

Integrating physiological threshold experiments with climate modeling to project mangrove species' range expansion

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Abstract

Predictions of climate-related shifts in species ranges have largely been based on correlative models. Due to limitations of these models, there is a need for more integration of experimental approaches when studying impacts of climate change on species distributions. Here, we used controlled experiments to identify physiological thresholds that control poleward range limits of three species of mangroves found in North America. We found that all three species exhibited a threshold response to extreme cold, but freeze tolerance thresholds varied among species. From these experiments, we developed a climate metric, freeze degree days (FDD), which incorporates both the intensity and the frequency of freezes. When included in distribution models, FDD accurately predicted mangrove presence/absence. Using 28 years of satellite imagery, we linked FDD to observed changes in mangrove abundance in Florida, further exemplifying the importance of extreme cold. We then used downscaled climate projections of FDD to project that these range limits will move northward by 2.2–3.2 km yr⁻¹ over the next 50 years.

Keywords: *Avicennia germinans*, climate change, ecological thresholds, freeze tolerance, *Laguncularia racemosa*, range expansion, *Rhizophora mangle*, species distribution modeling

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Introduction

A variety of studies have linked changing climate conditions to range shifts (Chen *et al.*, 2011), changes in phenology (Pau *et al.*, 2011), and altered community structure, function, and productivity in terrestrial, marine, and freshwater ecosystems (Parmesan, 2006). These changes highlight the need for accurate predictions of how species and ecosystems will respond to future climate change (Bellard *et al.*, 2012). In response to this need, correlative distribution modeling has become one of the most frequently used tools for predicting the future distributions of species. Correlative models link the presence/absence or abundance of species to environmental variables using statistical techniques (Guisan & Zimmermann, 2000). These types of models are easy to implement for a wide range of species as they do not require detailed knowledge of the mechanistic processes that control survival and reproduction, they use a commonly available data source, and they can be easily tailored to fit available data (Potter & Hargrove, 2013). However, correlative distribution models only

implicitly consider the processes that limit the potential range of a species. As a result, it is not always clear whether there is a causal relationship between the response and the predictor variables of the model (Meynard & Quinn, 2007). This can be particularly problematic when automated model selection strategies are applied to a large number of correlated predictor variables. Moreover, these models generally assume that the processes that set range limits are fixed in space and time, an assumption that may be violated when these models are used to predict climate-driven range shifts or to model species with nonequilibrium distributions (Kearney & Porter, 2009; Buckley *et al.*, 2010).

Another limitation of many distribution models is that they use environmental predictors based on climatic means. However, there is growing recognition that changes in the frequency and intensity of extreme weather events can profoundly impact ecosystems (Jentsch *et al.*, 2007; Lloret *et al.*, 2012; Cavanaugh *et al.*, 2014) and that incorporating measures of climate extremes can improve predictions of species distribution models (Zimmermann *et al.*, 2009). Moreover, including climate variability in species distribution models is important because climate change is expected to impact the frequency of extreme events (Easterling

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et al., 2000; IPCC, 2012). From an organism's point of view, extreme events occur when the acclimation capacities of the organism are exceeded (Gutschick & BassiriRad, 2003). Therefore, information about the organism's response to environmental conditions is needed to define what constitutes an extreme event.

Because of these limitations, there have been calls for more experimental approaches to studying the impacts of climate change (Thompson *et al.*, 2013; Kreyling *et al.*, 2014). Such experiments provide a mechanistic understanding of the processes controlling a species' distribution, which can be applied to nonequilibrium distributions and/or novel conditions (Kearney & Porter, 2009). This approach can be used to identify thresholds in the response of organisms to changes in environmental conditions (Kreyling *et al.*, 2014) so that the impacts of extreme events can be better understood. Ideally, these kinds of experiments will be incorporated in a framework that uses a variety of approaches, including distribution modeling, to formulate predictions about how climate change will impact species distributions (Dawson *et al.*, 2011). For example, mechanistic understanding gained from laboratory and field experiments can be used to develop highly proximal predictor variables for correlative models (Kearney & Porter, 2009).

Understanding how coastal foundation species will respond to climate change is particularly important. Seagrasses, kelps, salt marshes, and mangroves are all foundation species that structure communities by modifying the habitats and resources of other organisms, and therefore have a major influence on ecosystem structure and function (Ellison *et al.*, 2005). These coastal ecosystems are socially and economically important and provide trillions of dollars of value in ecosystem services each year (Costanza *et al.*, 1997).

In coastal wetlands near tropical–temperate transition zones, salt marshes have been displaced by a poleward expansion of tropical and subtropical mangrove trees in recent decades (Stevens *et al.*, 2006; Cavanaugh *et al.*, 2014; Saintilan *et al.*, 2014). Mangroves are limited to tropical and subtropical climates between ca. 30N and 40S, while salt marshes dominate more temperate coastlines (Duke *et al.*, 1998). While multiple environmental factors determine mangrove distributions (Duke *et al.*, 1998), cold temperatures are believed to control their poleward limits (Soares *et al.*, 2012). There is evidence that in some regions (e.g., southeast North America) the observed poleward expansion of mangroves is due to a decrease in the frequency of extreme cold events (Cavanaugh *et al.*, 2014). Correlative distribution modeling suggests that mangrove–salt marsh ecotones in many parts of the world may exist near temperature-related thresholds, where increasing winter tempera-

tures could lead to further mangrove expansion (Osland *et al.*, 2013; Record *et al.*, 2013).

