

Carbapenem-Resistant and Extended-Spectrum β -Lactamase-Producing Enterobacterales in Children, United States, 2016–2020

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Learning Objectives

Upon completion of this activity, participants will be able to:

- Distinguish the most common bacteria associated with carbapenem-resistant Enterobacterales (CRE) among US children and adolescents
- Distinguish the most common bacteria associated with extended-spectrum β -lactamase-producing Enterobacterales (ESBL-E) among US children and adolescents
- Analyze how age affects the risk for CRE and ESBL-E among children and adolescents
- Evaluate other characteristics of pediatric patients with CRE and ESBL-E in the United States

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We conducted surveillance for carbapenem-resistant Enterobacterales (CRE) during 2016–2020 at 10 US sites and extended-spectrum β -lactamase-producing Enterobacterales (ESBL-E) during 2019–2020 at 6 US sites. Among 159 CRE cases in children (median age 5 years), CRE was isolated from urine for 131 (82.4%) and blood from 20 (12.6%). Annual CRE incidence rate (cases/100,000 population) was 0.47–0.87. Among 207 ESBL-E cases in children (median age 6 years), ESBL-E was isolated from urine of 196 (94.7%) and blood of 8 (3.9%). Annual ESBL-E incidence rate was 26.5 in 2019 and 19.63 in 2020. CRE and ESBL-E rates were >2-fold higher among infants than other age groups. Most CRE and ESBL-E cases were healthcare-associated community-onset (68 [43.0%] for CRE vs. 40 [23.7%] for ESBL-E) or community-associated (43 [27.2%] for CRE vs. 109 [64.5%] for ESBL-E). Programs to detect, prevent, and treat multidrug-resistant infections must include pediatric populations (particularly the youngest) and outpatient settings.

Increasing antimicrobial resistance (AMR) remains a critical public health threat (1,2). Carbapenem-resistant Enterobacterales (CRE) have been identified as an urgent public health threat and extended-spectrum β -lactamase (ESBL)-producing Enterobacterales (ESBL-E) as a serious public health threat (1). Both bacteria types remain of concern because of transmissibility of the AMR genes they harbor and limited treatment options. Particularly concerning are plasmid-mediated resistance mechanisms in which genes encoding carbapenemases and ESBLs, as well as other resistance determinants, can disseminate between different organisms, thus furthering the spread of CRE and ESBL-E (3). Knowledge of the burden of these infections has implications for public health and the control strategies needed to prevent spread in adult and pediatric populations.

Most US studies have focused on risk factors for infection or colonization of CRE and ESBL-E in adults; national epidemiologic data for children are comparatively lacking (4–9). Moreover, attention to antimicrobial-resistant bacterial infections in

children has perhaps been further disrupted by the COVID-19 pandemic, and trends have potentially worsened over the past 3 years. Few antimicrobial drugs can treat CRE and ESBL-E infections (10), and limited pediatric-specific clinical trials of antimicrobial drugs contribute to a scarcity of knowledge with regard to children compared with adults (11). A small number of studies have described the continued emergence of AMR mechanisms in US children (12–16) and identified variations in epidemiology by hospital and bacteria species across multiple pediatric medical centers (17–19). However, most studies were conducted in earlier years and were not designed to characterize the clinical and molecular features of cases identified from community and hospital settings on a population level.

The Centers for Disease Control and Prevention (CDC) Emerging Infections Program (EIP) conducts laboratory and population-based surveillance for CRE and ESBL-E in diverse US sites through the Multi-site Gram-negative Surveillance Initiative (<https://www.cdc.gov/hai/eip/mugsi.html>). Using those data, we focused on the descriptive and comparative epidemiology of CRE and ESBL-E, 2 of the most pressing gram-negative bacteria resistance threats, in US children.

Our study activity was reviewed by CDC, deemed not research, and was conducted consistent with applicable federal law and CDC policy (e.g., 45 C.F.R. part 46.102(l)(2), 21 C.F.R. part 56; 42 U.S.C. §241(d); 5 U.S.C. §552a; 44 U.S.C. §3501 et seq.). Similarly, the protocol was reviewed by all participating EIP sites and either was deemed nonresearch or received institutional review board approval with a waiver of informed consent.

Methods

Surveillance Population

As of 2016, county-level CRE surveillance was conducted in selected US metropolitan counties at 8 EIP

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sites (Colorado, Georgia, Maryland, Minnesota, New Mexico, New York, Oregon, Tennessee); surveillance subsequently expanded to 2 additional sites (California in 2017 and Connecticut statewide in 2018) (20). The total population of the 10 participating areas under surveillance in 2020 was an estimated 23.2 million, of which an estimated 4.9 million were children (21).

County-level ESBL-E surveillance started in July 2019 in selected counties at 6 EIP sites (Colorado, Georgia, Maryland, New Mexico, New York, Tennessee). The total population of the 6 participating areas under surveillance in 2020 was an estimated 3.0 million, of which an estimated 626,000 were children (21) (Appendix, <https://wwwnc.cdc.gov/EID/article/30/6/23-1734-App1.pdf>).

Case Definitions and Data Collection

Beginning in 2016, we defined an incident pediatric CRE case as the first isolation during a 30-day period of *Klebsiella pneumoniae*, *K. oxytoca*, *K. aerogenes*, *Enterobacter cloacae* complex, or *Escherichia coli* resistant to ≥ 1 carbapenem (imipenem, meropenem, doripenem, ertapenem) from a normally sterile body site (Appendix) or urine specimen from a surveillance area resident < 18 years of age. We defined an incident pediatric ESBL-E case as the first isolation during a 30-day period of *E. coli*, *K. pneumoniae*, *K. variicola*, or *K. oxytoca* resistant to ≥ 1 extended-spectrum cephalosporin (ceftazidime, cefotaxime, or ceftriaxone) and nonresistant (i.e., susceptible or intermediate) to all tested carbapenems from a normally sterile body site or urine specimen from a surveillance area resident < 18 years of age. To prevent duplication with CRE surveillance, we excluded ESBL-E isolates that were carbapenem resistant. For both ESBL-E and CRE, if a new specimen meeting the case definition was collected > 30 days after the patient's last incident case with the same organism, it was also reported as an incident case. CRE and ESBL-E cases were identified through a query of automated testing instruments based on laboratory protocols (Appendix).

All incident CRE cases, as well as all incident ESBL-E cases from a sterile source, underwent medical record review by using a standardized case report form to collect patient demographics, underlying conditions, healthcare exposures and outcomes, location of specimen collection, associated infection types, and antimicrobial susceptibility testing results (20) (Appendix). For ESBL-E cases identified from urine sources, for each year, first incident cases per species in a patient were reviewed.

Cases were considered hospital onset if the incident culture was collected > 3 days after hospital

admission. All other cases were considered community onset and further classified as either 1) health-care-associated if the person had hospitalization, surgery, residence in a long-term care facility or long-term care acute care hospital, or chronic dialysis in the year before culture or had an indwelling device in the 2 days before culture; or 2) community-associated if none of those risk factors were identified.

Isolate Collection

We submitted a convenience sample of isolates from all EIP sites to CDC for confirmatory and molecular characterization. CRE and ESBL-E isolates underwent species identification by matrix-assisted laser desorption/ionization time-of-flight mass spectrometry (MALDI-TOF Biotyper 3.1; Bruker Daltronics, <https://www.bruker.com>). We conducted antimicrobial susceptibility testing of CRE isolates by using reference broth microdilution with a metallo- β -lactamase screen (22,23), screening for carbapenemases by using the modified carbapenem inactivation method (24), and real-time PCR testing for *bla*_{KPC}, *bla*_{NDM}, *bla*_{VIM}, *bla*_{IMP}, and *bla*_{OXA-48-like} genes (25–28). If a CRE isolate harbored a carbapenemase-producing gene according to PCR, it was classified as carbapenemase producing (CP). We used real-time PCR to screen all isolates with a colistin MIC ≥ 2 $\mu\text{g}/\text{mL}$ for plasmid-mediated colistin resistance genes (*mcr-1* and *mcr-2*) (29). We conducted antimicrobial susceptibility testing of ESBL-E by using reference broth microdilution and performed phenotypic screening for ESBL production with ceftazidime and cefotaxime alone and in combination with clavulanate (24). We conducted whole-genome sequencing on a subset of CRE isolates from 2016–2018 that were confirmed to be carbapenem resistant and on the subset of ESBL isolates received from CDC (Appendix).

