect to those in the annotation for said position"

<220>
<221> VARIANT
<222> (105)..(105)
<223> /replace="Met "

<220>
<221> VARIANT
<222> (106)..(106)
<223> /replace="Asn"

<220>
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<222> (105)..(106)
<223> /note="Residues given in the sequence have no preference
with respect to those in the annotation for said positions"

<220>
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<222> (116)..(116)
<223> /replace="Met "

<220>
<221> misc_feature
<222> (116)..(116)
<223> /note="Residue given in the sequence has no preference
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<220>  
<221> VARIANT  
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<223> /replace="Arg"

<220>  
<221> misc\_feature  
<222> (118)..(118)  
<223> /note="Residue given in the sequence has no preference  
with respect to those in the annotation for said position"

<220>  
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<222> (125)..(125)  
<223> /replace="Asn"

<220>  
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<220>  
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<220>  
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<223> /replace="Glu"

<220>  
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<220>  
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<223> /replace="Thr"

<220>  
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<220>  
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<220>
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<223> /replace="Asn"
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<223> /replace="Val"

<220>  
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<220>  
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<220>  
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<223> /replace="Thr"

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<223> /replace="Arg"

<220>  
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<220>
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<223> /replace="Gly"

<220>  
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<223> /replace="Ala"

<220>  
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<223> /replace="Glu"

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<222> (285)..(285)  
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<222> (286)..(286)  
<223> /replace="His"

<220>  
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<220>  
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<220>  
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<223> /replace="Leu"

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<220>  
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<223> /replace="Asn"

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<223> /replace="Gly"

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<220>
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<223> /replace="Arg"

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<223> /replace="Met "

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<223> /replace="His "

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<220>  
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<223> /replace="Glu"

<220>  
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<220>  
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<220>
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<222> (356)..(356)
<223> /replace="Thr"

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<220>  
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<223> /replace="Gly"

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<220>  
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<223> /replace="Gln"

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<223> /replace=" "

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 <223> /replace=" "

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Leu Pro Asp Asp Gly Thr Leu Trp Thr Lys Leu Ala Asn Asp Ala Asn  
 20 25 30



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Asn Leu Ser Ser Leu Gly Ile Ser Ala Leu Trp Ile Pro Pro Ala Trp  
35 40 45

Lys Gly Thr Ser Arg Ser Asp Val Gly Tyr Gly Val Tyr Asp Leu Tyr  
50 55 60

Asp Leu Gly Glu Phe Asn Gln Lys Gly Thr Ile Arg Thr Lys Tyr Gly  
65 70 75 80

Thr Lys Ala Gln Tyr Leu Gln Ala Ile Asn Ala Ala His Ala Ala Gly  
85 90 95

Ile Gln Val Tyr Ala Asp Val Val Phe Asp His Lys Gly Gly Ala Asp  
100 105 110

Ala Thr Glu Trp Val Asp Ala Val Glu Val Asn Pro Ser Asp Arg Asn  
115 120 125

Gln Glu Ile Ser Gly Thr Tyr Gln Ile Gln Ala Trp Thr Lys Phe Asp  
130 135 140

Phe Pro Gly Arg Gly Asn Thr His Ser Ser Phe Lys Trp Arg Trp Tyr  
145 150 155 160

His Phe Asp Gly Val Asp Trp Asp Glu Ser Arg Lys Leu Ser Asn Arg  
165 170 175

Ile Tyr Lys Phe Arg Gly Ile Gly Lys Ala Trp Asp Trp Glu Val Asp  
180 185 190

Thr Glu Asn Gly Asn Tyr Asp Tyr Leu Met Tyr Ala Asp Ile Asp Met  
195 200 205

Asp His Pro Glu Val Val Thr Glu Leu Lys Asn Trp Gly Lys Trp Tyr  
210 215 220

Val Asn Thr Thr Asn Ile Asp Gly Phe Arg Ile Asp Ala Val Lys His  
225 230 235 240

Ile Lys Phe Ser Phe Phe Pro Asp Trp Ile Ser His Val Arg Ser Gln  
245 250 255

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Thr Gly Lys Pro Leu Phe Thr Val Ala Glu Phe Trp Ser Tyr Asp Ile  
260 265 270

Asn Lys Ile His Asn Tyr Ile Thr Lys Thr Asn Gly Thr Met Ser Leu  
275 280 285

Phe Asp Ala Pro Leu His Asn Lys Phe Tyr Thr Ala Ser Lys Ser Gly  
290 295 300

Gly Ala Phe Asp Met Arg Thr Ile Met Thr Asn Thr Leu Met Lys Asp  
305 310 315 320

Gln Pro Thr Leu Ala Val Thr Phe Val Asp Asn His Asp Ser Glu Pro  
325 330 335

Gly Gln Ala Leu Gln Ser Phe Val Asp Pro Trp Phe Lys Pro Leu Ala  
340 345 350

Tyr Ala Phe Ile Leu Thr Arg Gln Glu Gly Tyr Pro Cys Val Phe Tyr  
355 360 365

Gly Asp Tyr Tyr Gly Ile Pro Gln His Asn Ile Pro Ala Leu Lys Ser  
370 375 380

Lys Ile Asp Pro Ile Leu Ile Ala Arg Arg Asp Tyr Ala Tyr Gly Thr  
385 390 395 400

Gln His Asp Tyr Leu Asp His Ser Asp Ile Ile Gly Trp Thr Arg Glu  
405 410 415

Gly Val Thr Glu Lys Pro Gly Ser Gly Leu Ala Ala Ile Ile Ser Asp  
420 425 430

Gly Pro Gly Gly Ser Lys Trp Met Phe Val Gly Lys Asn His Ala Gly  
435 440 445

Lys Val Phe Tyr Asp Ile Thr Gly Asn Arg Ala Asp Thr Val Thr Ile  
450 455 460

Asn Ala Asp Gly Trp Gly Glu Phe Lys Val Asn Gly Gly Ser Val Ser  
465 470 475 480

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Ile Trp Val Pro Lys Lys Thr Thr Val Ser Thr Ile Ala Arg Pro Ile  
485 490 495

Thr Thr Arg Pro Trp Thr Gly Glu Phe Val Arg Trp Thr Glu Pro Arg  
500 505 510

Leu Val Ala Trp Pro  
515

<210> 26  
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<220>  
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<223> /note="Description of artificial sequence: Synthetic  
consensus sequence"

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<223> /replace=" "

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<223> /replace="His"

<220>  
<221> VARIANT  
<222> (3)..(3)  
<223> /replace="Asn"

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<221> VARIANT  
<222> (4)..(4)  
<223> /replace="Gly"

<220>  
<221> VARIANT  
<222> (5)..(5)  
<223> /replace="Thr"

<220>  
<221> misc\_feature  
<222> (1)..(5)  
<223> /note="Residues given in the sequence have no preference  
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<220>  
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<222> (19)..(19)

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<223> /replace="Asn"

<220>
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<223> /note="Residue given in the sequence has no preference
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<223> /replace="Asn"

<220>
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<222> (23)..(23)
<223> /replace="His"

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<222> (22)..(23)
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<223> /replace="Asn"

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<222> (25)..(25)
<223> /note="Residue given in the sequence has no preference
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<220>
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<222> (28)..(28)
<223> /replace="Arg"

