



Synthesis of antibacterial 1,3-diyne-linked peptoids from an Ugi-4CR/Glaser coupling approach

Martin C. N. Brauer, Ricardo A. W. Neves Filho, Bernhard Westermann, Ramona Heinke and Ludger A. Wessjohann*§

Full Research Paper

Open Access

Address:

Department of Bioorganic Chemistry, Leibniz Institute of Plant Biochemistry, Weinberg 3, D-06120 Halle/Saale, Germany

Email:

Ludger A. Wessjohann* - wessjohann@ipb-halle.de

* Corresponding author

§ Fax: +49 345 5582 1309; Tel: +49 345 5582 1301

Keywords:

antibacterial; combinatorial; diynes; homodimerization; multicomponent reactions; peptoids; Ugi reaction

Beilstein J. Org. Chem. 2015, 11, 25–30.

doi:10.3762/bjoc.11.4

Received: 29 October 2014

Accepted: 23 December 2014

Published: 07 January 2015

This article is part of the Thematic Series "Multicomponent reactions II".

Guest Editor: T. J. J. Müller

© 2015 Brauer et al; licensee Beilstein-Institut.

License and terms: see end of document.

Abstract

A library of ten 1,3-diyne-linked peptoids has been synthesized through an Ugi four-component reaction (U-4CR) followed by a copper-catalysed alkyne homocoupling (Glaser reaction). The short and chemoselective reaction sequence allows generating diverse (pseudo) dimeric peptoids. A combinatorial version allows the one-pot preparation of, e.g., six-compound-libraries of homo- and heterodimers verified by ESI-MS and HPLC. In a preliminary evaluation, some compounds display moderate activity against the Gram-positive bacterium *Bacillus subtilis*.

Introduction

A re-occurring principle of nature to mediate or increase biological activity is dimerization [1]. Many protein receptors dimerize upon activation and recruit their active form by this transformation. This process is mainly initiated by dimeric natural products or symmetric bivalent ligands, which can be of peptidic origin [2,3]. As an example, Harran and co-workers synthesized a low-molecular weight *C*₂-symmetric 1,3-diyne-linked peptide **1** which was able to mimic the function of Smac (second mitochondria-derived activator of caspase) protein by triggering caspase 8 activation as well as apoptosis at concentrations as low as 100 pM. The higher activity of **1** in comparison to **2** (Figure 1) is possibly related to the ability of **1** to

interact simultaneously with adjacent baculovirus inhibitory repeat (Bir) domains in the human X chromosome that encodes IAP (inhibitor of apoptosis) [4]. In another study Chen and co-workers found a GLP-1R antagonist only because of an unexpected dimerization [5]; and a dimer of *S*-adenosylmethionine is up to 13-fold more active than the monomer for promoting the binding of *Escherichia coli* methionine repressor to its operator DNA [6].

Peptoids are compounds which are able to mimic peptide structures [7–9]. In addition to the mimetic function, these compounds also possess an enhanced resistance to proteolytic