While field observations (Cavanaugh *et al.*, 2014) and distribution models (Osland *et al.*, 2013) have suggested that mangroves exhibit a threshold response to freeze, only a handful of studies have experimentally examined freeze tolerance in mangroves (Markley *et al.*, 1982; Stuart *et al.*, 2007; Pickens & Hester, 2011). Markley *et al.* (1982) grew propagules of North American mangrove species in a common garden and found variability in freeze tolerance across species and between different populations of the same species. Stuart *et al.* (2007) performed controlled laboratory experiments that demonstrated variability in freeze tolerance across five species of North American and Australian mangroves. Neither of these studies was designed to identify freeze tolerance thresholds; they did not utilize a gradient of freeze intensities or durations (see Kreyling *et al.*, 2014). Pickens & Hester (2011) did examine the response of mangrove propagules and seedlings to three temperature treatments (5.7, 2.5, and -6.5 °C) and four durations (2, 6, 12, and 24 hours), but this study was limited to early life stages of one species of mangroves, *Avicennia germinans*.

In this study, we combined laboratory experiments, satellite-based observations, and climate modeling to (i) identify specific, temperature-related controls of mangrove range limits and (ii) forecast future poleward migration of three species of mangroves in Florida. We experimentally examined the freeze tolerance of the three dominant species of mangrove native to Florida, *A. germinans* (black mangrove), *Laguncularia racemosa* (white mangrove), and *Rhizophora mangle* (red mangrove). We then incorporated the mechanistic understanding derived from these experiments to identify the climate threshold controlling the range limit of each of the three species. We tested the relevance of these climate thresholds by examining the response of mangrove stands to past freeze events using 28 years of Landsat satellite imagery. Finally, we used downscaled climate model projections to forecast the future range limit and estimate the future rate of poleward migration for each species.

Materials and methods

Study area

Our study area included wetlands along the Atlantic coasts of Florida, Georgia, and South Carolina, USA from 26 to 33 N. The current northern limit of mangroves in eastern North America is located around 30 N, just north of St. Augustine FL (Williams *et al.*, 2014). Salt marshes dominate the more temperate climates to the north, whereas mangroves and salt marsh coexist in an ca. 2° ecotone to the south (28–30 N). South of 28 N, mangroves typically dominate coastal wetlands

(Kangas & Lugo, 1990). Mangrove wetlands in Florida are dominated by 1–3 species of short- to medium-sized trees, including black, red, and white mangrove. Recently, Williams *et al.* (2014) identified the northernmost individuals in Florida from each of the three mangrove species. The northernmost black mangrove was found at 30.11 N, 81.37 W; the northernmost red mangrove was found at 29.94 N, 81.32 W; and the northernmost white mangrove was found at 29.73 N, 81.24 W.

Determining physiological threshold responses of mangroves to severe cold

In June 2013, we collected branches from black, red, and white mangroves in Avalon State Park (27.55 N, 80.33 W). To maintain the physiological integrity of the branches, we removed a long branch from each tree, cut off the top ca. 15 cm of the branch under water, and placed this tip directly into floral water tubes. We then placed branches in a freezer for 4.5 hour trials, using a thermoregulator (Aqua Logic EC115R; Aqua Logic, Inc., San Diego, CA, USA) to maintain a steady temperature. We conducted a total of 21 different trials, with average minimum temperatures in each trial ranging from -0.2 to -11.0 °C (recorded with HOBO data loggers). We included branches from 20 individuals per species in almost all trials ($N = 420$ blacks, 418 reds, and 360 whites), except for the -5.3 °C trial, which had 18 red mangrove individuals and zero white mangroves, and the -4.2 °C and one of the -6.7 °C trials, which also had zero white mangroves.

To assess susceptibility to cold, we measured photoinactivation or the change in chlorophyll fluorescence yield (Y) after a freeze event. Change in yield is a common freeze tolerance metric (Cavender-Bares *et al.* 2005; Pérez *et al.*, 2014) that correlates with other freeze tolerance metrics (Boorse *et al.* 1998). We measured yield before and after the freeze treatment with a mini-PAM (Photosynthetic Yield Analyzer; Walz, Mess- und Regeltechnik, Effeltrich, Germany) and quantified photoinactivation as $1 - Y_{\text{post}}/Y_{\text{max}}$ (Pérez *et al.*, 2014). Photoinactivation thus ranged from zero (no loss of photosynthetic function) to one (complete loss of photosynthetic function). Because yield is sensitive to the light history of leaves, temperature, and current light conditions, we sampled analogous leaves across individuals. We marked leaves, so we could repeat measurements after the cold treatment at the exact same leaf location and orientation, and allowed branches to equilibrate to uniform laboratory light and temperature conditions for at least 1 hour before assessing yield. We analyzed photoinactivation with average minimum temperature and species identity as predictors using general linear models with logistic link functions (logistic regression). Species identity was incorporated into the models using dummy variables with white mangrove being the base case. We assessed the effect of species on freeze tolerance by examining the significance of the dummy variable coefficients.

Linking mangrove physiological thresholds to range limits

We created a series of models to relate mangrove presence/absence to modern climate. We characterized modern climate

using gridded daily minimum temperature (T_{min}) data for 1981–2010 from Maurer *et al.* (2002). The original resolution of the data was $1/8^\circ$, which we statistically downscaled to 30 arc second (ca. 800 m) geographic resolution (see Appendix S1 for more details). We then developed a mangrove presence/absence data set with the same spatial resolution as our climate data (800 m). Our analysis was restricted to cells that contained estuarine and marine wetlands according to the US Fish and Wildlife Service National Wetlands Inventory. Mangrove presence/absence in each cell was determined using data from two mangrove distribution data sets (Spalding *et al.*, 2010; Giri *et al.*, 2011b). A composite of the two data sets was used to minimize errors of omission. The available mangrove presence/absence data sets did not differentiate among species, and so all three species of mangroves were grouped together for this analysis. From the daily climate data, we calculated mean daily T_{min} , mean winter (December–February) T_{min} , mean winter minimum T_{min} , and the overall minimum T_{min} of the record for each grid cell for the time period from 1980 to 2010.