Statistical Analyses

We calculated crude incidence rates by using case counts and 2016–2020 US Census estimates of the surveillance area population < 18 years of age. For incidence rates presented by region, demographics, or age group, denominators represent the pediatric population also stratified by that subgroup. Analysis was limited to case report forms completed as of November 29, 2022. We performed descriptive and comparative analyses for CRE and ESBL-E cases by using the χ^2 test or the Fisher exact test (where applicable) for categorical variables and the Wilcoxon rank sum test for continuous variables. We used SAS version 9.4 (SAS Institute Inc, <https://www.sas.com>) to conduct data analyses.

Results

Cases and Incidence Rates

During 2016–2020, a total of 159 incident CRE cases were identified in 142 children across 10 EIP sites. Of the 159 cases, 83 (52.2%) isolates were *E. cloacae* complex, 50 (31.5%) *E. coli*, 17 (10.7%) *K. pneumoniae*, 5 (3.1%) *K. aerogenes*, and 4 (2.5%) *K. oxytoca* (Table 1). The number of CRE cases per EIP site ranged from 3 to 47. *E. coli* comprised half or most of the CRE cases in New Mexico (50.0%), Tennessee (71.4%), and California (85.7%), whereas *E. cloacae* complex were the most common organisms at the other sites (44.4%–71.4%). Of the 142 unique persons with CRE, during the 5-year surveillance period ≥2 incident cultures were obtained from 17 (12.0%) (range 2–6 episodes).

During 2019–2020, a total of 207 incident ESBL-E cases were identified in 184 children across the 6 participating EIP sites. Of the 207 cases, 182 (87.9%) isolates were *E. coli*, 23 (11.1%) *K. pneumoniae*, and 2 (1.0%) *K. oxytoca* (Table 1). The number of ESBL-E cases per EIP site ranged from 14 to 57 cases; at all 6 sites, the predominant organism was *E. coli* (82.5%–100.0%). Of the 184 unique persons with ESBL-E, during the 1.5-year surveillance period, ≥2 incident cultures were obtained from 23 (12.5%) (range 2–4 episodes).

The overall annual CRE incidence rate (cases/100,000 pediatric population) across EIP sites during the 5-year period was 0.70 (range 0.47–0.87). The overall annual ESBL-E incidence rate during the 1.5-year period was 23.08, decreasing from 26.54 in 2019 to 19.63 in 2020. Crude incidence rates for CRE and ESBL-E varied by geographic region and year (Table 2).

During 2016–2020, annual incidence rates for infants (children <1 year of age) were consistently higher than those for other age groups, ranging from 1.95 to 3.82 cases/100,000 pediatric population for CRE and 46.85 to 91.97 cases/100,000 pediatric population for ESBL-E (Figure 1). In addition, crude annual incidence rates for CRE and ESBL-E were nearly always higher for female than male children (Table 2), except in the youngest age group. During 2016–2020, average annual crude incidence rates for CRE cases were higher for male than for female children <1 year of age (3.38 vs. 2.35 cases/100,000 pediatric population).

Demographics and Clinical Characteristics

Of 159 CRE cases, 94 (59.1%) were in girls (Table 3), compared with 49.1% of the overall pediatric population. Median age was 5 (interquartile range [IQR] 1–10) years; 4 (2.5%) children were <1 month of age and 31 (19.5%) were 1–12 months of age (compared with 5.4% of the population <1 year of age). Most children with CRE were White (79 [50.0%]) and non-Hispanic (86 [54.1%]). Similarly, of 207 ESBL-E cases, most were in girls (165 [79.7%] compared with 49.1% of the overall pediatric population), White (92 [44.4%]), and non-Hispanic (96 [46.4%]). The median age was 6 (IQR 2–15) years; 3 (1.5%) children were <1 month of age, and 27 (13.0%) were 1–12 months of age (compared with 5.1% of the population <1 year of age).

Clinical characteristics were available for 158 CRE and 169 ESBL-E cases with completed case report forms (Table 3). Of those, a greater proportion of children with CRE than ESBL-E had a history of premature birth (20 [12.7%] vs. 11 [6.5%] among those with

Table 1. Incident CRE and ESBL-E cases in children, by organism, United States*

Organism	Incident CRE cases, 2016–2020						Incident ESBL-E cases, 2019–2020†		
	No. (%) cases	Isolates submitted for carbapenemase testing, no.	No. (%) CP isolates‡	No. (%) carbapenemase genes‡			No. (%) cases	No. isolates submitted for ESBL testing	No. (%) ESBL-producing organisms§
			<i>bla</i> _{KPC}	<i>bla</i> _{NDM}	<i>bla</i> _{OXA-48} -like				
<i>Escherichia coli</i>	50 (31.5)	26	5 (19.2)	0	2 (7.7)	3 (11.5)	182 (87.9)	16	15 (93.8)
<i>Enterobacter cloacae</i> complex	83 (52.2)	47	1 (2.1)	1 (2.1)	0	0	NA	NA	NA
<i>Klebsiella aerogenes</i>	5 (3.1)	4	1 (25.5)	0	1 (25.0)	0	NA	NA	NA
<i>K. oxytoca</i>	4 (2.5)	1	1 (100.0)	1 (100)	0	0	2 (1.0)	NA	NA
<i>K. pneumoniae</i>	17 (10.7)	8	1 (12.5)	0	1 (12.5)	0	23 (11.1)	3	3 (100.0)
Total	159	86	9 (10.5)	2 (2.3)	4 (4.7)	3 (3.5)	207	19	18 (94.7)

*All incident pediatric cases with available case report form data as of November 28, 2022, were included in this analysis. CP, carbapenemase-producing; CRE, carbapenem-resistant Enterobacterales; ESBL-E, extended-spectrum β-lactamase-producing Enterobacterales; NA, not applicable (no organisms tested); ESBL-E surveillance included *Escherichia coli*, *Klebsiella oxytoca*, *Klebsiella pneumoniae*.

†ESBL-E surveillance began in July 2019 at all participating sites. California, Connecticut, Minnesota, and Oregon do not participate in ESBL-E surveillance.

‡Percentages shown are of isolates submitted. Carbapenemases and colistin resistance genes not listed in the table were not detected for any isolates.

§Percentages shown are of isolates submitted. Phenotypic screening for ESBL-E production was performed by using ceftazidime and cefotaxime alone and in combination with clavulanate according to Clinical and Laboratory Standards Institute guidelines (24).

Table 2. Incident pediatric CRE and ESBL-E cases with annual crude incidence, by geographic regions and demographic characteristics, United States*

Category	Crude annual incidence rate						
	Incident CRE cases					Incident ESBL-E cases	
	2016	2017	2018	2019	2020	2019†	2020
EIP sites by geographic region†							
Northeast	0.63	0.00	1.24	0.91	1.38	26.15	24.39
Midwest	1.25	0.74	0.25	0.99	0.99	NA	NA
South	0.82	0.55	0.71	1.05	0.44	26.30	16.57
West	0.86	0.37	0.48	0.64	0.38	27.15	20.15
Total	0.87	0.47	0.68	0.87	0.63	26.54	19.63
Demographic characteristics							
Sex§							
M	0.44	0.32	0.75	0.67	0.52	13.04	6.28
F	1.26	0.62	0.61	1.07	0.74	39.89	33.43
Race							
White	0.68	0.31	0.55	0.78	0.50	23.85	13.96
Black	0.57	0.32	0.57	1.05	0.38	12.79	5.92
Other¶	0.41	0.26	0.35	0.24	0.56	20.26	12.29

*Crude annual incidence rate, cases/100,000 pediatric population. CRE, carbapenem-resistant Enterobacterales; EIP, Emerging Infections Program; ESBL-E, extended-spectrum β -lactamase-producing Enterobacterales; NA, not applicable.

†EIP sites grouped according to the 4 geographic regions defined by the US Census Bureau because of small case counts at some sites. California did not participate in CRE surveillance in 2016. Connecticut did not participate in CRE surveillance in 2016 or 2017. California, Connecticut, Minnesota, and Oregon do not participate in ESBL-E surveillance.

‡ESBL-E surveillance was completed for 6 months during 2019. Crude annual incidence rates estimated as the number of cases multiplied by 2 divided by population based on 2019 US Census for that year.

§One CRE and 1 ESBL-E case had sex reported as unknown.

¶Other represents all reported races not indicated as White or Black, including American Indian or Alaska Native, Asian, Native Hawaiian or other Pacific Islander, or some other race. This combination category was necessary because of small numbers of pediatric cases in persons of races other than White or Black. In addition, 37 CRE cases and 74 ESBL-E cases had race indicated as unknown (not included in this category).

term birth; $p = 0.06$) and any underlying condition (99 [62.7%] vs. 59 [34.9%] $p < 0.01$).