<220>
<221> VARIANT
<222> (29)..(29)
<223> /replace="Asp"

<220>
<221> misc_feature
<222> (28)..(29)
<223> /note="Residues given in the sequence have no preference
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<220>
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<223> /replace="Ser"

<220>
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<221> misc_feature
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<222> (35)..(35)
<223> /replace="Arg"

<220>
<221> VARIANT
<222> (36)..(36)
<223> /replace="Asn"

<220>
<221> VARIANT
<222> (37)..(37)
<223> /replace="Arg"

<220>
<221> misc_feature
<222> (35)..(37)
<223> /note="Residues given in the sequence have no preference
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<220>
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<222> (53)..(53)
<223> /replace="Gln"

<220>
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<222> (54)..(54)
<223> /replace="Asn"

<220>
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<222> (53)..(54)
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<222> (60)..(60)
<223> /replace="Ala"

<220>
<221> misc_feature
<222> (60)..(60)
<223> /note="Residue given in the sequence has no preference
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<220>
<221> VARIANT
<222> (85)..(85)
<223> /replace="Leu"
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<220>  
<221> VARIANT  
<222> (86)..(86)  
<223> /replace="Glu"

<220>  
<221> VARIANT  
<222> (87)..(87)  
<223> /replace="Ser"

<220>  
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<222> (85)..(87)  
<223> /note="Residues given in the sequence have no preference  
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<220>  
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<222> (90)..(90)  
<223> /replace="His"

<220>  
<221> misc\_feature  
<222> (90)..(90)  
<223> /note="Residue given in the sequence has no preference  
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<220>  
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<222> (92)..(92)  
<223> /replace="Leu"

<220>  
<221> VARIANT  
<222> (93)..(93)  
<223> /replace="Lys"

<220>  
<221> VARIANT  
<222> (94)..(95)  
<223> /replace="Asn"

<220>  
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<222> (92)..(95)  
<223> /note="Residues given in the sequence have no preference  
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<220>  
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<222> (105)..(105)  
<223> /replace="Met"

<220>  
<221> VARIANT  
<222> (106)..(106)

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<223> /replace="Asn"

<220>
<221> misc_feature
<222> (105)..(106)
<223> /note="Residues given in the sequence have no preference
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<220>
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<222> (116)..(116)
<223> /replace="Asn"

<220>
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<222> (116)..(116)
<223> /note="Residue given in the sequence has no preference
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<220>
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<222> (118)..(118)
<223> /replace="Leu"

<220>
<221> misc_feature
<222> (118)..(118)
<223> /note="Residue given in the sequence has no preference
with respect to those in the annotation for said position"

<220>
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<222> (125)..(125)
<223> /replace="Asn"

<220>
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<222> (126)..(126)
<223> /replace="Asn"

<220>
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<222> (125)..(126)
<223> /note="Residues given in the sequence have no preference
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<222> (134)..(134)
<223> /replace="Asp"

<220>
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<222> (134)..(134)
<223> /note="Residue given in the sequence has no preference
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<220>  
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<222> (136)..(136)  
<223> /replace="Thr"

<220>  
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<223> /note="Residue given in the sequence has no preference  
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<220>  
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<222> (138)..(138)  
<223> /replace="Glu"

<220>  
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<222> (138)..(138)  
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<220>  
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<222> (154)..(154)  
<223> /replace="Asp"

<220>  
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<222> (154)..(154)  
<223> /note="Residue given in the sequence has no preference  
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<220>  
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<222> (169)..(169)  
<223> /replace="Gln"

<220>  
<221> misc\_feature  
<222> (169)..(169)  
<223> /note="Residue given in the sequence has no preference  
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<220>  
<221> VARIANT  
<222> (172)..(172)  
<223> /replace=" "

<220>  
<221> VARIANT  
<222> (173)..(173)  
<223> /replace="Phe"

<220>  
<221> VARIANT  
<222> (174)..(174)



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<223> /replace="Gln"

<220>

<221> VARIANT

<222> (175)..(175)

<223> /replace="Asn"

<220>

<221> misc\_feature

<222> (172)..(175)

<223> /note="Residues given in the sequence have no preference  
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<220>

<221> VARIANT

<222> (183)..(183)

<223> /replace="Asp"

<220>

<221> misc\_feature

<222> (183)..(183)

<223> /note="Residue given in the sequence has no preference  
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<220>

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<222> (215)..(215)

<223> /replace="Asn"

<220>

<221> misc\_feature

<222> (215)..(215)

<223> /note="Residue given in the sequence has no preference  
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<220>

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<222> (219)..(219)

<223> /replace="Arg"

<220>

<221> misc\_feature

<222> (219)..(219)

<223> /note="Residue given in the sequence has no preference  
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<220>

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<222> (222)..(222)

<223> /replace="Glu"

<220>

<221> misc\_feature

<222> (222)..(222)

<223> /note="Residue given in the sequence has no preference  
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<220>  
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<223> /replace="Thr"

<220>  
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<222> (225)..(225)  
<223> /note="Residue given in the sequence has no preference  
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<220>  
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<222> (228)..(228)  
<223> /replace="Leu"

<220>  
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<222> (228)..(228)  
<223> /note="Residue given in the sequence has no preference  
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<220>  
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<222> (246)..(246)  
<223> /replace="Thr"

<220>  
<221> VARIANT  
<222> (247)..(247)  
<223> /replace="Arg"

<220>  
<221> misc\_feature  
<222> (246)..(247)  
<223> /note="Residues given in the sequence have no preference  
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<220>  
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<222> (255)..(255)  
<223> /replace="Asn"

<220>  
<221> VARIANT  
<222> (256)..(256)  
<223> /replace="Ala"

<220>  
<221> misc\_feature  
<222> (255)..(256)  
<223> /note="Residues given in the sequence have no preference  
with respect to those in the annotation for said positions"

<220>  
<221> VARIANT  
<222> (260)..(260)

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<223> /replace="Glu"

<220>
<221> misc_feature
<222> (260)..(260)
<223> /note="Residue given in the sequence has no preference
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<220>
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<222> (263)..(263)
<223> /replace="Ala"

<220>
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<222> (263)..(263)
<223> /note="Residue given in the sequence has no preference
with respect to those in the annotation for said position"

<220>
<221> VARIANT
<222> (269)..(269)
<223> /replace="Lys"

<220>
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<222> (270)..(270)
<223> /replace="Asn"

<220>
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<222> (269)..(270)
<223> /note="Residues given in the sequence have no preference
with respect to those in the annotation for said positions"

<220>
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<222> (273)..(273)
<223> /replace="Gly"

<220>
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<222> (274)..(274)
<223> /replace="Ala"

<220>
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<222> (273)..(274)
<223> /note="Residues given in the sequence have no preference
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<220>
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<222> (276)..(276)
<223> /replace="Glu"

<220>
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<221> misc_feature
<222> (276)..(276)
<223> /note="Residue given in the sequence has no preference
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<220>
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<222> (280)..(280)
<223> /replace="Asn"

<220>
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<222> (280)..(280)
<223> /note="Residue given in the sequence has no preference
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<220>
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<222> (284)..(284)
<223> /replace="Trp"

<220>
<221> VARIANT
<222> (285)..(285)
<223> /replace="Asn"