Based on the results from our freeze tolerance experiments, we also developed a new winter severity index, freeze degree days (FDD). First, we used the modeled logistic relationship between photoinactivation (from the laboratory analysis) and freeze intensity to identify the temperature that corresponded to 10% photoinactivation (Fig. 1). A cut-off of 10% photoinactivation was used to identify the base temperature because it would be more likely to capture the impacts of minor freezes than a threshold based on the inflection point of the sigmoid curve (i.e., the temperature corresponding to 50% photoinactivation). We used the threshold temperature of the most tolerant species, *A. germinans*, which resulted in a threshold of -3.2 °C. Selecting this base temperature is akin to calculating

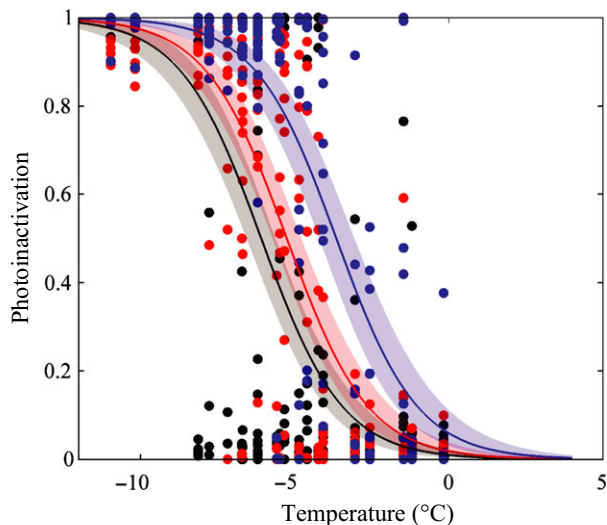


Fig. 1 Relationship between temperature treatment and photoinactivation for branches from three mangrove species from laboratory studies. Black lines represent *Avicennia germinans*, red lines *Rhizophora mangle*, and blue lines *Laguncularia racemosa*. Shaded regions represent 95% confidence interval.

a base temperature for calculating growing degree-days; however, in that case, the goal is to identify the temperature below which plant growth and development ceases (Yang *et al.*, 1995). FDD was calculated as,

$$\text{FDD} = \max([0T_{\text{base}} - T_{\text{min}}]), \quad (1)$$

with T_{base} being the temperature that corresponded to 10% photoinactivation in the above experiment (-3.2 °C). We examined the sensitivity of our results to varying T_{base} from 0 to -4 °C (0–35% photoinactivation). Because our mangrove presence/absence data did not differentiate species, we used the threshold temperature of the most tolerant species, *A. germinans*, to calculate FDD. The average annual FDD was calculated for each cell between 1980 and 2010. Finally, we calculated the annual number of days below the threshold to distinguish the impact of freeze frequency from that of freeze intensity. The daily T_{min} -based temperature metrics were highly correlated (the absolute value of the mean and minimum bivariate correlations were 0.93 and 0.84, respectively; Table S1), so we developed single predictor models and used Akaike's information criterion (AIC) to measure the relative quality of each model.

The FDD-based mangrove presence/absence model had the lowest AIC and so was selected as the optimal model (Fig. 2). Relative likelihood that another model would minimize information loss, calculated following (Burnham & Anderson, 2002),

$$\exp((\text{AIC}_{\text{min}} - \text{AIC}_i)/2), \quad (2)$$

was $\ll 0.001$ for all models. To further test the idea that FDD influences the northern range limit for mangroves, we compared changes in mangrove area between summers to the cumulative FDD during the intervening winter. We used time series of mangrove area from 1984 to 2011 derived from Landsat 5 Thematic Mapper (TM) imagery collected each summer (Cavanaugh *et al.*, 2014). Imagery for 1985, 1998, 2004, and 2010 was not available due to cloud cover. Using a handheld GPS, we identified areas in Avalon State Park (where branches were collected for laboratory analysis, 27.55 N, 80.33 W) and Sebastian Inlet State Park (27.85 N, 80.45 W) that contained large (>30 m \times 30 m), monospecific stands of red, black, and white mangroves in close proximity to each other. We then identified the Landsat pixels that were within each outlined stand such that the Landsat pixels represented pure species-specific stands of mangrove trees. We then calculated each year's cumulative FDD value for those pixels. For each of the three stands in each of the two sites, we compared the annual change in mangrove area between successive summers to the cumulative FDD of the intervening winter for each year that experienced a freeze event. Freeze events occurred at the two sites in nine of the years for which we had satellite imagery. However, minimum temperatures fell below the FDD base temperature (-3.2 °C) in only one (1989) of those 9 years. In that year, the minimum temperature was -4.5 °C at the Sebastian Inlet site and -5.3 °C at the Ft. Pierce site. In the other 8 years that experienced freezes, cumulative FDD was 0. As we did not have a range of FDD values, we could not justify fitting a curve relating change in mangrove area to FDD (i.e.,

the approach we used to analyze the laboratory experiment). Instead, we grouped the data from the two sites and performed an ANOVA to compare years that experienced a freeze event, but had zero FDD to years with FDD > 0 . We used Bonferroni-adjusted *t*-tests to compare species-specific responses.