Culture Sources and Associated Infection Types

For most CRE and ESBL-E cases, including those in children <1 year of age, organisms were isolated from urine (131 [82.4%] from CRE cases and 196 [94.7%] from ESBL-E cases) (Table 4). Accordingly, the most common reported infection type was lower urinary tract infection (89 [56.3%] for CRE and 125 [74.0%] for ESBL-E). For CRE and ESBL-E cases, the greatest number of isolates were collected in an emergency department or outpatient setting (108 [68.4%] for CRE and 154 [91.1%] for ESBL-E), although CRE cases were more likely than ESBL-E cases to be hospital onset (40 [25.3%] vs. 13 [7.7%]; $p < 0.01$) (Table 3). Healthcare-associated community onset (68 [43.0%] CRE vs. 40 [23.7%] ESBL-E) and community-associated (43 [27.2%] CRE vs. 109 [64.5%] ESBL-E) represented most CRE and ESBL-E cases.

Healthcare Exposures and Outcomes

Among cases with available case report form data, a greater proportion of children with CRE than ESBL-E underwent acute care hospitalization (74 [46.8%] CRE vs 38 [22.5%] ESBL-E; $p < 0.01$) or surgery (61 [38.6%] CRE vs. 16 [9.5%] ESBL-E; $p < 0.01$) within 1 year before specimen collection (Table 4). In the 2 days before specimen collection, CRE cases were also more likely

than ESBL-E cases to have a central venous catheter (37 [23.4%] CRE vs. 12 [7.1%] ESBL-E; $p < 0.01$) or other indwelling device (excluding urinary catheter) (60 [38.0%] CRE vs. 19 [11.2%] ESBL-E; $p < 0.01$). ESBL-E cases were significantly more likely than CRE cases to have no reported healthcare exposures (110 [65.1%] vs. 43 [27.2%]; $p < 0.01$). Among ESBL-E cases, antimicrobial use was documented in the 30 days before date of incident specimen collection for 52 (31.8%).

Hospitalization at the time of or within 30 days of specimen collection was required for a greater proportion of community-onset CRE (40 [36.0%]) versus ESBL-E (13 [8.7%]) cases ($p < 0.01$). Median duration of admission among all hospitalized community-onset and hospital-onset cases was 18 days (IQR 3–103 days) for those with CRE versus 10 days (IQR 4–43 days) for those with ESBL-E ($p = 0.34$).

Isolate Testing

The antimicrobial resistance profiles of incident CRE and ESBL-E cases from local clinical laboratories are shown elsewhere (Appendix Table 1). Among the 86 CRE isolates submitted for carbapenemase testing, 9 (10.5%) isolates from 6 of the 10 EIP sites harbored a carbapenemase: 4 bla_{NDM} , 3 bla_{OXA-48} -like, and 2 bla_{KPC} (Table 1). Distribution of CP-CRE varied by organism. The 9 CP-CRE isolates were from 9 children, fewer than half of whom were 1–3 years of age (3 [15.8%]) and 4–9 years of age (3 [13.0%]); 6 (18.8%) were from

children with no reported underlying conditions (Appendix Table 2). The most common source was urine (8 CP-CRE isolates, 11.6% of submitted). Among the 11 CRE isolates from 2016–2018 that were sequenced, identified multilocus sequence types (STs) were diverse (Appendix Table 3); 2 of 11 isolates harbored CP genes. Separately, of the 7 ESBL-E organisms that underwent whole-genome sequencing at CDC, 6 were *E. coli*; ST131 and potential acquired ESBL gene *bla*_{CTX-M-15} were most common (Appendix Table 4).

Discussion

Over a 5-year surveillance period, 159 incident pediatric CRE cases were reported across 10 EIP sites (representing >4 million children), resulting in an overall crude incidence of 0.70 cases/100,000 pediatric population. The CRE case estimate is lower than the 207 incident pediatric ESBL-E cases identified over 1.5 years across 6 EIP sites (>600,000 children), which corresponds to an average crude incidence of 23.08 cases/100,000 pediatric population. The burden of infections was higher among girls than boys, more were detected in urine than in sterile site cultures, and incidence was disproportionately high among children <1 year of age. We found variation in rates of infections by year, geographic region, and species and in the percentages of CRE organisms that produced carbapenemases.

Similar to findings of other studies (17,30), in our study, *E. coli* accounted for many resistant isolates and represented most ESBL-E species identified. Of note, *E. cloacae* complex comprised most of the incident CRE cases. Our finding differs from that

of another large, nationally representative pediatric study conducted during 1999–2012 (30), in which the most common organisms identified were *E. coli* and *Proteus mirabilis*. It also differs from that of an earlier EIP study of adult CRE cases conducted during 2012–2013, in which *K. pneumoniae* accounted for most CRE cases identified (5). The previous EIP study used a different case definition that was more specific for carbapenemase-producing CRE (i.e., excluded er-tapenem and required carbapenem nonsusceptibility and third-generation cephalosporin resistance) and thus may have excluded certain species that are less likely to produce carbapenemases, probably affecting the comparison to the cases in our study (31). From our sample of 47 pediatric CR-*E. cloacae* complex isolates submitted for carbapenemase testing, only 1 was confirmed to be a carbapenemase producer (*bla*_{KPC}). Thus, the variability in species predominance and limited number of carbapenemase genes identified in our study may result from differences in case definition.

Most CRE and ESBL-E cases in our study were healthcare-associated infections with community onset or were community-associated infections. Strikingly, nearly 65% of the ESBL-E cases were reported to be community associated, and patients had no reported history of healthcare exposure. In addition, most cases of CRE and ESBL-E were detected from cultures collected outside an acute care hospital, a finding that differs from previous studies reporting pediatric CRE infections more commonly isolated from hospitalized patients (30). The most common source of CRE and ESBL-E in this study was urine

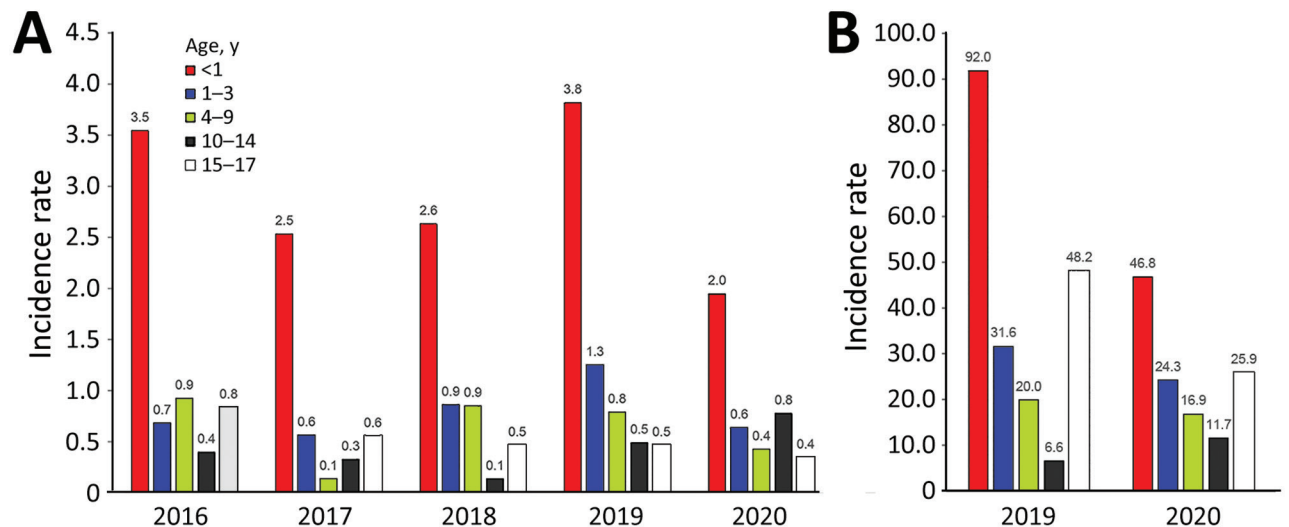


Figure. Annual crude incidence rates (cases/100,000 pediatric population) for incident pediatric carbapenem-resistant Enterobacterales (A) and extended-spectrum β -lactamase-producing Enterobacterales (B) cases, by age group, United States, 2016–2020. Incidence rate denominators are also stratified by age group.