<220>
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<222> (286)..(286)
<223> /replace="His"

<220>
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<222> (284)..(286)
<223> /note="Residues given in the sequence have no preference
with respect to those in the annotation for said positions"

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<223> /replace="Val"

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<222> (291)..(291)
<223> /note="Residue given in the sequence has no preference
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<222> (295)..(295)
<223> /replace="Tyr"

<220>
<221> VARIANT
<222> (296)..(296)
<223> /replace="Asn"
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<220>  
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<222> (297)..(297)  
<223> /replace="Leu"

<220>  
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<222> (295)..(297)  
<223> /note="Residues given in the sequence have no preference  
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<222> (299)..(299)  
<223> /replace="Asn"

<220>  
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<222> (299)..(299)  
<223> /note="Residue given in the sequence has no preference  
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<220>  
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<222> (302)..(302)  
<223> /replace="Asn"

<220>  
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<222> (302)..(302)  
<223> /note="Residue given in the sequence has no preference  
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<220>  
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<222> (306)..(306)  
<223> /replace="Asn"

<220>  
<221> misc\_feature  
<222> (306)..(306)  
<223> /note="Residue given in the sequence has no preference  
with respect to those in the annotation for said position"

<220>  
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<222> (310)..(310)  
<223> /replace="Ala"

<220>  
<221> VARIANT  
<222> (311)..(311)  
<223> /replace="Lys"

<220>  
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<222> (310)..(311)
<223> /note="Residues given in the sequence have no preference
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<220>
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<222> (314)..(314)
<223> /replace="Asn"

<220>
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<222> (315)..(315)
<223> /replace="Gly"

<220>
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<222> (314)..(315)
<223> /note="Residues given in the sequence have no preference
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<222> (319)..(319)
<223> /replace="Gln"

<220>
<221> VARIANT
<222> (320)..(320)
<223> /replace="Lys"

<220>
<221> VARIANT
<222> (321)..(321)
<223> /replace="His"

<220>
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<222> (319)..(321)
<223> /note="Residues given in the sequence have no preference
with respect to those in the annotation for said positions"

<220>
<221> VARIANT
<222> (323)..(323)
<223> /replace="Met"

<220>
<221> VARIANT
<222> (324)..(324)
<223> /replace="His"

<220>
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<222> (323)..(324)
<223> /note="Residues given in the sequence have no preference
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<223> /replace="Gln"

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<220>  
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<222> (338)..(338)  
<223> /replace="Glu"

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<222> (338)..(338)  
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<223> /replace="Glu"

<220>  
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<223> /note="Residue given in the sequence has no preference  
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<223> /replace="Gln"

<220>  
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<222> (346)..(346)  
<223> /replace="Glu"

<220>  
<221> misc\_feature  
<222> (345)..(346)  
<223> /note="Residues given in the sequence have no preference  
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<220>  
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<222> (355)..(355)  
<223> /replace="Leu"

<220>  
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<222> (355)..(355)

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<223> /note="Residue given in the sequence has no preference  
with respect to those in the annotation for said position"

<220>

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<222> (360)..(360)

<223> /replace="Glu"

<220>

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<222> (361)..(361)

<223> /replace="Gln"

<220>

<221> misc\_feature

<222> (360)..(361)

<223> /note="Residues given in the sequence have no preference  
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<220>

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<222> (365)..(365)

<223> /replace="Ser"

<220>

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<222> (365)..(365)

<223> /note="Residue given in the sequence has no preference  
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<222> (376)..(376)

<223> /replace="Thr"

<220>

<221> misc\_feature

<222> (376)..(376)

<223> /note="Residue given in the sequence has no preference  
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<222> (378)..(378)

<223> /replace="Ser"

<220>

<221> misc\_feature

<222> (378)..(378)

<223> /note="Residue given in the sequence has no preference  
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<220>

<221> VARIANT

<222> (391)..(391)

<223> /replace="Glu"



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<222> (391)..(391)
<223> /note="Residue given in the sequence has no preference
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<220>
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<222> (394)..(394)
<223> /replace="Gln"

<220>
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<222> (395)..(395)
<223> /replace="Asn"

<220>
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<223> /note="Residues given in the sequence have no preference
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<222> (405)..(405)
<223> /replace="Phe"

<220>
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<222> (405)..(405)
<223> /note="Residue given in the sequence has no preference
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<222> (408)..(408)
<223> /replace="His"

<220>
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<222> (409)..(409)
<223> /replace="Asn"

<220>
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<222> (408)..(409)
<223> /note="Residues given in the sequence have no preference
with respect to those in the annotation for said positions"

<220>
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<222> (418)..(418)
<223> /replace="Asn"

<220>
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<222> (418)..(418)
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<223> /note="Residue given in the sequence has no preference
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<220>
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<222> (420)..(420)
<223> /replace="Thr"

<220>
<221> VARIANT
<222> (421)..(421)
<223> /replace="His"

<220>
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<222> (420)..(421)
<223> /note="Residues given in the sequence have no preference
with respect to those in the annotation for said positions"

<220>
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<223> /replace="Asn"

<220>
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<222> (423)..(423)
<223> /note="Residue given in the sequence has no preference
with respect to those in the annotation for said position"

<220>
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<222> (428)..(428)
<223> /replace="Thr"

<220>
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<222> (428)..(428)
<223> /note="Residue given in the sequence has no preference
with respect to those in the annotation for said position"

<220>
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<222> (437)..(437)
<223> /replace="Glu"

<220>
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<222> (437)..(437)
<223> /note="Residue given in the sequence has no preference
with respect to those in the annotation for said position"

<220>
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<222> (444)..(444)
<223> /replace="Gln"
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<223> /note="Residue given in the sequence has no preference
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<222> (446)..(446)
<223> /replace="Lys"

<220>
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<222> (446)..(446)
<223> /note="Residue given in the sequence has no preference
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<220>
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<222> (449)..(449)
<223> /replace="Gln"

<220>
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<222> (449)..(449)
<223> /note="Residue given in the sequence has no preference
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<220>
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<222> (459)..(459)
<223> /replace="Pro"

<220>
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<222> (460)..(460)
<223> /replace="Gly"

<220>
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<222> (459)..(460)
<223> /note="Residues given in the sequence have no preference
with respect to those in the annotation for said positions"

<220>
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<222> (471)..(471)
<223> /replace="Asn"

<220>
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<222> (471)..(471)
<223> /note="Residue given in the sequence has no preference
with respect to those in the annotation for said position"

<220>
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<222> (473)..(473)  
<223> /replace="Ser"

<220>  
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<222> (473)..(473)  
<223> /note="Residue given in the sequence has no preference  
with respect to those in the annotation for said position"