The current latitudinal range limits for black, red, and white mangroves were used to infer the climate envelope for each species. We used field surveys to identify the northernmost stand of black, red, and white mangroves where stand equaled a contiguous group of >10 trees. Note that we did not use the range limits identified by Williams *et al.* (2014) as they represented the locations of isolated individuals. The FDD value at the location of the northernmost stands of black, red, and white mangroves was inferred to be the threshold for the presence of each of the three mangrove species.

Projections of future mangrove range limits

We used downscaled climate projections to forecast future range limits for each species. To characterize future climatic conditions, we used daily T_{min} from 2030 to 2060 from an ensemble of 21 general circulation models (GCMs) as part of the 5th phase of the Coupled Model Intercomparison Project (CMIP5), developed for the most recent report by the Intergovernmental Panel on Climate Change (IPCC). We considered the most severe emissions scenario (RCP 8.5) for each GCM to characterize the upper bound on projected trends (e.g., Scoccimarro *et al.*, 2013). We alternatively projected future range limits for each species using the RCP 4.5 scenario. As with the observed T_{min} data, we statistically downscaled the already downscaled $1/8^\circ$ data of each GCM (Maurer *et al.*, 2007) to a resolution of 30 arc seconds. We then identified grid cells where the projected climate data crossed the mangrove presence FDD threshold for each species. We also forecasted future range limits for each species using thresholds based on the other climate variables [mean daily T_{min} , mean winter (December–February) T_{min} , mean winter minimum T_{min} , minimum T_{min} and mean number of days less than 3.2 °C].

Results

All three species of mangroves demonstrated a threshold response to freezing temperatures, with large reductions in performance occurring between -2 and -6 °C (Fig. 1). Freeze tolerance varied among species in the laboratory experiments (Table 1). Black mangroves were the most freeze tolerant and white mangroves were the least tolerant. This result matched the observation that black mangroves have the most northward distribution in Florida, followed by red mangroves and then white mangroves (Williams *et al.*, 2014).

We used these freeze response curves to develop our mangrove-specific winter severity index, FDD, with $T_{\text{base}} = -3.2$ °C. There was a significant relationship between all five of our temperature metrics and

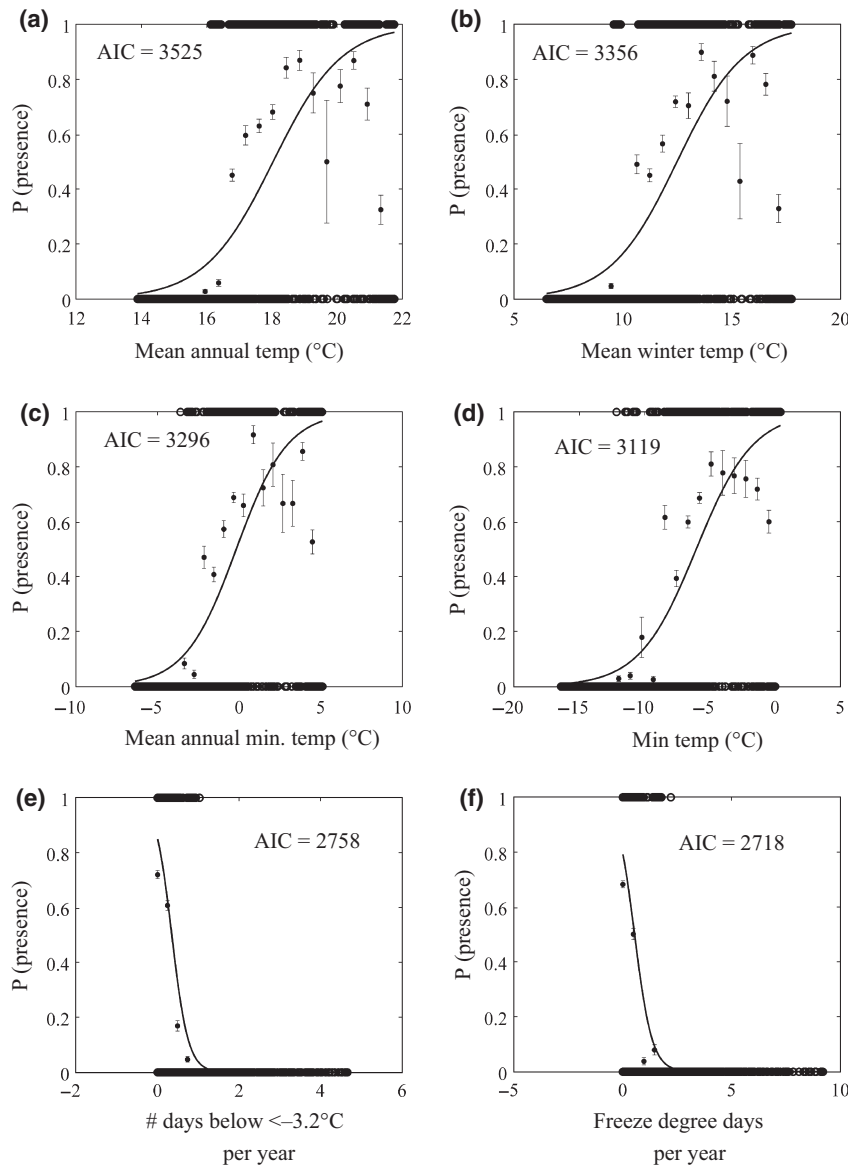


Fig. 2 Logistic relationships between mangrove presence/absence and (a) mean annual temperature, (b) mean winter temperature, (c) mean annual minimum temperature, (d) minimum temperature, (e) number of days less than $-3.2\text{ }^{\circ}\text{C}$ per year and (f) degree days less than $-3.2\text{ }^{\circ}\text{C}$ per year. Closed circles and error bars give the mean and standard error of presence/absence data separated into 20 bins. Lines give the logistic regressions for the relationships. Solid lines are significant at $P < 0.01$. Open circles give the raw presence/absence data.