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Table 3. Demographic and clinical characteristics of children with incident CRE and ESBL-E cases, United States*

Characteristic	CRE, 2016–2020 n = 159	ESBL-E, 2019–2020† n = 207	p value
Demographic			
Sex			<0.01
F	94 (59.1)	165 (79.7)	
M	64 (40.3)	41 (19.8)	
Unknown	1 (0.6)	1 (0.5)	
Age group, y			<0.01
Median, IQR	5 (1–10)	6 (2–15)	
<1	35 (22.0)	30 (14.5)	
1–3	30 (18.9)	40 (19.3)	
4–9	47 (29.6)	56 (27.1)	
10–14	28 (17.6)	27 (13.0)	
15–17	19 (12.0)	54 (26.1)	
Race			0.62
White	79 (50.0)	92 (44.4)	
Black	29 (18.2)	25 (12.1)	
Other	14 (8.8)	16 (7.7)	
Unknown	37 (23.3)	74 (35.8)	
Ethnicity			0.89
Hispanic	43 (27.0)	45 (21.7)	
Non-Hispanic	86 (54.1)	96 (46.4)	
Unknown	30 (18.9)	66 (31.9)	
Clinical‡			
Underlying conditions§	n = 158	n = 169	
Premature birth	20 (12.7)	11 (6.5)	0.06
Diabetes mellitus	0	2 (1.2)	0.27
Neurologic condition, any	34 (21.5)	20 (11.8)	0.02
Urinary tract problems/abnormalities	42 (26.6)	22 (13.0)	<0.01
Cardiovascular disease	13 (8.2)	3 (1.8)	<0.01
Chronic pulmonary disease	25 (15.8)	21 (12.4)	0.38
Chronic renal disease	24 (15.2)	3 (1.8)	<0.01
Gastrointestinal disease	3 (1.9)	1 (0.6)	0.23
Skin condition	12 (7.6)	8 (4.3)	0.28
Malignancy (hematologic or solid organ)	10 (6.3)	2 (1.2)	<0.01
Transplant (hematopoietic stem cell or solid organ)	15 (9.5)	1 (0.6)	<0.01
None	59 (37.3)	110 (65.1)	<0.01
Any condition	99 (62.7)	59 (34.9)	<0.01
Epidemiologic classification of incident cases			
Hospital onset	40 (25.3)	13 (7.7)	<0.01
Community-associated	43 (27.2)	109 (64.5)	<0.01
Healthcare-associated community onset	68 (43.0)	40 (23.7)	<0.01
Unknown	7 (4.4)	7 (4.1)	0.90

*Values are no. (%) except as indicated. Fisher exact test used for comparative statistics when >20% of expected cell counts were <5. Boldface indicates p<0.05. CRE, carbapenem-resistant Enterobacterales; ESBL-E, extended-spectrum β-lactamase-producing Enterobacterales.

†ESBL-E surveillance was completed for 6 months during 2019.

‡Clinical characteristics were available for cases with completed case report forms only.

§Cases could have >1 underlying condition associated with culture. Underlying conditions are further defined as follows: premature birth, birth before the week 37 of pregnancy, selected if medical record indicated premature birth and patient was <2 y of age; diabetes mellitus, includes both type I and type II; neurologic condition, any, includes cerebral palsy, chronic cognitive deficits, epilepsy/seizure/seizure disorders, multiple sclerosis, neuropathy, and others; urinary tract problems/abnormalities, a structural or functional urinary tract abnormality leading to obstruction or retention of urine as documented in the medical record; cardiovascular disease, includes congenital heart disease, congestive heart failure, prior cerebrovascular accident/stroke, peripheral vascular disease; chronic pulmonary disease, includes cystic fibrosis and any chronic respiratory condition resulting in chronic symptomatic dyspnea in medical record; chronic renal disease, includes chronic kidney disease (all stages), end-stage renal disease with or without dialysis; gastrointestinal disease, includes inflammatory bowel disease, liver disease, peptic ulcer disease, and short gut syndrome; skin condition, includes pressure ulcers, surgical wounds, other skin conditions such as eczema, psoriasis; malignancy, includes hematologic, metastatic and nonmetastatic solid organ; transplant, includes hematopoietic stem cell and solid organ.

(and lower urinary tract infections), which probably contributed to the high proportion of cases collected in outpatient settings. Increasing prevalence of community-associated ESBL-E urinary tract infections has been noted across patients of all ages (32,33). Our findings highlight a similar shift in the clinical epidemiology of multidrug-resistant infections in children, supported by a rising trend in community-acquired ESBL-E causing urinary tract

infections in children (34). Continued implementation of national programs to detect, prevent, and treat multidrug-resistant infections must increasingly include pediatric populations and outpatient settings.

Although incidence of CRE was lower than that of ESBL-E, children with CRE infection were generally hospitalized for longer durations, and rates of intensive care unit admission were higher. Children with CRE infection were also more likely to have ≥1 underlying

condition and prior healthcare exposure. Meropol et al. (19) and Logan et al. (30) also reported higher proportions of underlying conditions in children with multidrug-resistant gram-negative infections. However, our direct comparison of CRE and ESBL-E epidemiology revealed statistically significant clinical differences even among children with multidrug-resistant gram-negative infections. We also observed the proportion of CRE and ESBL-E in patients with no underlying conditions to be markedly higher than that found in

previous studies focused on adult populations with those infections (5,9).

CRE incidence rates fluctuated by region throughout the study period, partly reflecting variation in the small number of cases occurring on a yearly basis. Separately, the overall incidence of ESBL-E clearly decreased during 2020. That finding contrasts with reports of increased rates of ESBL-E infections among hospitalized patients, primarily adults, in 2020 (2). Our data were unadjusted and included nonhospitalized

Table 4. Isolate culture source, collection location, and infection types among incident pediatric CRE and ESBL-E cases, United States*

Category	CRE, 2016–2020	ESBL-E, 2019–2020†	p value
Culture source	n = 159	n = 207	
Urine‡	131 (82.4)	196 (94.7)	<0.01
Blood	20 (12.6)	8 (3.9)	<0.01
Other normally sterile site	8 (5.0)	3 (1.5)	0.05
Isolate collection location§	n = 158	n = 169	
Acute care hospital	49 (31.0)	15 (8.9)	<0.01
Outside acute care hospital	108 (68.4)	154 (91.1)	<0.01
Emergency department	16/108 (14.8)	0	<0.01
Outpatient setting	92/108 (85.2)	154/154 (100.0)	<0.01
Long-term care facility	0	0	NA
Long-term acute care facility	0	0	NA
Unknown	1 (0.6)	0	0.48
Infection types¶			
Lower urinary tract infection#	89 (56.3)	125 (74.0)	<0.01
Pyelonephritis	9 (5.7)	9 (5.3)	0.88
Bacteremia**	20 (12.7)	9 (5.3)	0.02
Other infection types	27 (17.1)	14 (8.3)	0.02
None	21 (13.3)	19 (11.2)	0.57
Unknown	9 (5.7)	8 (4.7)	0.70
Healthcare exposures in prior year			
Acute care hospitalization	74 (46.8)	38 (22.5)	<0.01
Resident of long-term care facility	0	2 (1.2)	0.27
Admission to long-term acute care hospital	0	0	NA
Inpatient or outpatient surgery††	61 (38.6)	16 (9.5)	<0.01
Chronic dialysis	5 (3.2)	0	0.03
Indwelling device in the 2 d before DISC			
Urinary catheter	29 (18.4)	20 (11.8)	0.10
Central venous catheter	37 (23.4)	12 (7.1)	<0.01
Any other device	60 (38.0)	19 (11.2)	<0.01
IV or oral antimicrobial use 30 d before DISC‡‡	N/A	52 (30.8)	N/A
None of the above healthcare exposures	43 (27.2)	110 (65.1)	<0.01
Outcomes, no. patients			
Hospitalization among community-onset cases§§	40/111 (36.0)	13/149 (8.7)	<0.01
ICU admission ≤6 d after DISC	10 (6.3)	5 (3.0)	0.15
30-d mortality			
Cases with an incident blood or sterile site specimen	4/27 (14.8)	2/9 (22.2)	0.29
Cases with an incident urine specimen	2/131 (1.5)	0	0.23

*Values are no. (%) except as indicated. Denominators are indicated when different from total number. Fisher exact test used for comparative statistics when >20% of expected cell counts were <5. Boldface indicates p<0.05. CRE, carbapenem-resistant Enterobacterales; DISC, date of incident specimen collection; ESBL-E, extended-spectrum β-lactamase-producing Enterobacterales; IV, intravenous.