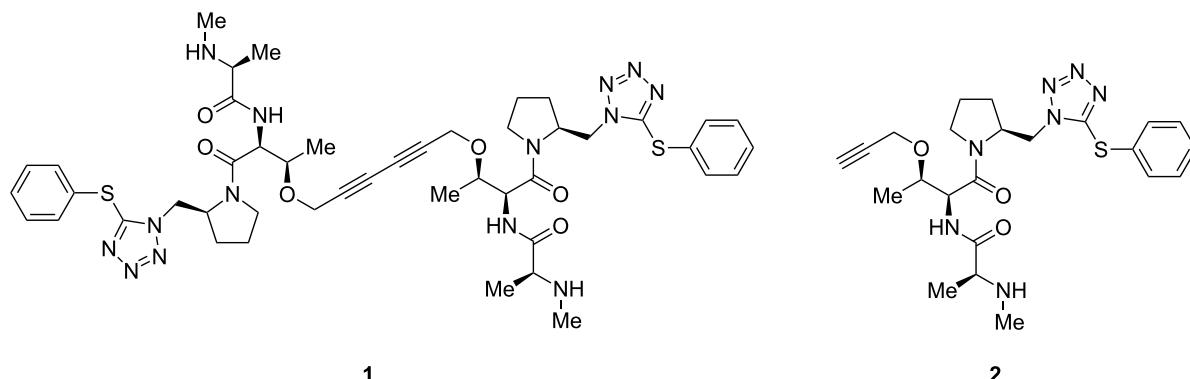


Figure 1: Apoptosis inducer C₂-symmetric 1,3-diyne-linked peptide **1** and its inactive monomer **2**.

enzymes. The fastest method for synthesizing peptoids is the Ugi four-component reaction (U-4CR) [10–12]. In combination with other protocols, this reaction has been used in the synthesis of bioactive peptides and pseudopeptides, e.g., tubulysin mimetics [13], julocrotine derivatives [14], architecturally complex peptoid macrocycles [15,16], building blocks for diversity-oriented synthesis [17], and heterocyclic compounds [18]. Complex structures as well as simple Ugi products exhibit promising biological profiles, e.g., cytotoxicity [13,19,20], fungicidal [21,22] and antibacterial properties [23–26], or inhibition of histone deacetylases [27]. The Ugi post-modification strategy has also been employed in the synthesis of heterocyclic and natural product inspired compounds [28–32]. Although several protocols of U-4CR followed by transition metal-catalysed reactions have been published so far [33], to the best of our knowledge, there are no reports about U-4CR/Glaser-type (homo) coupling combinations.

In view of an increasing interest to synthesize dimerized peptidomimetics with pharmacological properties through a step-efficient protocol that allows rapid access to highly diverse dimer libraries, we set out to develop a strategy based on an U-4CR/Glaser-type homocoupling sequence [34]. In comparison to popular cross linking reactions like, e.g., click reactions or amide bonds, the Glaser coupling allows the use of truly identical monomers. This decreases the number of steps for appropriate starting materials, and allows access to true homodimers in sensu strictu.

Results and Discussion

To achieve the synthesis of monomers eligible for dimerizations by Glaser coupling, equimolar amounts of propargylamine (**3**), aldehyde **4**, carboxylic acid **5**, and isocyanides **6** were reacted in methanol at room temperature over 24 h following well established Ugi protocols [12]. After flash

column chromatography *N*-propargyl peptoids **7a–j** were obtained in good yields. The next step was the copper-catalysed homocoupling (Glaser reaction) of the terminal alkyne functions. Albeit several protocols are reported for this reaction, the CuCl-catalyzed method recently described by Jia and co-workers was utilised to access the C₂-symmetric 1,3-diyne because it does not require expensive catalysts, ligands, or additives (Table 1) [34]. The coupling reaction was clean without notable side product formation as confirmed by TLC analysis, and the desired peptoid dimers **8a–j** could be obtained in high to quantitative yields. Aromatic as well as aliphatic carboxylic acids and aldehydes have been successfully employed in both multicomponent and coupling reactions. When performing the reaction with methyl isocyanoacetate (Table 1, entry 2) the desired products could be obtained in good yields with the ester group remaining untouched. It is important to note that different protecting groups can be used: Boc-, PhAc- and Cbz-protected peptoid derivatives (Table 1, entries 8–10) reacted to the corresponding dimers **7h–j** without complications. The structure of the compounds **7a–j**, as well as **8a–j**, have been confirmed by ¹H, ¹³C NMR spectra, and HRMS. In addition, HPLC analyses revealed that an adjacent stereocenter (Table 1, entry 8, **7h/8h**) does not racemize under the reaction conditions of both the MCR and the Glaser coupling.

Due to the high selectivity and high conversions found in the Glaser coupling step, our attention turned toward the development of a combinatorial version of the copper-catalysed homodimerization. In this strategy two or more alkyne peptoids should couple simultaneously in the same reaction vessel in order to generate small libraries of dimers. In contrast to parallel synthesis, the combinatorial approach easily generates non-symmetric dimers **9**, **10** and **11**. Thus, the peptoids **7f**, **7h** and **7j** were pooled to a Glaser reaction as depicted in Scheme 1.