Table 1 Results from dummy variable logistic regression comparing photoinactivation to temperature treatment and species type in laboratory studies

Variable	Estimate	Std. error	df	<i>t</i>	<i>P</i>
Intercept (species – white)	-2.77	0.35	594	-7.9	<0.01
Temperature	-0.76	0.06	594	-11.7	<0.01
Species – black	-4.62	0.29	594	-6.3	<0.01
Species – red	-3.99	0.29	594	-4.2	<0.01

mangrove presence ($P < 0.01$ for all logistic models; Fig. 2). However, mean annual FDD was the temperature variable that best predicted mangrove presence based on AIC (Fig. 2). As a result, we used FDD to infer the current temperature-related threshold for mangrove presence and to predict future mangrove range limits for black, red, and white mangroves. When the threshold for calculating FDD was varied from 0 to $-4.0\text{ }^{\circ}\text{C}$, AIC varied a minor amount, but still remained lower than for the other temperature variables (Table S2).

Our Landsat analysis demonstrated that years when FDD was >0 experienced significantly higher loss of mangrove area than years that experienced minor freezes where minimum temperature was greater than -3.2°C (Fig. 3; Table 2). Some species-specific variability in freeze tolerance was observed during the extreme freeze event, with black mangroves experiencing the least amount of loss (Fig. 3).

The northernmost stand of black mangroves was located at 29.92°N , 81.31°W ; the northernmost stand of red mangroves was located about 23 km further south at 29.73°N , 81.24°W ; and the northernmost stand of white mangroves was located about 7 km south of the red mangroves at 29.66°N , 81.22°W (Fig. 4). At these current range limits of black, red, and white mangroves, the average FDD per year between 1980 and 2010 were 2.1, 1.5, and 1.5, respectively (Fig. 4). Based on RCP 8.5 climate projections of where these FDD

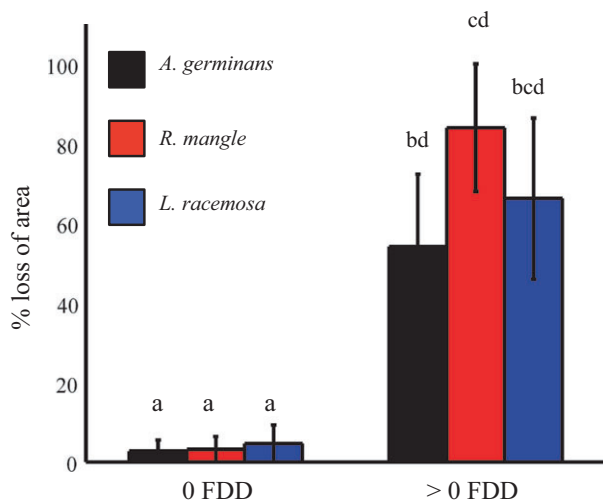


Fig. 3 Comparison of the impact of minor freezes with 0 freeze degree days (FDD) to hard freezes with FDD >0 for stands of black, red, and white mangroves. $n = 16$ for each species in the 0 FDD case and $n = 2$ for each species in the >0 FDD case. Unique letters denote significant differences based on Bonferroni-adjusted t -tests. Error bars give the interquartile range of the responses.

Table 2 Results from analysis of variance comparing the impact of minor freezes with 0 freeze degree days (FDD) to hard freezes with FDD >0 for stands of black, red, and white mangroves. Data for this analysis came from Landsat observations of change in mangrove cover between successive summers

Source	SS	df	MS	F	P
FDD	23133.79	5.00	4626.76	61.03	<0.01
Error	3638.83	48.00	75.81		
Total	26772.63	53.00			