<220>  
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&lt;400&gt; 26

His Ala Ala Pro Phe Asn Gly Thr Met Met Gln Tyr Phe Glu Trp His  
1 5 10 15Leu Pro Asp Asp Gly Thr Leu Trp Thr Lys Leu Ala Asn Asp Ala Asn  
20 25 30Asn Leu Ser Ser Leu Gly Ile Thr Ala Ile Trp Ile Pro Pro Ala Trp  
35 40 45Lys Gly Thr Ser Arg Ser Asp Val Gly Tyr Gly Val Tyr Asp Leu Tyr  
50 55 60Asp Leu Gly Glu Phe Asn Gln Lys Gly Thr Val Arg Thr Lys Tyr Gly  
65 70 75 80Thr Lys Ala Gln Tyr Leu Gln Ala Ile Gln Ala Ala His Ala Ala Gly  
85 90 95Met Gln Val Tyr Ala Asp Val Val Phe Asp His Lys Gly Gly Ala Asp  
100 105 110Ala Thr Glu Trp Val Asp Ala Val Glu Val Asn Pro Ser Asp Arg Asn  
115 120 125Gln Glu Ile Ser Gly Thr Tyr Gln Ile Gln Ala Trp Thr Lys Phe Asp  
130 135 140Phe Pro Gly Arg Gly Asn Thr Tyr Ser Ser Phe Lys Trp Arg Trp Tyr  
145 150 155 160His Phe Asp Gly Val Asp Trp Asp Glu Ser Arg Gln Lys Leu Ser Arg  
165 170 175Ile Tyr Lys Phe Arg Gly Ile Gly Lys Ala Trp Asp Trp Glu Val Asp  
180 185 190Ser Glu Asn Gly Asn Tyr Asp Tyr Leu Met Tyr Ala Asp Leu Asp Met  
195 200 205

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Asp His Pro Glu Val Val Thr Glu Leu Lys Asn Trp Gly Lys Trp Tyr  
210 215 220

Val Asn Thr Thr Asn Ile Asp Gly Phe Arg Ile Asp Ala Val Lys His  
225 230 235 240

Ile Lys Phe Ser Phe Phe Pro Asp Trp Leu Ser His Val Arg Ser Gln  
245 250 255

Thr Gly Lys Pro Leu Phe Thr Val Ala Glu Phe Trp Ser Tyr Asp Ile  
260 265 270

Asn Lys Leu His Asn Tyr Ile Thr Lys Thr Asn Gly Thr Met Ser Leu  
275 280 285

Phe Asp Ala Pro Leu His Asn Lys Phe Tyr Thr Ala Ser Lys Ser Gly  
290 295 300

Gly Ala Phe Asp Met Arg Thr Leu Leu Thr Asn Thr Leu Met Lys Asp  
305 310 315 320

Gln Pro Thr Leu Ala Val Thr Phe Val Asp Asn His Asp Ser Glu Pro  
325 330 335

Gly Gln Ala Leu Gln Ser Phe Val Asp Pro Trp Phe Lys Pro Leu Ala  
340 345 350

Tyr Ala Phe Ile Leu Thr Arg Gln Glu Gly Tyr Pro Cys Val Phe Tyr  
355 360 365

Gly Asp Tyr Tyr Gly Ile Pro Gln His Asn Ile Pro Ala Leu Lys Ala  
370 375 380

Lys Ile Asp Pro Ile Leu Ile Ala Arg Arg Asp Phe Ala Tyr Gly Thr  
385 390 395 400

Gln His Asp Tyr Leu Asp His Ser Asp Ile Ile Gly Trp Thr Arg Glu  
405 410 415

Gly Val Thr Glu Lys Pro Gly Ser Gly Leu Ala Ala Ile Ile Ser Asp  
420 425 430



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Gly Pro Gly Gly Ser Lys Trp Met Tyr Val Gly Lys Asn His Ala Gly  
435 440 445

Lys Val Phe His Asp Ile Thr Gly Asn Lys Ser Asp Thr Val Thr Ile  
450 455 460

Asn Ala Asp Gly Trp Ala Glu Phe Lys Val Asn Gly Gly Ser Val Ser  
465 470 475 480

Ile Trp Val Pro Arg Lys Thr Thr Val Ser Thr Ile Ala Arg Pro Ile  
485 490 495

Thr Thr Arg Pro Trp Thr Gly Glu Phe Val Arg Trp Thr Glu Pro Arg  
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Leu Val Ala Trp Pro  
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<223> /replace="Asn"

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<222> (173)..(173)
<223> /replace="Leu"

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<223> /replace="Ile"  
  
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<220>  
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<222> (246)..(247)  
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<223> /note="Residues given in the sequence have no preference  
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<223> /replace="Ala"

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<223> /replace="Ala"

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<223> /replace="Ala"

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<222> (284)..(284)

<223> /replace="Trp"

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<223> /replace="Asn"

<220>

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<222> (286)..(286)

<223> /replace="His"

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<221> misc\_feature

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<222> (283)..(286)
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<223> /replace="Gly"

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<223> /replace="Gln"

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<223> /replace="His"

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<220>  
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<220>  
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<223> /note="Residue given in the sequence has no preference with respect to those in the annotation for said position"

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<223> /replace="Thr"

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<222> (394)..(395)

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<223> /replace="Phe"

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<222> (421)..(421)
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<223> /replace="Lys"

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<222> (449)..(449)  
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<223> /replace="Arg"

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<222> (452)..(452)  
<223> /note="Residue given in the sequence has no preference  
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<223> /replace=" "

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1 5 10 15

Leu Pro Asp Asp Gly Thr Leu Trp Thr Lys Leu Ala Asn Asp Ala Asn  
20 25 30

Asn Leu Ser Ser Leu Gly Ile Thr Ala Leu Trp Ile Pro Pro Ala Trp  
35 40 45

Lys Gly Thr Ser Arg Ser Asp Val Gly Tyr Gly Val Tyr Asp Leu Tyr  
50 55 60

Asp Leu Gly Glu Phe Asn Gln Lys Gly Thr Val Arg Thr Lys Tyr Gly  
65 70 75 80

Thr Lys Ala Gln Tyr Leu Gln Ala Ile Gln Ala Ala His Ala Ala Gly  
85 90 95

Ile Gln Val Tyr Ala Asp Val Val Phe Asp His Lys Gly Gly Ala Asp  
100 105 110

Gly Thr Glu Trp Val Asp Ala Val Glu Val Asn Pro Ser Asp Arg Asn  
115 120 125

Gln Glu Ile Ser Gly Thr Tyr Gln Ile Gln Ala Trp Thr Lys Phe Asp  
130 135 140

Phe Pro Gly Arg Gly Asn Thr His Ser Ser Phe Lys Trp Arg Trp Tyr  
145 150 155 160

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His Phe Asp Gly Val Asp Trp Asp Glu Ser Arg Gln Lys Leu Ser Lys  
165 170 175

Ile Tyr Lys Phe Arg Gly Ile Gly Lys Ala Trp Asp Trp Glu Val Asp  
180 185 190

Thr Glu Asn Gly Asn Tyr Asp Tyr Leu Met Tyr Ala Asp Ile Asp Met  
195 200 205

Asp His Pro Glu Val Ile Thr Glu Leu Lys Asn Trp Gly Lys Trp Tyr  
210 215 220

Val Asn Thr Thr Asn Ile Asp Gly Phe Arg Ile Asp Ala Val Lys His  
225 230 235 240

Ile Lys Phe Ser Phe Phe Pro Asp Trp Leu Ser His Val Arg Ser Gln  
245 250 255

Thr Gly Lys Pro Leu Phe Thr Val Ala Glu Phe Trp Ser Tyr Asp Ile  
260 265 270

Asn Lys Ile His Asn Tyr Ile Thr Lys Thr Asn Gly Thr Met Ser Leu  
275 280 285

Phe Asp Ala Pro Leu His Asn Lys Phe Tyr Thr Ala Ser Lys Ser Gly  
290 295 300

Gly Ala Phe Asp Met Arg Thr Ile Leu Thr Asn Ser Leu Met Lys Asp  
305 310 315 320