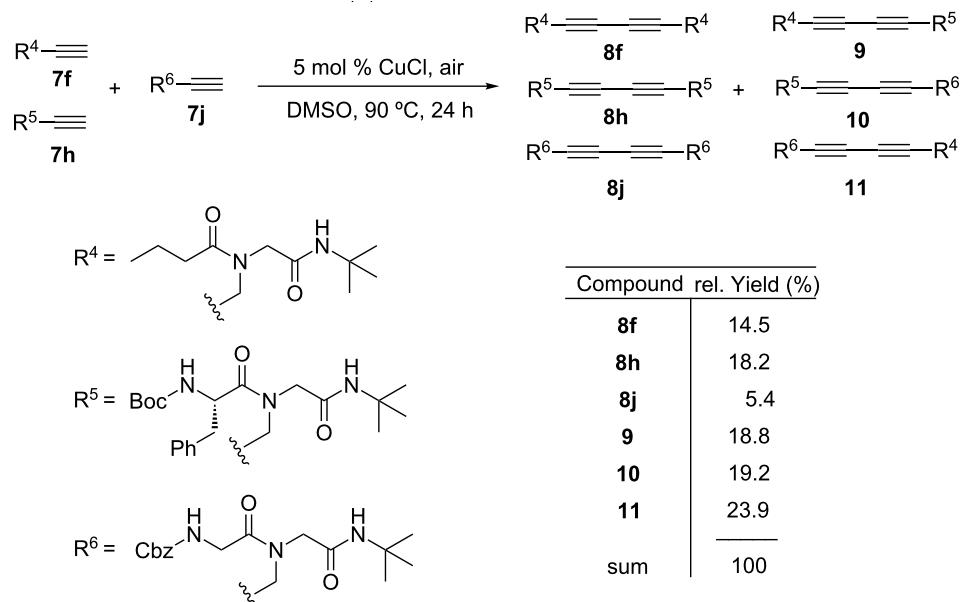
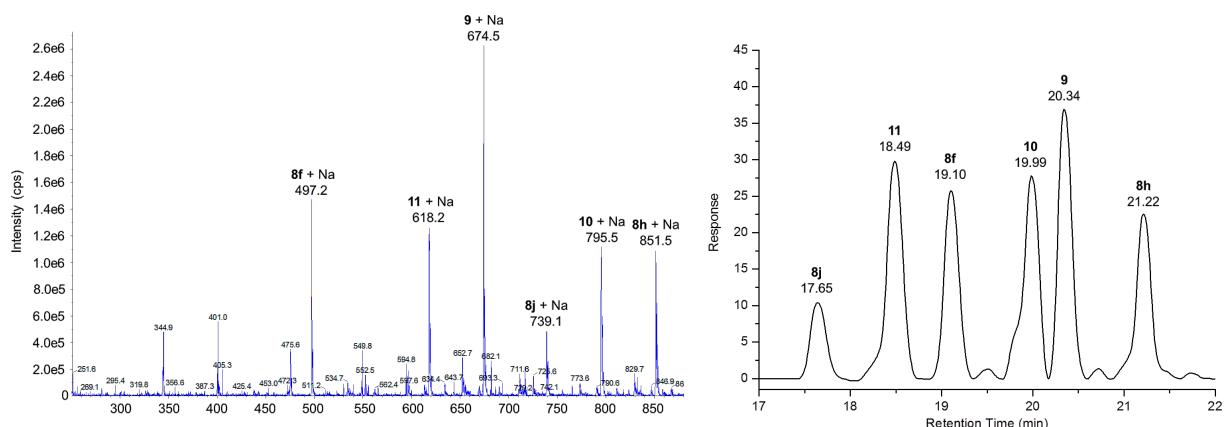
Table 1: Synthesis of compounds **7a–j** and **8a–j**.