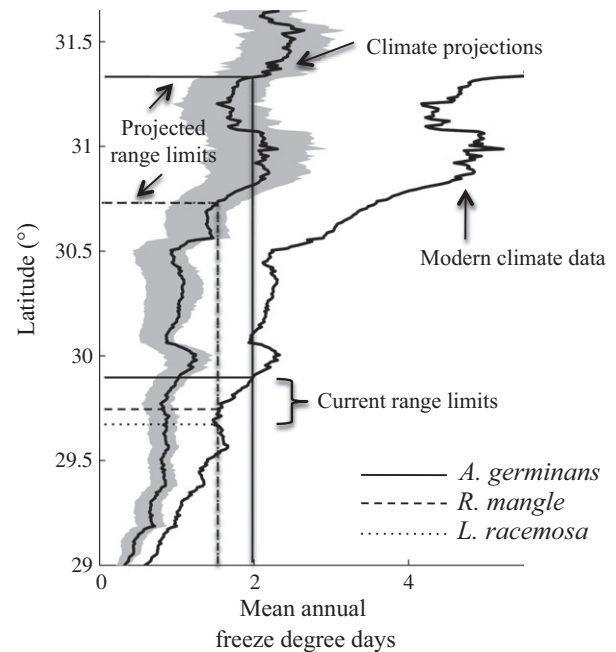


Fig. 4 The mean annual freeze degree days (FDD) for modern climate (1980–2010) and for climate projections (2030–2060) along the Atlantic coastlines of Florida and Georgia from 29° to 31.5°N . The current northernmost stands of *Avicennia germinans*, *Rhizophora mangle*, and *Laguncularia racemosa* are shown by the lower horizontal solid, dashed, and dotted lines. Based on the mean annual FDD at the current range limits (2.1 for *A. germinans* and 1.5 for *R. mangle* and *L. racemosa*), we project future range limits in 2060 (upper horizontal solid, dashed, and dotted lines). The upper dotted and dashed lines showing the projected range limits of *R. mangle* and *L. racemosa* overlap and so are difficult to distinguish from one another. The shaded area around the climate projection line gives the interquartile range of the 21 climate projection models we examined.

thresholds will occur in 2060, we project that the northern range limits for black, red, and white mangroves on the Atlantic coast of North America will be located at 31.33°N [interquartile projections: (31.20°N , 31.50°N)], 30.71°N (30.55°N , 31.35°N), and 30.71°N (30.55°N , 31.35°N), respectively (Figs 4 and 5). These projections correspond to northward migration rates of 3.2 (2.9 , 3.7), 2.2 (1.9 , 3.7), and 2.4 (2.0 , 3.8) km yr^{-1} . Using the other climate metrics [mean daily T_{min} , mean winter (December–February) T_{min} , mean winter minimum T_{min} , minimum T_{min} and mean number of days less than -3.2°C] resulted in range limit projections that ranged from 82 km south to 33 km north of the projections made using FDD (Table S3). As expected, range limit models built from RCP 4.5 climate projections predict slightly slower northward migration of range limits, reaching 31.23°N by 2060 [interquartile projections: (30.69°N , 31.50°N)], 30.55°N (30.50°N , 31.31°N), and 30.55°N (30.50°N , 31.31°N), respectively (Fig. S1).

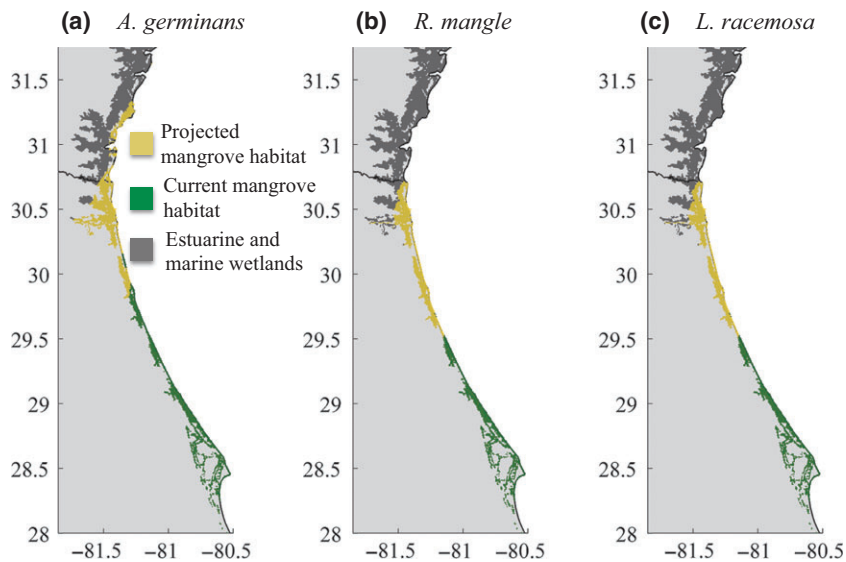


Fig. 5 Map of US southeastern Atlantic coastline showing the modeled current and projected (2060) extents of (a) *Avicennia germinans*, (b) *Rhizophora mangle*, and (c) *Laguncularia racemosa*. Projected range limits are based on the mean of the ensemble CMIP 5 RCP 8.5 projections.

Discussion

We used controlled laboratory studies to identify physiological thresholds in mangroves and develop a simple climate-based model to identify the climatic variable (FDD) that sets the poleward range limits for the three species of mangroves in North America. We then validated the role of FDD using Landsat observations of annual changes in mangrove area on landscape scales in response to cold winters. Finally, a simple climate-based modeling approach was used to project future movement of these range limits. This approach of integrating controlled laboratory studies with multi-decadal satellite observations and climate models has the potential to improve predictions of the impacts of climate change on ecosystems (Dawson *et al.*, 2011; Kreyling *et al.*, 2014). Biological responses to temperature changes can be nonlinear and characterized by thresholds (Gutschick & BassiriRad, 2003). Controlled laboratory experiments provide a means for identifying these thresholds. However, these kinds of experiments do not capture the complexity of real systems and so results can be difficult to scale up to the ecosystem level (Kreyling *et al.*, 2014). Field observations and experiments can provide a way to validate and scale up these laboratory results. These can consist of observations of how the system has responded to past changes (e.g., the retrospective satellite-based approach used here) or of manipulative field experiments (e.g., Beier *et al.*, 2004; Loik *et al.*, 2004; Lloret *et al.*, 2005). Modeled climate projections can then be used to infer the response of the ecosystem to future environmental changes.

However, it is important that climate projections, which typically have coarse spatial resolutions, are down-scaled so they can provide predictions at scales that are ecologically meaningful and relevant to local management (i.e., county and state levels).