Gln Pro Thr Leu Ala Val Thr Phe Val Asp Asn His Asp Ser Glu Pro  
325 330 335

Gly Gln Ala Leu Gln Ser Phe Val Asp Pro Trp Phe Lys Pro Leu Ala  
340 345 350

Tyr Ala Phe Ile Leu Thr Arg Gln Glu Gly Tyr Pro Cys Val Phe Tyr  
355 360 365

Gly Asp Tyr Tyr Gly Ile Pro Gln His Asn Ile Pro Ser Leu Lys Ser  
370 375 380

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Lys Ile Asp Pro Leu Leu Ile Ala Arg Arg Asp Tyr Ala Tyr Gly Thr  
385 390 395 400

Gln His Asp Tyr Leu Asp His Ser Asp Ile Ile Gly Trp Thr Arg Glu  
405 410 415

Gly Val Ser Glu Lys Pro Gly Ser Gly Leu Ala Ala Ile Ile Ser Asp  
420 425 430

Gly Pro Gly Gly Ser Lys Trp Met Tyr Val Gly Lys Gln His Ala Gly  
435 440 445

Lys Val Phe Tyr Asp Ile Thr Gly Asn Arg Ser Asp Thr Val Thr Ile  
450 455 460

Asn Ala Asp Gly Trp Gly Glu Phe Lys Val Asn Gly Gly Ala Val Ser  
465 470 475 480

Val Trp Val Pro Arg Lys Thr Thr Val Ser Thr Ile Ala Arg Pro Ile  
485 490 495

Thr Thr Arg Pro Trp Thr Gly Glu Phe Val Arg Trp Thr Glu Pro Arg  
500 505 510

Leu Val Ala Trp Pro  
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<210> 28  
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<223> /replace="Gly"

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<223> /note="Residue given in the sequence has no preference  
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<223> /replace="His"

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<221> VARIANT

<222> (28)..(28)

<223> /replace="Asp"

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<221> misc\_feature

<222> (27)..(28)

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<220>

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<223> /replace="Asp"

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<222> (36)..(36)

<223> /replace="Ala"

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<223> /replace="Leu"

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<222> (92)..(92)

<223> /replace="Lys"

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<222> (91)..(92)

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<222> (95)..(95)

<223> /replace="Asp"

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<223> /replace="Asn"

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<223> /replace="Ala"

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<223> /replace="Gln"

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<223> /replace="Trp"

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<223> /replace="Ala"

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<223> /replace="Asp"

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<223> /replace="Asn"

<220>  
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<223> /replace="Gln"

<220>  
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<222> (169)..(169)  
<223> /replace="Arg"

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<222> (170)..(170)  
<223> /replace="Tyr"

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<223> /replace="Gln"

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<222> (172)..(172)  
<223> /replace="Glu"

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<222> (173)..(173)  
<223> /replace="Asn"

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<223> /replace="His"

<220>  
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<222> (168)..(174)  
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<222> (179)..(179)  
<223> /replace="Ala"

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<222> (180)..(180)  
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<223> /replace="Trp"

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<223> /replace="Glu"

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with respect to those in the annotation for said positions"

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Pro Asp Asp Gly Thr Leu Trp Thr Lys Leu Ala Asn Asp Ala Asn Asn  
20 25 30

Leu Ser Ser Leu Gly Ile Thr Ala Ile Trp Ile Pro Pro Ala Tyr Lys  
35 40 45

Gly Thr Ser Arg Ala Asp Val Gly Tyr Gly Val Tyr Asp Leu Tyr Asp  
50 55 60

Leu Gly Glu Phe Asn Gln Lys Gly Thr Val Arg Thr Lys Tyr Gly Thr  
65 70 75 80

Lys Ala Gln Tyr Leu Gln Ala Ile Gln Ala Ala His Ala Ala Gly Ile  
85 90 95

Asn Val Tyr Ala Asp Val Val Phe Asp His Lys Gly Gly Ala Asp Gly  
100 105 110

Thr Glu Trp Val Asp Ala Val Glu Val Asn Pro Ser Asp Arg Asn Gln  
115 120 125

Asp Ile Ser Gly Thr Tyr Gln Ile Gln Ala Trp Thr Lys Phe Asp Phe  
130 135 140

Pro Gly Arg Gly Asn Thr Tyr Ser Ser Phe Lys Trp Arg Trp Phe His  
145 150 155 160

Phe Asp Gly Val Asp Trp Asp Glu Ser Arg Lys Leu Ser Arg Ile Phe  
165 170 175

Lys Phe Arg Gly Ile Gly Lys Ala Trp Asp Trp Glu Val Asp Thr Glu  
180 185 190

Asn Gly Asn Tyr Asp Tyr Leu Leu Tyr Ala Asp Ile Asp Met Asp His  
195 200 205

Pro Glu Val Val Thr Glu Leu Lys Asn Trp Gly Lys Trp Phe Val Asn  
210 215 220

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Thr Thr Asn Ile Asp Gly Phe Arg Leu Asp Ala Ile Lys His Ile Lys  
225 230 235 240

Phe Ser Phe Phe Pro Asp Trp Leu Ser His Val Arg Ser Gln Thr Gly  
245 250 255

Lys Pro Leu Phe Thr Val Gly Glu Tyr Trp Ser Tyr Asp Ile Asn Lys  
260 265 270

Leu His Asn Tyr Ile Thr Lys Thr Asn Gly Thr Met Ser Leu Phe Asp  
275 280 285

Ala Pro Leu His Asn Lys Phe Tyr Thr Ala Ser Lys Ser Gly Gly Ala  
290 295 300

Phe Asp Met Arg Thr Ile Leu Thr Asn Ser Leu Met Lys Asp Gln Pro  
305 310 315 320

Thr Leu Ala Val Thr Phe Val Asp Asn His Asp Thr Glu Pro Gly Gln  
325 330 335

Ala Leu Gln Ser Trp Val Asp Pro Trp Phe Lys Pro Leu Ala Tyr Ala  
340 345 350

Phe Ile Leu Thr Arg Gln Glu Gly Tyr Pro Cys Val Phe Tyr Gly Asp  
355 360 365

Tyr Tyr Gly Ile Pro Asn Tyr Asn Ile Pro Ala Leu Lys Ser Lys Ile  
370 375 380

Asp Pro Leu Leu Ile Ala Arg Arg Asp Tyr Ala Tyr Gly Thr Gln His  
385 390 395 400

Asp Tyr Leu Asp His Ser Asp Ile Ile Gly Trp Thr Arg Glu Gly Val  
405 410 415

Ser Glu Lys Pro Gly Ser Gly Leu Ala Ala Ile Ile Ser Asp Gly Pro  
420 425 430

Gly Gly Ser Lys Trp Met Tyr Val Gly Lys Gln His Ala Gly Lys Val  
435 440 445

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Phe Tyr Asp Leu Thr Gly Asn Arg Ser Asp Ser Val Thr Ile Asn Ser  
450 455 460