Entry	R ¹	R ²	R ³	Monomer 7 yield (%)	Dimer 8 yield (%)
1	CH ₃			7a 97	8a 88
2	CH ₃			7b 95	8b 80
3	CH ₃			7c 99	8c 99
4	Ph			7d 70	8d 91
5	n-C ₃ H ₇			7e 91	8e 99
6	n-C ₃ H ₇	H		7f 70	8f 99
7				7g 98	8g 97
8		H		7h 82	8h 99
9				7i 82	8i 96
10		H		7j 80	8j 99

The ESI-MS spectrum of the crude library confirmed the presence of all expected Glaser-coupled products **8f**, **8h**, **8j**, **9**, **10** and **11**. The HPLC–MS analysis of the composition resulted in six peaks with different retention times and intensities identified via MS as the six desired components of the library. Figure 2 illustrates the expanded region of the ESI-MS spectrum (positive mode) and the HPLC chromatogram with the respective assignments of the obtained peaks. The analysis of the obtained spectra revealed that the non-symmetric dimers **9**, **10** and **11** are formed preferentially. The abundance differences

observed are mostly lower than 2-fold, in one case up to ca. 4-fold. This is still acceptable for our initial bioactivity assays, as most screening setups cover several orders of magnitude of concentration anyhow. Therefore no further attempt to optimize for an equal product distribution was deemed necessary.

To gain insight into the antibiotic potential of the products, single compound dimers **8a–j** were subjected to a preliminary evaluation against *Bacillus subtilis* (Figure 3) [35,36]. The

**Scheme 1:** Combinatorial Glaser coupling involving acetylenes **7f**, **7j** and **7h**.

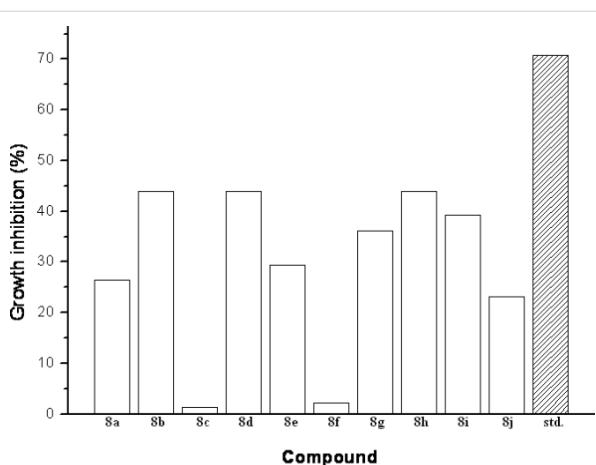


Figure 3: Growth inhibition of *Bacillus subtilis* by compounds 8a–j at 1 μ M (15 h), and standard erythromycin at 1 μ M (15 h).

displayed growth inhibitory activity against *Bacillus subtilis* in a preliminary assay.

Supporting Information

Supporting Information File 1

Complete experimental procedures, characterization and figures of ^1H and ^{13}C NMR spectra.

[<http://www.beilstein-journals.org/bjoc/content/supplementary/1860-5397-11-4-S1.pdf>]

Acknowledgements

The authors acknowledge support from the State of Saxony-Anhalt (MK-LSA, Projekt “Lipopeptide”). We thank Dr. Jürgen Schmidt and Ms. Anja Ehrlich for HRMS and HPLC support, respectively. M.C.N.B and R.A.W.N.F. thank the Brazilian National Research Council (CNPq) for Ph.D. fellowships; R.H. gratefully acknowledges support by the Studienstiftung des Deutschen Volkes.