†ESBL-E surveillance was completed for 6 mo during 2019.

‡Two incident urine CRE cases with subsequent nonincident blood cultures were identified. One incident urine ESBL-E case with subsequent nonincident blood culture was identified. Nonincident blood cultures are not counted here.

§Isolate collection location, epidemiologic class, infection types available for cases with completed case report forms only.

¶Cases could have >1 type of associated infection reported.

#Lower urinary tract infection includes cases involving infection of the bladder or urethra. Pyelonephritis, or infections involving the kidney(s), were counted separately.

**Bacteremia includes cases with a positive blood specimen or a documented diagnosis of sepsis, septicemia, bacteremia, or blood stream infection. 3 CRE cases of sepsis and 2 ESBL-E cases of sepsis (with or without blood cultures) are included in the bacteremia classification above.

††Surgery defined as procedures occurring in an operating room where a surgeon makes ≥1 incision through skin or mucous membrane, including laparoscopic approach. Ambulatory surgery centers may be included.

‡‡Antimicrobial use (intravenous or oral) in 30 d before DISC collected for ESBL-E cases only.

§§Hospitalization at time of or within 30 d after DISC.

patients, and it is possible that declines in outpatient healthcare use during the pandemic may have affected rates of ESBL-E among children. In addition, our data represent only mid-2019 through 2020, making the decline more difficult to interpret.

We also found annual CRE and ESBL-E incidence rates to be higher for female children and infants (most >1 month of age) compared with other age groups. We suspect that rates of antimicrobial-resistant infections were higher among girls in part because of increased testing in those populations resulting from a higher number of baseline urinary tract infections. The epidemiology among infants may differ from that among the overall pediatric population because of risk factors associated with infection acquired in neonatal intensive care units (35–37), vertical transmission (38,39), and higher rates of fecal colonization with antimicrobial-resistant *Enterobacteriaceae* (40). Recent evidence highlights how the human microbiome undergoes marked developmental progression over the first 2 years of life (41). Consistent with such a maturation process, Darda et al. observed spontaneous decolonization within 12 months among all neonates colonized with carbapenem-resistant gram-negative bacteria (42). Nonetheless, even when limited to prenatal and intrapartum exposures, antimicrobial drugs can profoundly affect the infant microbiome, involving expansion of gram-negative populations (proteobacteria) and supporting our observation of differences in the epidemiology among infants (43). A more focused look at that age group may be noteworthy for future studies.

Among the several limitations inherent in the use of surveillance systems, the case definitions for CRE and ESBL-E relied on susceptibility testing performed locally, and methods varied across laboratories. In addition, automated testing instruments at clinical laboratories may be more likely than other test methods to overdiagnose CRE. Second, data were retrospectively abstracted from medical records, and the quality of medical record documentation can vary between healthcare system and facility types, resulting in differences in reporting for some data elements. In addition, medical records were reviewed for the child only, and no maternal information was included in chart review, which may have limited identification of household risk factors (e.g., international travel by family, previously established as a risk factor for CRE and ESBL-E). Separately, because ≈97% of the incident ESBL-E cases from 1 site in 2020 did not have a case report form completed, no data beyond the clinical laboratory report were available. Third, isolate collection represents a convenience sample and may not be representative of all cases. In addition, we have

limited data on STs because not all isolates submitted to CDC were sequenced. Fourth, we acknowledge that data for ESBL-E in our study are limited to 1.5 years and collected from fewer sites than surveillance data for CRE. Last, although the surveillance system includes geographically diverse catchment areas, it is not designed to be representative of the entire US pediatric population.

In summary, we found that CRE infections occurred less frequently than ESBL-E infections among US children but were more often associated with healthcare risk factors and hospitalization. Despite annual and geographic variation in the incidence of CRE and ESBL-E, the rate of infection for both pathogens was consistently highest among infants. Our descriptive data about major antimicrobial-resistant pathogens among children support continued infection prevention and control practices and antimicrobial stewardship in pediatric healthcare settings, particularly for patients in the youngest age group.

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References

- Centers for Disease Control and Prevention. Antibiotic resistance threats in the United States, 2019 [cited 2024 Apr 22]. <https://www.cdc.gov/drugresistance/pdf/threats-report/2019-ar-threats-report-508.pdf>
- Centers for Disease Control and Prevention. COVID-19: US impact on antimicrobial resistance [cited 2024 Apr 22]. <https://www.cdc.gov/drugresistance/pdf/covid19-impact-report-508.pdf>
- Logan LK, Weinstein RA. The epidemiology of carbapenem-resistant *Enterobacteriaceae*: the impact and evolution of