Asp Gly Trp Gly Glu Phe Lys Val Asn Gly Gly Ser Val Ser Val Trp  
465 470 475 480

Val Pro Arg Lys Thr Thr Val Ser Thr Ile Ala Arg Pro Ile Thr Thr  
485 490 495

Arg Pro Trp Thr Gly Glu Phe Val Arg Trp Thr Glu Pro Arg Leu Val  
500 505 510

Ala Trp Pro  
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<210> 29

<211> 515

<212> PRT

<213> Artificial sequence

<220>

<221> source

<223> /note="Description of artificial sequence: Synthetic  
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<223> /replace=" "

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<222> (2)..(2)

<223> /replace="Asp"

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<221> VARIANT

<222> (3)..(3)

<223> /replace="Gly"

<220>

<221> VARIANT

<222> (4)..(4)

<223> /replace="Leu"

<220>

<221> misc\_feature

<222> (1)..(4)

<223> /note="Residues given in the sequence have no preference  
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<223> /replace="Glu"

<220>  
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<222> (18)..(18)  
<223> /replace="Asn"

<220>  
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<220>  
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<223> /replace="Gln"

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<222> (22)..(22)  
<223> /replace="His"

<220>  
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<223> /replace="Asn"

<220>  
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<222> (24)..(24)  
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<223> /replace="His"

<220>  
<221> VARIANT  
<222> (28)..(28)  
<223> /replace="Asp"

<220>  
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<222> (27)..(28)

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<220>
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<223> /replace="Glu"

<220>
<221> VARIANT
<222> (32)..(32)
<223> /replace="Ala"

<220>
<221> misc_feature
<222> (31)..(32)
<223> /note="Residues given in the sequence have no preference
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<220>
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<222> (35)..(35)
<223> /replace="Asn"

<220>
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<222> (36)..(36)
<223> /replace="Ala"

<220>
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<222> (35)..(36)
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<223> /replace="Asn"

<220>
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<222> (50)..(50)
<223> /note="Residue given in the sequence has no preference
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<220>
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<222> (52)..(52)
<223> /replace="Gln"

<220>
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<222> (52)..(52)
<223> /note="Residue given in the sequence has no preference
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<223> /replace="Ala"

<220>  
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<222> (59)..(59)  
<223> /note="Residue given in the sequence has no preference  
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<220>  
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<222> (84)..(84)  
<223> /replace="Leu"

<220>  
<221> VARIANT  
<222> (85)..(85)  
<223> /replace="Glu"

<220>  
<221> VARIANT  
<222> (86)..(86)  
<223> /replace="Arg"

<220>  
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<222> (84)..(86)  
<223> /note="Residues given in the sequence have no preference  
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<220>  
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<223> /replace="Gly"

<220>  
<221> misc\_feature  
<222> (89)..(89)  
<223> /note="Residue given in the sequence has no preference  
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<220>  
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<222> (91)..(91)  
<223> /replace="Leu"

<220>  
<221> VARIANT  
<222> (92)..(92)  
<223> /replace="Lys"

<220>  
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<222> (91)..(92)  
<223> /note="Residues given in the sequence have no preference

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with respect to those in the annotation for said positions"
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<223> /replace="Asn"

<220>
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<222> (95)..(95)
<223> /replace="Asp"

<220>
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<222> (94)..(95)
<223> /note="Residues given in the sequence have no preference
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<220>
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<222> (104)..(104)
<223> /replace="Met"

<220>
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<222> (105)..(105)
<223> /replace="Asn"

<220>
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<222> (104)..(105)
<223> /note="Residues given in the sequence have no preference
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<220>
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<222> (108)..(108)
<223> /replace="Leu"

<220>
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<222> (108)..(108)
<223> /note="Residue given in the sequence has no preference
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<220>
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<222> (112)..(112)
<223> /replace="Phe"

<220>
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<222> (112)..(112)
<223> /note="Residue given in the sequence has no preference
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<221> VARIANT  
<222> (115)..(115)  
<223> /replace="Ala"

<220>  
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<222> (115)..(115)  
<223> /note="Residues given in the sequence have no preference  
with respect to those in the annotation for said positions"

<220>  
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<222> (117)..(117)  
<223> /replace="Gln"

<220>  
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<222> (117)..(117)  
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<222> (120)..(120)  
<223> /replace="Gln"

<220>  
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<222> (120)..(120)  
<223> /note="Residue given in the sequence has no preference  
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<220>  
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<222> (125)..(125)  
<223> /replace="Asn"

<220>  
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<222> (125)..(125)  
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<220>  
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<222> (127)..(127)  
<223> /replace="Trp"

<220>  
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<222> (127)..(127)  
<223> /note="Residue given in the sequence has no preference  
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<220>  
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<223> /replace="Val"

<220>
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<222> (133)..(133)
<223> /note="Residue given in the sequence has no preference
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<222> (135)..(135)
<223> /replace="Thr"

<220>
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<222> (135)..(135)
<223> /note="Residue given in the sequence has no preference
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<220>
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<222> (137)..(137)
<223> /replace="Asp"

<220>
<221> misc_feature
<222> (137)..(137)
<223> /note="Residue given in the sequence has no preference
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<222> (141)..(141)
<223> /replace="Gly"

<220>
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<222> (141)..(141)
<223> /note="Residue given in the sequence has no preference
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<220>
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<222> (148)..(148)
<223> /replace="Asn"

<220>
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<222> (148)..(148)
<223> /note="Residue given in the sequence has no preference
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<220>
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<222> (150)..(150)
<223> /replace="Ala"
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<220>  
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<222> (150)..(150)  
<223> /note="Residue given in the sequence has no preference  
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<220>  
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<222> (153)..(153)  
<223> /replace="Asp"

<220>  
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<222> (153)..(153)  
<223> /note="Residue given in the sequence has no preference  
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<220>  
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<222> (162)..(162)  
<223> /replace="Asn"

<220>  
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<222> (162)..(162)  
<223> /note="Residue given in the sequence has no preference  
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<220>  
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<223> /replace="Gln"

<220>  
<221> VARIANT  
<222> (169)..(169)  
<223> /replace="Arg"

<220>  
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<222> (170)..(170)  
<223> /replace="Tyr"

<220>  
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<222> (171)..(171)  
<223> /replace="Gln"

<220>  
<221> VARIANT  
<222> (172)..(172)  
<223> /replace="Glu"

<220>  
<221> VARIANT  
<222> (173)..(173)  
<223> /replace="Asn"

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<221> VARIANT  
<222> (174)..(174)  
<223> /replace="His"

<220>  
<221> misc\_feature  
<222> (168)..(174)  
<223> /note="Residues given in the sequence have no preference  
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<220>  
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<222> (179)..(179)  
<223> /replace="Ala"

<220>  
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<222> (180)..(180)  
<223> /replace="Asn"

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<222> (181)..(181)  
<223> /replace="Thr"

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<222> (182)..(182)  
<223> /replace="Asn"

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<223> /replace=" "

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<223> /replace=" "

<220>  
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<222> (179)..(184)  
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<220>  
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<222> (186)..(186)  
<223> /replace="Asn"

<220>  
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<222> (186)..(186)  
<223> /note="Residue given in the sequence has no preference"