References

1. Marianayagan, N. J.; Sunde, M.; Mathews, J. M. *Trends Biochem. Sci.* **2004**, *29*, 618–625. doi:10.1016/j.tibs.2004.09.006
2. Hadden, M. K.; Blagg, B. S. J. *Anti-Cancer Agents Med. Chem.* **2008**, *8*, 807–816. doi:10.2174/187152008785914743
3. Lian, G.; Yu, B. *Chem. Biodiversity* **2010**, *7*, 2660–2691. doi:10.1002/cbdv.201000038
4. Li, L.; Thomas, R. M.; Suzuki, H.; De Brabander, J. K.; Wang, X.; Harran, P. G. *Science* **2004**, *305*, 1471–1474. doi:10.1126/science.1098231
5. Chen, D.; Liao, J.; Li, N.; Zhou, C.; Liu, Q.; Wang, G.; Zhang, R.; Zhang, S.; Lin, L.; Chen, K.; Xie, X.; Nan, F.; Young, A. A.; Wang, M.-W. *Proc. Natl. Acad. Sci. U. S. A.* **2007**, *104*, 943–948. doi:10.1073/pnas.0610173104
6. Joce, C.; White, R.; Stockley, P. G.; Warriner, S.; Turnbull, W. B.; Nelson, A. *Bioorg. Med. Chem. Lett.* **2012**, *22*, 278–284. doi:10.1016/j.bmcl.2011.11.017
7. Seo, J.; Lee, B.-C.; Zuckermann, R. N. *Comprehensive Biomaterials*; Elsevier: Amsterdam, 2011; Vol. 2, pp 53–76. doi:10.1016/B978-0-08-055294-1.00256-7
8. Vagner, J.; Qu, H.; Hruby, V. J. *Curr. Opin. Chem. Biol.* **2008**, *12*, 292–296. doi:10.1016/j.cbpa.2008.03.009 And references cited therein.
9. Zuckermann, R. N.; Kerr, J. M.; Kent, S. B. H.; Moos, W. H. *J. Am. Chem. Soc.* **1992**, *114*, 10646–10647. doi:10.1021/ja00052a076
10. Zhu, J.; Bienaymé, H., Eds. *Multicomponent Reactions*; Wiley-VCH: Weinheim, Germany, 2005. doi:10.1002/3527605118
11. Vercillo, O. E.; Andrade, C. K. L.; Wessjohann, L. A. *Org. Lett.* **2008**, *10*, 205–208. doi:10.1021/o10702521g
12. Wessjohann, L. A.; Kaluderovic, G.; Neves Filho, R. A. W.; Morejon, M. C.; Lemanski, G.; Ziegler, T. *Multicomponent Reactions 1: Further Components Carboxylic Acids and Amine (Ugi Reaction)*. In *Science of Synthesis*; Müller, T. J. J., Ed.; Thieme: New York, 2013; pp 415–497.
13. Pando, O.; Stark, S.; Denkert, A.; Porzel, A.; Preusentanz, R.; Wessjohann, L. A. *J. Am. Chem. Soc.* **2011**, *133*, 7692–7695. doi:10.1021/ja2022027
14. Neves Filho, R. A. W.; Westermann, B.; Wessjohann, L. A. *Beilstein J. Org. Chem.* **2011**, *7*, 1504–1507. doi:10.3762/bjoc.7.175
15. Rivera, D. G.; Wessjohann, L. A. *Molecules* **2007**, *12*, 1890–1899. doi:10.3390/12081890
16. Rivera, D. G.; Wessjohann, L. A. *J. Am. Chem. Soc.* **2009**, *131*, 3721–3732. doi:10.1021/ja809005k
17. Neves Filho, R. A. W.; Stark, S.; Morejon, M. C.; Westermann, B.; Wessjohann, L. A. *Tetrahedron Lett.* **2012**, *53*, 5360–5363. doi:10.1016/j.tetlet.2012.07.064
18. Akritopoulou-Zanke, I.; Djuric, S. W. *Heterocycles* **2007**, *73*, 125–147. doi:10.3987/REV-07-SR(U)3
19. Yamada, R.; Cao, X.; Butkevich, A. N.; Millard, M.; Odde, S.; Mordwinkin, N.; Gundla, R.; Zandi, E.; Louie, S. G.; Petasis, N. A.; Neamati, N. *J. Med. Chem.* **2011**, *54*, 2902–2914. doi:10.1021/jm101655d
20. Olsen, C. A.; Ziegler, H. L.; Nielsen, H. M.; Frimodt-Møller, N.; Jaroszewski, J. W.; Franzyk, H. *ChemBioChem* **2010**, *11*, 1356–1360. doi:10.1002/cbic.201000232
21. Galetti, M. D.; Cirigliano, A. M.; Cabrera, G. M.; Ramírez, J. A. *Mol. Diversity* **2012**, *16*, 113–119. doi:10.1007/s11030-011-9334-1
22. Lamberth, C.; Jeanguenat, A.; Cederbaum, F.; De Mesmaeker, A.; Zeller, M.; Kempf, H.-J.; Zeun, R. *Bioorg. Med. Chem.* **2008**, *16*, 1531–1545. doi:10.1016/j.bmcl.2007.10.019
23. Socha, A. M.; Tan, N. Y.; LaPlante, K. L.; Sello, J. K. *Bioorg. Med. Chem.* **2010**, *18*, 7193–7202. doi:10.1016/j.bmcl.2010.08.032
24. Neves Filho, R. A. W.; Stark, S.; Westermann, B.; Wessjohann, L. A. *Beilstein J. Org. Chem.* **2012**, *8*, 2085–2090. doi:10.3762/bjoc.8.234
25. Kodadek, T. *Chem. Biol.* **2013**, *20*, 1202–1203. doi:10.1016/j.chembiol.2013.10.006
26. Hu, Y.; Amin, M. N.; Padhee, S.; Wang, R. E.; Qiao, Q.; Bai, G.; Li, Y.; Mathew, A.; Cao, C.; Cai, J. *ACS Med. Chem. Lett.* **2012**, *3*, 683–686. doi:10.1021/ml3001215
27. Grolla, A. A.; Podestà, V.; Chini, M. G.; Di Micco, S.; Vallario, A.; Genazzani, A. A.; Canonico, P. L.; Bifulco, G.; Tron, G. C.; Sorba, G.; Pirali, T. *J. Med. Chem.* **2009**, *52*, 2776–2785. doi:10.1021/jm801529c