- a global menace. *J Infect Dis*. 2017;215(suppl_1):S28–36. <https://doi.org/10.1093/infdis/jiw282>
4. Braykov NP, Eber MR, Klein EY, Morgan DJ, Laxminarayan R. Trends in resistance to carbapenems and third-generation cephalosporins among clinical isolates of *Klebsiella pneumoniae* in the United States, 1999–2010. *Infect Control Hosp Epidemiol*. 2013;34:259–68. <https://doi.org/10.1086/669523>
 5. Guh AY, Bulens SN, Mu Y, Jacob JT, Reno J, Scott J, et al. Epidemiology of carbapenem-resistant Enterobacteriaceae in 7 US communities, 2012–2013. *JAMA*. 2015;314:1479–87. <https://doi.org/10.1001/jama.2015.12480>
 6. Kritsotakis EL, Tsioutis C, Roubelaki M, Christidou A, Gikas A. Antibiotic use and the risk of carbapenem-resistant extended-spectrum- β -lactamase-producing *Klebsiella pneumoniae* infection in hospitalized patients: results of a double case-control study. *J Antimicrob Chemother*. 2011;66:1383–91. <https://doi.org/10.1093/jac/dkr116>
 7. Swaminathan M, Sharma S, Poliansky Blash S, Patel G, Banach DB, Phillips M, et al. Prevalence and risk factors for acquisition of carbapenem-resistant Enterobacteriaceae in the setting of endemicity. *Infect Control Hosp Epidemiol*. 2013;34:809–17. <https://doi.org/10.1086/671270>
 8. Marchaim D, Chopra T, Bhargava A, Bogan C, Dhar S, Hayakawa K, et al. Recent exposure to antimicrobials and carbapenem-resistant Enterobacteriaceae: the role of antimicrobial stewardship. *Infect Control Hosp Epidemiol*. 2012;33:817–30. <https://doi.org/10.1086/666642>
 9. Duffy N, Karlsson M, Reses HE, Campbell D, Daniels J, Stanton RA, et al. Epidemiology of extended-spectrum β -lactamase-producing Enterobacteriales in five US sites participating in the Emerging Infections Program, 2017. *Infect Control Hosp Epidemiol*. 2022;43:1586–94.
 10. Tamma PD, Aitken SL, Bonomo RA, Mathers AJ, van Duin D, Clancy CJ. Infectious Diseases Society of America 2023 guidance on the treatment of antimicrobial resistant gram-negative infections. *Clin Infect Dis*. 2023; ciad428
 11. Romandini A, Pani A, Schenardi PA, Pattarino GAC, De Giacomo C, Scaglione F. Antibiotic resistance in pediatric infections: global emerging threats, predicting the near future. *Antibiotics (Basel)*. 2021;10:393. <https://doi.org/10.3390/antibiotics10040393>
 12. Logan LK, Bonomo RA. Metallo- β -lactamase (MBL)-producing Enterobacteriaceae in United States children. *Open Forum Infect Dis*. 2016;3:ofw090. <https://doi.org/10.1093/ofid/ofw090>
 13. Logan LK, Medernach RL, Domitrovic TN, Rispens JR, Hujer AM, Qureshi NK, et al. The clinical and molecular epidemiology of CTX-M-9 group producing Enterobacteriaceae infections in children. *Infect Dis Ther*. 2019;8:243–54. <https://doi.org/10.1007/s40121-019-0237-2>
 14. Logan LK, Hujer AM, Marshall SH, Domitrovic TN, Rudin SD, Zheng X, et al. Analysis of beta-lactamase resistance determinants in Enterobacteriaceae from Chicago children: a multicenter survey. *Antimicrob Agents Chemother*. 2016;60:3462–9. <https://doi.org/10.1128/AAC.00098-16>
 15. Logan LK, Rispens JR, Medernach RL, Domitrovic TN, Hujer AM, Marshall SH, et al. A multicentered study of the clinical and molecular epidemiology of TEM- and SHV-type extended-spectrum beta-lactamase producing Enterobacteriales infections in children. *Pediatr Infect Dis J*. 2021;40:39–43. <https://doi.org/10.1097/INF.0000000000002916>
 16. Logan LK, Medernach RL, Rispens JR, Marshall SH, Hujer AM, Domitrovic TN, et al. Community origins and regional differences highlight risk of plasmid-mediated fluoroquinolone resistant Enterobacteriaceae infections in children. *Pediatr Infect Dis J*. 2019;38:595–9. <https://doi.org/10.1097/INF.0000000000002205>
 17. Zerr DM, Weissman SJ, Zhou C, Kronman MP, Adler AL, Berry JE, et al. The molecular and clinical epidemiology of extended-spectrum cephalosporin- and carbapenem-resistant Enterobacteriaceae at 4 US pediatric hospitals. *J Pediatric Infect Dis Soc*. 2017;6:366–75. <https://doi.org/10.1093/jpids/piw076>
 18. Chiotos K, Tamma PD, Flett KB, Naumann M, Karandikar MV, Bilker WB, et al. Multicenter study of the risk factors for colonization or infection with carbapenem-resistant Enterobacteriaceae in children. *Antimicrob Agents Chemother*. 2017;61:e01440-17. <https://doi.org/10.1128/AAC.01440-17>
 19. Meropol SB, Haupt AA, Debanne SM. Incidence and outcomes of infections caused by multidrug-resistant Enterobacteriaceae in children, 2007–2015. *J Pediatric Infect Dis Soc*. 2018;7:36–45. <https://doi.org/10.1093/jpids/piw093>
 20. Centers for Disease Control and Prevention. Multi-site Gram-negative Surveillance Initiative [cited 2023 Apr 24]. https://www.cdc.gov/hai/eip/mugs.html#anchor_46658
 21. US Census Bureau. 2020 Population estimates 2020 [cited 2023 Jan 19]. <https://www.census.gov/quickfacts/fact/faq/US/PST045221>
 22. Migliavacca R, Docquier JD, Mugnaioli C, Amicosante G, Daturi R, Lee K, et al. Simple microdilution test for detection of metallo- β -lactamase production in *Pseudomonas aeruginosa*. *J Clin Microbiol*. 2002;40:4388–90. <https://doi.org/10.1128/JCM.40.11.4388-4390.2002>
 23. Karlsson M, Lutgring JD, Ansari U, Lawsin A, Albrecht V, McAllister G, et al. Molecular characterization of carbapenem-resistant Enterobacteriales collected in the United States. *Microb Drug Resist*. 2022;28:389–97. <https://doi.org/10.1089/mdr.2021.0106>
 24. Clinical and Laboratory Standards Institute. Performance standards for antimicrobial susceptibility testing; 33rd edition. Supplement M100. Wayne (PA): The Institute; 2023.
 25. Rasheed JK, Kitchel B, Zhu W, Anderson KF, Clark NC, Ferraro MJ, et al. New Delhi metallo- β -lactamase-producing Enterobacteriaceae, United States. *Emerg Infect Dis*. 2013;19:870–8. <https://doi.org/10.3201/eid1906.121515>
 26. Lutgring JD, Zhu W, de Man TJB, Avillan JJ, Anderson KF, Lonsway DR, et al. Phenotypic and genotypic characterization of Enterobacteriaceae producing oxacillinase-48-like carbapenemases, United States. *Emerg Infect Dis*. 2018;24:700–9. <https://doi.org/10.3201/eid2404.171377>
 27. Campbell D, Daniels J, Rasheed JK, Karlsson M. Development of a multiplex TaqMan probe-based real-time PCR assay for detection of bla_{MIP} variants. Poster presented at: ASM Microbe 2017; New Orleans, LA, USA; 2017 June 1–5.
 28. Prussing C, Canulla T, Singh N, McAuley P, Gosciminski M, King E, et al. Characterization of the first carbapenem-resistant *Pseudomonas aeruginosa* clinical isolate harboring bla_{SIM-1} from the United States. *Antimicrob Agents Chemother*. 2021;65:e0106621. <https://doi.org/10.1128/AAC.01066-21>
 29. Daniels JB, Campbell D, Boyd S, Ansari U, Lutgring J, Rasheed JK, et al. Development and validation of a Clinical Laboratory Improvement Amendments-compliant multiplex real-time PCR assay for detection of mcr genes. *Microb Drug Resist*. 2019;25:991–6. <https://doi.org/10.1089/mdr.2018.0417>
 30. Logan LK, Renschler JP, Gandra S, Weinstein RA, Laxminarayan R; Centers for Disease Control; Prevention

- Epicenters Program. Carbapenem-resistant *Enterobacteriaceae* in children, United States, 1999–2012. *Emerg Infect Dis*. 2015;21:2014–21. <https://doi.org/10.3201/eid2111.150548>
31. Chea N, Bulens SN, Kongphet-Tran T, Lynfield R, Shaw KM, Vagnone PS, et al. Improved phenotype-based definition for identifying carbapenemase producers among carbapenem-resistant *Enterobacteriaceae*. *Emerg Infect Dis*. 2015;21:1611–6. <https://doi.org/10.3201/eid2109.150198>
 32. Thaden JT, Fowler VG Jr, Sexton DJ, Anderson DJ. Increasing incidence of extended-spectrum beta-lactamase-producing *Escherichia coli* in community hospitals throughout the southeastern United States. *Infect Control Hosp Epidemiol*. 2016;37:49–54. <https://doi.org/10.1017/ice.2015.239>
 33. Horie A, Nariai A, Katou F, Abe Y, Saito Y, Koike D, et al. Increased community-acquired upper urinary tract infections caused by extended-spectrum beta-lactamase-producing *Escherichia coli* in children and the efficacy of flomoxef and cefmetazole. *Clin Exp Nephrol*. 2019;23:1306–14. <https://doi.org/10.1007/s10157-019-01775-w>
 34. Collingwood JD, Yarbrough AH, Boppana SB, Dangle PP. Increasing prevalence of pediatric community-acquired UTI by extended spectrum β -lactamase-producing *E. coli*: cause for concern. *Pediatr Infect Dis J*. 2023;42:106–9. <https://doi.org/10.1097/INF.0000000000003777>
 35. Yin L, He L, Miao J, Yang W, Wang X, Ma J, et al. Carbapenem-resistant Enterobacterales colonization and subsequent infection in a neonatal intensive care unit in Shanghai, China. *Infect Prev Pract*. 2021;3:100147. <https://doi.org/10.1016/j.infpip.2021.100147>
 36. Flannery DD, Chiotos K, Gerber JS, Puopolo KM. Neonatal multidrug-resistant gram-negative infection: epidemiology, mechanisms of resistance, and management. *Pediatr Res*. 2022;91:380–91. <https://doi.org/10.1038/s41390-021-01745-7>
 37. Yaffee AQ, Roser L, Daniels K, Humbaugh K, Brawley R, Thoroughman D, et al. Notes from the field: Verona integron-encoded metallo-beta-lactamase-producing carbapenem-resistant Enterobacteriaceae in a neonatal and adult intensive care unit – Kentucky, 2015. *MMWR Morb Mortal Wkly Rep*. 2016;65:190. <https://doi.org/10.15585/mmwr.mm6507a5>
 38. Principe L, Meroni E, Conte V, Mauri C, Di Pilato V, Giani T, et al. Mother-to-child transmission of KPC-producing *Klebsiella pneumoniae*: potential relevance of a low microbial urinary load for screening purposes. *J Hosp Infect*. 2018;98:314–6. <https://doi.org/10.1016/j.jhin.2017.10.008>
 39. Sotgiu G, Are BM, Pesapane L, Palmieri A, Muresu N, Cossu A, et al. Nosocomial transmission of carbapenem-resistant *Klebsiella pneumoniae* in an Italian university hospital: a molecular epidemiological study. *J Hosp Infect*. 2018;99:413–8. <https://doi.org/10.1016/j.jhin.2018.03.033>
 40. Islam S, Selvarangan R, Kanwar N, McHenry R, Chappell JD, Halasa N, et al. Intestinal carriage of third-generation cephalosporin-resistant and extended-spectrum β -lactamase-producing *Enterobacteriaceae* in healthy US children. *J Pediatric Infect Dis Soc*. 2018;7:234–40. <https://doi.org/10.1093/jpids/pix045>
 41. Wernroth ML, Peura S, Hedman AM, Hetty S, Vicenzi S, Kennedy B, et al. Development of gut microbiota during the first 2 years of life. *Sci Rep*. 2022;12:9080. <https://doi.org/10.1038/s41598-022-13009-3>
 42. Darda VM, Iosifidis E, Antachopoulos C, Kirvasilis F, Zarras C, Simitsopoulou M, et al. A longitudinal study of spontaneous gut decolonization of carbapenem-resistant gram-negative bacteria in neonatal and pediatric patients. *Pediatr Infect Dis J*. 2022;41:648–53. <https://doi.org/10.1097/INF.0000000000003562>
 43. Dierikx TH, Visser DH, Benninga MA, van Kaam AHLC, de Boer NKH, de Vries R, et al. The influence of prenatal and intrapartum antibiotics on intestinal microbiota colonisation in infants: a systematic review. *J Infect*. 2020;81:190–204. <https://doi.org/10.1016/j.jinf.2020.05.002>