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with respect to those in the annotation for said position"
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<222> (188)..(188)
<223> /replace="Arg"

<220>
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<222> (188)..(188)
<223> /note="Residue given in the sequence has no preference
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<222> (191)..(191)
<223> /replace="Glu"

<220>
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<222> (191)..(191)
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<222> (201)..(201)
<223> /replace="Gly"

<220>
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<222> (201)..(201)
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with respect to those in the annotation for said position"

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<222> (203)..(203)
<223> /replace="Asn"

<220>
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<222> (203)..(203)
<223> /note="Residue given in the sequence has no preference
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<223> /replace="Phe"

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<222> (207)..(207)
<223> /replace="Ser"

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<223> /note="Residues given in the sequence have no preference
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<222> (212)..(212)
<223> /replace="Gln"

<220>
<221> VARIANT
<222> (213)..(213)
<223> /replace="Glu"

<220>
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<223> /note="Residues given in the sequence have no preference
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<223> /replace="Asp"

<220>
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<223> /replace="Ser"

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<222> (220)..(220)
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<223> /replace="Thr"

<220>
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<222> (224)..(224)
<223> /replace="Asp"

<220>
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<222> (225)..(225)
<223> /replace="Glu"
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<220>  
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<223> /replace="Leu"

<220>  
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<223> /replace="Asp"

<220>  
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<222> (223)..(227)  
<223> /note="Residues given in the sequence have no preference  
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<223> /replace="Pro"

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<223> /replace="Trp"

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<222> (242)..(242)  
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<222> (244)..(244)  
<223> /replace="Thr"

<220>  
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<222> (245)..(245)  
<223> /replace="Ser"

<220>  
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<222> (244)..(245)  
<223> /note="Residues given in the sequence have no preference  
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<223> /replace="Arg"

<220>
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<222> (249)..(249)
<223> /note="Residue given in the sequence has no preference
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<223> /replace="Gln"

<220>
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<222> (251)..(251)
<223> /note="Residue given in the sequence has no preference
with respect to those in the annotation for said position"

<220>
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<222> (254)..(254)
<223> /replace="Glu"

<220>
<221> VARIANT
<222> (255)..(255)
<223> /replace="Ala"

<220>
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<222> (256)..(256)
<223> /replace="Asp"

<220>
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<223> /replace="Gln"

<220>
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<222> (258)..(258)
<223> /replace="Asp"

<220>
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<222> (254)..(258)
<223> /note="Residues given in the sequence have no preference
with respect to those in the annotation for said positions"

<220>
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<222> (261)..(261)
<223> /replace="Val"

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<222> (261)..(261)
<223> /note="Residue given in the sequence has no preference
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<220>
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<222> (267)..(267)
<223> /replace="Lys"

<220>
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<222> (268)..(268)
<223> /replace="Asp"

<220>
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<222> (267)..(268)
<223> /note="Residues given in the sequence have no preference
with respect to those in the annotation for said positions"

<220>
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<222> (271)..(271)
<223> /replace="Gly"

<220>
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<222> (272)..(272)
<223> /replace="Ala"

<220>
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<223> /note="Residues given in the sequence have no preference
with respect to those in the annotation for said positions"

<220>
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<222> (274)..(274)
<223> /replace="Glu"

<220>
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<222> (275)..(275)
<223> /replace="Phe"

<220>
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<223> /note="Residues given in the sequence have no preference
with respect to those in the annotation for said positions"

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<222> (278)..(278)
<223> /replace="Asp"
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<222> (279)..(279)  
<223> /replace="Glu"

<220>  
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<223> /replace="Met "

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<222> (278)..(280)  
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with respect to those in the annotation for said positions"

<220>  
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<222> (282)..(282)  
<223> /replace="Trp"

<220>  
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<222> (283)..(283)  
<223> /replace="Glu"

<220>  
<221> misc\_feature  
<222> (282)..(283)  
<223> /note="Residues given in the sequence have no preference  
with respect to those in the annotation for said positions"

<220>  
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<222> (289)..(289)  
<223> /replace="Val "

<220>  
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<222> (289)..(289)  
<223> /note="Residue given in the sequence has no preference  
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<220>  
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<222> (292)..(292)  
<223> /replace="Asn"

<220>  
<221> VARIANT  
<222> (293)..(293)  
<223> /replace="Tyr"

<220>  
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<222> (294)..(294)

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<223> /replace="Asn"

<220>
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<222> (292)..(294)
<223> /note="Residues given in the sequence have no preference
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<223> /replace="Arg"

<220>
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<222> (297)..(297)
<223> /note="Residue given in the sequence has no preference
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<223> /replace="Gln"

<220>
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<222> (301)..(301)
<223> /note="Residue given in the sequence has no preference
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<220>
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<222> (309)..(309)
<223> /replace="Asn"

<220>
<221> misc_feature
<222> (309)..(309)
<223> /note="Residue given in the sequence has no preference
      with respect to those in the annotation for said position"

<220>
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<222> (312)..(312)
<223> /replace="Arg"

<220>
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<222> (313)..(313)
<223> /replace="Gly"

<220>
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<222> (312)..(313)
<223> /note="Residues given in the sequence have no preference
      with respect to those in the annotation for said positions"
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<220>  
<221> VARIANT  
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<223> /replace="Glu"

<220>  
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<223> /replace="Asp"

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<223> /replace="Gly"

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<222> (382)..(383)
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<222> (403)..(403)
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<222> (456)..(456)

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<223> /note="Residue given in the sequence has no preference
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Ala Ala Pro Phe Asn Gly Thr Met Met Gln Tyr Phe Glu Trp His Leu  
1 5 10 15

Pro Asp Asp Gly Thr Leu Trp Thr Lys Leu Ala Asn Asp Ala Asn Asn  
20 25 30

Leu Ser Ser Leu Gly Ile Thr Ala Ile Trp Ile Pro Pro Ala Tyr Lys  
35 40 45

Gly Thr Ser Arg Ala Asp Val Gly Tyr Gly Val Tyr Asp Leu Tyr Asp  
50 55 60

Leu Gly Glu Phe Asn Gln Lys Gly Thr Val Arg Thr Lys Tyr Gly Thr  
65 70 75 80

Lys Ala Gln Tyr Leu Gln Ala Ile Gln Ala Ala His Ala Ala Gly Ile  
85 90 95

Asn Val Tyr Ala Asp Val Val Phe Asp His Lys Gly Gly Ala Asp Gly  
100 105 110

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Thr Glu Trp Val Asp Ala Val Glu Val Asn Pro Ser Asp Arg Asn Gln  
115 120 125

Asp Ile Ser Gly Thr Tyr Gln Ile Gln Ala Trp Thr Lys Phe Asp Phe  
130 135 140

Pro Gly Arg Gly Asn Thr Tyr Ser Ser Phe Lys Trp Arg Trp Phe His  
145 150 155 160

Phe Asp Gly Val Asp Trp Asp Glu Ser Arg Lys Leu Ser Arg Ile Phe  
165 170 175

Lys Phe Arg Gly Ile Gly Lys Ala Trp Asp Trp Glu Val Asp Thr Glu  
180 185 190

Asn Gly Asn Tyr Asp Tyr Leu Leu Tyr Ala Asp Ile Asp Met Asp His  
195 200 205

Pro Glu Val Val Thr Glu Leu Lys Asn Trp Gly Lys Trp Phe Val Asn  
210 215 220

Thr Thr Asn Ile Asp Gly Phe Arg Leu Asp Ala Ile Lys His Ile Lys  
225 230 235 240

