

Supplementary Information for

A pan-sarbecovirus vaccine based on RBD of SARS-CoV-2 original strain elicits potent neutralizing antibodies against XBB in NHPs

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1 **Materials and Methods**

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3 **Cell lines**

4 HEK-293T cells, Expi-293F cells, Vero-E6 cells and Huh-7 cells were obtained from the American
5 Type Culture Collection (ATCC). Dulbecco's Modified Eagle's Medium (DMEM) was used to culture
6 HEK-293T cells, Vero-E6 cells and Huh-7 cells with supplementation of 10% fetal bovine serum
7 (FBS) and 1% penicillin-streptomycin at 37°C, 5% CO₂. 293T II medium (Sino Biological) was used
8 to culture Expi-293F cells at 37 °C in 5% CO₂ with shaking at 100 rpm.

9

10 **Serum samples**

11 Sera from immunized rhesus macaques were from our previous studies (1). A total of six rhesus
12 macaques were assigned into two groups, and each group had three animals. Macaques in the
13 first group were immunized with 100 µg RBD-Fc protein formulated with 400 µg CF501. 100 µg
14 RBD-Fc protein formulated with an equal volume of Alum were used to immunize macaques in the
15 second group. Immunizations were conducted on days 0, 21 and 115, and sera were collected as
16 previously described.

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18 **Protein expression**

19 RBD expression was performed as previously reported (2). The plasmid of Pfuse-SARS-CoV-2-
20 BA.2.2-RBD, Pfuse-SARS-CoV-2-BA.2.9-RBD, Pfuse-SARS-CoV-2-BA.2.12.1-RBD, Pfuse-

21 SARS-CoV-2-BA.5-RBD, Pfuse-SARS-CoV-2-BA.2.75-RBD, Pfuse-SARS-CoV-2-BF.7-RBD,
22 Pfuse-SARS-CoV-2-BQ.1.1-RBD or Pfuse-SARS-CoV-2-XBB-RBD was separately transfected
23 into Expi-293F cells using EZ Trans transfection reagent (Life iLAB Bio-Technology, China). After
24 6 days, suspensions were harvested, and the RBDs were purified using Ni-NTA (Qiagen). The
25 purified proteins were identified by SDS-PAGE. All proteins were stored at -80 °C.

26 27 **ELISA**

28 Enzyme-linked immunosorbent assay (ELISA) was performed as previously described(1). Briefly,
29 a concentration of 1 µg/ml RBD was coated onto the ELISA plates overnight at 4 °C. Next, the
30 ELISA plates were blocked using a blocking buffer (PBS containing 5% BSA). Sera were serially
31 diluted and applied to the ELISA plates for incubation at 37 °C for 45 min. After that, the plates
32 were washed using PBST 5 times. HRP-conjugated goat anti-monkey IgG (Abcam, UK) (1:10,000)
33 was added into the plates and further incubated for 45 min at 37°C. Finally, the reactions were
34 visualized by adding 3,3',5,5'-tetramethylbenzidine (TMB), and H₂SO₄ was used to stop the
35 reaction. Absorbance at 450 nm (A450) was read by using a microplate reader (InfiniteM200PRO,
36 Switzerland). Endpoint titers were defined as the highest dilutions that showed A450>2.1-fold of
37 background values.

38 39 **Pseudovirus production**

40 The production of pseudoviruses was performed as previously described (1). Briefly, the plasmids
41 of pNL4-3.Luc.R-E and PcDNA3.1-BA.2.2-S/ PcDNA3.1-BA.2.9-S/ PcDNA3.1-BA.2.12.1-S/
42 PcDNA3.1-BA.5-S/ PcDNA3.1-BA.2.75-S/ PcDNA3.1-BF.7-S/ PcDNA3.1-BQ.1.1-S/ and
43 PcDNA3.1-XBB-S were co-transfected into HEK293T cells using the Vigofect transfect reagent.
44 After 12 h, fresh DMEM containing 10% FBS was used to replace supernatants. Finally,
45 supernatants were collected after further culture for 48 h and stored at -80 °C.

46 47 **Pseudovirus neutralization assay**

48 The pseudovirus neutralization assay was conducted as described previously (1). Briefly, Huh-7
49 cells were used as the target cells. A total of 1×10⁴ cells/well were seeded in a 96-well plate. After
50 8 h, serially diluted sera were incubated with pseudovirus for 30 min. The mixture of sera and
51 pseudovirus was applied into Huh-7 cells. After 60 h, luciferase activities were detected using the
52 Firefly Luciferase Assay Kit (Promega, USA). NT50 is defined as the dilution of sera that shows
53 50% of the luminescence units of viral control. Once the titer was below 1:100, the titer was then
54 recorded as 1:50.

55 56 **Authentic virus neutralization assay**

57 Vero-E6 cells were used as the target cells for live Omicron BA.2.2 infection. 1×10⁴ Vero-E6
58 cells/well were seeded in a 96-well plate. Diluted sera from immunized rhesus macaques were first
59 mixed with live Omicron BA.2.2 for 30 min. Mixtures of virus and serum were added into the Vero-
60 E6 cells. After 48 h, supernatants were collected to conduct the RT-qPCR assay. RT-qPCR was
61 conducted to quantify SARS-CoV-2 viral mRNA level by using the One-Step PrimeScrip RT-PCR
62 Kit (Takara, Japan). Cells were used to perform the immunofluorescence assay. First, the cells
63 were treated with paraformaldehyde and 0.1% Triton X. After blocking with 3% BSA, cells were
64 incubated with mouse anti-SARS-CoV-2 N antibody (1:1000) for 30 min at 37 °C. Finally, after
65 washing with PBS 5 times, cells were added to Alexa Fluor donkey anti-mouse IgG 488-labeled
66 antibody (Thermo). Fluorescence microscopy was used to capture SARS-CoV-2 N protein.

67 68 **References**

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