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Carbapenem-Resistant and Extended-Spectrum β -Lactamase–Producing Enterobacterales in Children, United States, 2016–2020

Appendix

Supplemental Methods

Total Surveillance Population

The total population of the 10 participating areas for CRE surveillance in 2020 was an estimated 23.2 million; this included Atlanta, Georgia area (8 counties, estimated population 4,202,188), Minneapolis/St. Paul, Minnesota (2 counties, estimated population 1,833,917), Portland, Oregon (3 counties, estimated population 1,837,201), Denver, Colorado (5 counties, estimated population 2,831,052), Baltimore, Maryland (4 counties, estimated population 1,945,451), Albuquerque, New Mexico (1 county, estimated population 676,444), Rochester, New York (1 county, estimated population 759,443), Nashville, Tennessee (8 counties, estimated population 1,817,304), San Francisco, California (3 counties, estimated population 3,722,245), and all of Connecticut (8 counties, estimated population 3,605,944).

The total population of the 6 participating areas for ESBL-producing Enterobacterales surveillance in 2020 was an estimated 3.0 million; this included counties of Atlanta, Georgia (2 counties, estimated population 441,832), Boulder, Colorado (1 county, estimated population 330,758), Baltimore Maryland (1 county, estimated population 585,708), Albuquerque, New Mexico (1 county, estimated population 676,444), Rochester, New York (1 county, estimated population 759,443), and middle Tennessee (4 counties, estimated population 164,106).

Normally Sterile Body Sites

For this surveillance, normally sterile sites included blood, cerebrospinal fluid, pleural fluid, pericardial fluid, peritoneal fluid, synovial fluid, bone, internal body tissue (lymph node, brain, heart, liver spleen, vitreous fluid, kidney, pancreas, or ovary), muscle, deep tissue, or samples from other normally sterile sites.

Data Collection

Both CRE and ESBL-E cases were identified through a query of automated testing instruments based on the protocols of the laboratories. Antimicrobial susceptibility test methods varied among the clinical laboratories although the majority reported the use of an automated test system (MicroScan, Beckman Coulter Diagnostics, Brea, CA; Vitek, bioMérieux, Marcy-l'Étoile, FR, or BD Phoenix, Becton Dickinson, Franklin Lakes, NJ). Kirby Bauer and E-tests were often used for confirmatory testing.

For cases that underwent a medical record review, one of the data elements collected included hospitalization in the year preceding the date of incident specimen collection. For cases ≤ 12 months of age, if the birth hospitalization included a neonatal intensive care unit (NICU) stay with discharge, or there were any additional hospitalizations after the birth hospitalization with discharge, then the infant would be considered to have had a prior hospitalization. If the birth hospitalization was otherwise in a well-newborn nursery or that was the only hospitalization prior to the initial invasive CRE or ESBL-E culture, then that would not count as a prior hospitalization. If the infant was in the NICU and had never been discharged home, this would also not count as a prior hospitalization.

Isolate Collection

A convenience sample of isolates from all EIP sites was submitted to CDC for confirmatory and molecular characterization. Sampling approach was determined separately by each of the participating clinical laboratories based on staff time and resources available for isolate selection and storage. Laboratories typically chose isolates meeting the case definition which were readily available and accessible for shipment to CDC.

Whole Genome Sequencing

Whole genome sequencing was conducted on a subset of CRE isolates that were confirmed carbapenem resistant using reference broth microdilution at CDC. CRE sequencing

criteria changed slightly each year to reduce the overall volume and burden on sequencing labs. For 2016 isolates, CDC sequenced all isolates that confirmed as CRE after antimicrobial susceptibility testing. For 2017 isolates, CDC sequenced isolates that confirmed as CRE after antimicrobial susceptibility testing, but excluded Ertapenem-mono-resistant *E. cloacae* complex and *K. aerogenes*. For 2018 isolates, CDC sequenced isolates that confirmed as CRE after antimicrobial susceptibility testing, but excluded Ertapenem-mono-resistant *E. cloacae* complex.

Whole genome sequencing was performed on all ESBL-E isolates received at CDC. Sequencing was performed using an Illumina MiSeq or NovSeq system (San Diego, CA). For MiSeq, genomic DNA was extracted using the Promega Maxwell 16 Low Elution Volume DNA Purification Kit and the Maxwell 16 MDx Instrument (Madison, WI, United States). For NovSeq, genomic DNA was sheared to a mean size of 600 bp using a Covaris LE220 focused ultrasonicator (Covaris Inc., Woburn, MA). DNA fragments were Ampure (Beckman Coulter Inc., Indianapolis, IN) cleaned and used to prepare dual-indexed sequencing libraries using NEBNext Ultra library prep reagents (New England Biolabs Inc., Ipswich, MA) and barcoding indices synthesized in the CDC Biotechnology Core Facility. Libraries were analyzed for size and concentration, normalized, pooled and denatured for loading onto flowcells for cluster generation. Sequencing was performed on a Novaseq using Novaseq 2x250bp paired-end sequencing kits. On completion, sequence reads were filtered for read quality, basecalled and demultiplexed using bcl2fastq (v2.19). All sequences were analyzed using the CDC laboratory's in-house QuAISAR-H pipeline (Quality, Assembly, species Identification, Sequence typing, Annotation, Resistance mechanisms for Hospital acquired infections; https://github.com/DHQP/QuAISAR_singularity). QuAISAR-H identified antibiotic resistant (AR) genes, including carbapenemase genes, beta-lactamase genes, and plasmid-mediated colistin resistance (*mcr*) genes, using a non-redundant combined database of acquired AR genes from ResFinder; ARG-ANNOT, and AMRFinderPlus; genes with a minimum of 98% identity and 90% coverage threshold were reported. Multilocus sequence type (MLST) was determined using the publicly available schemes curated by pubMLST (last accessed 07/15/2022).

Appendix Table 1. Antimicrobial resistance of incident carbapenem-resistant Enterobacteriaceae (CRE) and extended-spectrum beta-lactamase-producing Enterobacterales (ESBL-E) cases by organism based on testing by local clinical laboratories

Antimicrobial Agent	CRE (2016-2020)	ESBL (2019-2020)
	No. resistant / no. tested (%)	No. resistant / no. tested (%)
Aminoglycosides	29/151 (19.2%)	44/157 (28.0%)
Amikacin	6/115 (5.2%)	3/99 (3.0%)
Gentamicin	23/150 (15.3%)	38/156 (24.4%)
Tobramycin	25/142 (17.6%)	39/125 (31.2%)
Carbapenems	153/154 (99.4%) *	0/161 (0%)
Doripenem	0/21 (0%)	0/8 (0%)
Ertapenem	141/149 (94.6%) *	0/169 (0%)
Imipenem	28/95 (29.5%)	0/95 (0%)
Meropenem	22/114 (19.3%)	0/107 (0%)
Extended-spectrum cephalosporins	128/148 (86.5%)	164/164 (100%)
Cefotaxime	30/46 (65.2%)	26/26 (100%)
Ceftazidime	79/103 (76.7%)	82/111 (73.9%)
Ceftriaxone	122/142 (85.9%)	162/163 (99.4%)
Cefepime	53/140 (37.9%)	80/150 (53.3%)
Cefoxitin ^b	---	---
Fluoroquinolones	45/150 (30.0%)	79/147 (53.7%)
Ciprofloxacin	43/142 (30.3%)	73/139 (52.5%)
Levofloxacin	24/109 (22.0%)	49/107 (45.8%)
Beta-lactam and non-beta-lactam combination agents		
Amoxicillin-clavulanate	29/79 (36.7%)	28/76 (36.8%)
Piperacillin-tazobactam	96/138 (69.6%)	14/151 (9.3%)
Ceftazidime-avibactam	0/12 (0%)	0/2 (0%)
Meropenem-vaborbactam	0/3 (0%)	0/2 (0%)
Imipenem-relebactam *	---	---
Folate pathway antagonists		
Trimethoprim-sulfamethoxazole	29/148 (19.6%)	64/160 (40.0%)
Other antimicrobials		
Aztreonam	64/95 (67.4%)	69/90 (76.7%)
Colistin [†]	1/3 (33.3%)	---
Fosfomycin	0/2 (0%)	0/17 (0%)
Nitrofurantoin	14/134 (10.5%)	5/152 (3.3%)
Tigecycline	0/42 (0%)	0/15 (0%)

CRE: Carbapenem-Resistant Enterobacterales; ESBL-E: Extended-Spectrum β -Lactamase-Producing Enterobacterales.