Phe Ser Phe Phe Pro Asp Trp Leu Ser His Val Arg Ser Gln Thr Gly  
245 250 255

Lys Pro Leu Phe Thr Val Gly Glu Tyr Trp Ser Tyr Asp Ile Asn Lys  
260 265 270

Leu His Asn Tyr Ile Thr Lys Thr Asn Gly Thr Met Ser Leu Phe Asp  
275 280 285

Ala Pro Leu His Asn Lys Phe Tyr Thr Ala Ser Lys Ser Gly Gly Ala  
290 295 300

Phe Asp Met Arg Thr Ile Leu Thr Asn Ser Leu Met Lys Asp Gln Pro  
305 310 315 320

Thr Leu Ala Val Thr Phe Val Asp Asn His Asp Thr Glu Pro Gly Gln  
325 330 335

Ala Leu Gln Ser Trp Val Asp Pro Trp Phe Lys Pro Leu Ala Tyr Ala

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340 345 350

Phe Ile Leu Thr Arg Gln Glu Gly Tyr Pro Cys Val Phe Tyr Gly Asp  
355 360 365

Tyr Tyr Gly Ile Pro Asn Tyr Asn Ile Pro Ala Leu Lys Ser Lys Ile  
370 375 380

Asp Pro Leu Leu Ile Ala Arg Arg Asp Tyr Ala Tyr Gly Thr Gln His  
385 390 395 400

Asp Tyr Leu Asp His Ser Asp Ile Ile Gly Trp Thr Arg Glu Gly Val  
405 410 415

Ser Glu Lys Pro Gly Ser Gly Leu Ala Ala Ile Ile Ser Asp Gly Pro  
420 425 430

Gly Gly Ser Lys Trp Met Tyr Val Gly Lys Gln His Ala Gly Lys Val  
435 440 445

Phe Tyr Asp Leu Thr Gly Asn Arg Ala Asp Ser Val Thr Ile Asn Ser  
450 455 460

Asp Gly Trp Gly Glu Phe Lys Val Asn Gly Gly Ser Val Ser Val Trp  
465 470 475 480

Val Pro Arg Lys Thr Thr Val Ser Thr Ile Ala Arg Pro Ile Thr Thr  
485 490 495

Arg Pro Trp Thr Gly Glu Phe Val Arg Trp Thr Glu Pro Arg Leu Val  
500 505 510

Ala Trp Pro  
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<211> 515

<212> PRT

<213> Artificial sequence

<220>

<221> source

<223> /note="Description of artificial sequence: Synthetic  
consensus sequence"

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with respect to those in the annotation for said positions"

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Ala Ala Pro Phe Asn Gly Thr Met Met Gln Tyr Phe Glu Trp Tyr Leu  
1 5 10 15

Pro Asp Asp Gly Thr Leu Trp Thr Lys Val Ala Asn Glu Ala Asn Asn  
20 25 30

Leu Ser Ser Leu Gly Ile Thr Ala Leu Trp Leu Pro Pro Ala Tyr Lys  
35 40 45

Gly Thr Ser Arg Ser Asp Val Gly Tyr Gly Val Tyr Asp Leu Tyr Asp  
50 55 60

Leu Gly Glu Phe Asn Gln Lys Gly Thr Val Arg Thr Lys Tyr Gly Thr  
65 70 75 80

Lys Ala Gln Tyr Leu Gln Ala Ile Gln Ala Ala His Ala Ala Gly Met  
85 90 95

Gln Val Tyr Ala Asp Val Val Phe Asp His Lys Gly Gly Ala Asp Gly  
100 105 110

Thr Glu Trp Val Asp Ala Val Glu Val Asn Pro Ser Asp Arg Asn Gln  
115 120 125

Glu Ile Ser Gly Thr Tyr Gln Ile Gln Ala Trp Thr Lys Phe Asp Phe  
130 135 140

Pro Gly Arg Gly Asn Thr Tyr Ser Ser Phe Lys Trp Arg Trp Tyr His  
145 150 155 160

Phe Asp Gly Val Asp Trp Asp Glu Ser Arg Lys Leu Ser Arg Ile Tyr  
165 170 175

Lys Phe Arg Gly Ile Gly Lys Ala Trp Asp Trp Glu Val Asp Thr Glu  
180 185 190

Asn Gly Asn Tyr Asp Tyr Leu Met Tyr Ala Asp Leu Asp Met Asp His  
195 200 205

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Pro Glu Val Val Thr Glu Leu Lys Asn Trp Gly Lys Trp Tyr Val Asn  
210 215 220

Thr Thr Asn Ile Asp Gly Phe Arg Leu Asp Ala Val Lys His Ile Lys  
225 230 235 240

Phe Ser Phe Phe Pro Asp Trp Leu Ser Tyr Val Arg Ser Gln Thr Gly  
245 250 255

Lys Pro Leu Phe Thr Val Gly Glu Tyr Trp Ser Tyr Asp Ile Asn Lys  
260 265 270

Leu His Asn Tyr Ile Thr Lys Thr Asn Gly Thr Met Ser Leu Phe Asp  
275 280 285

Ala Pro Leu His Asn Lys Phe Tyr Thr Ala Ser Lys Ser Gly Gly Ala  
290 295 300

Phe Asp Met Arg Thr Leu Met Thr Asn Thr Leu Met Lys Asp Gln Pro  
305 310 315 320

Thr Leu Ala Val Thr Phe Val Asp Asn His Asp Thr Glu Pro Gly Gln  
325 330 335

Ala Leu Gln Ser Trp Val Asp Pro Trp Phe Lys Pro Leu Ala Tyr Ala  
340 345 350

Phe Ile Leu Thr Arg Gln Glu Gly Tyr Pro Cys Val Phe Tyr Gly Asp  
355 360 365

Tyr Tyr Gly Ile Pro Gln Tyr Asn Ile Pro Ser Leu Lys Ser Lys Ile  
370 375 380

Asp Pro Leu Leu Ile Ala Arg Arg Asp Tyr Ala Tyr Gly Thr Gln His  
385 390 395 400

Asp Tyr Leu Asp His Ser Asp Ile Ile Gly Trp Thr Arg Glu Gly Val  
405 410 415

Thr Glu Lys Pro Gly Ser Gly Leu Ala Ala Leu Ile Thr Asp Gly Pro  
420 425 430

Gly Gly Ser Lys Trp Met Tyr Val Gly Lys Gln His Ala Gly Lys Val

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435 440 445

Phe Tyr Asp Leu Thr Gly Asn Arg Ser Asp Thr Val Thr Ile Asn Ser  
450 455 460

Asp Gly Trp Gly Glu Phe Lys Val Asn Gly Gly Ser Val Ser Val Trp  
465 470 475 480

Val Pro Arg Lys Thr Thr Val Ser Thr Ile Ala Arg Pro Ile Thr Thr  
485 490 495

Arg Pro Trp Thr Gly Glu Phe Val Arg Trp Thr Glu Pro Arg Leu Val  
500 505 510

Ala Trp Pro  
515