We found that northern range limits of mangroves in southeast North America are controlled by both frequency and intensity of discrete, extreme cold events. Previous studies characterized the impacts of freeze events on mangrove structure and mortality on local scales (Stevens *et al.*, 2006; Ross *et al.*, 2009). A number of studies have documented the impacts of an unusual series of freezes that occurred in the 1980s and led to massive amounts of mangrove mortality across the southeastern United States (Lonard & Judd, 1991; Montague & Odum, 1997; Stevens *et al.*, 2006; Giri *et al.*, 2011a). More recent work has demonstrated that extreme cold is more important than mean temperature in controlling the abundance and distribution of mangroves on regional scales (Osland *et al.*, 2013; Cavanaugh *et al.*, 2014). For example, Cavanaugh *et al.* (2014) demonstrated that a decrease in the frequency of extreme cold events between 1984 and 2010 corresponded to regional increases in mangrove cover over that same time period. Osland *et al.* (2013) found that the average lowest temperature of the year, a measure of severity, was a strong predictor of mangrove presence and abundance. We built off of these studies by creating a new metric that integrates the frequency and intensity of extreme cold events, FDD. This metric was a slightly better predictor of mangrove presence/

absence than mean annual minimum temperature, a measure of freeze severity, or the average number of days below $-3.2\text{ }^{\circ}\text{C}$, our measure of freeze frequency. FDD is correlated with the other winter severity metrics evaluated; however, we observed differences of 11–73 km when we compared projections made with FDD to those made with mean annual minimum temperature and the average number of days below $-3.2\text{ }^{\circ}\text{C}$ (Table S3). These differences are small relative to the full distributional range of mangroves, but they are comparable to the differences between the current range limits of black, red, and white mangrove species. Although we do not have the data to evaluate the performance of these different predictions, our FDD model has a stronger theoretical grounding as it can capture the effect of years that experience multiple extreme cold events. Most of the winters that were characterized as ‘severe freeze seasons’ by the Florida Crop and Livestock Service experienced sustained cold spells or multiple distinct freeze events (Miller & Downton, 1993). Identifying the specific proximate controls of species distributions should facilitate more accurate predictions of how future climate change will alter species distributions (Buckley *et al.*, 2010), particularly for systems that demonstrate nonlinear and threshold dynamics.

Our FDD metric was developed using mechanistic knowledge gained from controlled laboratory studies and validated using field observations of landscape-scale responses of mangroves to past freeze events. Previous controlled experimental tests of mangrove freeze tolerance suggest that cold tolerance varies among species (Markley *et al.*, 1982; Stuart *et al.*, 2007; Cook-Patton *et al.*, in review), life stages (Pickens & Hester, 2011), and populations (Markley *et al.*, 1982; Cook-Patton *et al.*, in review). However, these studies did not examine the response of different species of adult mangroves along a gradient of temperature treatments, so they may not be optimally useful when deriving species-specific thresholds in mangrove freeze tolerance. In our experiments, all three species of mangroves exhibited a threshold response to decreasing temperatures (Fig. 1). Our satellite observations of the impacts of past freeze events also provide evidence for a threshold response of mangroves to freeze on landscape scales. Mangrove stands were relatively unaffected by minor freezes where temperatures remained above $-3.2\text{ }^{\circ}\text{C}$ (Fig. 3). However, stands experienced large losses in area when temperatures fell below this threshold.

The laboratory studies of species-level variability in freeze tolerance matched the latitudinal distribution of mangroves in our study area. The most freeze-tolerant species in our experiments, black mangroves, has the

northernmost distribution, while the least freeze-tolerant species, white mangroves, has the southernmost range limit. In addition, the black mangrove stands showed the highest levels of freeze tolerance in the Landsat analyses. However, we were not able to detect significant difference between red and white mangroves in the Landsat analyses. This may be due to the fact that there were very few extreme freeze events at our field sites (only one winter experienced a minimum temperature less than $-5\text{ }^{\circ}\text{C}$), so there are higher levels of uncertainty regarding the landscape-scale effects of these severe freezes. In addition, there is evidence that effect size, that is the impact of a given treatment, in climate manipulation experiments decreases with spatial scale due to an increasing number of influential processes (Leuzinger *et al.*, 2011). For example, microenvironments and microclimates make some locations more habitable than would be expected from a geographically gridded climate product. As a result, the impact of severe freeze events might be more spatially heterogeneous than would be expected from gridded climate data.

While red mangroves are found 7 km further north than white mangroves (ca. 7 km), the average FDD at both range limits is the same (1.5). The small difference in range limits of these species may be caused by the differences in freeze tolerance identified by our experimental results. It is possible that there is in fact a small difference in FDD between the locations of the red and white mangrove range limits, but our downscaled climate data did not have sufficient accuracy to detect a difference. On the other hand, if the climatic conditions at these two locations are the same, the difference in range limits could be due to a process such as variability in dispersal capabilities (Rabinowitz, 1978). The current range limit of black mangroves is about 23 and 30 km north of the respective range limits of red and white mangroves. However, the range limit of black mangroves was characterized by a higher FDD per year (2.1) than the range limit for reds and whites (1.5 in both cases). The nature of the threshold response of mangroves to freezes suggests that small differences in freeze frequency and intensity could dictate a given site’s suitability for individual species, highlighting the value of considering high-resolution climate data sets when possible. Figure 4 indicates a relatively steep gradient in FDD between the range limits of red (29.73 N) and black (29.92 N) mangroves. The shape of the current FDD profile (Fig. 4) provides insight into the potential for range limit migration (i.e., range infilling) under current climatic conditions. For example, at 30.1 N, the mean average FDD per year during 1981–2010 is below 2.1, suggesting that black mangrove stands may be able to persist at this latitude in the current climate if

they were able to disperse and establish in suitable habitat there. This process may be occurring, as there has been a recent account of an individual black mangrove shrub at 30.1 N (Williams *et al.*, 2014).