*A minimum inhibitory concentration (MIC) >1 for ertapenem was considered sufficient for meeting the phenotypic case definition for MuGSI CRE surveillance. This table reports cases based on MIC at the local clinical laboratories, not the reported interpretation. One clinical laboratory in this study uses an automated testing instrument (ATI) card with the highest MIC for ertapenem being 1. This result was reported as R to clinicians while awaiting confirmatory testing, but does not meet the MIC > 1 breakpoint used in this table.

[†]No CRE or ESBL-E isolates were tested for cefoxitin or imipenem-relebactam susceptibility and no ESBL-E isolates were tested for colistin susceptibility.

Appendix Table 2. Characteristics of pediatric carbapenemase-producing Enterobacterales (CP-CRE) Cases, 2016–2020

Characteristic	Total Cases	Isolates Submitted for Carbapenemase Testing	No. of Carbapenemase-Producing Isolates (%) *
Year of Specimen Collection			
2016	31	8	0
2017	20	10	0
2018	34	22	2 (9.1)
2019	43	25	6 (24.0)
2020	31	21	1 (4.8)
Age Category			
< 1 y	35	22	1 (4.6)
1–3 y	30	19	3 (15.8)
4–9 y	47	23	3 (13.0)
10–14 y	28	11	2 (18.2)
15–17 y	19	11	0
Gender			
Female	94	46	5 (10.9)
Male	64	40	4 (10.0)
Unknown	1	0	0
Race			
White	79	44	4 (9.1)
Black	29	20	1 (5.0)
Other/Unknown	51	22	4 (18.2)
Ethnicity			
Hispanic	43	16	2 (12.5)

Characteristic	Total Cases	Isolates Submitted for Carbapenemase Testing	No. of Carbapenemase-Producing Isolates (%) *
Non-Hispanic	86	54	7 (13.0)
Unknown	30	16	0
Source			
Blood	20	13	1 (7.7)
Other Sterile	8	4	0
Urine	131	69	8 (11.6)
Collection Location			
Acute Care Hospital	50	32	2 (6.25)
Outpatient setting or ER	108	54	7 (13.0)
Unknown	1	0	0
Any Underlying Conditions			
0	59	32	6 (18.8)
≥1	99	54	3 (5.6)
Unknown	1	0	0
Epidemiologic Classification			
Hospital Onset	41	28	2 (7.1)
Community Associated	43	24	4 (16.7)
Healthcare Associated	68	34	3 (8.8)
Community Onset			
Unknown	7	0	0
International Travel			
Yes	6	4	2 (50.0)
No / Unknown	153	82	7 (8.5)

CRE: Carbapenem-Resistant Enterobacterales; ESBL-E: Extended-Spectrum β-Lactamase-Producing Enterobacterales; ER: Emergency Room.

* Percent shown is of percent total isolates submitted to CDC for testing.

Appendix Table 3. Multilocus sequence types, carbapenemase, and beta-lactamase genes from pediatric carbapenem-resistant Enterobacteriaceae (CRE) isolates that underwent whole genome sequencing (n=11)

Organism	Multilocus sequence types	Carbapenemase genes	Non-carbapenemase beta-lactamase genes
<i>Enterobacter cloacae</i> complex (n=5)	ST133(Pasteur)	NA	<i>bla</i> _{ACT-86} *
	ST252(Pasteur)	NA	<i>bla</i> _{ACT-3}
	ST467(Pasteur)	NA	<i>bla</i> _{ACT-17}
	ST50(Pasteur)	NA	<i>bla</i> _{ACT-15}
	ST526(Pasteur)	NA	<i>bla</i> _{MIR-16}
<i>Escherichia coli</i> (n=3)	ST1123(Pasteur),	NA	<i>bla</i> _{AmpC1} , <i>bla</i> _{AmpH} , <i>bla</i> _{CMY-2} , <i>bla</i> _{EC-8} †
	ST11538(Achtman)		
	ST963(Pasteur),	NA	<i>bla</i> _{AmpC1} , <i>bla</i> _{AmpH} , <i>bla</i> _{CMY-2} , <i>bla</i> _{EC-8} †
	ST963(Achtman)		
	ND (Pasteur),	<i>bla</i> _{OXA-48-like}	<i>bla</i> _{AmpH} , <i>bla</i> _{CTX-M-15} , <i>bla</i> _{EC-19} **, <i>bla</i> _{SHV-12}
	ST13455(Achtman),		<i>bla</i> _{CMY2-MIR-ACT-EC} ‡
<i>Klebsiella aerogenes</i> (n=2)	ST205(Pasteur)	NA	<i>bla</i> _{CTX-M-15} , <i>bla</i> _{TEM-1a} , <i>bla</i> _{CMY2-MIR-ACT-EC} §
<i>Klebsiella pneumoniae</i> (n=1)	ST373(Pasteur)	<i>bla</i> _{NDM-1}	
	ST1426(Pasteur)	NA	<i>bla</i> _{AmpH} , <i>bla</i> _{CTX-M-15} , <i>bla</i> _{OXA-1} , <i>bla</i> _{SHV-11} , <i>bla</i> _{TEM1}

ESBL-E: Extended-spectrum beta-lactamase-producing Enterobacterales; ST: Sequence Type; ND: Not Determined.

* This call is a mutant/novel allele that has 5 amino acid changes from the listed gene.

† This call is a mutant/novel allele that has 7 amino acids different from the listed gene.

‡ This call is a mutant/novel allele that is 1 amino acid different from the listed gene.

§ This call is a mutant/novel allele that has 3 amino acids different from the listed gene.

Appendix Table 4. Multilocus sequence types, acquired extended-spectrum beta-lactamases (ESBL), and beta-lactamase genes identified from pediatric ESBL-E isolates that underwent whole genome sequencing (n=7)

Organism	Sequence Type (scheme)	Acquired ESBL genes	beta-lactamase genes
<i>Escherichia coli</i> (n=6)	ST43(Pasteur), ST131(Achtman)	<i>bla</i> _{CTX-M-15}	<i>bla</i> _{AmpH} , <i>bla</i> _{EC-5} , <i>bla</i> _{OXA-1}
	ST43(Pasteur), ST131(Achtman)	<i>bla</i> _{CTX-M-27}	<i>bla</i> _{AmpH} , <i>bla</i> _{EC-5}
	ST506(Pasteur), ST131(Achtman)	<i>bla</i> _{CTX-M-15}	<i>bla</i> _{AmpH} , <i>bla</i> _{EC-5} , <i>bla</i> _{TEM-1}
	ST2(Pasteur), ST167(Achtman)	<i>bla</i> _{CTX-M-55}	<i>bla</i> _{AmpC1} , <i>bla</i> _{AmpH} , <i>bla</i> _{EC-15}
	ST87(Pasteur), ST58(Achtman)	<i>bla</i> _{CTX-M-15}	<i>bla</i> _{AmpC1} , <i>bla</i> _{AmpH} , <i>bla</i> _{EC-18} , <i>bla</i> _{CMY-2}
	ND(Pasteur), ST648(Achtman)	<i>bla</i> _{CTX-M-15}	<i>bla</i> _{AmpC1} , <i>bla</i> _{AmpH} , <i>bla</i> _{EC-19}
<i>Klebsiella pneumoniae</i> (n=1)	ST307(Pasteur)	<i>bla</i> _{CTX-M-15}	<i>bla</i> _{AmpH} , <i>bla</i> _{OXA-1} , <i>bla</i> _{SHV-28} , <i>bla</i> _{TEM-1}

ST: Sequence Type; ND: Not Determined.