28. Keating, T. A.; Armstrong, R. W. *J. Am. Chem. Soc.* **1996**, *118*, 2574–2583. doi:10.1021/ja953868b
29. Hulme, C.; Gore, V. *Curr. Med. Chem.* **2003**, *10*, 51–80. doi:10.2174/0929867033368600
30. Hulme, C.; Morrisette, M. M.; Volz, F. A.; Burns, C. J. *Tetrahedron Lett.* **1998**, *39*, 1113–1116. doi:10.1016/S0040-4039(97)10795-X
31. Wessjohann, L. A.; Rhoden, C. R. B.; Rivera, D. G.; Vercillo, O. E. *Top. Heterocycl. Chem.* **2010**, *23*, 199–226. doi:10.1007/7081_2009_25
32. Wessjohann, L. A.; Ruijter, E. *Top. Curr. Chem.* **2005**, *243*, 137–184. doi:10.1007/b96883
33. Koopmanschap, G.; Ruijter, E.; Orru, R. V. A. *Beilstein J. Org. Chem.* **2014**, *10*, 544–598. doi:10.3762/bjoc.10.50
34. Yin, K.; Li, C.; Li, J.; Jia, X. *Green Chem.* **2011**, *13*, 591–593. doi:10.1039/c0gc00413h
35. Heinke, R.; Franke, K.; Porzel, A.; Wessjohann, L. A.; Ali, N. A. A.; Schmidt, J. *Phytochemistry* **2011**, *72*, 929–934. doi:10.1016/j.phytochem.2011.03.008
36. Michels, K.; Heinke, R.; Kuipers, O. P.; Arnold, N.; Wessjohann, L. A. *J. Antibiot.* submitted.

License and Terms

This is an Open Access article under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

The license is subject to the *Beilstein Journal of Organic Chemistry* terms and conditions: (<http://www.beilstein-journals.org/bjoc>)

The definitive version of this article is the electronic one which can be found at:
doi:10.3762/bjoc.11.4