We project that over the next 50 years, if not limited by dispersal or the availability of suitable habitat, the northern range limits of black, red, and white mangroves will move north by about 160, 110, and 120 km, respectively. This corresponds to migration rates of 3.2, 2.2, and 2.4 km yr⁻¹ for black, red, and white mangroves. Williams *et al.* (2014) compared observations of the northernmost mangrove individuals for each species from 2007 to surveys conducted in 2013 and estimated hypothetical migration rates of 4.5, 3.7, and 1.3 km yr⁻¹, respectively for black, red, and white mangroves. These are short-term changes that almost certainly do not reflect long-term migration rates due to local climate oscillations, dispersal variability, and other stochastic processes. In a meta-analysis of range shifts observed for a variety of animal species over the course of 8–50 years, Chen *et al.* (2011) found that the median rate of poleward movement was 1.69 km yr⁻¹. It is surprising that the observed (Williams *et al.*, 2014) and estimated (this study) migration rate of mangroves would be higher than that of many animal species, given that model predictions and observations suggest that changes to the range limits of trees in response to climate change are slow and largely stochastic (Clark *et al.*, 2003). One explanation for rapid mangrove expansion is that water-dispersed plants such as mangroves often can travel further than those dispersed by wind or animals (Ridley, 1930). There is also a large amount of suitable mangrove habitat (i.e., tidal saline wetlands) in our study area, which could facilitate mangrove expansion. Competition with existing salt marsh vegetation has the potential to modulate the poleward expansion of mangroves, but mangroves appear to be competitively dominant in areas where abiotic conditions are suitable (Kangas & Lugo, 1990). We do not address the impacts of future sea-level rise in this paper as we were focused on poleward rather than landward changes to mangrove ranges. However, that process has the potential to alter the availability of suitable habitat. Landward expansion of mangroves due to sea-level rise has already been documented in Baja California (López-Medellín *et al.*, 2011), the US Gulf Coast (Krauss *et al.*, 2011), and the east coast of Australia (Rogers *et al.*, 2006). If landward migration of mangroves keeps pace with shoreline erosion, then the poleward movement of mangroves' range limits may not be impacted directly by sea-level rise. However, coastal development may eliminate the potential for landward migration in some areas, which could reduce the availability of suitable habitat and slow migration

rates. While this paper focuses on regional changes in mangrove range limits, local abiotic factors such as salinity, inundation frequency, and nutrient availability will certainly control mangrove suitability on smaller scales.

Theoretically, including mechanistic understanding into distribution models should improve predictions of responses to climate change (Buckley *et al.*, 2010). A possible next step would be to develop a fully mechanistic model of mangrove distributions. To do this, more detailed data on the functional traits of mangroves are needed. For example, we have identified freeze tolerance thresholds at the branch and leaf level; however, this experimental gradient approach has not yet been used to identify temperature thresholds for seedlings or whole adult plants. Further insight into the physiological response of mangroves to cold could be gained through experiments that decouple the effects of freezing intensity and freezing duration. There is also variability in the mechanisms by which various species of mangroves recover from extreme cold events. For example, *A. germinans* has the ability to resprout after parts of the plant have been killed by freeze, but *R. mangle* does not (Sherrod & McMillan, 1985). This characteristic could impact the recovery of mangrove populations in ways not captured by branch and leaf-scale freeze tolerance experiments. *Rhizophora mangle* would have to rely on dispersal from surviving populations to recolonize areas after disturbance events, which could delay range infilling. There are also questions about how cold impacts plant growth and reproduction. There is evidence of geographic variability in traits related to freeze tolerance in some species of mangroves (Cook-Patton *et al.*, *in review*), which also could be incorporated into mechanistic models.

While the frequency and intensity of cold events appear to set the poleward range limit of mangroves in southeastern North America, it is likely that other factors such as precipitation, ocean circulation, sea surface temperatures, atmospheric aridity, geomorphic setting, and controls on dispersal will play important roles in other parts of the world (Stuart *et al.*, 2007; Quisthoudt *et al.*, 2012; Saintilan *et al.*, 2014). For example, freezes are rare at the arid poleward range limits of mangroves in Baja California and Peru (Quisthoudt *et al.*, 2012). In addition, (Wilson & Saintilan, 2012) found that the current poleward range limit of *Rhizophora stylosa* on the east coast of Australia is well within the thermal tolerance limits of that species. If mechanistic niche models are to be applied in other regions, we will need additional data on salinity tolerance, water, light, and nutrient requirements, and other biophysical characteristics across the different life stages of mangroves.

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Supporting Information

Additional Supporting Information may be found in the online version of this article:

Figure S1. Map of US southeastern Atlantic coastline showing the modeled current and projected extents of (a) *Avicennia germinans*, (b) *Rhizophora mangle*, and (c) *Laguncularia racemosa*. Projected range limits are based on the mean of the ensemble CMIP 5 RCP 4.5 projections.

Table S1. Correlation coefficients of temperature metrics.

Table S2. Logistic model results for predicting mangrove forest presence using freezing degree days calculated with varying threshold temperatures.

Table S3. Projected range limits for mangrove species from models based on alternative temperature metrics.

Appendix S1. Details about our downscaling methods and use of climate projection models.